

# **Detecting Polycystic Ovary Syndrome (PCOS) Using ML Techniques**

## **Machine Learning Paper Implementation Report**

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**Prof. Priyanka Verma**

# ABSTRACT

The present world women population is widely affected by preterm abortions, infertility, anovulation etc. It is observed that polycystic ovary syndrome (PCOS), a condition seen among the women of reproductive age, is having a major influence in the cause of infertility. Over five million women worldwide in their reproductive age PCOS. It is an endocrine disorder characterized by changes in the female hormone levels and the abnormal production of male hormones. This condition leads to ovarian dysfunction with increased risk of miscarriage and infertility. The symptoms of PCOS include obesity, irregular menstrual cycle, and excessive production of male hormone, acne, and hirsutism. It is extremely difficult to diagnose PCOS due to the heterogeneity of symptoms associated and the presence of a varying number of associated gynecological disorders. The time and cost involved in innumerable clinical tests and ovary scanning has become a burden to the patients with PCOS. To address this problem this paper proposes a system for the early detection and prediction of PCOS from optimal and minimal but promising clinical and metabolic parameters, which act as an early marker for this disease. The data sets required for this system development are obtained through patient surveys of 541 women during doctor consultations and clinical examinations. Out of the 23 features from clinical and metabolic test results, 8 potential features are identified using SPSS V 22.0 based on their significance. Classification of PCOS with the feature set transformed with Principal Component Analysis (PCA) is done using various machine learning techniques such as Naïve Bayes classifier method, logistic regression, K-Nearest neighbor (KNN), Random Forest Classifier, Support Vector Machine (SVM) in Python. Results revealed that the most suitable and accurate method for the PCOS prediction is RFC with an accuracy of 89.02%.

**Keywords**—Machine learning, polycystic ovary syndrome, Classifier, Diagnostic aid

# i-HOPE: Detection And Prediction System For Polycystic Ovary Syndrome (PCOS) Using Machine Learning Techniques

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**Abstract**—The present world women population is widely affected by preterm abortions, infertility, anovulation etc. It is observed that polycystic ovary syndrome (PCOS), a condition seen among the women of reproductive age is having a major influence in the cause of infertility. Over five million women worldwide in their reproductive age PCOS. It is an endocrine disorder characterized by changes in the female hormone levels and the abnormal production of male hormones. This condition leads to ovarian dysfunction with increased risk of miscarriage and infertility. The symptoms of PCOS include obesity, irregular menstrual cycle, and excessive production of male hormone, acne, and hirsutism. It is extremely difficult to diagnose PCOS due to the heterogeneity of symptoms associated and the presence of a varying number of associated gynecological disorders. The time and cost involved in innumerable clinical tests and ovary scanning has become a burden to the patients with PCOS. To address this problem this paper proposes system for the early detection and prediction of PCOS from an optimal and minimal but promising clinical and metabolic parameters, which act as an early marker for this disease. The data sets required for this system development are obtained through patient survey of 541 women during doctor consultations and clinical examinations. Out of the 23 features from clinical and metabolic test results, 8 potential features are identified using SPSS V 22.0 based on their significance. Classification of PCOS with the feature set transformed with Principal Component Analysis (PCA) is done using various machine learning techniques such as Naïve Bayes classifier method, logistic regression, K-Nearest neighbor (KNN), Classification and Regression Trees (CART), Random Forest Classifier, Support Vector Machine (SVM) in Spyder Python IDE. Results revealed that the most suitable and accurate method for the PCOS prediction is RFC with an accuracy of 89.02%.

**Keywords**—Machine learning, polycystic ovary syndrome, Classifier, Diagnostic aid.

## I. INTRODUCTION

Technology and mankind together hand in hand can make way towards better health care and services. Machine learning is a subset of artificial intelligence, in which it provides the system with the ability to automatically learn and improve without being programmed explicitly. It mainly focuses on

developing algorithms that can access the datasets provided and use data for the learning purposes of the network. Applications of Machine Learning bring about huge transformation in the health industry, which includes detection, data prediction, image recognition etc.

