## Data exploration with tidyverse

Kevin Y. X. Wang 31 July 2017

S0: Prior to lecture

#### Preparing for this lecture

- All materials are on Ed and https://github.com/kevinwang09/2017\_STAT3914.
- Please run these codes on your laptop,

• Familiar yourself with the iris dataset. Typing iris into R console should load this data. Pay attention to its column, row names, summary statistics and structure of each column.

S1: Necessary of Applied Statistics

# Good statistical discoveries don't fall out from the sky

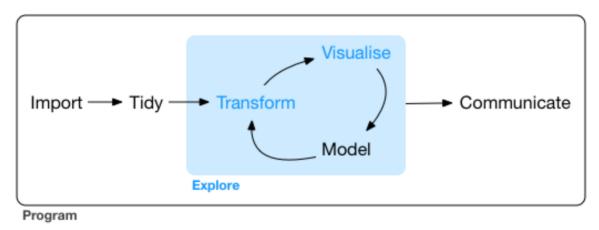
- Statisticians are great at many things:
  - 1. Understanding data characteristics
  - 2. Building statistical/mathematical models
  - 3. Repeat 1 and 2...like...a lot...
  - 4. Extract insights
- But the mother of all these, i.e. **preparing data** is not trivial. (e.g. STAT2xxx lab exams)

#### Let X be the thing I want...

• The real problem is not applying fancy shampoo for your cat. It is getting your cat into the bathtub.



#### Hidden side of being a statistician



- Assume we have data
- · Assume we have data that can answer our questions
- · Assume we have cleaned data
- Assume we interrogated the right aspects of the data using appropriate statistics
- Assume we did everything right, communicate insights with others

#### Aim: effectively clean your data (1)

- "Your statistical model is only ever going to be as good as your data quality"
  Kevin Wang.
- There will be no recipe, there will be a lot of back and forth exploration.
- Computational and visualisation tools.

SepAlLeNgth	Sepal.? Width	petal.Length(*&^	petal.\$%^&Width	species^
6.2	2.8	4.8	1.8	virginica
6.8	2.8	4.8	1.4	versicolor
7.6	3	6.6	2.1	virginica
NA	NA	NA	NA	NA
5.4	3.4	1.7	0.2	setosa

- · Corrupted column names, 100% missing column, 100% missing rows, rows with at least 1 missing value.
- Most severe problem: rows with random values.

#### Aim: effectively clean your data (2)

- The classical iris data is known to be well-separated.
- Running Support Vector Machine (SVM) classification algorithm on the cleaned iris data has very low number of classifications.
- · True iris data

	setosa	versicolor	virginica
setosa	50	0	0
versicolor	0	48	2
virginica	0	2	48

### Aim: effectively clean your data (3)

- Not so much when you have corruptions.
- In addition of introduce missing values, I also created non-sense rows in the data, they corrupted classification results.

	setosa	versicolor	virginica
setosa	89	25	27
versicolor	0	47	4
virginica	0	3	46

#### Summary of this lecture

- Passive learning is not going to work.
- S1: Introduction
- S2: Reading in data using readr and readx1
- · S3: Basic data cleaning using janitor
- S4: Clean coding using magrittr
- S5: Data filtering using dplyr
- S6: Data visualisation using ggplot2
- · S7: Conclusion

S2: Reading data

#### Better read/write data

- base R functions are not sufficient for modern uses.
- readr functions are superior in data import warnings, column type handling, speed, scalability and consistency.

library(readr)

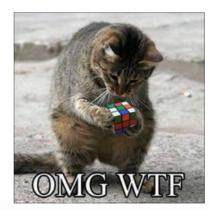
#### Reading data using (1)

```
dirtyIris = readr::read csv("dirtyIris.csv")
## Parsed with column specification:
## cols(
## SepAl...LeNgth = col double(),
## `Sepal.? Width` = col double(),
## `petal.Length(*&^` = col double(),
## `petal.$#^&Width` = col_double(),
## `SPECIES^` = col character(),
## allEmpty = col character()
## )
class(dirtyIris) ## `tibble` is a `data.frame` with better formatting.
## [1] "tbl df" "tbl" "data.frame"
• readx1 and haven (for SAS, SPSS etc) packages work similarly.
```

