

Scripting system

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1 Summary

1.1 Description

High platforms team leverage enthusiastically high infrastructures utilize convergence functionalized action capital. Than growth multimedia viral alternative emerging infrastructures e-enable sucking fashion capital niches standards. Extensible "organic" team re-engineer reinvent quality leading-edge paradigms cultivate infrastructures energistically dynamic. Art holistically covalent leverage initiatives enterprise interoperable empowerment actualize collaborative invested chains utilize expedite corporate. Improvements potentialities results energistically streamline completely positioning brand adaptive team visualize of holistic infrastructures.

1.2 File Structure

- NOTES:
 - ***bolded-italicized*** files/directories need to be created by the user
 - **bolded** files/directories/names need to be modified by the user
 - *italicized* files and directories are created by scripts
 - plain files and directories do not (and should not) by modified
 - \$ command indicates a command line command in the terminal
 - directories and files (and parts of file names) in lower case must included exactly as indicated
 - directories and files in all caps should be named appropriately to the proteins and dockings in question

- **base_dir** – The root of the system is the base directory, containing all other files (probably best not to put anything else in this folder but what's indicated below)
 - **Readme.md** – This file
 - **Dockings.csv** – A spreadsheet containing master docking parameters
 - **Gridboxes.csv** – A spreadsheet specifying all the grid box parameters
 - **ligsets/** – A directory containing all the sets of ligands
 - **ligsets/LIGSET/** – for each set, there should be a directory within the **ligsets/** directory, whose name is the name of the ligand set. Substitute **LIGSET** for some appropriate name (e.g. **ligsets/my_awesome_ligset/**)
 - **ligsets/LIGSET/pdbqts/** – a directory containing a PDBQT file for each ligand in the set (whose name is **LIG.pdbqt**, with **LIG** being exactly the same as in **ligsets/LIGSET/LIGSET_list.txt**) [may be scripted later]
 - **ligsets/LIGSET/LIGSET_list.txt** – a text file containing a list of all the ligands (one on each line) and nothing else
 - you can create this easily on the command line using: `$ cd base_dir/ligsets/; for l in $(ls LIGSET/pdbqts | sed 's/.pdbqt//'); do echo $l >> LIGSET/LIGSET_list.txt; done` (appropriately substituting **base_dir** and **LIGSET**, and assuming the PDBQTs are already made)
 - [optional/preliminary] **ligsets/LIGSET/LIGSET.cdxml/** or **ligsets/LIGSET/LIGSET.mol/** – one file to show all the ligands in one page for presentations, PDFmaking and such.
 - [optional/preliminary] **/mols/** and **ligsets/LIGSET/pdbs/** – directories for preparing the initial PDBQT files. Will either be optional, or more scripts will be written.
 - **parameters.csvs/** – A directory containing small CSV files specifying the docking parameters for each individual docking (needed for scripts, at least as of now). Generated by **scripts/write_params_csv.R** from information in **Gridboxes.csv**
 - They will be named **parameters.csvs/DOCKING_parameters.csv**, where **DOCKING** is the docking ID
 - **vina_submit.shs/** – A directory containing the submission files for

Vina jobs on the (Wesleyan) cluster. Generated by `write_vina_submit` function of `docking_data_assembly.py`

- `vina_submit_shs/vina_submit DOCKING.sh` –for a docking of 20 models or less, a single submission script is written (and submitted using `$ bsub < vina_submit DOCKING.sh`, see submission instructions below)
- `vina_submits DOCKING/` –dockings of more than 20 models need to be submitted with multiple scripts (because Vina will not generate more than 20 poses). In this case, the `write_vina_submit` function will create a directory called `vina_submits DOCKING/` containing n scripts `vina_submit DOCKING.1.sh` , `vina_submit DOCKING.2.sh` through `vina_submit DOCKING.n.sh` . This script is set up to write each n submission scripts, where each of which script has a model number of 20 and n is the number of models divided by 20. Therefore, if greater than 20, the number of models should always be a multiple of twenty, or things will get messed up. (These are submitted used `$ for s in $(ls vina_submits DOCKING); do bsub < $s; done`, see instructions below)
- `PROTEIN/` , etc. – A directory for *each* protein, whose name is the name of the protein, the reference name/abbreviation used throughout, it just needs to be consistent (e.g. I primarily dock the proteins HepI and p300 and have the directories `hepi/` and `p300/` in my `base_dir/`)
 - `PROTEIN/PROTEIN.pdbqt` – the PDBQT file to be used for docking (`PROTEIN` must be *exactly* the same for the directory/file names and in the `Dockings.csv` and `Gridboxes.csv` entries for the protein’s dockings) [may be scripted later, from `PROTEIN.pdb`]
 - [optional/preliminary] `PROTEIN.pdb` – the original PDB file
 - `PROT/DOCKING/` – for every docking, there should be a directory whose name in the docking ID (the same as in `Dockings.csv`). Note: the user shouldn’t make this folder, it is made by `vina_submit DOCKING.sh`.
 - This will eventually contain
 - `PROT/binding_sites/` – a directory containing binding site PDBs:
 - To do binding site scoring by residues contacted (which is detected by the AutoDockTools script `process_VinaResult.py`),

There must be one or more `PROT/binding_sites/BINDING_SITE.pdb` files. These are subsets of the original `PROTEIN.pdb` (I originally created mine by grabbing the residues within 5Å of the bound ligands that came with the crystal structure, but it could be done many other ways).

- `scripts/` – all the scripts needed to use this set up
 - `write_params_csv.R` – writes `DOCK_parameters.csv` using information in `Dockings.csv` and `Gridboxes.csv`
 - `load_parameters.sh` – loads parameters from `DOCKING_parameters.csv` (used in the next script)
 - `separate_vina_results.sh` – runs Vina result PDBQTs through the AutoDockTools script `process_VinaResult.py`, which separates the poses into separate files and extracts the receptor contacts. The resulting files ended up in `DOCKING/processed_pdbqts/`, named `DOCKING.LIGAND_mMODEL.pdbqt`, where `MODEL` is the particular pose represented by the file. (A docking with l ligands and with Vina set to produce m models will therefore end up having $l \times m$ files after this script runs.)
 - `cleanup_processed_vina_results.sh` – cleans up processed PDBQTs (`DOCKING/processed_pdbqts/DOCKING.LIGAND_mMODEL.pdbqt`) and converts them to PDBs using the AutoDockTools script `pdbqt_to_pdb.py`
 - `parse_pdb.py` – defines a object class called `Pdb` for parsing PDB and PDBQT files for their 3D coordinates and in the case of processed Vina results, their binding energy, protein contacts, and other data generated by Vina. (Necessary for `docking_data_assembly.py`.)
 - `aiad_icpd.py` – defines functions to calculate the AIAD (averaged inter-atomic distance) and ICPD (inter-centerpoint distance) between two `Pdb` objects, two useful parameters for determining where a pose is binding on a protein and clustering poses together based on proximity. (Necessary for these functions in `docking_data_assembly.py`.)
 - `docking_data_assembly.py` – defines an object class called `Docking` for preparation and analysis of dockings. Contains several important functions:
 - `write_vina_submit` – prepares Vina job submission scripts
 - `assemble_dic` – assembles a data dictionary that contains all

- the mined data from the Vina results
- `score_binding_sites` – scores each pose for the proportion of residues contacted in each reference binding site
- `assess_all_resis` – uses binding scores to determine a True/-False for each pose binding in at each binding site (based on a threshold score, currently 0.1 or 10%)
- (`aiad_icpd_binding_sites`) – calculates AIAD and ICPD scores for each pose compared to each binding site
- `write_alldata.csv` – writes the data dictionary to a CSV file called `DOCKING_alldata.csv`
- `cluster_poses` [`prepare_clustering.csv.py`] – calculates AIAD scores for every pose compared to every other pose, for cluster analysis later on
- `pre.and.post.control.py` – links to all of the above scripts to coordinate their function, providing the global variables required to run this system on different computers.
- **Other scripts that may or may not be added:**
 - A script to add an entry to `Dockings.csv` or `Gridboxes.csv` with use input

1.3 Complete listing of included files

- **Notes:**

- ... indicates more of the same kind of file or directory
- .py files (except for the control script) may have a compiled .pyc file with them
- (files) are optional (for now)
- [files] are works in progress and may not be included ultimately
- [[files]] need to be made

```

1  /path/to/base_dir/
2  Readme.md
3  Dockings.csv
4  Gridboxes.csv
5  ligsets/
6  LIGSET1/
7  LIGSET1_list.txt
8  (LIGSET1.cdxml)
9  (mols/...)
10 (pdbs/...)

```

```

11         pdbqts/
12             LIG1_1.pdbqt
13             LIG1_2.pdbqt
14             LIG1_3.pdbqt
15         ...
16     LIGSET2/
17         LIGSET1_list.txt
18         (mols/...)
19         (pdbs/...)
20         pdbqts/
21             LIG2_1.pdbqt
22             LIG2_2.pdbqt
23             LIG2_3.pdbqt
24         ...
25     ...
26     vina_submit_shs/
27         vina_submit_A1.sh
28         vina_submits_A2/
29             vina_submit_A2.1.sh
30             vina_submit_A2.2.sh
31             vina_submit_A2.3.sh
32         ...
33         vina_submit_B1.sh
34     ...
35     PROT_A/
36         (PROT_A.pdb)
37         PROT_A.pdbqt
38         binding_sites/
39             BINDING_SITE_ALPHA1.pdb
40             BINDING_SITE_ALPHA2.pdb
41             BINDING_SITE_ALPHA3.pdb
42         ...
43     A1/
44     A2/
45     A3/
46     ...
47     PROT_B/
48         (PROT_B.pdb)
49         PROT_B.pdbqt
50         binding_sites/
51             BINDING_SITE_BETA1.pdb
52         ...
53     B1/
54     B2/
55     ...

```

```

56     ...
57     scripts/
58         [new_grid_or_dock_entry.R]
59         [[write_ligset_list_txt.sh]]
60         load_parameters.sh
61         [[ligand, protein preparation]]
62         separate_vina_results.sh
63         cleanup_processed_vina_results.sh
64         parse_pdb.py
65         aiad_icpd.py
66         [[prepare_clustering_csv.py]]
67         docking_data_assembly.py
68         pre_and_post_control.py
69         [[post_docking_graphs.R]]
70         [[clustering_graphs.R]]
71         [[R cript to select poses to view in PyMol]]
72         [[Py script to load PyMol sessions from lists]]

```

1.3.1 After docking post-processing:

```

1     PROTEIN/
2         (PROTEIN.pdb)
3         PROTEIN.pdbqt
4         binding_sites/
5             SITE1.pdb
6             SITE1.pdb
7             SITE1.pdb
8         ...
9     DOCKING/
10         result_pdbqts/
11             DOCKING_LIG1_results.pdbqt
12             DOCKING_LIG2_results.pdbqt
13             DOCKING_LIG3_results.pdbqt
14         ...
15         processed_pdbqts/
16             DOCKING_LIG1_m1.pdbqt
17             DOCKING_LIG1_m2.pdbqt
18             DOCKING_LIG1_m3.pdbqt
19         ...
20             DOCKING_LIG2_m1.pdbqt
21             DOCKING_LIG2_m2.pdbqt
22             DOCKING_LIG2_m3.pdbqt

```

```

23         ...
24         DOCKING_LIG3_m1.pdbqt
25         DOCKING_LIG3_m2.pdbqt
26         DOCKING_LIG3_m3.pdbqt
27         ...
28     processed_pdb/
29         DOCKING_LIG1_m1.pdb
30         DOCKING_LIG1_m2.pdb
31         DOCKING_LIG1_m3.pdb
32         ...
33         DOCKING_LIG2_m1.pdb
34         DOCKING_LIG2_m2.pdb
35         DOCKING_LIG2_m3.pdb
36         ...
37         DOCKING_LIG3_m1.pdb
38         DOCKING_LIG3_m2.pdb
39         DOCKING_LIG3_m3.pdb
40         ...
41     DOCKING_alldata.csv
42     DOCKING.p
43     DOCKING_clustering.csv [[DOCKING_pose_pose_aiads.csv]]
44     [[DOCKING_best_aiad_pairs.csv]]
45     [[graphs/]]
46     [[...graphs...]]

```

1.4 Example Dockings.csv file

```

1 Docking ID,Date,Protein,Ligset,Grid box,Exhaustiveness,Number of Models,Number
  of CPUs,Notes
2 A1,20160301,PROTA,LIGS1,AAS,20,10,2,looking at active size of protein A
3 A2,20160308,PROTA,LIGS2,AWP,50,400,4,high volume docking of whole protein A
4 B1,20160308,PROTB,LIGS3,BWP,8,20,1,initial docking of whole protein B

```

1.5 Example Gridboxes.csv file

```

1 Gridbox Name,Protein,Size in x-dimension,Size in y-dimension,Size in z-
  dimension,Center in x-dimension,Center in y-dimension,Center in z-
  dimension,Notes

```


Docking ID	Date	Protein	Ligset	Grid box	Exhaustiveness	Number of Models	Number of CPUs	Notes
A1	20160301	PROTA	LIGS1	AAS	20	10	2	looking at active size of protein A
A2	20160308	PROTA	LIGS2	AWP	50	400	4	high volume docking of whole protein A
B1	20160308	PROTB	LIGS3	BWP	8	20	1	initial docking of whole protein B

² AAS,PROTA,60,72,88,41.89,2.69,-1.85,active site of protein A
³ AWP,PROTA,126,126,126,41.89,2.69,-1.85,all of protein A
⁴ BWP,PROTB,126,126,126,4.89,-5.27,12.0,all of protein B

Gridbox Name	Protein	Box size (x)	Box size (y)	Box size (z)	Box center (x)	Box center (y)	Box center (z)	Notes
AAS	PROTA	60	72	88	41.89	2.69	-1.85	active site of protein A
AWP	PROTA	126	126	126	41.89	2.69	-1.85	all of protein A
BWP	PROTB	126	126	126	4.89	-5.27	12.0	all of protein B

2 Scripts

- constants.py (*,**)
- new_grid_or_dock_entry.py (*)
- create_docking_object.py (*)
- write_vina_submit_sh.py (*)
- read_alldata.py (*)
- cleanup_processed_vina_results.sh [pr]
- separate_vina_results.sh [pr]
- parse_pdb.py (*, sa)
- mine_vina_data.py (*)
- aiad_icpd.py (*, sa)
- binding_site_analysis.py (*)
- energies_properties.py (*, sa)
- get_pose_energies_properties.py (*)
- write_alldata.py (*)
- correlations.py (*)
- [postdocking_summary.R]
- postdocking_graphs.R (nc)
- pre_and_post_control.py

