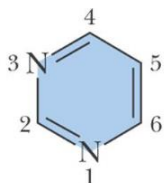


Unit 2 Exam Study Guide

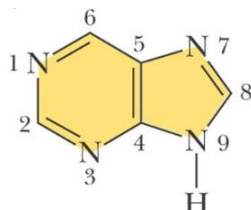
DNA Structure and Organization

Nitro Bases

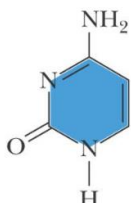
- Insol in h₂O bc arom



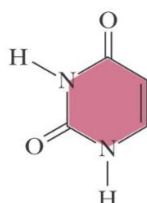
The pyrimidine ring



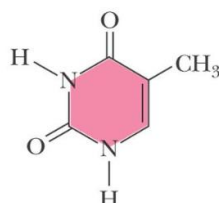
The purine ring system



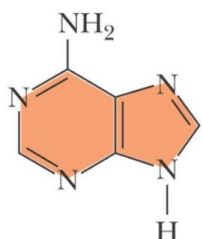
Cytosine
(2-oxy-4-amino
pyrimidine)



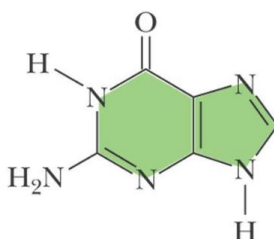
Uracil
(2-oxy-4-oxy
pyrimidine)



Thymine
(2-oxy-4-oxy
5-methyl pyrimidine)



Adenine
(6-amino purine)



Guanine
(2-amino-6-oxy purine)

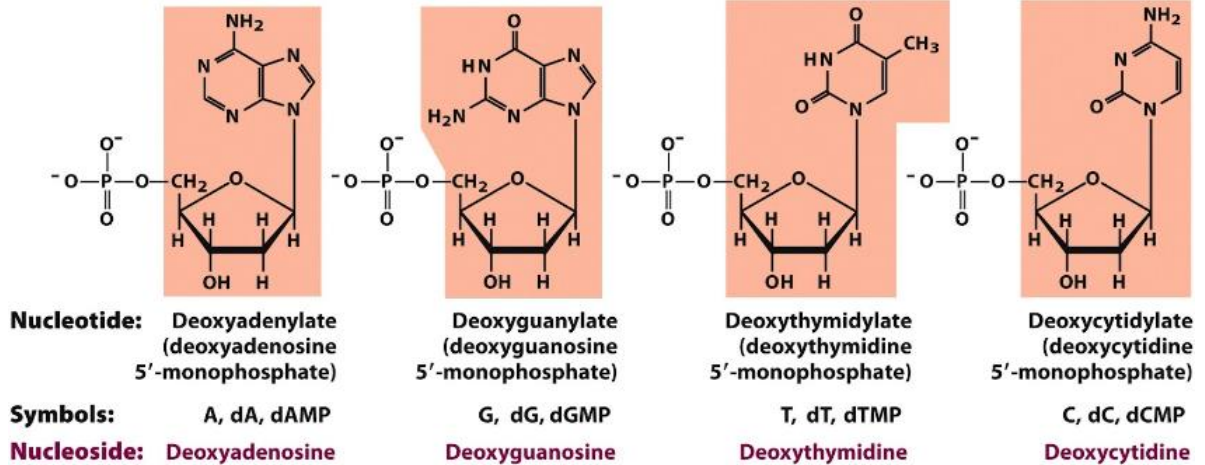
Nucleoside

- 9'N of pur or 1'N of pyr + glyco bond + 1'C deoxy/ribose sugar
- Sugar addition makes molec more water sol
- DNA more stable since deoxy; OH at 2' (on ribose) has effect on 2ndry struc & hydrolysis
- Syn conf favored by pur more than pyr bc steric hind
- Common ribonucleosides (ribose+nitro base): CYTidine, URidine, ADENosine, GUANosine → pyr get -dine while pur get -sine