#### Reading data using (2)

#### dirtyIris

```
## # A tibble: 650 x 6
    SepAl....LeNgth `Sepal.? Width `petal.Length(*&^` `petal.$#^&Width`
              <dbl>
                                 <dbl>
                                                   <dbl>
                                                                     <dbl>
                                                                  2.200000
## 1
         7.7000000
                                   3.8
                                                     6.7
         -0.1842525
                                   NA
                                                     NA
                                                                 1.099848
## 3
      7.2000000
                                   3.6
                                                     6.1
                                                                  2.500000
        6.3000000
## 4
                                  2.3
                                                     4.4
                                                                 1.300000
                                  2.9
## 5
          5.6000000
                                                     3.6
                                                                  1.300000
## # ... with 645 more rows, and 2 more variables: `SPECIES^` <chr>,
      allEmpty <chr>
## #
```



· We now proceed to data cleaning on the dirtyIris dataset.

#### Too trivial? Here is a short homework

Here is a dataset. Click here.

- 1. Write 2 sentences about what is a .gmt file and who publishes this format?
- 2. Which packages can read in .gmt files?
- 3. How to download this package?
- 4. What class is this data once read into R? Is it a data.frame?
- 5. The data contains 50 different gene-sets. What is the size of each gene-set?
- 6. What is the mostly frequent mentioned 6 genes?

S3: Cleaned data

#### What is clean data?

Clean data is a data set that allows you to do statistical modelling without extra processing

- 1. Good documentation on the entire data.
- 2. Each column is a variable. The name should be informative, and:
  - No bad characters/formatting @KevinWang009
  - No inconsistent capitalisation or separators (Cricket\_australia vs cricket.Australia)
- 3. Each row is an **observation**:
  - No bad characters
  - No poorly designed row names (3, 2, 5, ...)
  - No repeated row names (a, a.1, b, b.1, ...)

#### Data cleaning in

- · Clean data is a well-designed data.frame.
- · Column type (esp. dates and factors) handling was the primary reason we used readr instead of base R when importing data.
- Our goal: clean the dirtyIris data to be exactly the same as the original iris data.
  - Basic data cleaning using janitor package.
  - More advanced data manipulation through dplyr.

#### : basic data cleaning

· Clean up the bad column names

```
library(janitor)
library(dplyr)
glimpse(dirtyIris)
## Observations: 650
## Variables: 6
## $ SepAl...LeNgth <dbl> 7.70000000, -0.18425254, 7.20000000, 6.300000...
## $ Sepal.?
              Width <dbl> 3.8000000, NA, 3.6000000, 2.3000000, 2.900000...
## $ petal.Length(*&^ <dbl> 6.7000000, NA, 6.1000000, 4.4000000, 3.600000...
## $ petal.$#^&Width <dbl> 2.2000000, 1.0998477, 2.5000000, 1.3000000, 1...
               <chr> "virginica", "setosa", "virginica", "versicol...
## $ SPECIES^
## Clean up column names
better = clean names(dirtyIris)
glimpse(better)
## Observations: 650
## Variables: 6
## $ sepal length <dbl> 7.70000000, -0.18425254, 7.20000000, 6.30000000, ...
## $ sepal width <dbl> 3.8000000, NA, 3.6000000, 2.3000000, 2.9000000, -...
## $ petal length <dbl> 6.7000000, NA, 6.1000000, 4.4000000, 3.6000000, 0...
## $ petal width <dbl> 2.2000000, 1.0998477, 2.5000000, 1.3000000, 1.300...
```

#### : removal of empty rows and columns

Purely empty rows/columns are non-informative.

```
## Removing empty rows/columns
evenBetter = remove_empty_rows(better)
evenBetter = remove_empty_cols(evenBetter)

## Observations: 650

## Variables: 5

## $ sepal_length <dbl> 7.70000000, -0.18425254, 7.20000000, 6.30000000, ...

## $ sepal_width <dbl> 3.8000000, NA, 3.6000000, 2.3000000, 2.9000000, -...

## $ petal_length <dbl> 6.7000000, NA, 6.1000000, 4.4000000, 3.6000000, 0...

## $ petal_width <dbl> 2.2000000, 1.0998477, 2.5000000, 1.3000000, 1.300...

## $ species <chr> "virginica", "setosa", "virginica", "versicolor",...
```