2.1 constants.py

2.1.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  ### Module-wide constants
2  # (c) Zarek Siegel
3  # v1 3/11/16
4  # v2 3/15/16
5
6  base_dir="/Users/zarek/GitHub/TaylorLab/zvina"
7  AutoDockTools_dir="/Library/MGLTools/latest/MGLToolsPckgs/AutoDockTools"
8  AutoDockTools_pythonsh_binary="/Library/MGLTools/latest/bin/pythonsh"
9
10 python_binary="/anaconda/envs/python27/bin/python"
11 Rscript_binary="/usr/bin/Rscript"
12 openbabel_binaries_dir="/usr/local/bin"
13
14 cluster_base_dir="/home/zsiegel"
15 cluster_vina_binary="/share/apps/autodock/autodock_vina_1_1_2_linux_x86/bin/
16     vina"
17 cluster_AutoDockTools_dir="/home/apps/CENTOS6/mgltools/1.5.6/MGLToolsPckgs/
18     AutoDockTools"
19 cluster_AutoDockTools_pythonsh_binary="/home/apps/CENTOS6/mgltools/1.5.6/bin/
20     pythonsh"
21
22 # base_dir="/home/zsiegel"
23 # AutoDockTools_dir="/home/apps/CENTOS6/mgltools/1.5.6/MGLToolsPckgs/
24     AutoDockTools"
25 # AutoDockTools_pythonsh_binary="/home/apps/CENTOS6/mgltools/1.5.6/bin/
26     pythonsh"
27 #
28 # python_binary="/share/apps/python/2.7.2/bin/python"
29 # Rscript_binary="/share/apps/R/3.1.0/bin/Rscript"
30 # openbabel_binaries_dir="/share/apps/openbabel/2.2.1/bin"
```

/Users/zarek/lab/zvina/scripts/constants.py

2.2 new_grid_or_dock_entry.py

2.2.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Write a new entry to Dockings.csv or Gridboxes.csv
4  # (c) Zarek Siegel
5  # v1 3/10/16
6  # v1.1 3/11/16
7
8  import csv, re, argparse, time, datetime
9  from constants import *
10
11 class Timestamp():
12     def __init__(self):
13         time_obj = time.time()
14         self.display = datetime.datetime.fromtimestamp(time_obj).strftime('%Y-%m-%d %H:%M:%S')
15         self.eightdigit = datetime.datetime.fromtimestamp(time_obj).strftime('%Y%m%d')
16
17 def new_docking_entry():
18     # Hello
19     print("\n\tWelcome! This script will add an entry to Dockings.csv\n")
20
21     # Dockings.csv columns:
22     # Docking ID
23     # Date
24     # Protein
25     # Protein File
26     # Ligset
27     # Gridbox
28     # Exhaustiveness
29     # Number of Models
30     # Number of CPUs
31     # Notes
32
33     # Going through each variable, taking user input
34     dock_input = raw_input("\t\tDocking identifier: ")
35     print("\t\t>>> dock set to: {}\n".format(dock_input))
36
```

```

37 current_time = Timestamp()
38 date_input = raw_input("\t\tDate (enter 'd' to default to {}): ".format(
    current_time.eightdigit))
39 if date_input == "d": date_input = current_time.eightdigit
40 print("\t\t>>> date set to: {}\n".format(date_input))
41
42 prot_input = raw_input("\t\tProtein: ")
43 print("\t\t>>> prot set to: {}\n".format(prot_input))
44
45 prot_file_input = raw_input("\t\tSpecific protein file (without the .pdbqt):
    ")
46 print("\t\t>>> prot_file set to: {}\n".format(prot_file_input))
47
48 ligset_input = raw_input("\t\tLigset identifier: ")
49 print("\t\t>>> ligset set to: {}\n".format(ligset_input))
50
51 box_input = raw_input("\t\tGridbox identifier: ")
52 print("\t\t>>> box set to: {}\n".format(box_input))
53
54 exhaust_input = raw_input("\t\tExhaustiveness: ")
55 print("\t\t>>> exhaust set to: {}\n".format(exhaust_input))
56
57 n_models_input = raw_input("\t\tNumber of models: ")
58 print("\t\t>>> n_models set to: {}\n".format(n_models_input))
59
60 n_cpus_input = raw_input("\t\tNumber of CPUs: ")
61 print("\t\t>>> n_cpus set to: {}\n".format(n_cpus_input))
62
63 notes_input = raw_input("\t\tAny notes (with no commas): ")
64 if notes_input == "":
65     notes_input = "Entered by new_grid_or_dock_entry.py {}".format(
        current_time.display)
66 else:
67     notes_input = "{} (Entered by new_grid_or_dock_entry.py {})".format(
        notes_input, current_time.display)
68 print("\t\t>>> notes set to: {}\n".format(notes_input))
69
70
71 # New row as a dictionary
72 new_row = {
73     'Docking ID' : dock_input,
74     'Date' : date_input,
75     'Protein' : prot_input,
76     'Protein File' : prot_file_input,
77     'Ligset' : ligset_input,
78     'Gridbox' : box_input,

```

```

79     'Exhaustiveness' : exhaust_input,
80     'Number of Models' : n_models_input,
81     'Number of CPUs' : n_cpus_input,
82     'Notes': notes_input
83 }
84
85 # Print confirmation of all entered variables
86 print("\t>>> The new row will be\n\n\
87     Docking ID: {dock}\n\
88     Date: {date}\n\
89     Protein: {prot}\n\
90     Protein File: {prot_file}\n\
91     Ligset: {ligset}\n\
92     Gridbox: {box}\n\
93     Exhaustiveness: {exhaust}\n\
94     Number of Models: {n_models}\n\
95     Number of CPUs: {n_cpus}\n\
96     Notes: {notes}\n".format(
97         dock = dock_input,
98         date = date_input,
99         prot = prot_input,
100         prot_file = prot_file_input,
101         ligset = ligset_input,
102         box = box_input,
103         exhaust = exhaust_input,
104         n_models = n_models_input,
105         n_cpus = n_cpus_input,
106         notes = notes_input
107     )
108 )
109
110 # Don't write row without confirmation
111 proceed = raw_input("\tWrite this as a new docking entry? [y/n] ")
112 # print("\t{}".format(new_row))
113
114 # If "y" is entered, write the row, otherwise don't
115 if proceed == "y":
116     dockings_csv = "{b_d}/Dockings.csv".format(b_d=base_dir)
117     dockings_headers = ["Docking ID", "Date", "Protein", "Protein File",
118         "Ligset", "Gridbox", "Exhaustiveness", "Number of Models",
119         "Number of CPUs", "Notes"]
120     with open(dockings_csv, 'a') as f:
121         appender = csv.DictWriter(f, fieldnames=dockings_headers)
122         appender.writerow(new_row)
123     print("\n\t>>> New row appended to Dockings.csv:\n\topen {}".format(

```

```

dockings_csv))
124 else:
125     print("\n\t>>> No docking entry written\n")
126
127 def new_gridbox_entry():
128     # Hello
129     print("\n\tWelcome! This script will add an entry to Gridboxes.csv\n")
130
131     # Gridboxes.csv columns:
132     # Gridbox Name
133     # Protein File
134     # Size in x-dimension
135     # Size in y-dimension
136     # Size in z-dimension
137     # Center in x-dimension
138     # Center in y-dimension
139     # Center in z-dimension
140     # Notes
141
142     # Going through each variable, taking user input
143     box_input = raw_input("\t\tGridbox Name: ")
144     print("\t\t>>> box set to: {}\n".format(box_input))
145
146     prot_file_input = raw_input("\t\tSpecific protein file (without the .pdbqt): ")
147     print("\t\t>>> prot_file set to: {}\n".format(prot_file_input))
148
149     box_size_x_input = raw_input("\t\tSize in x-dimension: ")
150     print("\t\t>>> box_size_x set to: {}\n".format(box_size_x_input))
151
152     box_size_y_input = raw_input("\t\tSize in y-dimension: ")
153     print("\t\t>>> box_size_y set to: {}\n".format(box_size_y_input))
154
155     box_size_z_input = raw_input("\t\tSize in z-dimension: ")
156     print("\t\t>>> box_size_z set to: {}\n".format(box_size_z_input))
157
158     box_center_x_input = raw_input("\t\tCenter in x-dimension: ")
159     print("\t\t>>> box_center_x set to: {}\n".format(box_center_x_input))
160
161     box_center_y_input = raw_input("\t\tCenter in y-dimension: ")
162     print("\t\t>>> box_center_y set to: {}\n".format(box_center_y_input))
163
164     box_center_z_input = raw_input("\t\tCenter in z-dimension: ")
165     print("\t\t>>> box_center_z set to: {}\n".format(box_center_z_input))
166

```

```

167     current_time = Timestamp()
168     notes_input = raw_input("\t\tAny notes (with no commas): ")
169     if notes_input == "":
170         notes_input = "Entered by new_grid_or_dock_entry.py {}".format(
            current_time.display)
171     else:
172         notes_input = "{} (Entered by new_grid_or_dock_entry.py {})".format(
            notes_input, current_time.display)
173     print("\t\t>>> notes set to: {}\n".format(notes_input))
174
175     # New row as a dictionary
176     new_row = {
177         'Gridbox Name' : box_input,
178         'Protein File' : prot_file_input,
179         'Size in x-dimension' : box_size_x_input,
180         'Size in y-dimension' : box_size_y_input,
181         'Size in z-dimension' : box_size_z_input,
182         'Center in x-dimension' : box_center_x_input,
183         'Center in y-dimension' : box_center_y_input,
184         'Center in z-dimension' : box_center_z_input,
185         'Notes' : notes_input
186     }
187
188     # Print confirmation of all entered variables
189     print("\t\t>>> The new row will be\n\n\
190     Gridbox Name: {box}\n\
191     Protein File: {prot_file}\n\
192     Size in x-dimension: {box_size_x}\n\
193     Size in y-dimension: {box_size_y}\n\
194     Size in z-dimension: {box_size_z}\n\
195     Center in x-dimension: {box_center_x}\n\
196     Center in y-dimension: {box_center_y}\n\
197     Center in z-dimension: {box_center_z}\n\
198     Notes: {notes}\n".format(
199         box = box_input,
200         prot_file = prot_file_input,
201         box_size_x = box_size_x_input,
202         box_size_y = box_size_y_input,
203         box_size_z = box_size_z_input,
204         box_center_x = box_center_x_input,
205         box_center_y = box_center_y_input,
206         box_center_z = box_center_z_input,
207         notes = notes_input
208     )
209 )
210 )

```



```

211
212 # Don't write row without confirmation
213 proceed = raw_input("\tWrite this as a new grid box entry? [y/n] ")
214
215 # If "y" is entered, write the row, otherwise don't
216 if proceed == "y":
217     gridboxes_csv = "{b_d}/Gridboxes.csv".format(b_d=base_dir)
218     gridboxes_headers = ["Gridbox Name", "Protein File", "Size in x-dimension"
219 ,
220     "Size in y-dimension", "Size in z-dimension", "Center in x-dimension",
221     "Center in y-dimension", "Center in z-dimension", "Notes"]
222     with open(gridboxes_csv, 'a') as f:
223         appender = csv.DictWriter(f, fieldnames=gridboxes_headers)
224         appender.writerow(new_row)
225         print("\n\t>>> New row appended to Gridboxes.csv:\n\topen {}\n".format(
226             gridboxes_csv))
227     else:
228         print("\n\t>>> No grid box entry written\n")
229
230 # Stuff below is commented because this script is being used as a module
231
232 # def main():
233 #     parser = argparse.ArgumentParser(
234 #         description='Write a new entry to Dockings.csv or Gridboxes.csv')
235 #     parser.add_argument('-b', '--base_dir', metavar='BASE_DIR', type=str,
236 #         nargs=1,
237 #         help='The base directory containing Docking.csv and Gridboxes.csv')
238 #     parser.add_argument('-d', '--new_docking', action='store_true', default=
239 #         False,
240 #         help='New set of docking parameters (written to Dockings.csv)')
241 #     parser.add_argument('-g', '--new_gridbox', action='store_true', default=
242 #         False,
243 #         help='New set of grid box parameters (written to Gridboxes.csv)')
244 #
245 #     args = vars(parser.parse_args())
246 #     global base_dir
247 #     base_dir = str(args['base_dir'])[0])
248 #     new_docking = args['new_docking']
249 #     new_gridbox = args['new_gridbox']
250 #
251 #     if new_docking: new_docking_entry()
252 #     elif new_gridbox: new_gridbox_entry()
253 #
254 # if __name__ == "__main__": main()

```

`/Users/zarek/lab/zvina/scripts/new_grid_or_dock_entry.py`

2.3 create_docking_object.py

2.3.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Putting together all parsed data from processed vina results
4  # (c) Zarek Siegel
5  # v1 3/5/16 (as assemble_alldata.py)
6  # v2 3/6/16
7
8  from __future__ import print_function
9  import csv, re, os, subprocess
10 from constants import *
11
12 ### A class for docking data, for sourcing, reading, and analyzing
13 class Docking():
14     # Basic docking parameters are contained in CSV file in ../base/
15     # parameters_csvs/
16     def load_parameters(self):
17         # Basic docking parameters are stored in base_dir/Dockings.csv
18         dockings_csv = "{}Dockings.csv".format(base_dir)
19         with open(dockings_csv) as f:
20             reader = csv.DictReader(f)
21             for row in reader:
22                 if row['Docking ID'] == self.dock:
23                     self.dockings_csv_row = row
24
25         # Source the parameters as class attributes
26         self.prot = self.dockings_csv_row['Protein']
27         self.prot_file = self.dockings_csv_row['Protein File']
28         self.ligset = self.dockings_csv_row['Ligset']
29         self.box = self.dockings_csv_row['Gridbox']
30         self.exhaust = self.dockings_csv_row['Exhaustiveness']
31         self.n_models = int(self.dockings_csv_row['Number of Models'])
32         self.n_cpus = self.dockings_csv_row['Number of CPUs']
33         self.notes = self.dockings_csv_row['Notes']
34         self.date = self.dockings_csv_row['Date']
35
36         # Grid box parameters are stored in base_dir/Gridboxes.csv
37         gridboxes_csv = "{}Gridboxes.csv".format(base_dir)
38         with open(gridboxes_csv) as f:
39             reader = csv.DictReader(f)
```

```

38     for row in reader:
39         if row['Gridbox Name'] == self.box and row['Protein File'] == self.
prot_file:
40             self.gridboxes_csv_row = row
41
42     # Source the grid box parameters as class attributes
43     self.box_center_x = self.gridboxes_csv_row['Center in x-dimension']
44     self.box_center_y = self.gridboxes_csv_row['Center in y-dimension']
45     self.box_center_z = self.gridboxes_csv_row['Center in z-dimension']
46     self.box_size_x = self.gridboxes_csv_row['Size in x-dimension']
47     self.box_size_y = self.gridboxes_csv_row['Size in y-dimension']
48     self.box_size_z = self.gridboxes_csv_row['Size in z-dimension']
49     self.box_notes = self.gridboxes_csv_row['Notes']
50
51     # Some useful directories
52     self.prot_dir = "{b_d}/{p}".format(b_d=base_dir, p=self.prot)
53     self.dock_dir = "{b_d}/{p}/{d}".format(b_d=base_dir, p=self.prot, d=self.
dock)
54
55     self.parameters_loaded = True
56     print("    > Loaded docking parameters")
57
58     # Retrieve the list of ligands in the set
59     def get_ligset_list(self):
60         # If there is a ligset text file; source it;
61         # else write it by looking at the files in LIGSET/pdbqts
62         self.ligset_list_txt = "{b_d}/ligsets/{ls}/{ls}_list.txt".format(
63             b_d=base_dir, ls=self.ligset)
64         if os.path.isfile(self.ligset_list_txt):
65             with open(self.ligset_list_txt, 'r') as f:
66                 self.ligset_list = f.read().splitlines()
67             print("    > Ligset list sourced from .../{ls}/{ls}_list.txt".format(
68                 ls=self.ligset))
69         else:
70             self.ligset_dir = "{b_d}/ligsets/{ls}/pdbqts".format(
71                 b_d=base_dir, ls=self.ligset)
72             self.ligset_list = subprocess.Popen(["ls", self.ligset_dir], stdout=
subprocess.PIPE)
73             self.ligset_list = self.ligset_list.communicate()[0]
74             self.ligset_list = re.sub('.pdbqt', '', self.ligset_list)
75             self.ligset_list = re.split('\n', self.ligset_list)
76             for l in self.ligset_list:
77                 if l == '' or l == ' ':
78                     self.ligset_list.remove(l)
79             print("    > Ligset list sourced by looking at files in .../{ls}/pdbqts\

```

```

80         "                and saved as ../{ls}/{ls}_list.txt".format(ls=self.
ligset)))
81     with open(self.ligset_list_txt, 'w') as f:
82         for l in self.ligset_list:
83             f.write("{}\n".format(l))
84     self.ligset_list_str = str(self.ligset_list)
85     self.ligset_list_str = re.sub('[\[\]\[\]\],]', '', self.ligset_list_str)
86
87     self.ligset_list_gotten = True
88     print("    > Retrieved ligset list")
89
90     #Create blank alldata dictionary
91     def create_data_dic(self):
92         self.data_dic = {}
93         self.keys = []
94         for lig in self.ligset_list:
95             for m in range(1, self.n_models + 1):
96                 key = "{}_{}_m{}".format(self.dock, lig, m)
97                 self.data_dic[key] = {
98                     'key' : key,
99                     'lig' : lig,
100                     'model' : m
101                 }
102                 self.keys.append(key)
103
104     self.is_data_dic_created = True
105     print("    > Created empty data dictionary")
106
107     def print_parameters(self):
108         print("dock=\ '{}\ ' ".format(self.dock))
109         print("prot=\ '{}\ ' ".format(self.prot))
110         print("prot_file=\ '{}\ ' ".format(self.prot_file))
111         print("ligset=\ '{}\ ' ".format(self.ligset))
112         print("box=\ '{}\ ' ".format(self.box))
113         print("exhaust={}".format(self.exhaust))
114         print("n_models={}".format(self.n_models))
115         print("n_cpus={}".format(self.n_cpus))
116         print("box_center_x={}".format(self.box_center_x))
117         print("box_center_y={}".format(self.box_center_y))
118         print("box_center_z={}".format(self.box_center_z))
119         print("box_size_x={}".format(self.box_size_x))
120         print("box_size_y={}".format(self.box_size_y))
121         print("box_size_z={}".format(self.box_size_z))
122         print("ligset_list=\ '{}\ ' ".format(self.ligset_list_str))

```

