

# Linnorm Package

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**Type** Package

**Title** Linear model and normality based normalization method (Linnorm)

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**Description** A Normalization method for RNA-seq Expression data or large scale count data for parametric tests. This package also includes several functions for the simulation of RNA-seq count data based on known distributions.

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**Imports** Rcpp (>= 0.12.2),  
RcppArmadillo,  
MASS,  
limma

**LinkingTo** Rcpp, RcppArmadillo

**Suggests** BiocStyle, knitr, rmarkdown, seqc

**VignetteBuilder** knitr

**biocViews** Sequencing, ChIPSeq, RNASeq, DifferentialExpression,  
GeneExpression, Genetics, Normalization, Software, Transcription, BatchEffect

**NeedsCompilation** yes

**RoxygenNote** 5.0.1

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Linnorm

*Linnorm Normalization Function***Description**

This function performs the Linear model and normality based normalization method (Linnorm) for RNA-seq expression data or large scale count data.

**Usage**

```
Linnorm(datamatrix, showinfo = FALSE, method = "default",
        perturbation = 10, minZeroPortion = 2/3)
```

**Arguments**

datamatrix	The matrix or data frame that contains your dataset. Each row is a feature (or Gene) and each column is a sample (or replicate). Undefined values such as NA are not supported.
showinfo	Logical. Show lambda value calculated. Defaults to FALSE.
method	"default" or "lambda" The program will output the transformed matrix if the method is "default". If the method is "lambda", the program will output a lambda value.
perturbation	Integer $\geq 2$ . To search for an optimal minimal deviation parameter (please see the article), Linnorm uses the iterated local search algorithm which perturbs away from the initial local minimum. The range of the area searched in each perturbation is exponentially increased as the area get further away from the initial local minimum, which is determined by their index. This range is calculated by $10 * (\text{perturbation}^{\text{index}})$ .
minZeroPortion	Double $\geq 0, \leq 1$ . Features without at least this portion of non-zero values will not be used in the calculation of normalizing parameter. Defaults to 2/3.

**Details**

If method is default, Linnorm outputs a transformed expression matrix. For users who wish to work with lambda instead, the output is a single lambda value. Please note that users with the lambda value can obtain a normalized Linnorm dataset by: `log1p(lambda * datamatrix)`. There is no need to rerun the program if a lambda is already calculated.

**Value**

This function returns either a transformed data matrix or a lambda value.

## Examples

```
#Obtain example matrix:
library(seqc)
SampleA <- ILM_aceview_gene_BGI[,grepl("A_",colnames(ILM_aceview_gene_BGI))]
rownames(SampleA) <- ILM_aceview_gene_BGI[,2]
#Extract a portion of the matrix for an example
expMatrix <- SampleA[,1:3]
normalizedExp <- Linnorm(expMatrix)
normalizedExp <- Linnorm(expMatrix, method = "lambda")
```

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Linnorm.limma

*Linnorm-limma pipeline for Differentially Expression Analysis*


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## Description

This function first performs Linnorm normalization on the dataset. Then, it will perform limma for DEG analysis. Finally, it will correct fold change outputs from limma results, that will be wrong otherwise. Please cite both Linnorm and limma when you use this function for publications.

## Usage

```
Linnorm.limma(datamatrix, design = NULL, output = "DEResults",
  noINF = TRUE, showinfo = FALSE, perturbation = 10,
  minZeroPortion = 2/3, robust = TRUE)
```

## Arguments

datamatrix	The matrix or data frame that contains your dataset. Each row is a feature (or Gene) and each column is a sample (or replicate). Undefined values such as NA are not supported.
design	A design matrix required for limma. Please see limma's documentation or our vignettes for more detail.
output	Character. "DEResults" or "Both". Set to "DEResults" to output a matrix that contains Differential Expression Analysis Results. Set to "Both" to output a list that contains both Differential Expression Analysis Results and the transformed data matrix.
noINF	Logical. Prevent generating INF in the fold change column by using Linnorm's lambda and adding one. If it is set to FALSE, INF will be generated if one of the conditions has zero expression. Defaults to TRUE.
showinfo	Logical. Show lambda value calculated. Defaults to FALSE.
perturbation	Integer $\geq 2$ . To search for an optimal minimal deviation parameter (please see the article), Linnorm uses the iterated local search algorithm which perturbs away from the initial local minimum. The range of the area searched in each perturbation is exponentially increased as the area get further away from the initial local minimum, which is determined by their index. This range is calculated by $10 * (\text{perturbation}^{\text{index}})$ .