Polycystic ovary syndrome (PCOS), is one of the relevant, most prevalent hormonal disorder seen among the women of childbearing age. This is a heterogeneous endocrine disorder which is highly prone to infertility, anovulation, cardiovascular disease, type 2 diabetes, obesity etc. PCOS is a common condition detected in nearly 12-21% of women of reproductive age and among them 70% is remain undiagnosed. PCOS condition can be treated to some extent by controlled medication and bringing alterations in life style. This includes the treatment methods with pills for birth control, diabetes, fertility, anti-androgen medicines and scanning procedures like ultrasound scan. When such interventions fail, invasive treatment procedures like surgical drilling of ovaries is also used for improving the ovulation ability of the ovary by reducing the male hormone level.

The aetiology of PCOS is underpinned by both insulin resistance and hyperandrogenism. Clinically it is characterized by reproductive, metabolic and psychological features and represents a major health burden to women. Diagnosis is recommended based on clinical or biochemical and radiological test results. PCOS is diagnosed by exclusion of irrelevant symptoms or test results, mainly because of lack of knowledge of its complex patho-mechanism. The diverse symptoms of this condition force medical practitioners to call for large number of clinical test results and unnecessary radiological imaging procedures. The early detection and diagnosis of PCOS with minimal tests and imaging procedures is of utmost importance and of great significance as the condition directly leads to ovarian dysfunction with an increased risk of miscarriage, infertility or even gynaecological cancer and mental agony for the patients due to wastage of time and money.

## II. LITERATURE SURVEY

Among the in-numerous problems that exist around us, the problems that are related to the reproductive health of women

was selected as an area of our interest, due to its importance in this contemporary society. A detailed survey of studies on PCOS and systems to support its diagnosis was carried out. Literature says that about 5-10% of Indian women in reproductive age are affected by the multifaceted endocrine disorder called Polycystic Ovary Syndrome (PCOS) [16]. It is a major cause of anovulatory infertility and increases the risk for insulin resistance, obesity, cardiovascular disease and psychosocial disorders [17]. The symptoms for PCOS might be varying from patient to patient. Some of them are irregularity in menstrual periods, acne, overweight, increased tendency for infertility, intense hair fall, balding of front head, increased facial hair growth [1]. Traditionally the PCOS can be suspected when number of follicles in an ovary is more than 12 per unit area and visible in radiological scan [15]. Some authors have proposed changing the cutoff from 12 follicles to 20 or abandoning ultrasound altogether in favor of other biomarkers, such as serum anti-Mullerian hormone (AMH) [1,10]. The diagnosis of PCOS is uncomplicated, requiring only the careful application of a few well-standardized diagnostic methods. PCOS diagnosis is often delayed and this affects patients' well-being negatively [21]. Escobar *et al.* [11] suggests that treatment should be symptom-oriented, long term and dynamic and adapted to the changing circumstances, personal needs and expectations of the individual patient. Joham *et al.* in [6] considered the relation of PCOS and infertility rate of women in this community and use of fertility hormone treatment was significantly higher in women reporting PCOS. Considering the prevalence of PCOS and the health and economic burden of infertility, strategies to optimize diagnosis of PCOS and the factors leading to fertility are important. This is because infertility is reported to be 15-fold higher in women reporting PCOS, independent of BMI [18]. There is a bi-directional relationship between obesity and PCOS. Both exacerbate each other in a never-ending cyclical manner. Essah, P.A. and Nestler, J.E suggests that the prevalence of obesity in PCOS women is 30–75% [19]. Clinical validation of PCOS is usually done by Rotterdam criteria [8] or standards set by societies involved in PCOS

research. Pictorial depiction of three popular criteria can be seen in Table I.

TABLE I. CRITERIA FOR DIAGNOSIS OF PCOS

Clinical Finding	National Institutes of Health criteria, 1990 ( Must have both of the findings marked below)	Rotterdam Criteria, 2003(must have any two of the findings marked below)	Androgen Excess and PCOS Society, 2009(must have A plus either B or C)
<i>Hyperandrogenism*</i>	X	X	A
<i>Oligomenorrhea</i>	X	X	B
<i>Polycystic ovaries</i>		X	C

\* Clinical or biochemical evidence of excess androgen.