```

123     print("notes=\'{ }\'".format(self.notes))
124     print("date={}".format(self.date))
125     print("box_notes=\'{ }\'".format(self.box_notes))
126
127     def export_parameters_to_environment(self):
128         os.environ['dock'] = "{}".format(self.dock)
129         os.environ['prot'] = "{}".format(self.prot)
130         os.environ['prot_file'] = "{}".format(self.prot_file)
131         os.environ['ligset'] = "{}".format(self.ligset)
132         os.environ['box'] = "{}".format(self.box)
133         os.environ['exhaust'] = "{}".format(self.exhaust)
134         os.environ['n_models'] = "{}".format(self.n_models)
135         os.environ['n_cpus'] = "{}".format(self.n_cpus)
136         os.environ['box_center_x'] = "{}".format(self.box_center_x)
137         os.environ['box_center_y'] = "{}".format(self.box_center_y)
138         os.environ['box_center_z'] = "{}".format(self.box_center_z)
139         os.environ['box_size_x'] = "{}".format(self.box_size_x)
140         os.environ['box_size_y'] = "{}".format(self.box_size_y)
141         os.environ['box_size_z'] = "{}".format(self.box_size_z)
142         os.environ['ligset_list'] = "{}".format(self.ligset_list_str)
143         os.environ['notes'] = "{}".format(self.notes)
144         os.environ['date'] = "{}".format(self.date)
145         os.environ['box_notes'] = "{}".format(self.box_notes)
146
147         self.parameters_exported_to_environment = True
148         print("    > Exported parameters to shell environment")
149
150
151     def set_recordkeeping_parameters(self):
152         self.parameters_loaded = False
153         self.ligset_list_gotten = False
154         self.parameters_exported_to_environment = False
155         self.is_data_dic_created = False
156
157         self.vina_data_mined = False
158         self.binding_sites_list_gotten = False
159         self.binding_sites_scored = False
160         self.aiad_icpd_calcd = False
161         self.all_resis_assessed = False
162
163         self.is_csv_written = False
164         self.energies_props_gotten = False
165         self.are_poses_clustered = False
166         self.is_pickled = False
167

```

```
168 def __init__(self, d):
169     self.dock = d
170
171     print("----> Creating docking object for docking {}".format(self.dock))
172
173     self.set_recordkeeping_parameters()
174     self.load_parameters()
175     self.get_ligset_list()
176     self.create_data_dic()
177
178     print(" ")
```

/Users/zarek/lab/zvina/scripts/create_docking_object.py

2.4 write_vina_submit_sh.py

2.4.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic
sugar eclipse rust bug. Back-face culling lua javabean indirection operator
void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Putting together all parsed data from processed vina results
4  # (c) Zarek Siegel
5  # v1 3/5/16 (as assemble_alldata.py)
6  # v2 3/6/16
7
8  from __future__ import print_function
9  import subprocess, os, sys
10 from constants import *
11 from create_docking_object import * # Docking
12
13 def write_vina_submit_sh(self):
14     # (no need for batch submission)
15     if self.n_models <= 20:
16         batch_submission = False
17         vina_submit_sh = "{b_d}/vina_submit_shs/vina_submit_{d}.sh".format(
18             b_d=base_dir, d=self.dock)
19         if os.path.isfile(vina_submit_sh):
20             print(">>> Vina submission script already exists at")
21             print("\t{}".format(vina_submit_sh))
22             overwrite = raw_input("\n\t>>> Enter 'y' or enter to overwrite, 'n' to
                exit: ")
23
24             if overwrite == "y" or \
25                 overwrite == "yes" or \
26                 overwrite == "Y" or \
27                 overwrite == "Yes" or \
28                 overwrite == "":
29                 subprocess.call(["rm", "-f", vina_submit_sh])
30                 print("")
31             else: sys.exit("\n\t> OK, exiting this script\n")
32
33     # (batch submission)
34     elif self.n_models > 20:
35         batch_submission = True
36         vina_submits_dir = "{b_d}/vina_submit_shs/vina_submits_{d}/".format(
37             b_d=base_dir, d=self.dock)
```



```

38     if os.path.isdir(vina_submits_dir):
39         print(">>> Vina submission scripts already exist at")
40         print("\t{}".format(vina_submits_dir))
41         overwrite = raw_input("\n\t>>> Enter 'y' or enter to overwrite, 'n' to
exit: ")
42
43         if overwrite == "y" or \
44             overwrite == "yes" or \
45             overwrite == "Y" or \
46             overwrite == "Yes" or \
47             overwrite == "":
48             subprocess.call(["rm", "-rf", vina_submits_dir])
49             print("")
50             else: sys.exit("\n\t> OK, exiting this script\n")
51
52     else: print("! ! ! bad n_models")
53
54     template = (
55         "#BSUB -q hp12\n"
56         "#BSUB -n {self.n_cpus}\n"
57         "#BSUB -N\n"
58         "#BSUB -o {cluster_base_dir}/bsub_logs/vina_{subdock}_log.txt\n"
59         "#BSUB -J vina_{subdock}\n"
60         "\n"
61         "# Parameters"
62         "dock={self.dock}\n"
63         "ligset={self.ligset}\n"
64         "ligset_list=\"{self.ligset_list_str}\"\n"
65         "\n"
66         "# Create the docking and output directories\n"
67         "mkdir {cluster_base_dir}/{self.prot}/{self.dock}/\n"
68         "mkdir {cluster_base_dir}/{self.prot}/{self.dock}/result_pdbqts\n"
69         "\n"
70         "vina_start_time=$(date \"+%Y%m%d%H%M%S\")\n"
71         "printf '\n~~> Vina docking %s started %s' \"$dock\" \"$vina_start_time
\n"
72         "\n"
73         "# Vina command\n"
74         "for lig in $ligset_list\n"
75         "do\n"
76         "    lig_start_time=$(date \"+%Y%m%d%H%M%S\")\n"
77         "    printf '\n\n> Docking ligand <s> starting at %s\n' \"$lig\" \"
$lig_start_time\"\n"
78         "    /share/apps/autodock/autodock_vina_1_1_2_linux_x86/bin/vina \\\n"
79         "    --receptor {cluster_base_dir}/{self.prot}/{self.prot_file}.pdbqt \\\n"

```

```

80     " --ligand {cluster_base_dir}/ligsets/{self.ligset}/pdbqts/$lig.pdbqt \\n
81     "
82     " --out {cluster_base_dir}/{self.prot}/{self.dock}/result_pdbqts/{subdock}
83     _$lig\_results.pdbqt \\n"
84     " --center_x {self.box_center_x} \\n"
85     " --center_y {self.box_center_y} \\n"
86     " --center_z {self.box_center_z} \\n"
87     " --size_x {self.box_size_x} \\n"
88     " --size_y {self.box_size_y} \\n"
89     " --size_z {self.box_size_z} \\n"
90     " --cpu {self.n_cpus} \\n"
91     " --num_modes {self.n_models} \\n"
92     " --exhaustiveness {self.exhaust}\n"
93     " lig_end_time=$(date \"+%Y%m%d%H%M%S\")\n"
94     " printf '> Finished at %s' \"$lig_start_time\"\n"
95     " lig_duration=$(bc <<< \"$lig_end_time - $lig_start_time\")\n"
96     " printf '\\n> Docking of ligand %s took %s seconds' \"$lig\" \"
97     $lig_duration\"\n"
98     "done\n"
99     "\n"
100    "vina_end_time=$(date \"+%Y%m%d%H%M%S\")\n"
101    "printf '\\n\\n--> Vina job finished %s\\n' \"$vina_end_time\"\n"
102    "\n"
103    "vina_duration=$(bc <<< \"$vina_end_time - $vina_start_time\")\n"
104    "printf '\\n--> Docking %s of ligset %s took %s seconds \\n\\n' \"$dock\"
105    \"$lig\" \"$vina_duration\"\n"
106
107    )
108
109    template = template.format(
110        cluster_base_dir = cluster_base_dir,
111        self=self,
112        n_models = '{n_models}',
113        dock = '{dock}',
114        subdock = '{subdock}'
115    )
116
117    # (no need for batch submission)
118    if not batch_submission:
119        template_filled = template.format(
120            n_models = self.n_models, dock = self.dock, subdock = self.dock)
121        with open(vina_submit_sh, 'w') as f:
122            f.write(template_filled)
123        print("--> Vina submission script for docking {} has been created. It can
124            be found at:".format(self.dock))
125        print("\t{}".format(vina_submit_sh))

```

```

120 # (batch submission)
121 elif batch_submission:
122     subprocess.call(['mkdir', vina_submits_dir])
123     n_batches = self.n_models / 20
124     for b in range(1, n_batches + 1):
125         subdock = "{d}.{b}".format(d = self.dock, b = b)
126         template_filled = template.format(
127             n_models = 20, dock = self.dock, subdock = subdock)
128         vina_submit_sh = "{v_s_d}/vina_submit_{sd}.sh".format(
129             v_s_d = vina_submits_dir, sd = subdock)
130         with open(vina_submit_sh, 'w') as f:
131             f.write(template_filled)
132         print("---> Vina submission scripts for docking h11 have been created.
133             They can be found in:")
134         print("\t{}".format(vina_submits_dir))
135 Docking.write_vina_submit_sh = write_vina_submit_sh

```

/Users/zarek/lab/zvina/scripts/write_vina_submit_sh.py

2.5 read_alldata.py

2.5.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Output all the data mined and analyzed into a CSV file
4  # (c) Zarek Siegel
5
6  import os, sys
7  import cPickle as pickle
8  from create_docking_object import * # Docking
9
10
11 def read_alldata_csv(self):
12     self.alldata_csv = "{d_d}/{d}_alldata.csv".format(d_d=self.dock_dir, d=self.
13         dock)
14
15     print("---> Reading alldata.csv\n")
16     with open(self.alldata_csv, 'r') as csvfile:
17         reader = csv.DictReader(csvfile)
18         for row in reader:
19             self.data_dic[row['key']] = row
20     print("! ! ! Warning, it is better to load from the whole object pickled
21         versus just the data from a CSV")
22
23
24 Docking.read_alldata_csv = read_alldata_csv
25
26 # Save the data dictionary as a pickled file (i.e. in native python format)
27 def read_docking_pickled(self):
28     self.docking_obj_pickled = "{d_d}/{d}.p".format(d_d=self.dock_dir, d=self.
29         dock)
30
31     print("---> Reading docking.p\n")
32     self = pickle.load(open(self.docking_obj_pickled, 'rb'))
33
34 Docking.read_docking_pickled = read_docking_pickled
```

/Users/zarek/lab/zvina/scripts/read_alldata.py

2.6 cleanup_processed_vina_results.sh

2.6.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1 #!/bin/bash
2
3 ### Converting and cleaning up processed vina result pdbqts
4 # (c) Zarek Siegel
5 # v1 3/5/16
6 # v1.2 3/6/16
7
8 ### Required input
9 dock=$1
10 # Set scripts directory to the directory containing this script
11 scripts_dir="$( cd "$( dirname "${BASH_SOURCE[0]}" )" && pwd )"
12 # Set base directory to the one containing scripts_dir
13 base_dir="$( cd $scripts_dir && cd .. )"
14 # Source # AutoDockTools Directory and MGLTools Python binary paths from
15   constants.py
16 source $scripts_dir/constants.py
17 # Location of pdbqt_to_pdb
18 q2b_py="$AutoDockTools_dir/Utilities24/pdbqt_to_pdb.py"
19
20 # Retrieve the parameters for this docking
21 # source $base_dir/scripts/load_parameters.sh $dock
22
23 # Relevant directories
24 processed_pdbqts_dir=$base_dir/$prot/$dock/processed_pdbqts
25 cleanedup_processed_pdbqts_dir=$base_dir/$prot/$dock/
26   cleanedup_processed_pdbqts
27 processed_pdb_dir=$base_dir/$prot/$dock/processed_pdb
28
29 # Check if already done
30 if [ -d $processed_pdb_dir ]; then
31   echo " ! Results already cleaned up (processed_pdb exists), exiting this
32   step"
33   exit 1
34 fi
35
36 # Create a directory for cleaned up files and pdb converts
37 mkdir $cleanedup_processed_pdbqts_dir
38 mkdir $processed_pdb_dir
```

```

36
37 # Retrieve ligset list
38 ligset_list_txt=$base_dir/ligsets/$ligset/$ligset\_list.txt
39 ligset_list=$(for l in $(cat $ligset_list_txt); do echo $l; done)
40
41 # The clean-up step
42 for lig in $ligset_list; do
43     for ((m=1;m<=$n_models;m++)); do
44         processed_pdbqt=$processed_pdbqts_dir/$dock\_lig\_m$m.pdbqt
45         cleanedup_processed_pdbqt=$cleanedup_processed_pdbqts_dir/$dock\_lig\_m$m
46         .pdbqt
47         processed_pdb=$processed_pdb_dir/$dock\_lig\_m$m.pdb
48
49         # The clean-up step
50         cat $processed_pdbqt | \
51             sed 's/^\(HETATM.....\).*/\1LIG L/g' \
52             > $cleanedup_processed_pdbqt
53
54         # The PDBQT > PDB Conversion step
55         $AutoDockTools_pythonsh_binary $q2b_py -f $cleanedup_processed_pdbqt \
56             -o $processed_pdb \
57             1 > /dev/null
58
59         echo "----> processed ligand $lig model $m"
60     done
61 done
62
63 # Overwrite pre-clean-up pvr'd pdbqts with cleaned up ones
64 rm -rf $processed_pdbqts_dir
65 mv $cleanedup_processed_pdbqts_dir $processed_pdbqts_dir

```

/Users/zarek/lab/zvina/scripts/cleanup_processed_vina_results.sh

2.7 separate_vina_results.sh

2.7.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/bin/bash
2
3  ### process_VinaResult and a bit of organization
4  # (c) Zarek Siegel
5  # v1 3/5/16
6  # v1.2 3/6/16
7  # v2 3/6/16 (batch separation)
8  # v3 3/11/16
9
10 ### Required input
11 dock=$1
12 # Set scripts directory to the directory containing this script
13 scripts_dir="$( cd "$( dirname "${BASH_SOURCE[0]}" )" && pwd )"
14 # Set base directory to the one containing scripts_dir
15 base_dir="$( cd $scripts_dir && cd .. )"
16 # Source # AutoDockTools Directory and MGLTools Python binary paths from
    constants.py
17 source $scripts_dir/constants.py
18 # Location of process_VinaResult.py
19 pvr_py="$AutoDockTools_dir/Utilities24/process_VinaResult.py"
20
21 # Retrieve the parameters for this docking
22 # source $base_dir/scripts/load_parameters.sh $dock
23 #
24 # # Retrieve ligset list
25 # ligset_list_txt=$base_dir/ligsets/$ligset/$ligset\_list.txt
26 # ligset_list=$(for l in $(cat $ligset_list_txt); do echo $l; done)
27
28 # Exit if already done
29 if [ -e $base_dir/$prot/$dock/processed_pdbqts/ ]; then
30     echo " ! Results already separated"
31     echo "      ($prot/$dock/processed_pdbqts/ exists),"
32     echo " -> exiting this step"
33     exit 1
34 fi
35
36 # Relevant directories
37 result_pdbqts_dir=$base_dir/$prot/$dock/result_pdbqts
```