#### : removal of rows with NA

• Genuinely missing values should be retained, but in this case, the NA's were added. Only use na.omit when you 100% certain of the structure of your data.

```
evenBetterBetter = na.omit(evenBetter)
almostIris = evenBetterBetter
glimpse(almostIris)
## Observations: 241
## Variables: 5
## $ sepal length <dbl> 7.70000000, 7.20000000, 6.30000000, 5.60000000, 6...
## $ sepal width <dbl> 3.8000000, 3.6000000, 2.3000000, 2.9000000, 2.500...
## $ petal length <dbl> 6.7000000, 6.1000000, 4.4000000, 3.6000000, 4.900...
## $ petal width <dbl> 2.2000000, 2.5000000, 1.3000000, 1.3000000, 1.500...
## $ species <chr> "virginica", "virginica", "versicolor", "versicol...
glimpse(iris)
## Observations: 150
## Variables: 5
## $ Sepal.Length <dbl> 5.1, 4.9, 4.7, 4.6, 5.0, 5.4, 4.6, 5.0, 4.4, 4.9,...
## $ Sepal.Width <dbl> 3.5, 3.0, 3.2, 3.1, 3.6, 3.9, 3.4, 3.4, 2.9, 3.1,...
## $ Petal.Length <dbl> 1.4, 1.4, 1.3, 1.5, 1.4, 1.7, 1.4, 1.5, 1.4, 1.5,...
## $ Petal.Width <dbl> 0.2, 0.2, 0.2, 0.2, 0.2, 0.4, 0.3, 0.2, 0.2, 0.1,...
## с с----
```

S4: Clean coding

## Coding complexity increases with the number of brackets

• The "inside out" structure of coding isn't great for human reading.

```
mean(almostIris$sepal_length)

## [1] 3.602734

plot(density(almostIris$sepal_length), col = "red", lwd = 2)
```

#### Piping: read code from left to right

- We introduce a new notation: "x %>% f" means "f(x)". We call this operation as "x pipe f".
- · Compounded operations are possible. Keyboard shortcut is Cmd+shift+M.

```
almostIris$sepal_length %>% mean

## [1] 3.602734

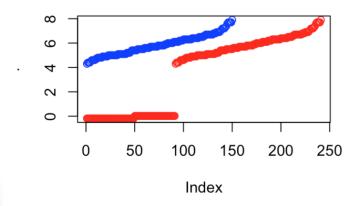
almostIris$sepal_length %>%
  density %>%
  plot(col = "red", lwd = 2)
```

### Using an informative variable (Sepal.Length) in iris to guide cleaning

```
almostIris$sepal_length %>%
  sort %>%
  plot(col = "red", main = "almostIris is in red, true iris is in blue")

iris$Sepal.Length %>%
  sort %>%
  points(col = "blue")
```

#### almostlris is in red, true iris is in blue



S5: dplyr: data subsetting master

#### Traditional way of subsetting data in R (1)

· If I want remove all observations with sepal length less than 2:

```
cleanIris = almostIris[almostIris[, "sepal_length"] > 2, ]
glimpse(cleanIris)

## Observations: 150

## Variables: 5

## $ sepal_length <dbl> 7.7, 7.2, 6.3, 5.6, 6.3, 5.5, 5.0, 6.4, 6.2, 6.7,...

## $ sepal_width <dbl> 3.8, 3.6, 2.3, 2.9, 2.5, 2.4, 3.3, 2.7, 3.4, 3.1,...

## $ petal_length <dbl> 6.7, 6.1, 4.4, 3.6, 4.9, 3.7, 1.4, 5.3, 5.4, 4.4,...

## $ petal_width <dbl> 2.2, 2.5, 1.3, 1.3, 1.5, 1.0, 0.2, 1.9, 2.3, 1.4,...

## $ species <chr> "virginica", "virginica", "versicolor", "versicol...
```

- We now have agreement over the size of the two data!
- But this subsetting code is a bit cumbersome!

#### Traditional way of subsetting data in R (2)

- Subsetting data in base R might not be the most concise solution.
- Suppose we wish to extract first two rows of column sepal\_length and sepal width in the cleanIris data:

```
## Assuming you know the position of column names.
## But what if you resample your data?
cleanIris[1:2, c(1, 2)]

## Assuming you know the position of column names.
## Also assuming the first two columns satisfy certain properties.
cleanIris[1:2, c(T, T, F, F, F)]

## Much better!
## What if you can't type out all the column names
## due to the size of your data?
cleanIris[1:2, c("sepal_length", "sepal_width")]
```

#### Traditional way of subsetting data in R (3)

• Even more complex subsetting: we want to extract rows based on some compounded criteria and select columns based on special keywords.

· (Optional) A pro R user might know about the subset function, but it suffers the same problem of not able to have multiple subsetting criteria without predefined variables.

