

Random loops model can explain the appearance of Topologically Associating Domains (TADs)

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Introduction

- 1 The spatio-temporal organization of a chromosome has significant implication to cellular activities, such as gene expression and regulation.
- 2 However, the spatial organization of the chromatin is not entirely known.
- 3 It is known that packing of the chromatin allows for long range gene regulation.
- 4 Furthermore, DNA looping has shown to be a mechanism for long range gene regulation.
- 5 Here we show that using random polymer looping model and fitting it to the experimental chromatin looping data we can explain the appearance of conserved structures in chromatin called TADs.
- 6 Using this model, we will be able to say 'something' about the mean structure.

Agenda

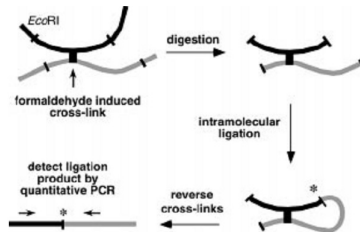
- 1 Introduction
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 - The encounter probability
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Chromosome Conformation Capture Experiments

A set of methods to simultaneously record millions of looping events occurring within the genome (specific or unspecific).

The general steps are:

- 1 intact nuclei are extracted from millions of cells
- 2 Formaldehyde induces protein-DNA and protein-protein cross-links
- 3 restriction enzymes digest the cross-linked DNA
- 4 cross-linked DNA is purified, diluted and ligated
- 5 cross-links are reversed
- 6 PCR to amplify ligation junctions
- 7 histogram of segment encounter is produced



The experimental data

- Two replicate of the CC experiments were conducted by Nora et. al 2012.
- Cells taken from undifferentiated mouse embryonic stem cells.
- We focus on a 920,432 bp subset of the data, around the X inactivation center of the X chromosome.
- The region harbors the Xist enhancer and Tisx promoter.
- We have the segments' encounter frequencies from the two experimental replicates.

Topologically Associating Domains (TADs)

Conserved structures showing chromosome interactions on the Mb scale, with higher intra than inter-region interactions. It is believed that the TAD forms a 'regulatory unit' for regulating gene expression, as can be seen by the correlation of gene expression located on the same TAD.

However, it is not yet clear what is its role in a single cell.

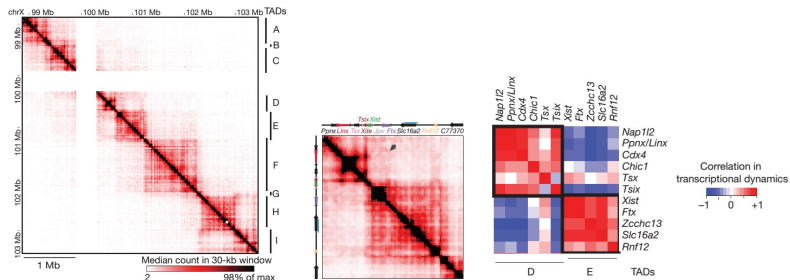
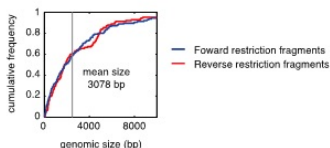


Figure: The 4.5 Mb region (left), enlargement of TAD D and E (center). Displayed median count in a 30kb window every 6kb, gene expression correlation (right)

From restriction segments to beads

- To coarse-grain the data, we choose a bead-size of 3000 bp, corresponding to the mean segment length resulted from the digestion of EcoRIII enzyme.



- the genomic section was evenly partitioned. Each bead receives a start and end index according to the segment it covers.

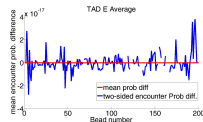
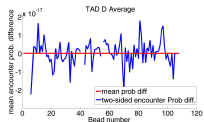
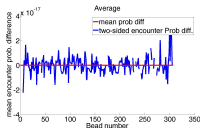
- for example,

bp range	start ind	end ind
500-3500	1	2
4000-4500	2	2
5000-12001	2	4

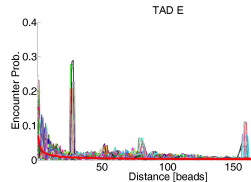
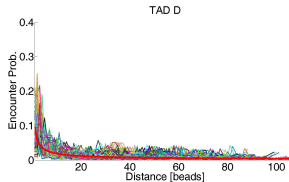
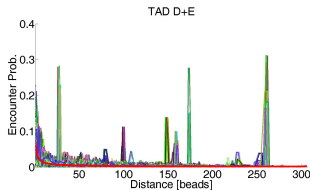
Bead encounter frequency