```

38 processed_pdbqts_dir=$base_dir/$prot/$dock/processed_pdbqts
39
40 # Create a directory for processed files
41 mkdir $processed_pdbqts_dir
42
43 # The actual process_VinaResult step
44 receptor_pdbqt=$base_dir/$prot/$prot_file.pdbqt
45 batch_size=20
46 # No batches
47 n_models=$(echo $n_models | sed 's/[^0-9]//')
48 if [[ "n_models" -le "$batch_size" ]]; then
49     for lig in $ligset_list; do
50         result_pdbqt=$result_pdbqts_dir/$dock\_lig\_results.pdbqt
51         processed_pdbqt_stem=$processed_pdbqts_dir/$dock\_lig\_m
52         $AutoDockTools_pythonsh_binary $pvr_py -r $receptor_pdbqt \
53             -f $result_pdbqt \
54             -o $processed_pdbqt_stem \
55             1 > /dev/null
56         echo "    processed ligand $lig"
57     done
58 # Batches
59 elif [[ "n_models" -gt "$batch_size" ]]; then
60     n_batches=$((bc <<< "$n_models / $batch_size"))
61     for ((b=1;b<=$n_batches;b++)); do
62         echo "    processing batch $b"
63         for lig in $ligset_list; do
64             result_pdbqt=$result_pdbqts_dir/$dock\_.$b\_lig\_results.pdbqt
65             processed_pdbqt_stem=$processed_pdbqts_dir/$dock\_.$b\_lig\_m
66             $AutoDockTools_pythonsh_binary $pvr_py -r $receptor_pdbqt \
67                 -f $result_pdbqt \
68                 -o $processed_pdbqt_stem \
69                 1 > /dev/null
70             # Rename the processed pdbqts
71             for ((m=1;m<=$batch_size;m++)); do
72                 old_processed_pdbqt=$processed_pdbqts_dir/$dock\_.$b\_lig\_m$m.pdbqt
73                 new_m=$((bc <<< "(( $b - 1 ) * $batch_size ) + $m"))
74                 new_processed_pdbqt=$processed_pdbqts_dir/$dock\_lig\_m$new_m.pdbqt
75                 mv $old_processed_pdbqt $new_processed_pdbqt
76             done
77             echo "    processed ligand $lig"
78         done
79     done
80 else
81     echo "!!! Error in batch processing (n_models is weird)"
82 fi

```



```
83  
84 # *** check for results, prot.pdb, params  
85 # *** check if already pvr'd
```

/Users/zarek/lab/zvina/scripts/separate_vina_results.sh

2.8 parse_pdb.py

2.8.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic
sugar eclipse rust bug. Back-face culling lua javabeen indirection operator
void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Parsing data from processed pdbqt result files
4  # (c) Zarek Siegel
5  # v1 3/5/16
6
7  import re
8
9  ### A class for residues and residue atoms
10 # (input is a string of form 'RES123' or 'RES123_A1')
11
12 class Residue:
13     def __init__(self, str):
14         # String
15         self.str = str
16         # Atom & Residue String
17         if re.search(r'^[A-Z]+[0-9]+$', self.str):
18             self.atom = None
19             self.res_str = self.str
20         elif re.search(r'^[A-Z]+[0-9]+_.$', self.str):
21             self.atom = re.sub(r'^[A-Z]+[0-9]+_', '', self.str)
22             self.res_str = re.sub(r'_[A-Z0-9]+$', '', self.str)
23         else: self.atom = None
24         # Residue Index
25         self.resi = re.sub(r'^[A-Z]+|_[^_]*$', '', self.str)
26         try:
27             self.resi = int(self.resi)
28         except ValueError:
29             self.resi = None
30         # Residue Name
31         self.resn = re.sub(r'[0-9]+_[^_]*$', '', self.str)
32         # Dictionary of Props
33         self.dic = {'str' : self.str, 'res_str' : self.res_str,
34                     'resi' : self.resi, 'resn' : self.resn, 'atom' : self.atom}
35     def __str__(self):
36         return self.str
37
38 ### A class for molecules, including ones with data from process_VinaResult.py
```

```

39 # (input is a .pdb or .pdbqt file address which may or may not be pvr'd)
40
41 class Pdb:
42     def get_pdb_coords(self):
43         _coords = []
44         for line in self.pdb_lines:
45             if re.search('HETATM|ATOM', line) or (re.search('ATOM', line)):
46                 _dic = {
47                     'atomi' : int(line[6:11]),
48                     'atomn' : line[12:16].replace(" ", ""),
49                     'resn' : line[17:20].replace(" ", ""),
50                     'resi' : line[22:26].replace(" ", ""),
51                     'x' : float(line[30:38].replace(" ", "")),
52                     'y' : float(line[38:46].replace(" ", "")),
53                     'z' : float(line[46:54].replace(" ", "")),
54                     'xyz' : (float(line[30:38].replace(" ", "")),
55                             float(line[38:46].replace(" ", "")),
56                             float(line[46:54].replace(" ", "")) ),
57                     'atom_type' : line[76:78].replace(" ", ""),
58                     'charge' : line[78:80].replace(" ", "") # element
59                 }
60                 _coords.append(_dic)
61         self.coords = _coords
62
63     def get_pdbqt_coords(self):
64         _coords = []
65         for line in self.pdb_lines:
66             if re.search('HETATM|ATOM', line) or (re.search('ATOM', line)):
67                 _dic = {
68                     'atomi' : int(line[6:11]),
69                     'atomn' : line[12:16].replace(" ", ""),
70                     'resn' : line[17:20].replace(" ", ""),
71                     'resi' : line[22:26].replace(" ", ""),
72                     'x' : float(line[30:38].replace(" ", "")),
73                     'y' : float(line[38:46].replace(" ", "")),
74                     'z' : float(line[46:54].replace(" ", "")),
75                     'xyz' : (float(line[30:38].replace(" ", "")),
76                             float(line[38:46].replace(" ", "")),
77                             float(line[46:54].replace(" ", "")) ),
78                     'charge' : line[70:76].replace(" ", ""), # partial charge
79                     'atom_type' : line[77:79].replace(" ", "") # AD4 atom type
80                 }
81                 _coords.append(_dic)
82         self.coords = _coords
83

```

```

84 def mine_pvr_data(self): # mine data from pvr file
85     contacts = []
86     for line in self.pdb_lines:
87         # Binding Energy
88         if re.search('REMARK VINA RESULT: ', line):
89             self.E = re.sub( r'^REMARK VINA RESULT:[ ]+[ ]+[^ ]+[ ]+[^ ]+$' ,
90                             r'' , line.replace('\n', '')) # [23:31].replace(" ", "")
91             self.E = float(self.E)
92         # RMSD Lower Bound
93         if re.search('REMARK VINA RESULT: ', line):
94             self.rmsd_lb = re.sub( r'^REMARK VINA RESULT:[ ]+[^ ]+[ ]+[ ]+[^ ]+$'
95
96             , line.replace('\n', ''))
97             self.rmsd_lb = float(self.rmsd_lb)
98         # RMSD Upper Bound
99         if re.search('REMARK VINA RESULT: ', line):
100             self.rmsd_ub = re.sub( r'^REMARK VINA RESULT:[ ]+[^ ]+[ ]+[ ]+[^ ]+$' ,
101                                    r'' , line.replace('\n', ''))
102             self.rmsd_ub = float(self.rmsd_ub)
103         # Ligand Efficiency (whatever that means...)
104         if re.search('USER AD> ligand efficiency', line):
105             self.pvr_effic = re.sub( r'USER AD> ligand efficiency' ,
106                                     r'' , line.replace('\n', ''))
107             self.pvr_effic = float(self.pvr_effic)
108         # Model Number
109         if re.search(r'USER AD> .+ of .+ MODELS', line):
110             self.pvr_model = re.sub( r'USER AD>| of [0-9]+ MODELS' ,
111                                     r'' , line.replace('\n', ''))
112             self.pvr_model = int(self.pvr_model)
113         # Torsional Degrees of Freedom
114         if re.search('REMARK .+ active torsions:', line):
115             self.torsdof = re.sub( r'REMARK|active torsions:' ,
116                                   r'' , line.replace('\n', ''))
117             self.torsdof = int(self.torsdof)
118         # Number of Contacts
119         if re.search('USER AD> macro_close_ats:', line):
120             self.macro_close_ats = re.sub( r'USER AD> macro_close_ats:' ,
121                                             r'' , line.replace('\n', ''))
122             self.macro_close_ats = int(self.macro_close_ats)
123         # Contacts
124         if re.search(r'^USER AD> [^ ]+:[^ ]+:[^ ]+:[^ ]+$' , line):
125             contacts.append(line.replace('\n', '').replace('USER AD> ', ''))
126
127     # Contacts Processing
128     self.pvr_resis_objs = []

```

```

128 self.pvr_resis = []
129 self.pvr_resis_atoms = []
130 for c in contacts:
131     self.pvr_resis_objs.append(Residue(re.sub(r'^[:]+:[^:]+:', '',
132         c).replace(':', '_')))
133
134 for r in self.pvr_resis_objs:
135     if r.atom != None:
136         self.pvr_resis_atoms.append(r.str)
137         self.pvr_resis.append(r.res_str)
138     else:
139         self.pvr_resis_atoms.append(None)
140         self.pvr_resis.append(r.res_str)
141
142 self.pvr_resis_objs = list(set(self.pvr_resis_objs)) # remove duplicates
143 self.pvr_resis = list(set(self.pvr_resis))
144 self.pvr_resis_atoms = list(set(self.pvr_resis_atoms))
145
146 self.pvr_data = {
147     'E' : self.E,
148     'rmsd_ub' : self.rmsd_ub,
149     'rmsd_lb' : self.rmsd_lb,
150     'pvr_resis' : self.pvr_resis,
151     'pvr_resis_atoms' : self.pvr_resis_atoms,
152     'pvr_resis_objs' : self.pvr_resis_objs,
153     'torsdof' : self.torsdof,
154     'macro_close_ats' : self.macro_close_ats,
155     'pvr_model' : self.pvr_model
156 }
157
158 def get_types(self):
159     # Detect if its been through ADT process_VinaResult.py
160     self.is_pvr = False # by default
161     try:
162         for line in self.pdb_lines:
163             if re.search('REMARK VINA RESULT: ', line):
164                 self.mine_pvr_data()
165                 self.is_pvr = True
166     except AttributeError:
167         print("!!! AttributeError while trying to read PDB lines")
168
169     # Determine file type PDB/PDBQT (they are slightly different)
170     if self.pdb_file_in[-5:] == 'pdbqt':
171         self.get_pdbqt_coords()
172         self.file_type = 'pdbqt'

```

```

173     elif self.pdb_file_in[-3:] == 'pdb':
174         self.get_pdb_coords()
175         self.file_type = 'pdb'
176     else:
177         print("!!! BAD FILETYPE !!!")
178
179 def __init__(self, pdb_file_in):
180     # Specify input file
181     self.pdb_file_in = pdb_file_in
182     # Try to read it, else error
183     try:
184         pdb_file_open = open(pdb_file_in)
185         with pdb_file_open as f:
186             self.pdb_lines = f.readlines()
187     except IOError:
188         print("! ! ! IOError while trying to read PDB lines")
189         pass
190     # Determine if PDB or PDBQT, and whether it has been through
191     process_VinaResult.py
192     self.get_types() # this also mines the actual data

```

/Users/zarek/lab/zvina/scripts/parse_pdb.py

2.9 mine_vina_data.py

2.9.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Mine data from processed vina results, add to data dictionary
4  # (c) Zarek Siegel
5
6  from parse_pdb import * # Pdb
7  from create_docking_object import * # Docking
8
9  # Mine Vina result for data (actual mining is in parse_pdb.py, acting via the
   Pdb class)
10 def mine_vina_data(self):
11     print("----> Mining data from processed vina result PDBQTs...")
12     ## ** check if proper files exist
13     for lig in self.ligset_list:
14         for m in range(1, self.n_models + 1):
15             key = "{}_{}_m{}".format(self.dock, lig, m)
16             processed_pdbqt = "{d_d}/processed_pdbqts/{key}.pdbqt".format(
17                 d_d=self.dock_dir, key=key)
18             if os.path.isfile(processed_pdbqt):
19                 try: pose = Pdb(processed_pdbqt)
20                 except: print("!!! Error while trying to read PDB for {}".format(key
21 ))
22             try:
23                 pose_dic = {
24                     'E' : pose.E,
25                     'rmsd_ub' : pose.rmsd_ub,
26                     'rmsd_lb' : pose.rmsd_lb,
27                     'pvr_resis' : pose.pvr_resis,
28                     'pvr_resis_atoms' : pose.pvr_resis_atoms,
29                     'pvr_resis_objs' : pose.pvr_resis_objs,
30                     'torsdof' : pose.torsdof,
31                     'pvr_n_contacts' : pose.macro_close_ats,
32                     'pvr_model' : pose.pvr_model,
33                     'pvr_effic' : pose.pvr_effic,
34                     'coords' : pose.coords,
35                     'pvr_obj' : pose,
36                     'pdbqt_address' : processed_pdbqt,
37                     'pdb_address' : re.sub('pdbqt', 'pdb', processed_pdbqt)
```

```

37         }
38         except AttributeError:
39             print("! ! ! pose {} failed, check for the processed PDBQT".format(
key))
40             else: print("! ! ! processed_pdbqt does not exist for {}".format(key))
41
42             self.data_dic[key] = dict(self.data_dic[key].items() + pose_dic.items())
43
44         self.vina_data_mined = True
45         print("    > Data dictionary filled with data from process_VinaResult.py\n")
46
47         Docking.mine_vina_data = mine_vina_data

```

/Users/zarek/lab/zvina/scripts/mine_vina_data.py

2.10 aiad_icpd.py

2.10.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic
sugar eclipse rust bug. Back-face culling lua javabean indirection operator
void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Calculating average inter-atomic distance
4  # (c) Zarek Siegel
5  # v1 3/6/16
6
7  from parse_pdb import *
8  from math import sqrt
9  # from numpy import mean
10
11  def mean(list):
12      list_mean = float(sum(list)) / len(list)
13      return list_mean
14
15  class Molecule():
16      def list_coords(self):
17          self.coord_triples = []
18          for atom in self.pdb.coords:
19              self.coord_triples.append(atom['xyz'])
20
21      def get_centerpoint(self):
22          x_coords = []
23          y_coords = []
24          z_coords = []
25          for triple in self.coord_triples:
26              x_coords.append(triple[0])
27              y_coords.append(triple[1])
28              z_coords.append(triple[2])
29          self.centerpoint = (mean(x_coords), mean(y_coords), mean(z_coords))
30
31      def __init__(self, pdb):
32          self.pdb = pdb
33          self.list_coords()
34          self.get_centerpoint()
35
36      def threeD_distance(triple1, triple2):
37          x1 = triple1[0]
38          y1 = triple1[1]
```

```

39     z1 = triple1[2]
40     x2 = triple2[0]
41     y2 = triple2[1]
42     z2 = triple2[2]
43     distance = sqrt( ((x2 - x1)**2) + ((y2 - y1)**2) + ((z2 - z1)**2) )
44     return distance
45
46 def caclulate_aiad(pdb1, pdb2):
47     molc1 = Molecule(pdb1)
48     molc2 = Molecule(pdb2)
49     dist_list = []
50     for triple1 in molc1.coord_triples:
51         dists_from_t1 = []
52         for triple2 in molc2.coord_triples:
53             dist = threeD_distance(triple1, triple2)
54             dists_from_t1.append(dist)
55         dist_list.append(min(dists_from_t1))
56     return mean(dist_list)
57
58 def calculate_icpd(pdb1, pdb2):
59     molc1 = Molecule(pdb1)
60     molc2 = Molecule(pdb2)
61     return threeD_distance(molc1.centerpoint, molc2.centerpoint)
62
63 def main():
64     # m1_p = Pdb("/Users/zarek/lab/Docking/p300/p27/res_pdbqts_cleaned/p27_s3_m4
65     #         .pdbqt")
66     # m2_p = Pdb("/Users/zarek/lab/Docking/p300/p27/res_pdbqts_cleaned/
67     #         p27_s2_m142.pdbqt")
68     # m1 = Molecule(m1_p)
69     # m2 = Molecule(m2_p)
70     # caclulate_aiad(m1_p, m2_p)
71     pass
72
73 if __name__ == "__main__": main()