A cross sectional study by Brower *et al.* [14] suggested that the presence of clinically evident menstrual dysfunction can be used to predict the presence and possibly the degree of insulin resistance in women with PCOS. Many of the technical studies carried out in PCOS diagnosis are using features of the ultrasound scan and image processing techniques for the diagnosis of the PCOS [1-2]. Some studies used clinical and metabolic features of the disease [3]. Few recent studies are diverted in fundamental research direction, investigating the associated factors such as obesity [4] and genetic factors [5]. A summary of such studies are given in Table II. Studies are also carried out in directions of analyzing urinary steroid hormone metabolites and enzyme activities in women with and without PCOS in order to test their value for diagnosing PCOS [20].

TABLE II. SUMMARY OF FEW STUDIES ADDRESSING THE ISSUE OF PCOS

Authors	Technique used	Objective of the study	Year
Cheng et al. [1]	Rule based classifier and Gradient boosted Tree classifiers	PCOS determination from Ultrasound Images	2016
Dewi, R.M. and Wisesty [2]	Gabor wavelet based feature extraction and CNN	PCOS determination from Ultrasound Images.	2018
Mehrotra et al. [3]	2 sample-test, Bayesian and Logistic Regression (LR) classifier	PCOS determination from clinical and metabolic parameters.	2011
Sachdev et al. [4]	Prospective observational study : Obese vs non-obese PCOS	Obese PCOS patients have a higher risk of adverse outcomes.	2019
Zhang et al. [5]	Machine-learning algorithms	PCOS prediction from identification of new PCOS genes.	2019
Joham et al. [6]	Logistic regression on data from cross-sectional analysis of a longitudinal cohort	Examination of factors associated with infertility and use of fertility treatment.	2015
K. Meena, M. Manimekalai, and S. Rethinavalli [7]	Information Gain Subset Evaluation and Neuro Fuzzy methods of feature selection and Decision Tree classifier	Framework for Filtering the PCOS Attributes	2015

### III. METHODOLOGY

For the development of an appropriate machine learning model based diagnostic aid for PCOS, a comparison of performance of various existing algorithms in our data set need to be presented. Preparation of the model is the most crucial step that provides the outline of the research. Steps that are included in the development of an appropriate model and tuning it for obtaining possibly the best result, is detailed below with the help of a work flow diagram, Figure 1. Along with that the effective tools and available platforms utilized for the development of the system must be mentioned. The following section describe both aspects.

#### A. Defining problem

The most important step is to define the problem appropriately including the inputs provided into the model and the output expected out of it. It is based on the assumptions like the outputs can be predicted from the inputs provided.

#### B. Data collection

The critical step that will decide about how good the model will be and also as the number of data collected increases the accuracy of the model also increases and hence better will be its performance. There are many ways for data gathering like real time data gathering or from repository platforms like kaggle and UCI machine learning repository which is one of the most frequently visited one.

#### C. Selection of implementation platform

Tools for the efficient running of machine learning methods and also platforms for statistical analysis should be properly opted. For this research, Syder python for model formation, HTML with SQL for designing a proper user interface, whereby the patient data can be input to the system and PCOS status can be obtained as output, and SPSS V22.0 for establishing the relevance of features, are used.

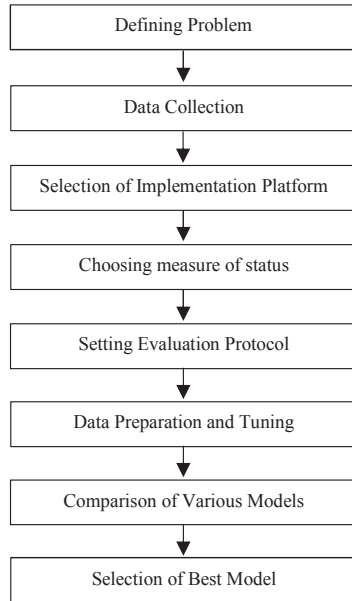


Fig. 1. Workflow of machine learning model

#### D. Choosing measure of status

In case classification problems, success is measured using the calculation of accuracy and precision of the model. In this study we have considered the following evaluation metrics:

- Accuracy :  $\frac{(TP+TN)}{(TP+TN+FP+FN)}$
- Precision (P) :  $\frac{(TP)}{(TP+FP)}$
- Sensitivity / Recall (R) :  $\frac{(TP)}{(TP+FN)}$
- Specificity :  $\frac{(TN)}{(TN+FP)}$
- F1 score :  $2 * \left( \frac{P * R}{P + R} \right)$

where TP, TN, FP, FN implies True Positive, True Negative, False Positive, False Negative respectively.