TAD D and E

- We work with the average of the two experimental replicates.
- A total length of 920,432 bp - resulting in 307 beads (TAD D 107 beads, TAD E 200 beads)
- We calculate the 'one-sided' encounter probability vs. distance (bead units) for each bead
- the mean encounter probability difference, shows that the data is left-right symmetric



- TAD E has several strong specific interactions. TAD D has almost no specific interactions. Strong inter-TAD specific interactions



Peaks of the encounter data

- About half of the peaks in the encounter data result from specific interactions **between TADs**.
- The other half comes from specific internal interactions of **TAD E**.
- To get an impression, a manual marking of the peaks shows:

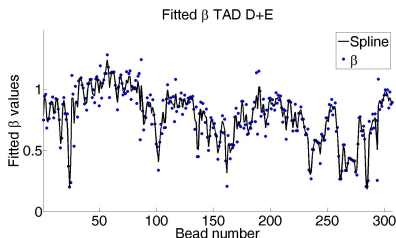
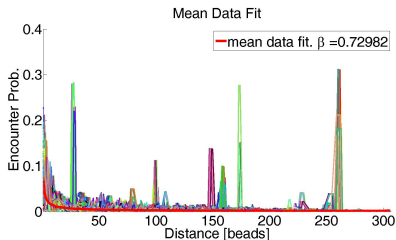
Bead numbers	Encountered beads	TAD
23-26	280-290	<i>D ↔ E</i>
49-53	148-155	<i>D ↔ E</i>
56-59	80-90	<i>D ↔ D</i>
115-117	165-170	<i>E ↔ E</i>
161-162	187-190	<i>E ↔ E</i>
182-184	260-264	<i>E ↔ E</i>
185-186	253-255	<i>E ↔ E</i>
234-236	184-189	<i>E ↔ E</i>
234-236	4-11	<i>E ↔ D</i>
243	88	<i>E ↔ D</i>
264	89-90	<i>E ↔ D</i>
274-277	113-120	<i>E ↔ E</i>

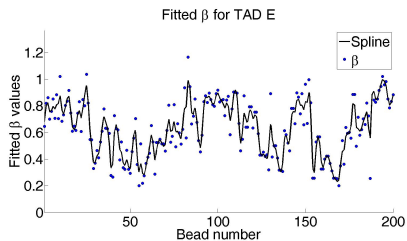
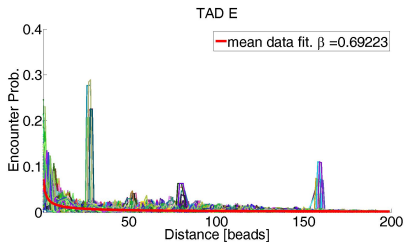
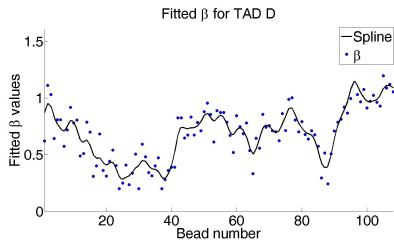
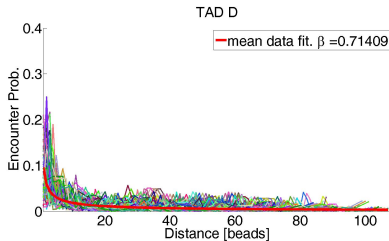
The encounter probability

For the case of TAD D, TAD E, and the two together, we estimate the bead encounter probability, p , and fit it with a function of the form

$$p_n(d) = \alpha d^{-\beta}$$

where, d is the distance in [bead] units, n is the bead number, $\alpha = \frac{1}{\sum_{j=1}^{d_{max}} j^{-\beta}}$ and β is a parameter to be estimated. We report the values of β for each bead in each case





Theoretical model

The Rouse model

We start with the classical and most simple model, the Rouse chain.