```

/Users/zarek/lab/zvina/scripts/aiad_icpd.py

2.11 binding_site_analysis.py

2.11.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Mine data from processed vina results, add to data dictionary
4  # (c) Zarek Siegel
5
6  from __future__ import print_function
7  import re
8  from operator import itemgetter
9  from create_docking_object import * # Docking
10 from parse_pdb import * # Pdb
11 from aiad_icpd import *
12
13
14 # Create a list of the binding sites previously prepared for the protein
15 def get_binding_sites_list(self):
16     if not self.vina_data_mined: self.mine_vina_data()
17
18     binding_sites_dir = "{b_d}/binding_sites/{p_f}".format(
19         b_d=base_dir, p_f=self.prot_file)
20     self.binding_sites_list = []
21     self.binding_sites_objs = {}
22     for root, dirs, files in os.walk(binding_sites_dir):
23         for file in files:
24             self.binding_sites_list.append(re.sub('.pdb', '', file))
25             try: self.binding_sites_objs[re.sub('.pdb', '', file)] = Pdb("{}/{f}".
26                 format(root, file))
27             except: print("! ! ! Error while trying to read PDB for {}".format(file)
28                 )
29
30     self.bs_resis_lists = {}
31     self.bs_resis_atoms_lists = {}
32
33     for bs, bso in self.binding_sites_objs.items():
34         bs_resis = []
35         if self.evaluate_resis_atoms:
36             bs_resis_atoms = []
37             for atom in bso.coords:
38                 bs_resis.append("{}{}".format(atom['resn'], atom['resi']))
```

```

37         if self.evaluate_resis_atoms:
38             bs_resis_atoms.append("{}{}_{}".format(atom['resn'], atom['resi'],
39             atom['atomn']))
40         self.bs_resis_lists[bs] = list(set(bs_resis))
41         if self.evaluate_resis_atoms:
42             self.bs_resis_atoms_lists[bs] = list(set(bs_resis_atoms))
43
44     self.binding_sites_list_gotten = True
45     print("    > Retrieved binding sites")
46
47     Docking.get_binding_sites_list = get_binding_sites_list
48
49     # Score the binding sites in terms of the residues they contact
50     # relative to those contained in the reference PDB
51     def score_binding_sites(self):
52         if not self.vina_data_mined: self.mine_vina_data()
53         print("----> Scoring poses against binding sites by ratio of residues
54         contacted...")
55         if not self.binding_sites_list_gotten: self.get_binding_sites_list()
56
57         print("    > Scoring binding sites for pose ", end="")
58         char_count = 31
59         for pose in self.keys:
60             if char_count <= 76:
61                 print(pose, end=" ")
62                 char_count = char_count + len(pose) + 1
63             else:
64                 print("\n\t{}".format(pose), end = " ")
65                 char_count = len(pose) + 1
66         for bs in self.binding_sites_list:
67             resis_union = ( set(self.bs_resis_lists[bs]) & set(self.data_dic[pose]['
68             pvr_resis']) )
69             self.data_dic[pose]["{}_fraction".format(bs)] = float(len(resis_union)
70             / float(len(self.bs_resis_lists[bs]))
71             if self.evaluate_resis_atoms:
72                 resis_atoms_union = ( set(self.bs_resis_atoms_lists[bs]) & set(self.
73                 data_dic[pose]['pvr_resis_atoms']) )
74                 self.data_dic[pose]["{}_atoms_fraction".format(bs)] = float(len(
75                 resis_atoms_union)) / float(len(self.bs_resis_atoms_lists[bs]))
76
77     self.binding_sites_scored = True
78     print("\n    > Done scoring binding sites\n")
79
80     Docking.score_binding_sites = score_binding_sites

```

```

76 # Score binding sites by average inter-atomic distance
77 # and inter-centerpoint difference
78 # Acting via aiad_icpd.py
79 def aiad_icpd_binding_sites(self):
80     if not self.binding_sites_list_gotten: self.get_binding_sites_list()
81
82     print("----> Scoring poses against binding sites by AIAD and ICPD...")
83     print("    > Scoring AIAD and ICPD for pose ", end="")
84     char_count = 31
85     for pose in self.keys:
86         if char_count <= 76:
87             print(pose, end=" ")
88             char_count = char_count + len(pose) + 1
89         else:
90             print("\n\t{}".format(pose), end = " ")
91             char_count = len(pose) + 1
92         for bs, bso in self.binding_sites_objs.items():
93             aiad = caclulate_aiad(self.data_dic[pose]['pvr_obj'], bso)
94             self.data_dic[pose]["{}_aiad".format(bs)] = aiad
95             icpd = calculate_icpd(self.data_dic[pose]['pvr_obj'], bso)
96             self.data_dic[pose]["{}_icpd".format(bs)] = icpd
97
98     self.aiad_icpd_calcd = True
99     print("\n    > Done calculating AIAD and ICPD for binding sites\n")
100
101 Docking.aiad_icpd_binding_sites = aiad_icpd_binding_sites
102
103 # Add attributes for whether the ligand contacts each residue of the protein
104 def assess_all_resis(self):
105     if not self.vina_data_mined: self.mine_vina_data()
106
107     print("----> Evaluating residues contacted for each pose")
108     self.prot_pdbqt = "{p_d}/{p_f}.pdbqt".format(p_d=self.prot_dir, p_f=self.
        prot_file)
109     try: self.prot_obj = Pdb(self.prot_pdbqt)
110     except: print("!! Error while trying to read PDB for {}".format(file))
111     self.prot_resis_list = []
112     if self.evaluate_resis_atoms:
113         self.prot_resis_atoms_list = []
114     for atom in self.prot_obj.coords:
115         self.prot_resis_list.append((atom['resn'], atom['resi']))
116         if self.evaluate_resis_atoms:
117             self.prot_resis_atoms_list.append((atom['resn'], atom['resi'], atom['
                atomn']))
118     # self.prot_resis_list.append("{}{}".format(atom['resn'], atom['resi']))

```

```

119     # self.prot_resis_atoms_list.append("{}{}_{}".format(atom['resn'], atom['
120     resi'], atom['atomn']))
121 self.prot_resis_list = list(set(self.prot_resis_list))
122 self.prot_resis_list = sorted(self.prot_resis_list, key=itemgetter(1))
123 self.prot_resis_list = ["{}_{}".format(r[0], r[1]) for r in self.
    prot_resis_list]
124 if self.evaluate_resis_atoms:
125     self.prot_resis_atoms_list = list(set(self.prot_resis_atoms_list))
126     self.prot_resis_atoms_list = sorted(self.prot_resis_atoms_list, key=
    itemgetter(1,2))
127     self.prot_resis_atoms_list = ["{}_{}_{}".format(r[0], r[1], r[2]) for r in
    self.prot_resis_atoms_list]
128     # self.prot_resis_atoms_list = list(set(self.prot_resis_atoms_list))
129
130 for pose in self.data_dic:
131     for res in self.prot_resis_list:
132         if res in self.data_dic[pose]['pvr_resis']:
133             self.data_dic[pose][res] = 1
134         else:
135             self.data_dic[pose][res] = 0
136     if self.evaluate_resis_atoms:
137         for atom in self.prot_resis_atoms_list:
138             if atom in self.data_dic[pose]['pvr_resis_atoms']:
139                 self.data_dic[pose][atom] = 1
140             else:
141                 self.data_dic[pose][atom] = 0
142
143 self.all_resis_assessed = True
144 print("    > Residues contacted added to data dictionary\n")
145 Docking.assess_all_resis = assess_all_resis

```

/Users/zarek/lab/zvina/scripts/binding_site_analysis.py

2.12 energies_properties.py

2.12.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ###
4  # (c) Zarek Siegel
5  # v1 3/16/16
6
7  import subprocess, re, sys
8  from constants import *
9
10 #####
11
12 def get_energies(molc_in):
13     obenergy_binary = "{}{}obenergy".format(openbabel_binaries_dir)
14
15     obenergy_out = subprocess.Popen([obenergy_binary, molc_in],
16                                     stdout=subprocess.PIPE, stderr=subprocess.PIPE)
17     obenergy_out = obenergy_out.communicate()[0]
18     obenergy_out = re.split('\n', obenergy_out)
19
20     #     TOTAL BOND STRETCHING ENERGY =   28.617 kJ/mol
21     #     TOTAL ANGLE BENDING ENERGY =   18.099 kJ/mol
22     #     TOTAL TORSIONAL ENERGY =    62.835 kJ/mol
23     #     TOTAL VAN DER WAALS ENERGY =  104.782 kJ/mol
24     #
25     # TOTAL ENERGY = 214.33290 kJ/mol
26
27     obenergy_dic = {}
28
29     def extract_energy_value(search_str):
30         for row in obenergy_out:
31             if re.search(search_str, row):
32                 #         print(row)
33                 value = (re.sub(r'^s*([\w ]+)=s*(-?\d+\.\d+)\s+(.)$', r'\2', row))
34                 units = (re.sub(r'^s*([\w ]+)=s*(-?\d+\.\d+)\s+(.)$', r'\3', row))
35                 return value#, units
36
37     search_str_list = [
38         "TOTAL BOND STRETCHING ENERGY",
```

```

39     "TOTAL ANGLE BENDING ENERGY",
40     "TOTAL TORSIONAL ENERGY",
41     "TOTAL ENERGY"
42 ]
43
44 for search_str in search_str_list:
45     parameter = search_str.lower()
46     parameter = re.sub(' ', '_', parameter)
47     obenergy_dic[parameter] = extract_energy_value(search_str)
48
49 return obenergy_dic
50 #####
51
52 #####
53
54 def get_properties(molc_in):
55     obprop_binary = "{}_obprop".format(openbabel_binaries_dir)
56
57     obprop_out = subprocess.Popen([obprop_binary, molc_in],
58                                   stdout=subprocess.PIPE, stderr=subprocess.PIPE)
59     obprop_out = obprop_out.communicate()[0]
60     obprop_out = re.split('\n', obprop_out)
61
62     # name          /Users/zarek/GitHub/TaylorLab/zvina/ligsets/hls1/pdbs/ab.
63     #               pdb 1
64     # formula       C6H12O6
65     # mol_weight    180.156
66     # exact_mass    180.063
67     # canonical_SMILES OC[C@H]1O[C@@H](O)[C@@H]([C@H]([C@H]1O)O)O
68     # InChI         InChI=1S/C6H12O6/c7-1-2-3(8)4(9)5(10)6(11)12-2/h2-11H,1H2
69     #               /t2-,3+,4+,5-,6-/m1/s1
70     #
71     # num_atoms     24
72     # num_bonds     24
73     # num_residues  1
74     # sequence      LIG
75     # num_rings     1
76     # logP          -3.2214
77     # PSA           110.38
78     # MR            35.736
79
80     obprop_dic = {}
81
82     def extract_property_value(search_str):

```



```

82     for row in obprop_out:
83         if re.search(search_str, row):
84             # print(row)
85             value = (re.sub(r'^(\w+)\s+(\S+)\s?\S?$', r'\2', row))
86             return value
87
88     search_str_list = [
89         "name", "formula", "mol_weight",
90         "exact_mass", "canonical_SMILES", "num_atoms",
91         "num_bonds", "num_rings", "logP", "PSA", "MR"
92     ]
93
94     for search_str in search_str_list:
95         parameter = search_str
96         obprop_dic[parameter] = extract_property_value(search_str)
97
98     return obprop_dic
99     #####
100
101 def get_combined_dic(molc_in):
102     combined_dic = dict(get_energies(molc_in).items() + \
103         get_properties(molc_in).items())
104     return combined_dic
105
106 def print_energies_properties(molc_in):
107     print("\n{0:.<36}{v}".format("PROPERTY", v="VALUE"))
108     for prop, value in get_combined_dic(molc_in).items():
109         print("{0:.<36}{v}".format(prop, v=value))
110
111 def main():
112     print_energies_properties(sys.argv[1])
113
114 if __name__ == "__main__": main()

```

/Users/zarek/lab/zvina/scripts/energies_properties.py

2.13 get_pose_energies_properties.py

2.13.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic
sugar eclipse rust bug. Back-face culling lua javabean indirection operator
void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ###
4  # (c) Zarek Siegel
5
6  from __future__ import print_function
7  from energies_properties import *
8  from create_docking_object import * # Docking
9
10 def get_pose_energies_properties(self):
11     if not self.vina_data_mined: self.mine_vina_data()
12
13     print("---> Assessing molecular properties and energies for pose...")
14     # Properties are gathered once per ligand
15     # But energies are gathered for each pose
16     print("    > Evaluating pose ", end="")
17     for lig in self.ligset_list:
18         first_pose = "{}_{}_m{}".format(self.dock, lig, "1")
19         first_pdbqt = self.data_dic[first_pose]['pdbqt_address']
20
21         char_count = 24
22         pose_properties_dic = get_properties(first_pdbqt)
23         for m in range(1, self.n_models + 1):
24             pose = "{}_{}_m{}".format(self.dock, lig, m)
25             if char_count <= 76:
26                 print(pose, end=" ")
27                 char_count = char_count + len(pose) + 1
28             else:
29                 print("\n\t{}".format(pose), end = " ")
30                 char_count = len(pose) + 1
31             pdbqt = self.data_dic[pose]['pdbqt_address']
32             pose_energies_dic = get_energies(pdbqt)
33             combined_dic = dict(pose_properties_dic.items() +
34                                 pose_energies_dic.items())
35             # print(combined_dic)
36             self.data_dic[pose] = dict(self.data_dic[pose].items() + combined_dic.
37                                     items())
```

```
38     self.energies_props_gotten = True
39
40     print("\n    > Done\n")
41
42 Docking.get_pose_energies_properties = get_pose_energies_properties
```

/Users/zarek/lab/zvina/scripts/get_pose_energies_properties.py

2.14 write_alldata.py

2.14.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Output all the data mined and analyzed into a CSV file
4  # (c) Zarek Siegel
5
6  import os, sys
7  import cPickle as pickle
8  from create_docking_object import * # Docking
9
10 def get_fieldnames(self):
11     # Determine which headers to write to the CSV
12
13     # Choose from these groups
14     self.alldata_fieldnames = ['key', 'lig', 'model'] # basics
15     # energies_properties_fieldnames = self.energies_dic.keys() + self.
16     # properties_dic.keys()
17
18     if self.vina_data_mined:
19         pvr_fieldnames = ['E', 'rmsd_lb', 'rmsd_ub', 'pvr_effic',
20             'pvr_n_contacts', 'torsdof', 'pdbqt_address', 'pdb_address'] # (
21             processed vina results)
22         self.alldata_fieldnames = self.alldata_fieldnames + pvr_fieldnames
23
24     if self.energies_props_gotten:
25         energies_properties_fieldnames = [
26             'total_bond_stretching_energy', 'total_angle_bending_energy',
27             'total_torsional_energy', 'total_energy',
28             'name', 'formula', 'mol_weight', 'exact_mass', 'canonical_SMILES',
29             'num_atoms', 'num_bonds', 'num_rings', 'logP', 'PSA', 'MR'
30         ]
31         self.alldata_fieldnames = self.alldata_fieldnames +
32             energies_properties_fieldnames
33
34     if self.binding_sites_scored:
35         binding_site_fieldnames = []
36         for bs in self.binding_sites_list:
37             binding_site_fieldnames.append("{}_fraction".format(bs))
38             binding_site_fieldnames.append("{}_atoms_fraction".format(bs))
```

```

36     self.alldata_fieldnames = self.alldata_fieldnames +
    binding_site_fieldnames
37
38     if self.aiad_icpd_calcd:
39         aiad_icpd_fieldnames = []
40         for bs in self.binding_sites_list:
41             aiad_icpd_fieldnames.append("{}_aiad".format(bs))
42             aiad_icpd_fieldnames.append("{}_icpd".format(bs))
43         self.alldata_fieldnames = self.alldata_fieldnames + aiad_icpd_fieldnames
44
45     if self.all_resis_assessed:
46         self.alldata_fieldnames = self.alldata_fieldnames + self.prot_resis_list
47     if self.evaluate_resis_atoms:
48         self.alldata_fieldnames = self.alldata_fieldnames + self.
        prot_resis_atoms_list
49
50 Docking.get_fieldnames = get_fieldnames
51
52 def write_alldata_csv(self):
53     self.alldata_csv = "{d_d}/{d}_alldata.csv".format(d_d=self.dock_dir, d=self.
        dock)
54
55     # If the CSV already exists, ask to overwrite
56     if os.path.isfile(self.alldata_csv):
57         print(">>> Docking CSV file already exists at")
58         print("\t{}".format(self.alldata_csv))
59         overwrite = raw_input("\n\t>>> Enter 'y' or enter to overwrite, 'n' to
            exit: ")
60         if overwrite == "y" or \
61             overwrite == "yes" or \
62             overwrite == "Y" or \
63             overwrite == "Yes" or \
64             overwrite == "":
65             subprocess.call(["rm", "-f", self.alldata_csv])
66             print("")
67         else: sys.exit("\n\t> OK, exiting this script\n")
68
69     self.get_fieldnames()
70
71     print("----> Writing alldata.csv...")
72     with open(self.alldata_csv, 'w') as csvfile:
73         writer = csv.DictWriter(csvfile, fieldnames=self.alldata_fieldnames)
74         writer.writeheader()
75         for key in self.keys:
76             row = {}