#### E. Setting validation protocol

Maintaining a hold out validation set, i.e., in this method some portion of the data is set apart for the purpose of testing as test data and remaining as train data.

Usually the data is split in the ratio 8:2 as train data to test data. It can be depicted as in fig.2:

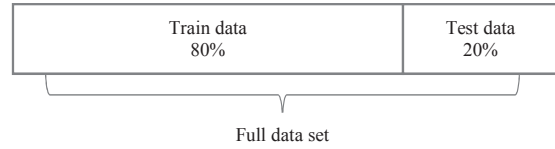


Fig. 2. Depiction of Hold-out validation set

#### F. Data preparation

This tiring process includes dealing with missing data, handling categorical data, feature scaling and selection of meaningful features. The missing values in the dataset is replaced by 'NaN'. Due to a model's inability to read a missing value, before confronting the model, the samples with missing values will be extirpated or else will be replaced with some pre-built estimators. Likewise before feeding the data into the model, ordinal and nominal data need to be considered accordingly. If the nature of the data demands, the dataset should undergo normalization and standardization. Finally, the overfitting can be avoided by reducing the dimensionality of data. This is done by reducing the number of feature sets present in the dataset. It is performed using Principal Component Analysis (PCA) in Spyder Python IDE, which works by identifying the patterns in the datasets and the correlations present between the features. The correlated data are then eliminated by directly removing such features. The optimal features identified by the PCA algorithm are verified for their potential in discriminating PCOS status, with SPSS V22.0. This is done by a procedure called independent sample t-test and the level of statistical significance across the two classes of PCOS and Non PCOS patients. Those features with a significance less than 0.01 are potential ones.

#### G. Comparison of various models

This step is to serve as a baseline. Study is carried out with selected set of features in a number of classifier algorithms. Among the existing innumerable machine learning algorithms some of them, which are proven to give best result in the



detection of PCOS and Non-PCOS condition from the literature survey, is used and listed below:

- **Logistic Regression (LR)**
- Linear Discriminant Analysis (LDA)
- K-nearest neighbors (KNN)
- Classification and Regression Trees (CART)
- Random Forest Classifier
- **Naïve Bayes Classifier**
- Support Vector Machine

#### IV. DESIGN AND DEVELOPMENT

Data acquired from various hospitals and clinics which includes both physiological and metabolic parameters that are the contributors towards PCOS and thereby infertility, are fed into the mathematical framework of i-HOPE. The proposed system of diagnostic aid is illustrated Figure 3.

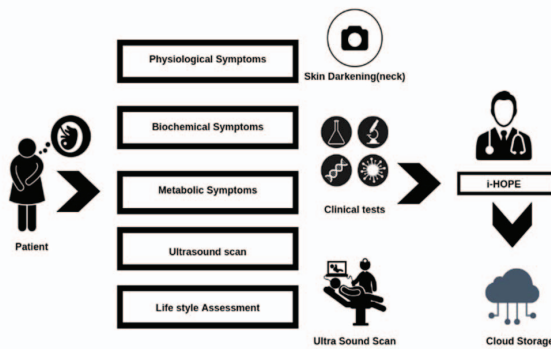


Fig. 3. Proposed model of i-HOPE

##### A. Data Collection :

More the number of features and samples, more distinguishing and accurate the model be. In this research we have considered a total of 541 samples which were collected from various clinics and hospitals in and around the district of Thrissur. Informed consent was collected from each patient, promising the anonymity of the data collected.

##### 1) Patient History Collection :

One proforma is assigned for each patient so that all the data about a single patient will be in contained in a Proforma. These Proforma were carried along for data collection into clinics and for patient survey. Later on the collected data from the Proforma is consolidated together in to a single data sheet. The features which were included within the proforma was inferred from various literature studies and clinical surveys conducted, whose identity was kept anonymous during the entire process.

