# droplet-spatial: supporting materila to Computational analysis of single cell RNAseq data chapter

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# Contents

#### Introduction

This github contains all the steps for an examplary analysis of droplet-based RNAseq and spatial-transcriptomics data, described in chapter *Computational analysis of single cell RNAseq data* in Methods in Molecular Biology, (vol. XX, 202X)

### 3.1 From fastq to counts table

#### 10XGenomics example

```
home <- getwd()
###################################
#cloning in your working folder
system("git clone https://github.com/kendomaniac/droplet-spatial.git")
#if it was already cloned the first time
setwd(paste(home, "droplet-spatial", sep="/"))
system("git pull")
###################################
# downloading the genome reference for human
setwd(paste(home, "droplet-spatial/genomes", sep="/"))
system("wget http://cf.10xgenomics.com/supp/cell-exp/refdata-cellranger-GRCh38-3.0.0.tar.gz")
system("gzip -d refdata-cellranger-GRCh38-3.0.0.tar.gz")
system("tar xvf refdata-cellranger-GRCh38-3.0.0.tar")
system("rm refdata-cellranger-GRCh38-3.0.0.tar")
hg38reference <-
 paste(home, "droplet-spatial/genomes/refdata-cellranger-GRCh38-3.0.0", sep="/")
scratch <-
  paste(home, "droplet-spatial/scratch", sep="/")
# scratch folder should be located on a SSD disk
#####################################
library(rCASC)
```

#### Spatial transcriptomics example

```
home <- getwd()
setwd(paste(home, "droplet-spatial/genomes", sep="/"))
system("wget http://cf.10xgenomics.com/supp/spatial-exp/refdata-cellranger-mm10-3.0.0.tar.gz")
system("gzip -d refdata-cellranger-mm10-3.0.0.tar.gz")
system("tar xvf refdata-cellranger-mm10-3.0.0.tar")
system("rm refdata-cellranger-mm10-3.0.0.tar")
mm10reference <-
 paste(home, "droplet-spatial/genomes/refdata-cellranger-mm10-3.0.0", sep="/")
scratch <- paste(home, "droplet-spatial/scratch", sep="/")</pre>
# scratch folder should be located on a SSD disk
library(rCASC)
setwd(paste(home, "droplet-spatial/data", sep="/"))
dir.create("st")
setwd(paste(home, "droplet-spatial/data/st", sep="/"))
dataset <- paste(home, "droplet-spatial/data/st, sep="/")</pre>
system("wget http://s3-us-west-2.amazonaws.com/10x.files/samples/
spatial-exp/1.0.0/V1_Mouse_Kidney/V1_Mouse_Kidney_fastqs.tar")
system("tar xvf V1_Mouse_Kidney_fastqs.tar")
```

```
system("rm V1_Mouse_Kidney_fastqs.tar")
fastqs <- paste(home, "droplet-spatial/data/V1_Mouse_Kidney_fastqs", sep="/")

system("wget http://cf.10xgenomics.com/samples/spatial-exp/1.0.0/
V1_Mouse_Kidney/V1_Mouse_Kidney_image.tif")
image <- paste(home, "droplet-spatial/data/st/V1_Mouse_Kidney_image.tif", sep="/")

stpipeline(group="docker", scratch.folder=scratch, data.folder=dataset,
genome.folder=mm10reference, fastqPathFolder=fastqs,
ID="kidneyst",imgNameAndPath=image, slide="V19L29-096",area="B1")</pre>
```

# 3.2 Cells QC

```
home <- getwd()</pre>
###################################
#cloning in your working folder
system("git clone https://github.com/kendomaniac/droplet-spatial.git")
#if it was already cloned the first time
setwd(paste(home, "droplet-spatial", sep="/"))
system("git pull")
library(rCASC)
setwd(paste(home, "droplet-spatial/data", sep="/"))
unzip("setA_5x100cells.txt.zip")
system("wget ftp://ftp.ensembl.org/pub/release-98/gtf/homo_sapiens/Homo_sapiens.GRCh38.98.gtf.gz")
system("gzip -d homo_sapiens/Homo_sapiens.GRCh38.98.gtf.gz")
mitoRiboUmi(group="docker", file=paste(getwd(), "setA_5x100cells.txt", sep="/"),
           scratch.folder=scratch, separator="\t", umiXgene=3,
           gtf.name="Homo_sapiens.GRCh38.98.gtf", bio.type="protein_coding")
```

# 3.3 Annotation and filtering

```
home <- getwd()</pre>
####################################
#cloning in your working folder
system("git clone https://github.com/kendomaniac/droplet-spatial.git")
#if it was already cloned the first time
setwd(paste(home, "droplet-spatial", sep="/"))
system("git pull")
#####################################
library(rCASC)
setwd(paste(home, "droplet-spatial/data", sep="/"))
unzip("setA_5x100cells.txt.zip")
system("wget ftp://ftp.ensembl.org/pub/release-98/gtf/homo_sapiens/Homo_sapiens.GRCh38.98.gtf.gz")
system("gzip -d homo_sapiens/Homo_sapiens.GRCh38.98.gtf.gz")
scannobyGtf(group="docker", file=paste(getwd(),"testSCumi_mm10.csv",sep="/"),
            gtf.name="Homo_sapiens.GRCh38.98.gtf", biotype="protein_coding",
            mt=TRUE, ribo.proteins=TRUE, umiXgene=3, riboStart.percentage=20,
            riboEnd.percentage=40, mitoStart.percentage=1,
            mitoEnd.percentage=20, thresholdGenes=100)
```

## 3.4 Selecting top ranked genes

## 3.5 Clustering

#### 3.5.1 Clustering with tSne

#### 3.5.2 Clustering with SIMLR

#### 3.5.3 Clustering with Griph

#### 3.5.4 Clustering with Seurat

```
home <- getwd()</pre>
#####################################
#cloning in your working folder
system("git clone https://github.com/kendomaniac/droplet-spatial.git")
#if it was already cloned the first time
setwd(paste(home, "droplet-spatial", sep="/"))
system("git pull")
#####################################
library(rCASC)
setwd(paste(home, "droplet-spatial/data", sep="/"))
seuratPCAEval(group="docker", scratch.folder=scratch, file=paste(getwd(),
              "filtered_expression_filtered_variance_setA_5x100cells.txt", sep="/"),
              separator="\t", logTen = 0, seed = 111, format="NULL")
#optimal threshold of PCs is 5
seuratBootstrap(group="docker",scratch.folder=scratch, file=paste(getwd(),
                "filtered_expression_filtered_variance_setA_5x100cells.txt", sep="/"),
                nPerm=80, permAtTime=8, percent=10, separator="\t", logTen=0,
                pcaDimensions=5, seed=111)
```

## 3.6 Discovering cluster-specific markers