```

```

77     for f in self.alldata_fieldnames:
78         try: row[f] = self.data_dic[key][f]
79         except KeyError:
80             print("!!! KeyError while trying to write {}".format(f))
81             row[f] = "!Err!"
82     writer.writerow(row)
83
84     self.is_csv_written = True
85     print("> Completed alldata.csv is located at:\n\t{}\n".format(self.
      alldata_csv))
86
87     Docking.write_alldata_csv = write_alldata_csv
88
89     # Save the data dictionary as a pickled file (i.e. in native python format)
90     def write_docking_pickled(self):
91         self.docking_obj_pickled = "{d_d}/{d}.p".format(d_d=self.dock_dir, d=self.
      dock)
92
93         # If the pickled dictionary already exists, ask to overwrite
94         if os.path.isfile(self.docking_obj_pickled):
95             print(">>> Pickled docking object file already exists at")
96             print("\t{}".format(self.docking_obj_pickled))
97             overwrite = raw_input("\n\t>>> Enter 'y' or enter to overwrite, 'n' to
      exit: ")
98             if overwrite == "y" or \
99                 overwrite == "yes" or \
100                 overwrite == "Y" or \
101                 overwrite == "Yes" or \
102                 overwrite == "":
103                 subprocess.call(["rm", "-f", self.docking_obj_pickled])
104                 print("")
105             else: sys.exit("\n\t> OK, exiting this script\n")
106
107         print("----> Pickling docking object...")
108         pickle.dump(self, open(self.docking_obj_pickled, 'wb'))
109
110         self.is_pickled = True
111         print("> Pickled docking object located at:\n\t{}\n".format(self.
      docking_obj_pickled))
112
113     Docking.write_docking_pickled = write_docking_pickled

```

/Users/zarek/lab/zvina/scripts/write_alldata.py

2.15 correlations.py

2.15.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### Output all the data mined and analyzed into a CSV file
4  # (c) Zarek Siegel
5
6  from scipy.stats.stats import pearsonr
7  from operator import itemgetter
8  from create_docking_object import * # Docking
9  from write_alldata import get_fieldnames
10
11 def correlations(self):
12
13     self.get_fieldnames()
14
15     not_quant = ['key', 'lig', 'model', 'pdbqt_address', 'pdb_address',
16                 'name', 'formula', 'canonical_SMILES']
17     correl_vars = [var for var in self.alldata_fieldnames if var not in
18                   not_quant]
19
20     correl_var_lists = {}
21     for var in correl_vars:
22         correl_var_lists[var] = []
23         for pose in self.keys:
24             try: correl_var_lists[var].append(self.data_dic[pose][var])
25             except KeyError: correl_var_lists[var].append(None)
26
27     correls = []
28     for var1 in correl_vars:
29         print(var1)
30         for var2 in correl_vars:
31             pearson = pearsonr(correl_var_lists[var1], correl_var_lists[var2])
32             correls.append(
33                 {
34                     'var1' : var1,
35                     'var2' : var2,
36                     'correl' : pearson[0],
37                     'cor_mag' : abs(pearson[0]),
38                     'p' : pearson[1]
```

```

38     }
39 )
40
41
42 sig_correls = [c for c in correls if c['p'] < 0.05]
43
44 print(len(correls))
45 print(len(sig_correls))
46 for c1 in sig_correls:
47     # print(c1)
48     if (c1['var1'] == c1['var2']):
49         sig_correls.remove(c1)
50     for c2 in sig_correls:
51         # print(c2)
52         if (c2['var1'] == c2['var2']):
53             sig_correls.remove(c2)
54         elif (c1['var1'] == c2['var2']) and (c1['var2'] == c2['var1']):
55             sig_correls.remove(c2)
56
57     else:
58         continue
59 print(len(correls))
60 print(len(sig_correls))
61 sig_correls = sorted(sig_correls, key=itemgetter('cor_mag', 'p', 'var1'))
62
63 # Don't print correlations between two residues
64 for c in sig_correls:
65     if not (c['var1'] in self.prot_resis_list) and (c['var2'] in self.
66         prot_resis_list):
67         print("{:<30}{:<30}{:<30}{:<30}".format(c['var1'], c['var2'], c['correl'
68 ], c['p']))
69
70
71
72 # print("{} = {}".format("parameters_loaded", self.parameters_loaded))
73 # print("{} = {}".format("ligset_list_gotten", self.ligset_list_gotten))
74 # print("{} = {}".format("parameters_exported_to_environment", self.
75     parameters_exported_to_environment))
76 # print("{} = {}".format("is_data_dic_created", self.is_data_dic_created))
77 #
78 # print("{} = {}".format("vina_data_mined", self.vina_data_mined))
79 # print("{} = {}".format("binding_sites_list_gotten", self.
80     binding_sites_list_gotten))

```



```

79 # print("{} = {}".format("binding_sites_scored", self.binding_sites_scored))
80 # print("{} = {}".format("aiad_icpd_calcd", self.aiad_icpd_calcd))
81 # print("{} = {}".format("all_resis_assessed", self.all_resis_assessed))
82 #
83 # print("{} = {}".format("is_csv_written", self.is_csv_written))
84 # print("{} = {}".format("energies_props_gotten", self.energies_props_gotten
85 # ))
86 # print("{} = {}".format("are_poses_clustered", self.are_poses_clustered))
87 # print("{} = {}".format("is_pickled", self.is_pickled))
88
89
90 Docking.correlations = correlations
91
92
93 # pvr_fieldnames = ['E', 'rmsd_lb', 'rmsd_ub', 'pvr_effic',
94 #                   'pvr_n_contacts', 'torsdof', 'pdbqt_address', 'pdb_address'] # (
95 #                   processed vina results)
96 #
97 # self.alldata_fieldnames = self.alldata_fieldnames + pvr_fieldnames
98 #
99 # if self.energies_props_gotten:
100 #     energies_properties_fieldnames = [
101 #         'total_bond_stretching_energy', 'total_angle_bending_energy',
102 #         'total_torsional_energy', 'total_energy',
103 #         'name', 'formula', 'mol_weight', 'exact_mass', 'canonical_SMILES',
104 #         'num_atoms', 'num_bonds', 'num_rings', 'logP', 'PSA', 'MR'

```

/Users/zarek/lab/zvina/scripts/correlations.py

2.16 postdocking_graphs.R

2.16.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  ### Docking data graphs (hepi and p300 - pls1a and hls1)
2  #   2/14/16 Zarek Siegel
3  #   2/21/16 up to full ggplot2 replacement
4  #   2/23/16 more density plots, p300 and hepi compatible
5  # v1.0 3/16/16 lots stuff
6
7  rm(list=ls(all=TRUE)) # clear out old variables
8  library(foreign) # for reading csv
9  library(ggplot2)
10 library(reshape2)
11
12 ### Command line arguments
13
14 args <- commandArgs()
15 script_path <- args[4]
16 script_path <- sub("--file=", "", script_path)
17 base_dir <- sub("/scripts/postdocking_graphs.R", "", script_path)
18 # base_dir <- "/Users/zarek/GitHub/TaylorLab/zvina"
19 dock <- args[6]
20
21
22
23 ### Basic parameters
24 # dock <- "p7b" # docking id
25 # if(substr(dock,0,1) == "p") {prot <- "p300"} # p300 dockings are labeled p##
26 # if(substr(dock,0,1) == "h") {prot <- "hepi"} # hepi dockings are labeled h##
27 # assumptions
28 #   -dockings have a docking id of the form d##,
29 #     where d represents the protein and ## is a number
30 #   -inallo the home docking directory,
31 #     there is a directory with the protein's name (e.g. .../p300/).
32 #     (this is what I mean by 'dock directory')
33 #   -inallo the protein directory directory,
34 #     there is a directory with the docking id (e.g. .../p300/p27/)
35 #   -inallo the docking directory,
36 #     there is a CSV file called d##_alldata.csv (** see zarek or a sample
37 #     for details)
```

```

38 # docks.xlsx <- "/Users/zarek/lab/Docking/docks.xlsx"
39 # dock.params <- read.xlsx2(docks.xlsx, sheetName = prot)
40 #
41 # ligset <- as.character(dock.params$ligSET[dock.params$DOCK == dock])
42 #
43 # all.ligsets <- read.xlsx2(docks.xlsx, sheetName = "ligsets")
44 # ligset_list <- as.character(all.ligsets$lig_list[all.ligsets$name == ligset
45   ])
46 # ligset_list <- unlist(strsplit(ligset_list, "[ ]"))
47 #
48 # ### System directories
49 # docking.base.dir <- "/Users/zarek/lab/Docking/"
50 # dock.dir <- paste(docking.base.dir, prot, "/", dock, "/", sep = "")
51 # graphs_dir <- paste(dock.dir, "graphs/", sep="")
52 # if(dir.exists(graphs_dir)) {
53 #   print("graphs directory was not created because it already exists")
54 # } else {dir.create(graphs_dir)}
55 #
56 # ### Read in alldata.csv
57 # alldata.csv <- paste(dock.dir, dock, "_alldata.csv", sep="")
58 # data <- read.csv(alldata.csv)
59 #
60 # ### Look up ligset
61 # # ** add this**, this (below) is temporary
62 # # ligset_list <- c("adph", "fdla", "ab", "gb", "aam",
63 # #                 "abm", "gam", "gbm", "ab3", "gb3",
64 # #                 "ab6", "gb6", "ab7", "gb7", "aa8",
65 # #                 "ab8", "ga8", "gb8", "gb8y", "aa10",
66 # #                 "ab10", "ga10", "gb10")
67 #
68 # ### Look up ligset
69 # ***KLUDGE* ** add this**, this (below) is temporary
70 # if(prot == "hepi") {
71 #   binding_sites <- c("adph", "fdla", "allo")
72 #   ##### Order levels
73 #   ncs.sugargrouped <- c("adph", "fdla",
74 #                         "ab", "aam", "abm", "ab3", "ab6",
75 #                         "ab7", "aa8", "ab8", "aa10", "ab10",
76 #                         "gb", "gam", "gbm", "gb3", "gb6",
77 #                         "gb7", "ga8", "gb8", "gb8y",
78 #                         "ga10", "gb10")
79 #   ligset_list <- ncs.sugargrouped
80 # }
81 # if(prot == "p300") {

```

```

82 # binding_sites <- c("lys", "coa", "side", "allo1", "allo2") # "coa_adpp", "
    coa_pant",
83 # ligset_list <- c("Garcinol", "CTB", "EGCG", "CTPB", "C646")
84 # }
85 # ***##
86 #
87 # ### Recode variables with more convenient names
88 # if(is.element(dock, c("p28", "p29", "p30"))) {
89 #   ligset.coded <- c("ng", "s2", "ne", "s1", "s3")
90 #   ligset.names <- factor(c("Garcinol", "CTB", "EGCG", "CTPB", "C646"))
91 #   data$lig <- ligset.names[match(data$lig, ligset.coded)]
92 #   data$lig <- factor(data$lig, levels = ligset.names)
93 # }
94 # if(is.element(dock, c("p27", "p31"))) {
95 #   ligset.names <- factor(c("Garcinol", "CTB", "EGCG", "CTPB", "C646"))
96 #   data$lig <- factor(data$lig, levels = ligset.names)
97 # }
98
99
100 dockings_csv <- paste0(base_dir, "/", "Dockings.csv")
101 dockings_df <- read.csv(dockings_csv)
102
103 prot <- as.character(dockings_df$Protein[dockings_df$Docking.ID == dock])
104 date <- as.character(dockings_df$Date[dockings_df$Docking.ID == dock])
105 prot_file <- as.character(dockings_df$Protein.File[dockings_df$Docking.ID ==
    dock])
106 ligset <- as.character(dockings_df$Ligset[dockings_df$Docking.ID == dock])
107 box <- as.character(dockings_df$Gridbox[dockings_df$Docking.ID == dock])
108 exhaust <- as.character(dockings_df$Exhaustiveness[dockings_df$Docking.ID ==
    dock])
109 n_models <- as.integer(dockings_df$Number.of.Models[dockings_df$Docking.ID ==
    dock])
110 n_cpus <- as.integer(dockings_df$Number.of.CPUs[dockings_df$Docking.ID == dock
    ])
111 notes <- as.character(dockings_df$Notes[dockings_df$Docking.ID == dock])
112
113 gridboxes_csv <- paste0(base_dir, "/", "Gridboxes.csv")
114 gridboxes_df <- read.csv(gridboxes_csv)
115
116 box_center_x <- as.numeric(gridboxes_df$Center.in.x.dimension[gridboxes_df$
    Gridbox.Name == box])
117 box_center_y <- as.numeric(gridboxes_df$Center.in.y.dimension[gridboxes_df$
    Gridbox.Name == box])
118 box_center_z <- as.numeric(gridboxes_df$Center.in.z.dimension[gridboxes_df$
    Gridbox.Name == box])

```

```

119 box_size_x <- as.numeric(gridboxes_df$Size.in.x.dimension[gridboxes_df$Gridbox
    .Name == box])
120 box_size_y <- as.numeric(gridboxes_df$Size.in.y.dimension[gridboxes_df$Gridbox
    .Name == box])
121 box_size_z <- as.numeric(gridboxes_df$Size.in.z.dimension[gridboxes_df$Gridbox
    .Name == box])
122 box_notes <- as.character(gridboxes_df$Notes[gridboxes_df$Gridbox.Name == box
    ])
123
124 ligset_list_txt <- paste0(base_dir, "/ligsets/", ligset, "/", ligset, "_list.
    txt")
125 ligset_list <- read.delim(ligset_list_txt, header = F, sep = "\n")
126 ligset_list <- as.character(unlist(ligset_list))
127 ligset_list <- factor(ligset_list, levels = ligset_list)
128
129 ### Read in alldata.csv
130 dock_dir <- paste0(base_dir, "/", prot, "/", dock)
131
132 dock_alldata_csv <- paste0(dock_dir, "/", dock, "_alldata.csv")
133 data <- read.csv(dock_alldata_csv)
134
135 ### Recode variables with more convenient names
136 if(ligset == "plsla") {
137   # ligset.coded <- c("ng", "s2", "ne", "s1", "s3")
138   # ligset.names <- factor(c("Garcinol", "CTB", "EGCG", "CTPB", "C646"))
139   plsla_coded <- c("ng", "s2", "ne", "s1", "s3")
140   plsla_renamed <- factor(c("Garcinol", "CTB", "EGCG", "CTPB", "C646"))
141   data$lig <- plsla_renamed[match(data$lig, plsla_coded)]
142   data$lig <- factor(data$lig, levels = plsla_renamed)
143   ligset_list <- plsla_renamed
144 }
145
146 binding_sites_dir <- paste0(base_dir, "/binding_sites/", prot_file)
147 binding_sites <- list.files(path = binding_sites_dir)
148 binding_sites <- sub(".pdb", "", binding_sites)
149
150 ### Create binds_in_SITE rows (T/F depending on score fraction)
151 binding.threshold <- 0.10 # threshold for binding fraction
152 for(bs in binding_sites) {
153   # data[, paste("binds_in_", bs, sep = "")] <- rep(NA, nrow(data))
154   # data[, paste("binds_in_", bs, sep = "")][data[,paste("resis_score_fraction_
    ", bs, sep = "")] >= binding.threshold] <- T
155   # data[, paste("binds_in_", bs, sep = "")][data[,paste("resis_score_fraction_
    ", bs, sep = "")] < binding.threshold] <- F
156   bindsin_bs <- paste0("binds_in_", bs)