**minZeroPortion** Double  $\geq 0$ ,  $\leq 1$ . Features without at least this portion of non-zero values will not be used in the calculation of normalizing parameter. Defaults to 2/3.

**robust** Logical. In the eBayes function of Limma, run with robust setting with TRUE or FALSE. Defaults to TRUE.

## Details

This function performs both Linnorm and limma for users who are interested in differential expression analysis. Please note that if you directly use a Linnorm Normalized dataset with limma, the output fold change and average expression will be wrong. (p values and adj.pvalues will be fine.) This is because the voom-limma pipeline assumes input to be in raw counts. This function is written to fix this problem.

## Value

If output is set to "DEResults", this function will output a matrix with Differential Expression Analysis Results with the following columns:

- logFC: Log 2 Fold Change
- XPM: Average Expression. If input is raw count or CPM, this column has the CPM unit. If input is RPKM, FPKM or TPM, this column has the TPM unit.
- t: moderated t-statistic
- P.Value: p value
- adj.P.Val: Adjusted p value. This is also called False Discovery Rate or q value.
- B: log odds that the feature is differential

If output is set to Both, this function will output a list with the following objects:

- DEResults: Differential Expression Analysis Results as described above.
- TMatrix: A Linnorm Normalized Expression Matrix.

## Examples

```
#Obtain example matrix:
library(seqc)
SampleA <- ILM_aceview_gene_BGI[,grepl("A_",colnames(ILM_aceview_gene_BGI))]
rownames(SampleA) <- ILM_aceview_gene_BGI[,2]
SampleB <- ILM_aceview_gene_BGI[,grepl("B_",colnames(ILM_aceview_gene_BGI))]
rownames(SampleB) <- ILM_aceview_gene_BGI[,2]
#Extract a portion of the matrix for an example
expMatrix <- cbind(SampleA[,1:3], SampleB[,1:3])
designmatrix <- c(1,1,1,2,2,2)
designmatrix <- model.matrix(~ 0+factor(designmatrix))
colnames(designmatrix) <- c("group1", "group2")
rownames(designmatrix) <- colnames(expMatrix)

#Example 1
DEGResults <- Linnorm.limma(expMatrix, designmatrix)
#Example 2
DEGResults <- Linnorm.limma(expMatrix, designmatrix, output="Both")
```

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RnaXSim	<i>This function simulates a RNA-seq dataset based on a given distribution.</i>
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## Description

This function simulates a RNA-seq dataset based on a given distribution.

## Usage

```
RnaXSim(thisdata, distribution = "Poisson", NumRep = 3, NumDiff = 5000,
        NumFea = 20000, showinfo = FALSE, MaxLibSizeLog2FC = 0.5)
```

## Arguments

thisdata	Matrix: The matrix or data frame that contains your dataset. Each row is a gene and each column is a replicate. Undefined values such as NA are not supported. This program assumes that all columns are replicates of the same sample.
distribution	Character: Defaults to "Poisson". This parameter controls the output distribution of the simulated RNA-seq dataset. It can be one of "Gamma" (Gamma distribution), "Poisson" (Poisson distribution), "LogNorm" (Log Normal distribution) or "NB" (Negative Binomial distribution).
NumRep	Integer: The number of replicates. This is half of the number of output samples. Defaults to 3.
NumDiff	Integer: The number of Differentially Changed Features. Defaults to 5000.
NumFea	Integer: The number of Total Features. Defaults to 20000.
showinfo	Logical: should we show data information on the console? Defaults to FALSE.
MaxLibSizeLog2FC	Double: The maximum library size difference from the mean that is allowed, in terms of log 2 fold change. Set to 0 to prevent program from generating library size differences. Defaults to 0.5.

## Value

This function returns a list that contains a matrix of count data in integer raw count and a vector that shows which genes are differentially expressed. In the matrix, each row is a gene and each column is a replicate. The first NumRep (see parameter) of the columns belong to sample 1, and the last NumRep (see parameter) of the columns belong to sample 2. There will be NumFea (see parameter) number of rows.

## Examples

```
#Obtain example matrix:
library(seqc)
SampleA <- ILM_aceview_gene_BGI[,grep("A_",colnames(ILM_aceview_gene_BGI))]
rownames(SampleA) <- ILM_aceview_gene_BGI[,2]
```

```
#Extract a portion of the matrix for an example
expMatrix <- SampleA[,1:10]
simulateddata <- RnaXSim(expMatrix, distribution="NB", NumRep=3, NumDiff = 500, NumFea = 3000)
```

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