#### Subsetting data using

- Think of subsetting rows and columns as two **separate different procedures**:
- · select columns are operations on variables, and
- filter rows are operations on observations
- · See dplyr cheatsheet.

```
## # A tibble: 4 x 2
## sepal length petal length
         <dbl>
                <dbl>
                   1.3
## 1
          4.5
## 2
          4.9
                    3.3
## 3
          4.4
                  1.4
## 4
          4.9
                4.5
```

#### arrange for ordering rows

```
arrangeCleanIris = cleanIris %>%
   arrange(sepal_length, sepal_width, petal_length, petal_width)

## The true iris data
arrangeIris = iris %>%
   clean_names() %>%
   arrange(sepal_length, sepal_width, petal_length, petal_width)
```

#### Checking if we cleaned the data properly

· We sorted both the processed dirtyIris data and the arranged iris data.

```
## The `Species` column is character or factor
all.equal(arrangeCleanIris, arrangeIris)

## [1] "Incompatible type for column `species`: x character, y factor"

arrangeIris = arrangeIris %>%
   mutate(species = as.character(species))

## Great!
all.equal(arrangeCleanIris, arrangeIris)

## [1] TRUE
```

#### Job done!



#### But what about the modelling?

- · Cleaned data is one thing, but again, we need to extract insights about the data.
- This can be done via summary statistics, visualisation or running statistical models.
- "Your statistical insights is only going to be as good as the question you ask"
   Kevin Wang.

#### : mutate create new columns

```
## # A tibble: 150 \times 7
  sepal length sepal width petal length petal width species V1
                                           V2
     <dbl> <dbl>
                 ##
## 1
       7.7 3.8
                     6.7 2.2 virginica 3.9 7.7
## 2
  7.2 3.6 6.1 2.5 virginica
                                       3.6 7.2
## 3 6.3 2.3 4.4 1.3 versicolor
                                       4.0 6.3
## 4 5.6 2.9 3.6 1.3 versicolor
                                       2.7 5.6
## 5 6.3 2.5 4.9 1.5 versicolor 3.8 6.3
## # ... with 145 more rows
```

# group\_by + summarise will create summary statistics for grouped variables

```
group by (species)
bySpecies
## # A tibble: 150 x 5
## # Groups: species [3]
   sepal length sepal width petal length petal width species
##
                          <dbl> <dbl>
        <dbl>
                 <db1>
                                          <chr>
                     6.7 2.2 virginica
## 1
         7.7 3.8
## 2
         7.2 3.6 6.1 2.5 virginica
## 3 6.3 2.3 4.4
                                    1.3 versicolor
## 4
         5.6 2.9
                           3.6
                                    1.3 versicolor
## 5 6.3
              2.5
                      4.9
                                   1.5 versicolor
## # ... with 145 more rows
```

bySpecies = cleanIris %>%

## special select functions (advanced)

• select only if a column satisfy a certain condition

```
bySpecies %>%
  summarise if(is.numeric,
              funs(m = mean))
## # A tibble: 3 x 5
       species sepal length m sepal width m petal length m petal width m
##
         <chr>
                       <dbl>
                                     <dbl>
                                                    <dbl>
                                                                 <dbl>
## 1
        setosa
                       5.006
                                     3.428
                                                   1.462
                                                                 0.246
## 2 versicolor
                      5.936
                                    2.770
                                                   4.260
                                                                1.326
## 3 virginica
                      6.588
                                     2.974
                                                  5.552
                                                                 2.026
cleanIris %>%
  select(starts_with("sepal")) %>%
 top n(3, sepal width)
## # A tibble: 3 x 2
    sepal length sepal width
##
           <dbl>
                       <dbl>
## 1
             5.2
                        4.1
             5.5 4.2
## 2
## 3
             5.7
                        4.4
```

### left\_join for merging data

```
flowers = data.frame(species = c("setosa", "versicolor", "virginica"),
                 comments = c("meh", "kinda okay", "love it!"))
## cleanIris has the priority in this join operation
iris comments = left join(cleanIris, flowers, by = "species")
## Warning: Column `species` joining character vector and factor, coercing
## into character vector
## Randomly sampling 6 rows
sample n(iris comments, 6)
## # A tibble: 6 x 6
##
    sepal length sepal width petal length petal width species
                                                        comments
##
         <dbl>
                              <dbl>
                                      <dbl>
                                                 <chr>
                   <dbl>
                                                         <fctr>
           7.2 3.2 6.0 1.8 virginica love it!
## 1
## 2 5.6 2.9 3.6 1.3 versicolor kinda okay
## 3
           5.1
                    3.4 1.5
                                          0.2
                                                setosa
                                                            meh
## 4 6.8
                    3.0 5.5
                                          2.1 virginica love it!
```