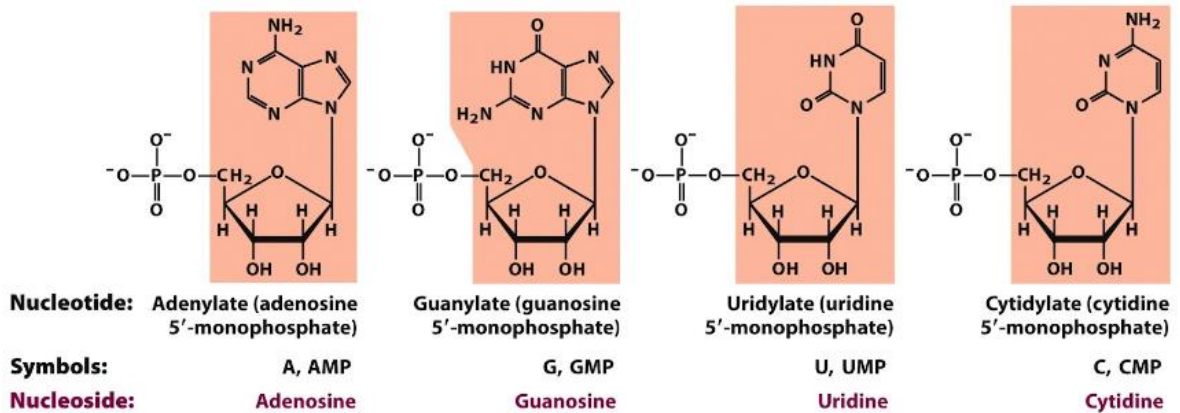
Nucleotide

- 5'C Nucleoside + esterification bond + phosphate
- DNA nuc = deoxyribonucleotides; RNA nuc = ribonucleotides

1) Deoxyribonuc Types:



2) Ribonuc Types:



3) Other Nuc Types

- Used as epigenetic marker (pro: mark own DNA to degrade foreign; eu: mark which genes should be active)
- DNA Only
 - a) 5-Methylcytidine: found in eu/ bac
 - b) N6-Methyladenosine: found in only bac
- RNA Only
 - a) Inosine: (adenosine w/o amine) gives rich gen code and found in wobble pos in anticodon
 - b) Pseudouridine: helps fold rRNA, stabilize tRNA, found more in eu than eubac

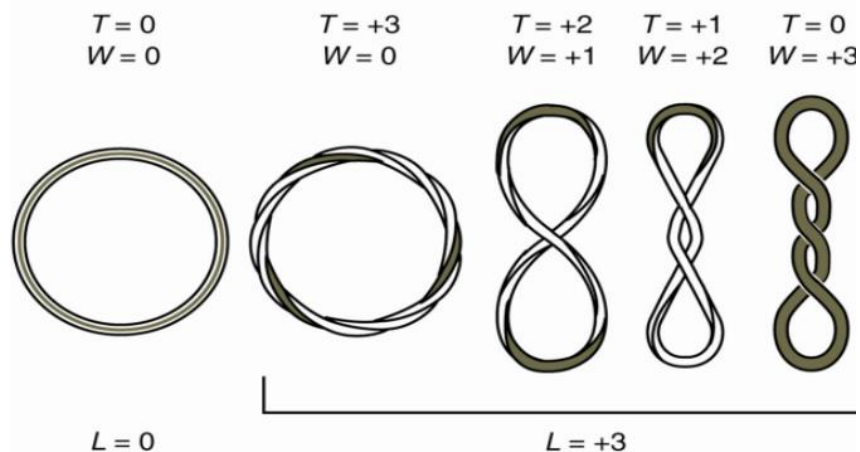
Nucleoside 5' Triphosphates

- Types: ATP (energy), CTP (lipid syn), UTP (carb met), GTP (prot syn)

- Function: E carriers; E stored in phosphoric bonds (explains E tranf in ADP to ATP)
- Major bchem rxn of nuc: pyro/phosphoryl group transf (ADP \rightarrow ATP, GDP \rightarrow GTP, etc)
- Cyclic nucleotides: sig molec/ regulators of cell met of all cells (cAMP, cGMP etc)
- Bases make nuc unique and can be used as recognition units; not involved in the biochem of met

Nucleic Acids

- Cov bond via phosphodiester link 3'C \rightarrow 5'C
 - 1) Primary Struc \rightarrow Sequencing
 - a) Chain Termination Method (F. Sanger)
 - DNA pol adds artificial primer and adds nuc to ssDNA
 - Last one will be dideoxynuc to terminate strand
 - Similar to PCR
 - b) Base-Specific Chemical Cleavage (Maxam & Gilbert)
 - Using chem agent, cleave strand at certain places of ssDNA with P-32 at end
 - Produces fragments that are read in electrophoresis based on ACTG
 - 2) Secondary Struc: reassociation is inversely prop to genome complexity (the more complex the organism, the less time it would take to anneal after denaturing)
 - 3) Tertiary Struc
 - Duplex DNA (dsDNAO = 10bp / turn)
 - Linking Number (L) = Twist (T) + Writhe (W).
 - Twist = helical turns, writhe = # times double helix crosses over itself
- Ex of pos supercoiling:

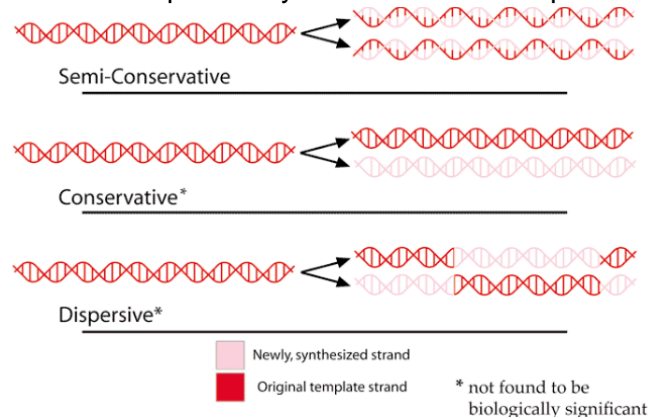


- DNA gyrase creates neg supercoiling so easy unwind \rightarrow A-subunit of gyrase cuts dna, holds ends, and ligates to form new supercoil

DNA Replication

Models

- Semiconserv model proved by Meselson/ Stahl exp



Elongation

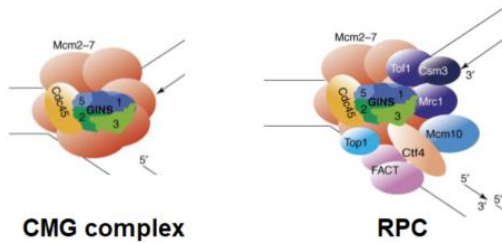
- Primase lays primer, gyrase undoes supercoil, helicase unwinds, SSB prot attached
- DNA pol 3 adds new nuc: dNTP w/ 3 PO₄ undergoes nuc attack on 3'OH to rid 2 PO₄
 - Pol 3 subunits: alpha (polymerase), epsilon (3' exonuclease), theta (holoenzyme assembly)
 - Beta-subunit acts as a sliding clamp holds all subunits together
- Pol 1 (5' exonuclease) in pro only and is used for repair and primer removal
- DNA ligase glues 2gthr nicks made by pol 1 via phosphodiester linkage

Replication Experiments

- Rep is slow (50 nuc/sec) and activates in clusters 30k-300k bp apart (not all at same t) to create rep bubble pairs
- High condensed rep late, low rep early
- Exp to identify rep origins in yeast → yeast DNA seq called ARS (autonomously replicating sequence)

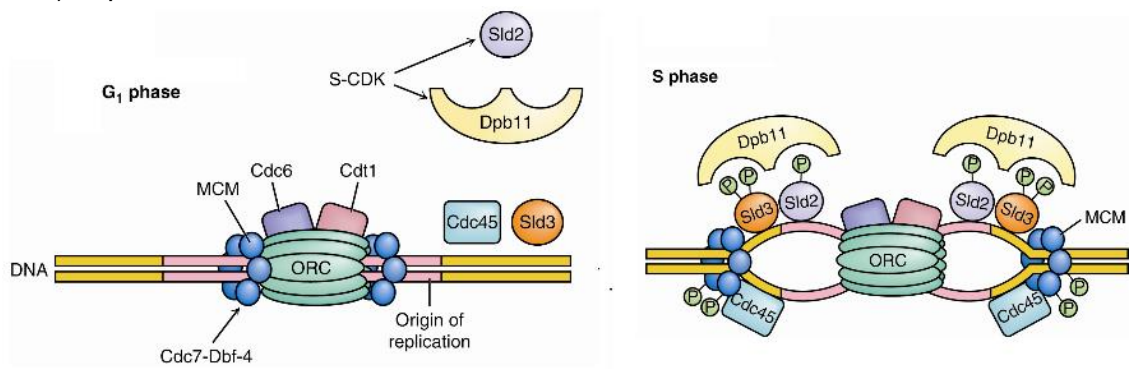
Origin Replication Complex

- Pre-replication complex (pre-RC) assembled via addition of Cdc6, Cdt1, MCM and ends in post-RC state
- *At rep fork of yeast*, Cdc45 + MCM + GINS form CMG complex
- CMG complex + other factors = RPC (replisome progression complex)



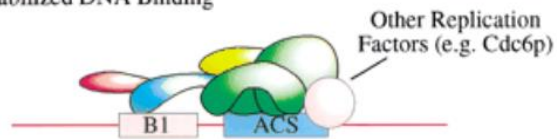
- Overall Process:

- ORC binds to origin and recruits Cdc6, MCM (rep licensing factor), and Cdt1
- Ppl (phosphorylation) mediated by S-CDK
- Cdc7-Dbf4 trigger the switch from G1 to S phase
- Ppl of Sld2 and Sld3 leads to recruit pol to origins
- Ppl of MCM serve as helicase activ



- Mechanisms that may control ORC chrom binding sites:

A. Stabilized DNA Binding



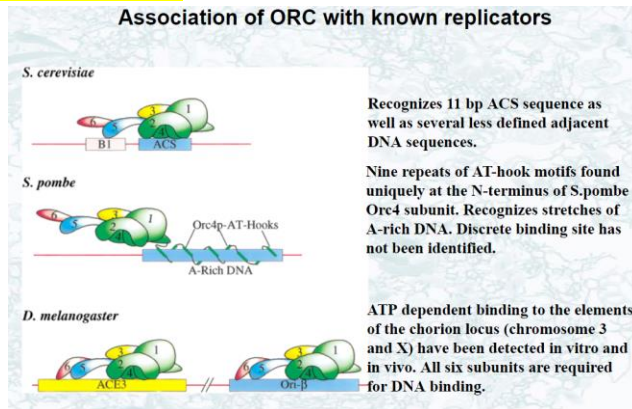
B. Recruitment



C. Local Chromatin Structure

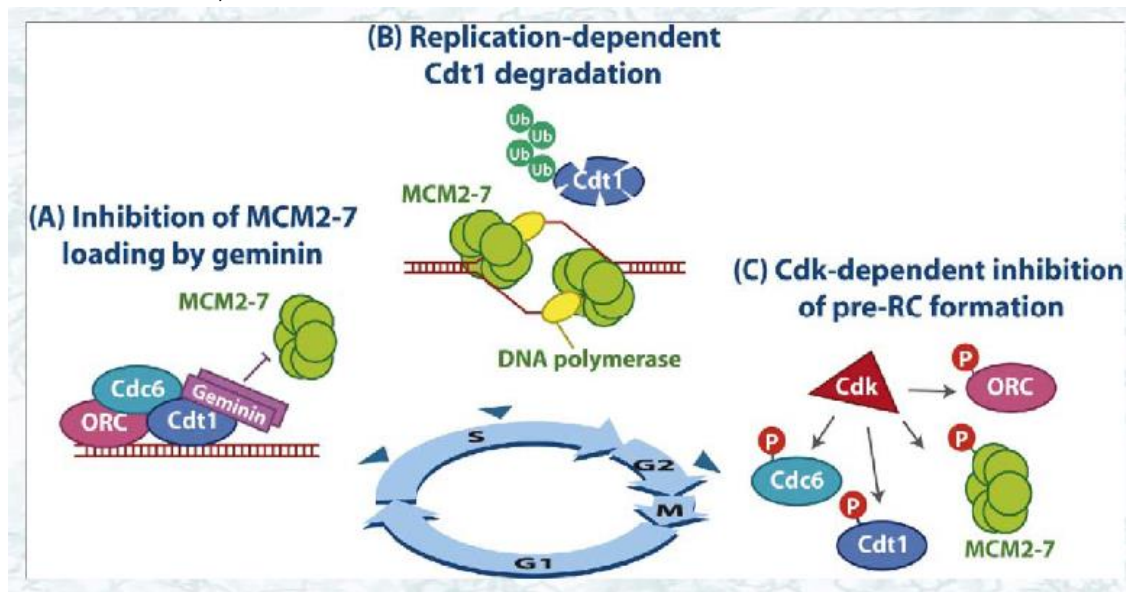


- Rev this slide



Re-Replication Prevention

- Prevention by pre-RC Formation:
 - a) Inhibitory complex between Cdt1 and geminin
 - Geminin is dna inhibitor that appears in S phase and disappears in M phase)
 - Geminin prevents MCM from forming in pre-RC
 - b) Cdt1 degradation in S phase by Skp2
 - c) High activity of cdk in G2/ M phases inactivates and degrades Cdc6, Cdt1, MCM2-7, and ORC