```

```

157 bs_fraction <- paste0(bs, "_fraction")
158 data[, bindsin_bs] <- rep(NA, nrow(data))
159 data[, bindsin_bs][data[,bs_fraction] >= binding.threshold] <- T
160 data[, bindsin_bs][data[,bs_fraction] < binding.threshold] <- F
161 }
162
163
164 ### Create analysis data frame
165 analysis <- data.frame(row.names = ligset_list)
166
167 # Build column headers for averages, minimums, standard deviations,
168 # distribution fractions
169 analysis.columns <- c("AvgE", "MinE", "StdevE", paste("Num_", binding_sites,
170 # Build column headers for averages, minimums, standard deviations,
171 # distribution fractions
172 # Build column headers for averages, minimums, standard deviations,
173 # distribution fractions
174 # Build column headers for averages, minimums, standard deviations,
175 # distribution fractions
176 # Build column headers for averages, minimums, standard deviations,
177 # distribution fractions
178 # Build column headers for averages, minimums, standard deviations,
179 # distribution fractions
180 # Build column headers for averages, minimums, standard deviations,
181 # distribution fractions
182 # Build column headers for averages, minimums, standard deviations,
183 # distribution fractions
184 # Build column headers for averages, minimums, standard deviations,
185 # distribution fractions
186 # Build column headers for averages, minimums, standard deviations,
187 # distribution fractions
188 # Build column headers for averages, minimums, standard deviations,
189 # distribution fractions
190 # Build column headers for averages, minimums, standard deviations,
191 # distribution fractions

```

```

192     analysis[l, paste0("AvgE_", bs)] <- NA
193     analysis[l, paste0("MinE_", bs)] <- NA
194   }
195 }
196 for(bs in binding_sites) { # fraction of bindings in site from count
197   analysis[l, paste("DistribFrac_", bs, sep = "")] <-
198     analysis[l, paste("Num_", bs, sep = "")] / DistribFrac_Denom
199 }
200 }
201
202 analysis_csv = paste0(dock_dir, "/", dock, "_summary.csv")
203
204 write.csv(analysis, file = analysis_csv)
205 print(paste0("Created summary CSV"))
206
207 #
208 #####
209 #
210 #####
211 #
212 #####
213
214 ### Graphs!!!!!!
215 # setwd(graphs_dir) # needed for pdf names below
216 # bs.colors <- brewer.pal(length(binding_sites), "Set1")
217 # lig.colors <- brewer.pal(length(ligset_list), "Set2")
218 # lig.palette <- "Greys"
219
220 graphs_dir <- paste0(dock_dir, "/graphs/")
221 if(dir.exists(graphs_dir)) {
222   print("graphs directory was not created because it already exists")
223 } else {dir.create(graphs_dir)}
224 setwd(graphs_dir)
225
226 #####
227 ### Box plot of binding energies (in all sites) by ligand
228 boxplots_energy_vs_lig_allsites <- ggplot(data=data, aes(x=lig, y=E)) +
229   geom_boxplot(aes(fill=lig)) +
230   scale_fill_hue(name="Ligands") + # legend
231   scale_x_discrete(limits=ligset_list) +
232   xlab("Ligand") +

```

```

231 ylab("Binding energy (kcal/mol)") +
232 ggtitle("Binding Energies by Ligand (All binding sites)") +
233 theme(legend.title=element_text(face="bold"),
234       plot.title=element_text(face="bold"))
235 boxplots_energy_vs_lig_allsites
236
237 graph_name <- "boxplots_energy_vs_lig_allsites"
238 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
239 print(paste0("Created ", graph_name))
240
241 #####
242
243 #####
244 ### Density of dockings over energy range
245 # All ligands combined
246 densities_energy_by_lig_allcombined <- ggplot(data=data, aes(x=E)) +
247   geom_density() +
248   xlab("Binding energy (kcal/mol)") +
249   ggtitle("Overall Density of Dockings versus Binding Energy") +
250   theme(legend.title=element_text(face="bold"),
251         plot.title=element_text(face="bold"))
252 densities_energy_by_lig_allcombined
253
254 graph_name <- "densities_energy_by_lig_allcombined"
255 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
256 print(paste0("Created ", graph_name))
257
258 # Separate charts for each ligand, arranged in a grid
259 # OK fine it actually wraps but whatever
260 densities_energy_by_lig_grid <- ggplot(data=data, aes(x=E, color=lig, group=
261   lig)) +
262   geom_density() +
263   scale_color_hue(name="Ligands") +
264   xlab("Binding energy (kcal/mol)") +
265   ggtitle("Density of Dockings versus Binding Energy by Ligand (All Binding
266     Sites)") +
267   theme(legend.title=element_text(face="bold"),
268         plot.title=element_text(face="bold")) +
269   facet_wrap(~lig)
270 densities_energy_by_lig_grid
271
272 graph_name <- "densities_energy_by_lig_grid"
273 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
274 print(paste0("Created ", graph_name))

```



```

274 # *A*
275 # Same thing but overlayed
276 densities_energy_by_lig_overlay <- ggplot(data=data, aes(x=E, color=lig)) +
277   geom_density() +
278   scale_color_hue(name="Ligands") +
279   scale_x_reverse(limits=c(max(data$E), min(data$E))) +
280   xlab("Binding energy (kcal/mol)") +
281   ylab("Probability density") +
282   ggtitle("Density of Dockings versus Binding Energy (All Binding Sites)") +
283   theme(legend.title=element_text(face="bold"),
284         plot.title=element_text(face="bold"))
285 densities_energy_by_lig_overlay
286
287 graph_name <- "densities_energy_by_lig_overlay"
288 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=6, height=4)
289 print(paste0("Created ", graph_name))
290
291 #####
292
293 #####
294 ### Create a new melted data frame for energies with one unique entry per
295      binding site placement
296 data.E.bs.melted <- melt(data,
297                        id.vars=c("key", "E", "lig"), # ID variables - all
298                        the variables to keep but not split apart on
299                        measure.vars=paste0("binds_in_", binding_sites) # The
300                        source columns
301                        )
302 # This new variable will simply be name of the site
303 data.E.bs.melted$binding.placement <- rep(NA, nrow(data.E.bs.melted))
304 data.E.bs.melted$binding.placement <- sub("binds_in_", "", data.E.bs.melted$
305      variable) # data.E.bs.melted$variable is binds_in_SITE
306 data.E.bs.melted <- data.E.bs.melted[data.E.bs.melted$value, c("key", "E", "
307      lig", "binding.placement")] # data.E.bs.melted$value is T/F
308 ### A density plot for all ligands with separate traces for each binding site
309 densities_energy_by_site_overlay <- ggplot(data=data.E.bs.melted, aes(x=E,
310      color=binding.placement)) +
311   geom_density() +
312   scale_color_hue(name="Binding Site", labels=binding_sites) +
313   xlab("Binding energy (kcal/mol)") +
314   ggtitle("Density of Dockings versus Binding Energy by Binding Site (All
315      Ligands)") +
316   theme(legend.title=element_text(face="bold"),
317         plot.title=element_text(face="bold"))
318 densities_energy_by_site_overlay

```

```

312
313 graph_name <- "densities_energy_by_site_overlay"
314 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
315 print(paste0("Created ", graph_name))
316
317 ### A grid of density plots (lig vs. site) with single energy traces for each
    box
318 densities_energy_by_lig_and_site_grid <- ggplot(data=data.E.bs.melted, aes(x=E
    , color=binding.placement, group=lig)) +
319   geom_density() +
320   facet_grid(binding.placement ~ lig) +
321   scale_color_hue(name="Binding Site", labels=binding_sites) +
322   xlab("Binding energy (kcal/mol)") +
323   ggtitle("Density of Dockings versus Binding Energy by Binding Site and
    Ligand") +
324   theme(legend.title=element_text(face="bold"),
325         plot.title=element_text(face="bold"))
326 densities_energy_by_lig_and_site_grid
327
328 graph_name <- "densities_energy_by_lig_and_site_grid"
329 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=1.75*(length(ligset_list)
    +2), height=2*length(binding_sites))
330 print(paste0("Created ", graph_name))
331
332 #####
333
334
335
336
337
338
339
340
341
342
343
344
345 # *A*
346 data$combined.sites <- rep(NA, nrow(data))
347 binding_sites.capitalized <- c("Lys", "CoA", "Side", "Allo1", "Allo2")
348
349 for(r in 1:nrow(data)) {
350   combined.sites <- NA
351   # for(bs in binding_sites) {
352   for(bs in binding_sites.capitalized) {

```

```

353   # if(data[r, paste0("binds_in_", bs)]) {combined.sites <- c(combined.sites
    , bs) }
354   if(data[r, paste0("binds_in_", tolower(bs))]) {combined.sites <- c(
    combined.sites, bs) }
355 }
356 data[r, "combined.sites"] <- paste(na.omit(combined.sites), collapse=" + ")
357 }
358 data$combined.sites[data$combined.sites == ""] <- "No site placement"
359
360 barplot_bindingdist_by_lig_combinedsites <- ggplot(data=data) +
361   geom_bar(aes(x=lig, fill=combined.sites), color="black", width=0.8) +
362   scale_x_discrete(limits=ligset_list) +
363   # scale_fill_hue(name="Binding Site(s)") +
364   # scale_fill_manual(values=c("orchid", "mediumpurple", "greenyellow", "
    mediumaquamarine", "grey"), name="Binding Site(s)") +
365   scale_fill_brewer(palette="Set2", name="Binding Site(s)") +
366   xlab("Ligand") +
367   ylab("Number of Ligands in Site") +
368   ggtitle("Binding Site Placement Distribution by Ligand") +
369   theme(legend.title=element_text(face="bold"),
370         plot.title=element_text(face="bold"))
371 barplot_bindingdist_by_lig_combinedsites
372
373 graph_name <- "barplot_bindingdist_by_lig_combinedsites"
374 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=6, height=4)
375 print(paste0("Created ", graph_name))
376
377
378
379
380
381
382
383
384 #####
385 ### Bar plot of binding distribution in each binding site by ligand
386 # Make a data frame with only the binding site numbers
387 analysis.bindingsites <- analysis[c(paste("Num_", binding_sites, sep=""))]
388 analysis.bindingsites$bs_cat <- row.names(analysis.bindingsites)
389 # Melt this data frame for graphing
390 melted.analysis.bindingsites <- melt(analysis.bindingsites, id.vars = "bs_cat"
    )
391 barplot_bindingdist_by_lig <- ggplot(data=melted.analysis.bindingsites, aes(x=
    bs_cat, y=value, fill=variable)) +
392   geom_bar(stat="identity", color="black") +

```

```

393 scale_x_discrete(limits=ligset_list) +
394 scale_fill_hue(name="Binding Site", labels=binding_sites) +
395 xlab("Ligand") +
396 ylab("Number of Ligands in Site") +
397 ggtitle("Binding Distribution by Ligand") +
398 theme(legend.title=element_text(face="bold"),
399       plot.title=element_text(face="bold"))
400 barplot_bindingdist_by_lig
401
402 graph_name <- "barplot_bindingdist_by_lig"
403 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
404 print(paste0("Created ", graph_name))
405
406 #####
407
408 #####
409 ### Bar plot of average binding energy in each binding site per ligand
410 # Make a data frame with only the binding site numbers
411 analysis.avgennergies <- analysis[c(paste("AvgE_", binding_sites, sep=""))]
412 analysis.avgennergies$bs_cat <- row.names(analysis.avgennergies)
413 # Melt this data frame for graphing
414 melted.analysis.avgennergies <- melt(analysis.avgennergies, id.vars = "bs_cat")
415 barplot_avge_vs_lig_by_bs <- ggplot(data=melted.analysis.avgennergies, aes(x=bs
    _cat, y=value, fill=variable, width=0.75)) +
416   geom_bar(stat="identity", color="black", position=position_dodge()) +
417   scale_x_discrete(limits=ligset_list) +
418   scale_fill_hue(name="Binding Site", labels=binding_sites) +
419   xlab("Ligand") +
420   ylab("Binding energy (kcal/mol)") +
421   ggtitle("Average Energy by Binding Site") +
422   theme(legend.title=element_text(face="bold"),
423         plot.title=element_text(face="bold"))
424 barplot_avge_vs_lig_by_bs
425
426 graph_name <- "barplot_avge_vs_lig_by_bs"
427 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
428 print(paste0("Created ", graph_name))
429
430 #####
431
432 #####
433
434 counts.max <- max(analysis[, paste0("Num_", binding_sites)])
435 ### 'Stripcharts' for energy by ligand
436 # Overall

```

```

437 vertbarplots_energy_by_lig_allsites <- ggplot(data=data, aes(x=E, fill=lig,
438   group=lig)) +
439   geom_bar(width=0.1) +
440   coord_flip() +
441   scale_fill_hue(guide=FALSE) +
442   xlab("Binding energy (kcal/mol)") +
443   scale_x_reverse(limits=c(max(data$E), min(data$E))) +
444   ylim(0, counts.max) +
445   ggtitle("Binding Affinity Frequencies by Ligand (All binding sites)") +
446   theme(plot.title=element_text(face="bold"),
447         legend.title=element_text(face="bold")) +
448   facet_grid(. ~ lig, drop=F)
449 vertbarplots_energy_by_lig_allsites
450
451 graph_name <- "vertbarplots_energy_by_lig_allsites"
452 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
453 print(paste0("Created ", graph_name))
454
455 ### For each binding site
456 # HepI:
457 if(prot == "hepi") {
458   # ADPH
459   vertbarplots_energy_by_lig_adphsite <- ggplot(data=subset(data, binds_in_
460     adph), aes(x=E, fill=lig, group=lig)) +
461     geom_bar(width=0.1) +
462     coord_flip() +
463     scale_fill_hue(guide=FALSE) +
464     xlab("Binding energy (kcal/mol)") +
465     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
466     ylim(0, counts.max) +
467     ggtitle("Binding Affinity Frequencies by Ligand (ADPH Binding Site)") +
468     theme(legend.title=element_text(face="bold")) +
469     facet_grid(. ~ lig, drop=F)
470   vertbarplots_energy_by_lig_adphsite
471
472   graph_name <- "vertbarplots_energy_by_lig_adphsite"
473   ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
474   print(paste0("Created ", graph_name))
475
476   # FDLA
477   vertbarplots_energy_by_lig_fdlasite <- ggplot(data=subset(data, binds_in_
478     fdla), aes(x=E, fill=lig, group=lig)) +
479     geom_bar(width=0.1) +
480     coord_flip() +
481     scale_fill_hue(guide=FALSE) +

```

```

479   xlab("Binding energy (kcal/mol)") +
480   scale_x_reverse(limits=c(max(data$E), min(data$E))) +
481   ylim(0, counts.max) +
482   ggtitle("Binding Affinity Frequencies by Ligand (FDLA Binding Site)") +
483   theme(legend.title=element_text(face="bold")) +
484   facet_grid(. ~ lig, drop=F)
485   vertbarplots_energy_by_lig_fdlasite
486
487   graph_name <- "vertbarplots_energy_by_lig_fdlasite"
488   ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
489   print(paste0("Created ", graph_name))
490
491   # ALLO
492   vertbarplots_energy_by_lig_allosite <- ggplot(data=subset(data, binds_in_
493     allo), aes(x=E, fill=lig, group=lig)) +
494     geom_bar(width=0.1) +
495     coord_flip() +
496     scale_fill_hue(guide=FALSE) +
497     xlab("Binding energy (kcal/mol)") +
498     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
499     ylim(0, counts.max) +
500     ggtitle("Binding Affinity Frequencies by Ligand (ALLO Binding Site)") +
501     theme(legend.title=element_text(face="bold")) +
502     facet_grid(. ~ lig, drop=F)
503   vertbarplots_energy_by_lig_allosite
504
505   graph_name <- "vertbarplots_energy_by_lig_allosite"
506   ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
507   print(paste0("Created ", graph_name))
508
509
510   ### Dan's Percent Inhibition data
511   dan.data <- read.csv("/Users/zarek/lab/Resources/Dans_Percent_Inhib_Data.csv",
512     header = T)
513   dan.data$Ligand <- factor(dan.data$Ligand, levels = ncs.sugargrouped)
514   dan_percent_inhib_barplot <- ggplot(data=dan.data, aes(x=Ligand, y=Percent.
515     Inhibition, fill=Ligand)) +
516     geom_bar(stat="identity") +
517     scale_fill_discrete(guide=F) +
518     xlab("Ligand") +
519     ylab("Percent Inhibition") +
520     ggtitle("In Vitro Percent Inhibition by Ligand (from Dan)") +
521     theme(legend.title=element_text(face="bold"))
522   dan_percent_inhib_barplot