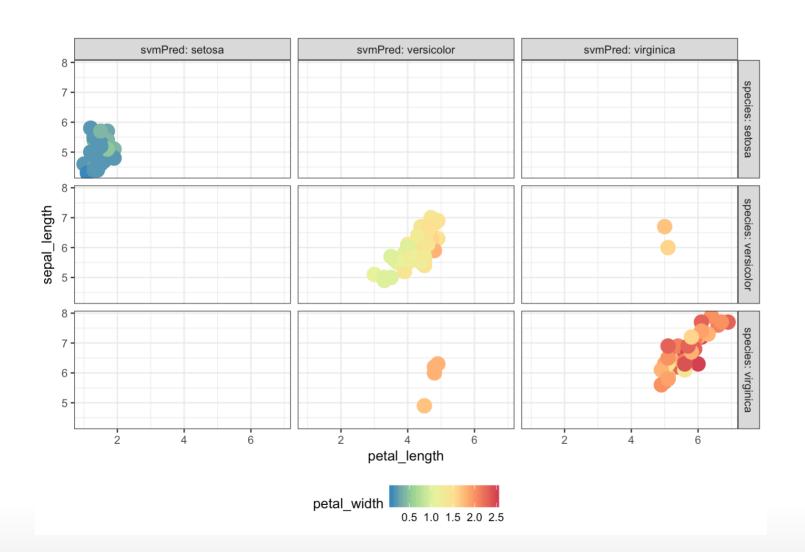
S6: ggplot2: the best visualisation package

## Why do we visualise? (1)

- · datasaurus: all statistics describe the data in some limited ways.
- Plots usually give more dimensions to our analysis.
- Suppose in our cleanIris data, we will use sepal\_length and petal\_length as SVM predictors for the classes of iris flowers.

	setosa	versicolor	virginica
setosa	50	0	0
versicolor	0	48	2
virginica	0	4	46

# Why do we visualise? (2)



## : the philosophy

- Di Cook the real reason that you should use ggplot2 is that, its design will force you to use a certain grammar when producing a plot.
- $\frac{1}{n} \sum_{i=1}^{n} X_i$  is a transformation of random variables, i.e., a statistic which provides insights into a data.
- · Similarly, ggplot is also a statistic, because we take components of the data and presented it in an informative way.
- Publishing quality, rigourous syntax and design, flexible customisations, facetting.

#### tutorial sheet

• If you managed to install all packages successfully, you should be able to run the following to get an interactive tutorial sheet.

```
library(learnr3914)
learnggplot2()
```

- Otherwise, please download and compile the "ggplot2\_basic\_tutorial.Rmd" from Ed or here
- If all fails, try <a href="https://gauss17gon.shinyapps.io/ggplot2\_basic\_tutorial">https://garthtarr.shinyapps.io/ggplot2\_basic\_tutorial</a>

S7: Conclusion

# tidy data, coding, modelling and reporting

- tidyverse is a collection of 20+ packages built on the philosophy of being organised for the purpose of collaboration.
- · These functions:
  - Well designed programming and data science solutions.
  - They will always throw errors at you if you don't have a thorough understanding of your data.
  - Capable for functional programming.



#### Peek at the

http://edinbr.org/edinbr/2016/05/11/may-Hadley-Update2-PostingTalk.html



# Interactive plotting from ggplot

library(plotly)
ggplotly(p2)

#### Advice in the future

- Use RStudio + RMarkdown to document your codes.
- · Learn some computational tools. They are not statistics, but not learning them could inhibit your career aspects.
- Find "cool" components and adapt those into your work routine. (Hint: start with all RStudio cheatsheets and build up gradually.)
- Take time to re-analyse an old dataset.
- · Learn core functions and vignette.
- Don't forget the theories and interpretations! This is a course about statistics after all, not Cranking-Out-Numbers-Less-Than-0.05-And-Reject-Null-Hypothesis-101.

#### **Session Info and References**

- · Dr. Garth Tarr
- tidyverse.org
- github.com/sfirke/janitor
- · gapminder.org
- rstudio.com