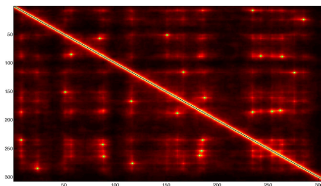
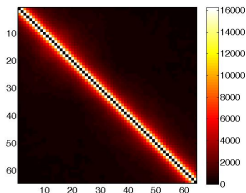
- A Rouse chain describes polymer dynamics as a stochastic motion of a collection of microscopic "beads" connected by harmonic springs
- the 3D motion of bead n in the chain of N beads

$$\frac{dR_n}{dt} = -\frac{3D}{b^2}(2R_n(t) - R_{n+1}(t) - R_{n-1}(t)) + f_n(t)$$

- R_n - the position of bead n
 b - the standard deviation of the distance between adjacent beads
 D - the diffusion constant
 f_n - white Gaussian noise
- From the theory, $Pr(\|R_n - R_m\| < \epsilon) \sim |n - m|^{-1.5}$

Simulation with simple rouse chain

- A simple Rouse model cannot reproduce the TAD, as expected.
- placing loops corresponding to the peaks of the encounter data does not reproduce the TADs.



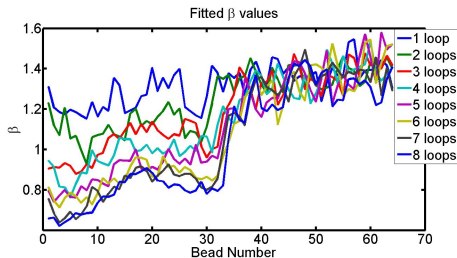
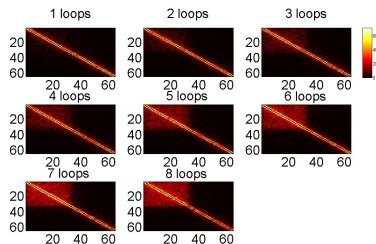
Random Fixed Loop Model

- The enhancer-transcription factor elements' encounter motivates the simulation of a chain with randomly placed loops
- these encounters might not be as frequent as 'stable' loops in the chromatin and therefore not shown significantly in the encounter maps
- simulate a chain of 64 beads, having a random loop in a bounded genomic region
- increasing number of loops at random position is simulated

Random loops in a bounded region

One TAD

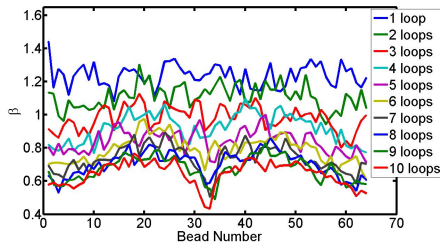
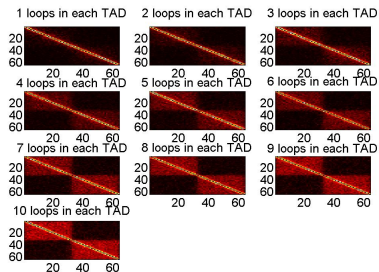
We examine the behavior of the encounter probability when we restrict the loops to be in one region of the polymer, increasing the number of loops from 1 to 8.



Random loops in a bounded region

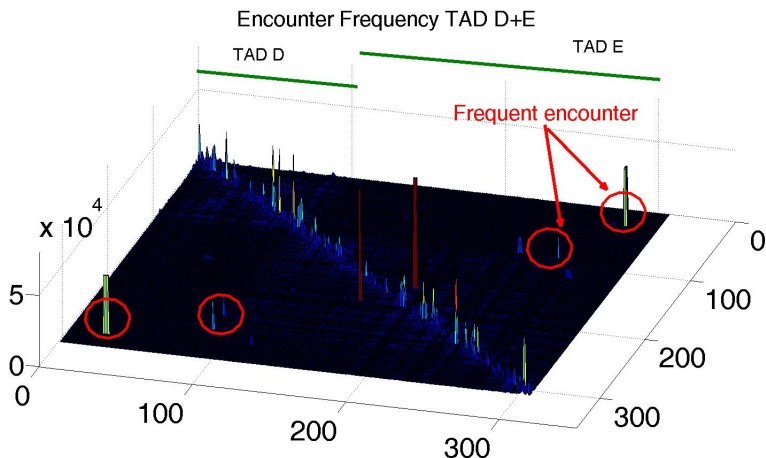
Two TADs

The same can be done with two bounded regions, where the number of loops is increased from 1 to 10 in a 64 beads chain.



Peaks at the edges of TADs

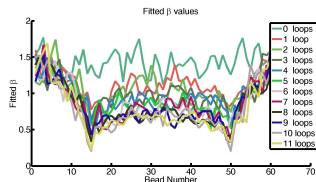
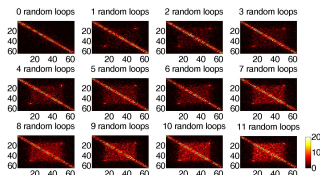
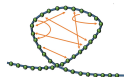
At the edges of TADs there are significant peaks of the encounter frequencies. The encounter data of Nora et al. was smoothed using a median filter. Therefore, those peaks were not clearly shown. Such peaks might indicate a stable loop in the structure of the genomic region and should be taken into account.