##### 2) Real-time data collection :

For the purpose of real time detection of PCOS, a patient interface is required to input the patient data. Data to be entered may include the personal data of the patient for further references and the parameters that determine the existence of PCOS. This entered data will be inputting in to the created algorithm for data processing. To create the interface, a front end is generated using HTML and the data entered in the front end is being written in to an excel sheet using SQL.

TABLE III. FINALIZED PARAMETERS

SI No	Parameters	Value
1	Age	15-35
2	BMI	<24(normal), >24(abnormal)
3	Cycle Length and regularity	Long, normal or short Regular/Irregular
4	LH : FSH Ratio	Normal/abnormal
5	Waist : Hip Ratio	Normal/abnormal
6	Weight gain	Yes(y)/No(n)
7	Excess facial or body hair	Yes(y)/No(n)
8	Dark areas on skin	Yes(y)/No(n)
9	Pimples	Yes(y)/No(n)
10	Blood Pressure	Normal/abnormal
11	Diabetes (before and after food)	Normal/abnormal
12	Fast food intake	Yes(y)/No(n)
13	Regular exercise	Yes(y)/No(n)
14	Loss of hair	Yes(y)/No(n)
15	No. of follicles (L <sup>a</sup> and R <sup>b</sup> )	High ,medium, low
16	Size of follicles (L <sup>a</sup> and R <sup>b</sup> ) (mm)	>10
17	TSH (mIU/L)	0.4-4(Normal)
18	AMH	1-4(Normal) >1(Abnormal)
19	PRL	2-29 (Non-Pregnant Females) 10-209 (Pregnant Females)
20	Vit D3	20-50 (Normal) >12 (Abnormal)
21	PRG	1.5-12.4 (Normal)

<sup>a</sup> Left ovary, <sup>b</sup> Right ovary

##### B. Parameter Selection

The proposed model i-Hope was fed with the data obtained from the survey conducted. Parameter finalization was done with the support of expert opinions and considering the contemporary researches that in a way or other affected PCOS. The features are transformed for removing correlating among themselves which might adversely affect the classification results. The finalized list of parameters before transformation are listed in Table III. These parameters include physiological, metabolic and biochemical attributes. Also considered the values of the result obtained from ultra sound scan which consists of the information about the cyst formed such as the number of cyst present in the right and left



ovary and their size. The significance of parameters were studied individually based on the independent sample test, Pearson and Spearman's rho correlation of parameters with the help of SPSS software by IBM. The correlation was proved to be significant at 0.01 level (2-tailed). Table IV shows the important features for the final design of the system and their statistical significance.

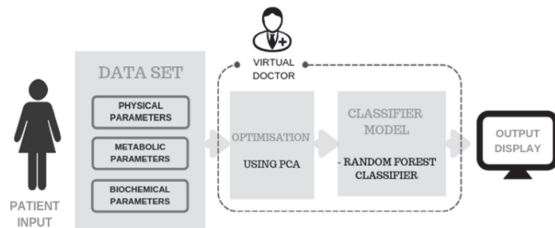


Fig. 4 The automated PCOS diagnosis aid i-Hope

### C. Moulding Model

1) *Setting model* : Through Synder python the evaluation metrics of the model is calculated. After that the evaluation protocol is implemented from the sklearn library of the python platform. The most discriminating and contributing feature set is selected by extirpating the redundant data set through implementation of Principal Component Analysis (PCA) which is a type of unsupervised learning.

2) *Model Selection* : Comparing various selected machine learning algorithms to find the best performing model using spot-check algorithm, which is a precursor for the selection of best and appropriate algorithm that suites the model. Selecting the best model with better performance by analyzing the confusion matrix of each algorithm. Among the models RFC provided the best result. Consolidating all the above noted details, the final design is improvised and modeled as shown in the Figure 4.

## V. RESULT AND DISCUSSIONS

A total of 541 cases were available for study, which was collected from various infertility treatment centers at Thrissur. The data comprised the women reproductive age group i.e., in between 18-40 years. Among the data collected 364 cases were normal and non-PCOS, the remaining 177 cases reported PCOS. Altogether there were 23 features, including the reports on transvaginal Ultrasound scan, hormone profile and

lifestyle of the patient with impressions on physical fitness and some listed in Table III. The optimal features after PCA is statistically analyzed to see their significance and are listed in Table IV.