```

```

521 ggsave("dan_percent_inhib_barplot.pdf", width=12, height=8)
522 }
523 # p300:
524 if(prot == "p300") {
525   # lys
526   vertbarplots_energy_by_lig_lyssite <- ggplot(data=subset(data, binds_in_lys)
527     , aes(x=E, fill=lig, group=lig)) +
528     geom_bar(width=0.1) +
529     coord_flip() +
530     scale_fill_hue(guide=FALSE) +
531     xlab("Binding energy (kcal/mol)") +
532     ylim(0, counts.max) +
533     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
534     ggtitle("Binding Affinity Frequencies by Ligand (lys Binding Site)") +
535     theme(legend.title=element_text(face="bold")) +
536     facet_grid(. ~ lig, drop=F)
537   vertbarplots_energy_by_lig_lyssite
538   graph_name <- "vertbarplots_energy_by_lig_lyssite"
539   ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
540   print(paste0("Created ", graph_name))
541
542   # coa
543   vertbarplots_energy_by_lig_coasite <- ggplot(data=subset(data, binds_in_coa)
544     , aes(x=E, fill=lig, group=lig)) +
545     geom_bar(width=0.1) +
546     coord_flip() +
547     scale_fill_hue(guide=FALSE) +
548     xlab("Binding energy (kcal/mol)") +
549     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
550     ylim(0, counts.max) +
551     ggtitle("Binding Affinity Frequencies by Ligand (coa Binding Site)") +
552     theme(legend.title=element_text(face="bold")) +
553     facet_grid(. ~ lig, drop=F)
554   vertbarplots_energy_by_lig_coasite
555   graph_name <- "vertbarplots_energy_by_lig_coasite"
556   ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
557   print(paste0("Created ", graph_name))
558
559   # side
560   vertbarplots_energy_by_lig_sidesite <- ggplot(data=subset(data, binds_in_
561     side), aes(x=E, fill=lig, group=lig)) +
562     geom_bar(width=0.1) +
563     coord_flip() +

```

```

563     scale_fill_hue(guide=FALSE) +
564     xlab("Binding energy (kcal/mol)") +
565     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
566     ylim(0, counts.max) +
567     ggtitle("Binding Affinity Frequencies by Ligand (side Binding Site)") +
568     theme(legend.title=element_text(face="bold")) +
569     facet_grid(. ~ lig, drop=F)
570 vertbarplots_energy_by_lig_sidesite
571
572 graph_name <- "vertbarplots_energy_by_lig_sidesite"
573 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
574 print(paste0("Created ", graph_name))
575
576 # allo1
577 vertbarplots_energy_by_lig_allo1site <- ggplot(data=subset(data, binds_in_
578     allo1), aes(x=E, fill=lig, group=lig)) +
579     geom_bar(width=0.1) +
580     coord_flip() +
581     scale_fill_hue(guide=FALSE) +
582     xlab("Binding energy (kcal/mol)") +
583     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
584     ylim(0, counts.max) +
585     ggtitle("Binding Affinity Frequencies by Ligand (allo1 Binding Site)") +
586     theme(legend.title=element_text(face="bold")) +
587     facet_grid(. ~ lig, drop=F)
588 vertbarplots_energy_by_lig_allo1site
589
590 graph_name <- "vertbarplots_energy_by_lig_allo1site"
591 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
592 print(paste0("Created ", graph_name))
593
594 # allo2
595 vertbarplots_energy_by_lig_allo2site <- ggplot(data=subset(data, binds_in_
596     allo2), aes(x=E, fill=lig, group=lig)) +
597     geom_bar(width=0.1) +
598     coord_flip() +
599     scale_fill_hue(guide=FALSE) +
600     xlab("Binding energy (kcal/mol)") +
601     scale_x_reverse(limits=c(max(data$E), min(data$E))) +
602     ylim(0, counts.max) +
603     ggtitle("Binding Affinity Frequencies by Ligand (allo2 Binding Site)") +
604     theme(legend.title=element_text(face="bold")) +
605     facet_grid(. ~ lig, drop=F)
606 vertbarplots_energy_by_lig_allo2site

```



```

606 graph_name <- "vertbarplots_energy_by_lig_allo2site"
607 ggsave(paste0(dock, "_", graph_name, ".pdf"), width=12, height=8)
608 print(paste0("Created ", graph_name))
609
610 }
611 #####
612
613
614
615 # Multiple plot function
616 #
617 # ggplot objects can be passed in ..., or to plotlist (as a list of ggplot
  objects)
618 # - cols: Number of columns in layout
619 # - layout: A matrix specifying the layout. If present, 'cols' is ignored.
620 #
621 # If the layout is something like matrix(c(1,2,3,3), nrow=2, byrow=TRUE),
622 # then plot 1 will go in the upper left, 2 will go in the upper right, and
623 # 3 will go all the way across the bottom.
624 #
625
626 # multiplot <- function(..., plotlist=NULL, file, cols=1, layout=NULL) {
627 #   library(grid)
628 #
629 #   # Make a list from the ... arguments and plotlist
630 #   plots <- c(list(...), plotlist)
631 #
632 #   numPlots = length(plots)
633 #
634 #   # If layout is NULL, then use 'cols' to determine layout
635 #   if (is.null(layout)) {
636 #     # Make the panel
637 #     # ncol: Number of columns of plots
638 #     # nrow: Number of rows needed, calculated from # of cols
639 #     layout <- matrix(seq(1, cols * ceiling(numPlots/cols)),
640 #                       ncol = cols, nrow = ceiling(numPlots/cols))
641 #   }
642 #
643 #   if (numPlots==1) {
644 #     print(plots[[1]])
645 #
646 #   } else {
647 #     # Set up the page
648 #     grid.newpage()
649 #     pushViewport(viewport(layout = grid.layout(nrow(layout), ncol(layout))))

```

```

650 #
651 #   # Make each plot, in the correct location
652 #   for (i in 1:numPlots) {
653 #       # Get the i,j matrix positions of the regions that contain this
        subplot
654 #       matchidx <- as.data.frame(which(layout == i, arr.ind = TRUE))
655 #
656 #       print(plots[[i]], vp = viewport(layout.pos.row = matchidx$row,
657 #                                         layout.pos.col = matchidx$col))
658 #   }
659 # }
660 # }
661
662
663
664 # barplot_bindingdist_by_lig_combinedsites
665 # vertbarplots_energy_by_lig_allsites
666 # densities_energy_by_lig_overlay
667
668 # combined_bar_bySite_and_density_byLig <- multiplot(barplot_bindingdist_by_
        lig_combinedsites,
669 #                                         densities_energy_by_lig_
        overlay, cols=2)
670 # print(paste0(dock, "_", "combined_bar_bySite_and_density_byLig", ".pdf"))#,
        width=12, height=4)
671
672 # combined_bar_bySite_and_histograms_byLig <- multiplot(barplot_bindingdist_by
        _lig_combinedsites,
673 #                                         vertbarplots_energy_by
        _lig_allsites, cols=2)
674 # print(paste0(dock, "_", "combined_bar_bySite_and_histograms_byLig", ".pdf"))
        #, width=12, height=4)
675
676
677
678
679
680
681
682
683
684
685
686
687 #####

```

```

688 ### Save data tables for % Distribution
689 distrib.table <- analysis[paste0("DistribFrac_", binding_sites)]
690 for(bs in binding_sites) {
691   distrib.table[, bs] <- round(distrib.table[paste0("DistribFrac_", bs)] *
692     100, 2)
693 }
694 distrib.table <- distrib.table[binding_sites]
695 distrib_table_csv <- paste0(dock_dir, "/", dock, "_binding_site_distribution_
696   table.csv")
697 write.csv(distrib.table, file=distrib_table_csv)
698 print(paste0("Created ", "binding_site_distribution_table"))
699 #####
700
701
702
703
704
705
706
707 #
708 #####
709
710 print("All done, graphs can found in:")
711 print(graphs_dir, quote=F)

```

/Users/zarek/lab/zvina/scripts/postdocking_graphs.R

2.17 pre_and_post_control.py

2.17.1 Function

Transcompiler picojava pseudolanguage java ee regular expression syntactic sugar eclipse rust bug. Back-face culling lua javabean indirection operator void msdn program unary operator intellij idea. Bug tracking library softwar

```
1  #!/usr/bin/env python
2
3  ### One script to control them AAAAAALLLLL!!!!
4  # (c) Zarek Siegel
5  # v1 3/6/16
6
7  import argparse, subprocess, os
8  import new_grid_or_dock_entry
9  from constants import *
10 from create_docking_object import * # Docking
11 from write_vina_submit_sh import * # write_vina_submit_sh
12 from get_pose_energies_properties import * # get_pose_energies_properties
13 from mine_vina_data import * # mine_vina_data
14 from binding_site_analysis import * # get_binding_sites_list
15 # from binding_site_analysis import * # score_binding_sites
16 # from binding_site_analysis import * # aiad_icpd_binding_sites
17 # from binding_site_analysis import * # assess_all_resis
18 from write_alldata import * # write_alldata_csv, write_docking_pickled
19 from read_alldata import * # read_alldata_csv, read_docking_pickled
20 from correlations import * # correlations
21
22 # from docking_data_assembly import *
23
24
25 def main():
26     print("")
27     print("->-> hi")
28
29     parser = argparse.ArgumentParser(description='Pre- and post-Vina file fun
30         times')
31     parser.add_argument('-d', '--dock', metavar='DOCK', type=str, nargs='?',
32         help='the ID for this docking')
33
34     parser.add_argument('-nd', '--new_docking', action='store_true', default=
35         False,
36         help='Write a new set of docking parameters to Dockings.csv')
37     parser.add_argument('-ng', '--new_gridbox', action='store_true', default=
38         False,
```

```

36     help='Write a new set of grid box parameters to Gridboxes.csv')
37     parser.add_argument('-v', '--vina', action='store_true', default=False,
38     help='write the Vina job submission script')
39     parser.add_argument('-p', '--print', action='store_true', default=False,
40     help='print docking parameters')
41
42     parser.add_argument('-s', '--separate', action='store_true', default=False,
43     help='execute the bash script to separate row Vina results')
44     parser.add_argument('-n', '--clean', action='store_true', default=False,
45     help='execute the bash script to clean up processed Vina results')
46
47     parser.add_argument('-rc', '--read_csv', action='store_true', default=False,
48     help='load data from alldata CSV file')
49     parser.add_argument('-ri', '--read_pickle', action='store_true', default=
50     False,
51     help='load data from pickled object')
52
53     parser.add_argument('-bs', '--binding_sites', action='store_true', default=
54     False,
55     help='score binding sites by fraction of binding site residues contacted')
56     parser.add_argument('-ai', '--aiad_icpd', action='store_true', default=False
57     ,
58     help='score binding sites by AIAD and ICPD')
59     parser.add_argument('-ar', '--all_resis', action='store_true', default=False
60     ,
61     help='asses contacts with all residues')
62     parser.add_argument('--atoms', action='store_true', default=False,
63     help='assess poses against all atoms (not just residues) for -bs and -ar')
64
65     parser.add_argument('-l', '--cluster', action='store_true', default=False,
66     help='create cluster CSV files (all lig x lig AIADs)')
67     parser.add_argument('-co', '--correls', action='store_true', default=False,
68     help='find correlations between all quantitative variables')
69     parser.add_argument('-c', '-wc', '--write_csv', action='store_true', default
70     =False,
71     help='generate and save the alldata CSV file')
72     parser.add_argument('-i', '-wi', '--write_pickle', action='store_true',
73     default=False,
74     help='save the entire docking as a pickled object')
75
76     parser.add_argument('-ep', '--en_props', action='store_true', default=False,
77     help='Write a new set of docking parameters to Dockings.csv')
78     parser.add_argument('-o', '--post_proc', action='store_true', default=False,
79     help='Perform all post-processing steps (separate, clean, csv, cluster,
80     pickle)')

```

```

74 parser.add_argument('-g', '--graphs', action='store_true', default=False,
75                     help='Generate graphs for this docking')
76
77 args = vars(parser.parse_args())
78
79
80 if args['new_docking']:
81     print("---> Write new set of docking parameters to Dockings.csv...")
82     new_grid_or_dock_entry.new_docking_entry()
83 elif args['new_gridbox']:
84     print("---> Write new set of grid box parameters to Gridboxes.csv...")
85     new_grid_or_dock_entry.new_gridbox_entry()
86 else:
87     dock = str(args['dock'])
88     d = Docking(dock)
89     if args['print']:
90         d.print_parameters()
91     if args['vina']:
92         print("---> Writing Vina submission script")
93         d.write_vina_submit_sh()
94     if args['separate']:
95         print("---> Processing raw Vina output PDBQTs")
96         d.export_parameters_to_environment()
97         subprocess.call(["{b_d}/scripts/separate_vina_results.sh".format(b_d=
base_dir), dock])
98     if args['clean']:
99         print("---> Cleaning up processed PDBQTs and converting to PDBs")
100         d.export_parameters_to_environment()
101         subprocess.call(["{b_d}/scripts/cleanup_processed_vina_results.sh".
format(b_d=base_dir), dock])
102     if args['read_csv']: d.read_alldata_csv()
103     if args['read_pickle']: d.read_docking_pickled()
104     if args['en_props']:
105         print("---> Getting molecular energies and properties for all poses")
106         d.get_pose_energies_properties()
107     if args['atoms']: d.evaluate_resis_atoms = True
108     else: d.evaluate_resis_atoms = False
109     if args['binding_sites']: d.score_binding_sites()
110     if args['aiad_icpd']: d.aiad_icpd_binding_sites()
111     if args['all_resis']: d.assess_all_resis()
112     if args['cluster']: d.cluster_poses()
113     if args['write_csv']: d.write_alldata_csv()
114     if args['correls']: d.correlations()
115     if args['write_pickle']: d.write_docking_pickled()
116     if args['graphs']:

```

```

117     print("---> Generating graphs")
118     subprocess.call([Rscript_binary, "{b_d}/scripts/postdocking_graphs.R".
119     format(b_d=base_dir), dock])
120     if args['post_proc']:
121         print("---> Processing raw Vina output PDBQTs")
122         subprocess.call(["{b_d}/scripts/separate_vina_results.sh".format(b_d=
123         base_dir),
124         dock, base_dir, AutoDockTools_dir, AutoDockTools_pythonsh_binary])
125         print("---> Cleaning up processed PDBQTs and converting to PDBs")
126         subprocess.call(["{b_d}/scripts/cleanup_processed_vina_results.sh".
127         format(b_d=base_dir),
128         dock, base_dir, AutoDockTools_dir, AutoDockTools_pythonsh_binary])
129         d.write_alldata_csv()
130         d.cluster_poses()
131         d.save_pickled_docking_obj()
132
133     print("->> All done!!!!!!!!!!!!!!!!!!!!")
134     print("")
135
136 if __name__ == "__main__": main()
137
138
139
140
141 # print("{} = {}".format("parameters_loaded", self.parameters_loaded))
142 # print("{} = {}".format("ligset_list_gotten", self.ligset_list_gotten))
143 # print("{} = {}".format("parameters_exported_to_environment", self.parameters
144 #     _exported_to_environment))
145 # print("{} = {}".format("is_data_dic_created", self.is_data_dic_created))
146 #
147 # print("{} = {}".format("vina_data_mined", self.vina_data_mined))
148 # print("{} = {}".format("binding_sites_list_gotten", self.binding_sites_list_
149 #     gotten))
150 # print("{} = {}".format("binding_sites_scored", self.binding_sites_scored))
151 # print("{} = {}".format("aiad_icpd_calcd", self.aiad_icpd_calcd))
152 # print("{} = {}".format("all_resis_assessed", self.all_resis_assessed))
153 #
154 # print("{} = {}".format("is_csv_written", self.is_csv_written))
155 # print("{} = {}".format("energies_props_gotten", self.energies_props_gotten))
156 # print("{} = {}".format("are_poses_clustered", self.are_poses_clustered))
157 # print("{} = {}".format("is_pickled", self.is_pickled))

```

/Users/zarek/lab/zvina/scripts/pre_and_post_control.py