Stable loop with random loops within

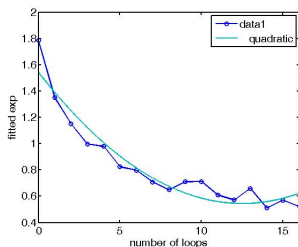
one TAD with 'tails'

- 1 Following the peaks observation, we form a mix of 'stable' big loop with internal random loops
- 2 in a 64 beads chain, beads 15 and 50 were connected
- 3 we iteratively add 11 loops within the stable loop
- 4 we can start seeing the emergence of β curve pattern as in the experimental data



β as a function of the number of loops

- 1 Through simulation we observe a quadratic relationship between the mean fitted β value and the number of random loops.
- 2 This enables us to say something about the mean structure.
- 3 The mean number of loops in a given genomic region can be extracted from experimental data.



Literature support for random enhancer promoter interactions

- 1 Job Dekker, Marc A Marti-Renom, and Leonid A Mirny. Exploring the three-dimensional organization of genomes: interpreting chromatin interaction data. *Nature Reviews Genetics*, 14(6):390403, 2013
- 2 Job Dekker, Karsten Rippe, Martijn Dekker, and Nancy Kleckner. Capturing chromosome conformation. *science*, 295(5558):13061311, 2002.
- 3 Josee Dostie, Todd A Richmond, Ramy A Arnaout, Rebecca R Selzer, William L Lee, Tracey A Honan, Eric D Rubio, Anton Krumm, Justin Lamb, Chad Nusbaum, et al. Chromosome conformation capture carbon copy (5c): a massively parallel solution for mapping interactions between genomic elements. *Genome research*, 16(10):12991309, 2006
- 4 Peter Fraser and Wendy Bickmore. Nuclear organization of the genome and the potential for gene regulation. *Nature*, 447(7143):413417, 2007
- 5 Johan H Gibcus and Job Dekker. The hierarchy of the 3d genome. *Molecular cell*, 49(5):773782, 2013
- 6 Suchit Jhunjhunwala, Menno C van Zelm, Mandy M Peak, Steve Cutchin, Roy Riblet, Jacques JM van Dongen, Frank G Grosveld, Tobias A Knoch, and Cornelis Murre. The 3d structure of the immunoglobulin heavy-chain locus: implications for long-range genomic interactions. *Cell*, 133(2):265279, 2008
- 7 Erez Lieberman-Aiden, Nynke L van Berkum, Louise Williams, Maxim Imakaev, Tobias Ragoczy, Agnes Telling, Ido Amit, Bryan R Lajoie, Peter J Sabo, Michael O Dorschner, et al. Comprehensive mapping of long-range interactions reveals folding principles of the human genome. *science*, 326(5950):289293, 2009.
- 8 Erez Lieberman-Aiden, Nynke L van Berkum, Louise Williams, Maxim Imakaev, Tobias Ragoczy, Agnes Telling, Ido Amit, Bryan R Lajoie, Peter J Sabo, Michael O Dorschner, et al. Comprehensive mapping of long-range interactions reveals folding principles of the human genome. *science*, 326(5950):289293, 2009.
- 9 Erez Lieberman-Aiden, Nynke L van Berkum, Louise Williams, Maxim Imakaev, Tobias Ragoczy, Agnes Telling, Ido Amit, Bryan R Lajoie, Peter J Sabo, Michael O Dorschner, et al. Comprehensive mapping of long-range interactions reveals folding principles of the human genome. *science*, 326(5950):289293, 2009.

etc..

Summary

- We sowed a plausible and simple model to explain the appearance of the conserved TAD domains
- A relationship between the mean number of loops and the encounter probability was shown by simulations
- The parameter used for our analysis, namely β , can easily calculated by experimentalists.

- ① Calculation of the mean first encounter time between beads in the Rouse ring
- ② Showing analytically the relationship between number of loops and decrease in β
- ③ Put in more rigorous form the relationship between the pattern of *beta* and the mean observed chromatin structure.
- ④ Calculating the equilibrium distribution of the random loop model
- ⑤ Simulating a model with random dynamic loops, in which loops can form and dissociate.