The algorithms opted comprises a mixture of simple linear and non-linear algorithms. The simple linear algorithms are LR and LDA. Non-linear methods are KNN, CART, RFC, NB, SVM. We reset the random number seed in each run to the data split. Accuracy estimations of each of the models were carried out with hold out validation and the estimated accuracy scores of each model was obtained. Table shows the results of evaluation metrics of the research, through this the performance of the models can be analyzed. From which we can arrive at a conclusion that the best performance was given by Random Forest Classifier model, where an accuracy of 89 % was achieved after data optimization.

TABLE IV. OPTIMAL FEATURES IDENTIFIED IN SPSS

Parameters contributing towards PCOS		Parameters contributing towards infertility	
Feature	Significance	Feature	Significance
Cycle irregularity	0.000	No. of abortion	0.002
Cycle Length	0.000		
FSH,LH Ratio	0.006	Thickness of Endometrium	0.660
AMH	0.000		
Follicle no.	0.000	Vitamin D3	0.050
Follicle size	0.002		
BMI	0.000	AMH	0.002
Weight gain	0.000		

Therefore, it can be concluded that either biochemical profile alone or USG result alone can't serve as a diagnostic tool for the treating PCOS. Because both the factors that relate PCOS and infertility falls in both categories. AMH turns out to be a very promising feature to detect PCOS and infertility as per our results. Comparing the accuracies obtained for other studies in PCOS detection, 97% is the highest accuracy obtained in [1], 82% in [2], 93.9% in [3] and 90% in [5]. Our results are lesser than these, even though a direct comparison is meaningless. With optimization of weight parameters of classifiers, performance of the system might improve.

TABLE V. ACCURACY SCORE, SENSITIVITY, SPECIFICITY AND PRECISION OF VARIOUS MODELS

Algorithm Used	Accuracy score	Sensitivity	Specificity	Precision	F1 score
Logistic Regression ( LR )	0.8536	0.6451	0.98039	0.952380	0.3845
K- Nearest Neighbors (KNN )	0.8658	0.8064	0.90196	0.83333	0.4098
Classification and Regression Trees (CART)	0.8292	0.8387	0.82352	0.74285	0.3939
Random Forest Classifier (RFC)	0.8902	0.7419	0.98039	0.95833	0.4182
Gaussian Naïve Bayes (NB)	0.8414	0.7419	0.90196	0.82142	0.3898
Support Vector Machines (SVM )	0.8292	0.5483	1.0	1.0	0.3541

## VI. CONCLUSION

Polycystic Ovary Syndrome (PCOS) is one of the most common type of endocrine disorder in reproductive age women. This may result in infertility and anovulation. The diagnostic criterion includes the clinical and metabolic parameters which are biomarker for the disease. We developed a system that automates the PCOS detection based on minimal set of potential markers. Our methodology involves the formulation of a feature vector based on real time data from patients during clinical and radiological investigation while they visit a healthcare facility. The eight metabolic and Ultrasound image features identified with the PCA feature transform and statistical significance is found promising for discriminating between normal and PCOS patients. Among the various algorithms used, RF algorithm is found superior in performance. This automated system can act as an assistive tool for the doctor for saving considerable time in examining the patients and hence reducing the delay in diagnosing the risk of PCOS. Implications from the clinical expert survey for this work suggests that, innovations that uphold the medical ethics are always welcome in the field of medicine and healthcare. Researches that could bring out useful innovative methodologies like the effect of Vitamin D on PCOS, studies that put forth the impact of PCOS on preterm labor/abortions, attempt to unveil the number of lean PCOS patients etc. need to be held in future.

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**DATA  
CLEANING  
&  
PRE-  
PROCESSING**

# PCOS\_ML\_Part\_1

October 29, 2021

## 1 Data Cleaning And Data Preprocessing

This notebook is written to clean our raw dataset and for solving unwanted issues so that we can use the data well structured dataset.