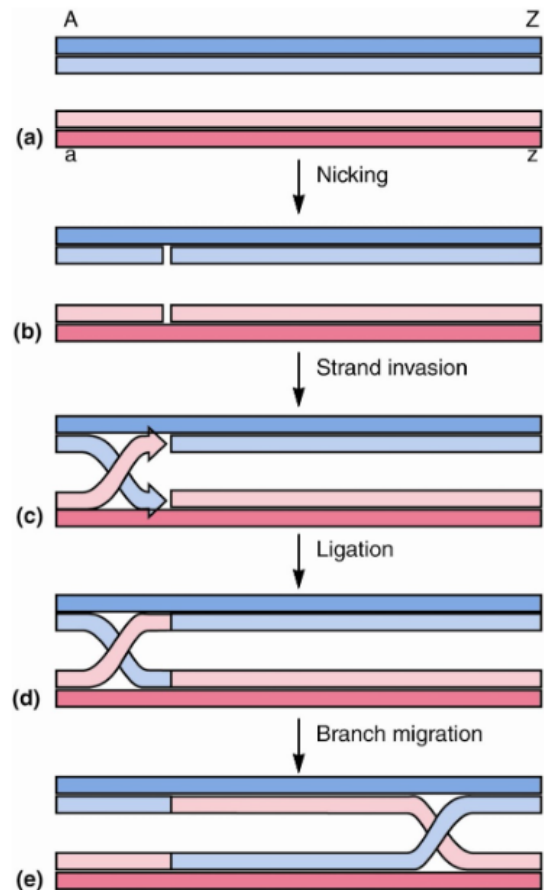
- Prevention by CDK-Mediation:
 - a) Ppl of Cdc6 and Cdt1 renders them unstable/ degrades them
 - b) MCM mod by cdk inactivates it
 - c) ORC ppl inhibits rep

DNA Mutation, Repair, and Recombination

Homologous Recombination

The Holliday model for homologous recombination.

- (A) Two homologous DNA duplexes are aligned – **synapsis**.
- (B) Recombination begins with the introduction of **single-stranded nicks** at homologous sites on two chromosomes
- (C) **Strand invasion** occurs through partial unwinding and base-pairing with the intact strand in the other duplex
- (D) Free ends from different duplexes are ligated resulting in **cross-stranded intermediate** – Holliday junction
- (E) **Branches can migrate** by unwinding and rewinding of two duplexes



- Bacterial
 - RecBCD: initiates recombination, helicase (D)/ nuclease (B) active, cuts chi site
 - RecA: strand invasion and homologous pairing
- Eukaryotic
 - RuvA tetramer: helicase, branch migration, fits into Holliday junction
 - RuvB hexameric rings: helicase, branch migration, assemble on DNA sides and act as motor
 - RuvC resolvase: nuclease active, branch migration

Transposons

- Class I (retrotransposons)
 - copy/paste
 - DNA → RNA → DNA via reverse transcriptase
 - 40% of human genome
- Class II (DNA transposons)

- cut/paste
- No RNA intermediate
- Uses many transposon enzymes

DNA Repairs

- 150 genes involved

1) Double strand break (ds DNA)

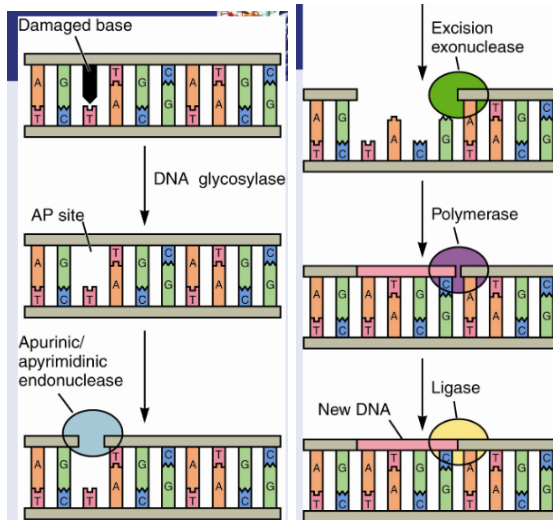
- Nonhom DNA: break in dsDNA (looks like =) → Ku 70/80 holds ends → kinase, enzymes, and ligase fix break
- Homo: break in dsDNA (looks like __--) → D-loop like RecA in homo recomb → either noncross or crossover recomb

2) Direct reversal

3) Mismatch repair (ss DNA)

4) Base excision repair (ss DNA):

- Dna glycosylases remove damaged base (creating an **apurinic/apyrimidinic or "AP" site**)
- AP endonuclease cleaves backbone
- Exonuclease removes AP site/ several nuc
- Exp repaired by DNA pol/ ligase



5) Nuc excision repair (ss DNA):

- Used instead of base excision if repair region too large
- Lesion can be like pyr dimer (UV light bonds nuc, so use light to break them)
- Uses exinucleases which break backbone in 2 places
 - **Bacteria: ABC exinuclease (UvrA, UvrB, UvrC) hydrolyzes 5th bond on 3' and 8th bond on 5' and removes ~12 nuc**
 - **Eu: exinuclease hydrolyzes 6th bond on 3' and 22nd bond on 5 and removes ~28 nuc**