## 2 Importing Necessary Modules

```
[ ]: import sys
import pandas as pd
import numpy as np
import matplotlib.pyplot as plt
%matplotlib inline
plt.style.use("ggplot")
plt.rcParams['figure.figsize'] = (12, 8)
import seaborn as sns
sns.set(style='whitegrid', color_codes=True)
import warnings
warnings.filterwarnings('ignore')
```

## 3 Load Dataset

```
[ ]: from google.colab import drive
drive.mount('/content/drive')
```

Mounted at /content/drive

```
[ ]: df_f = pd.read_csv('/content/drive/MyDrive/PCOS/PCOS_data_without_infertility.
    ↪ csv')
df_f.head(12).T
```

```
[ ]: df_f.info()
```

<class 'pandas.core.frame.DataFrame'>

RangeIndex: 541 entries, 0 to 540

Data columns (total 45 columns):

#	Column	Non-Null Count	Dtype
---	-----	-----	-----
0	Sl. No	541 non-null	int64

1	Patient File No.	541 non-null	int64
2	PCOS (Y/N)	541 non-null	int64
3	Age (yrs)	541 non-null	int64
4	Weight (Kg)	541 non-null	float64
5	Height(Cm)	541 non-null	float64
6	BMI	541 non-null	object
7	Blood Group	541 non-null	int64
8	Pulse rate(bpm)	541 non-null	int64
9	RR (breaths/min)	541 non-null	int64
10	Hb(g/dl)	541 non-null	float64
11	Cycle(R/I)	541 non-null	int64
12	Cycle length(days)	541 non-null	int64
13	Marraige Status (Yrs)	540 non-null	float64
14	Pregnant(Y/N)	541 non-null	int64
15	No. of absorptions	541 non-null	int64
16	I beta-HCG(mIU/mL)	541 non-null	float64
17	II beta-HCG(mIU/mL)	541 non-null	object
18	FSH(mIU/mL)	541 non-null	float64
19	LH(mIU/mL)	541 non-null	float64
20	FSH/LH	541 non-null	object
21	Hip(inch)	541 non-null	int64
22	Waist(inch)	541 non-null	int64
23	Waist:Hip Ratio	541 non-null	object
24	TSH (mIU/L)	541 non-null	float64
25	AMH(ng/mL)	541 non-null	object
26	PRL(ng/mL)	541 non-null	float64
27	Vit D3 (ng/mL)	541 non-null	float64
28	PRG(ng/mL)	541 non-null	float64
29	RBS(mg/dl)	541 non-null	float64
30	Weight gain(Y/N)	541 non-null	int64
31	hair growth(Y/N)	541 non-null	int64
32	Skin darkening (Y/N)	541 non-null	int64
33	Hair loss(Y/N)	541 non-null	int64
34	Pimples(Y/N)	541 non-null	int64
35	Fast food (Y/N)	540 non-null	float64
36	Reg.Exercise(Y/N)	541 non-null	int64
37	BP _Systolic (mmHg)	541 non-null	int64
38	BP _Diastolic (mmHg)	541 non-null	int64
39	Follicle No. (L)	541 non-null	int64
40	Follicle No. (R)	541 non-null	int64
41	Avg. F size (L) (mm)	541 non-null	float64
42	Avg. F size (R) (mm)	541 non-null	float64
43	Endometrium (mm)	541 non-null	float64
44	Unnamed: 44	2 non-null	object

dtypes: float64(16), int64(23), object(6)

memory usage: 190.3+ KB

## 4 Explore Dataset

### 5 Check For Null Values

```
[ ]: df_f.isnull()
```

```
[ ]:      Sl. No  Patient File No.  ...  Endometrium (mm)  Unnamed: 44
0      False                False  ...                False                True
1      False                False  ...                False                True
2      False                False  ...                False                True
3      False                False  ...                False                True
4      False                False  ...                False                True
..      ...                  ...  ...                  ...                  ...
536     False                False  ...                False                True
537     False                False  ...                False                True
538     False                False  ...                False                True
539     False                False  ...                False                True
540     False                False  ...                False                True
```

[541 rows x 45 columns]

```
[ ]: df_f.isnull().sum()
```

```
[ ]: Sl. No                0
Patient File No.         0
PCOS (Y/N)               0
Age (yrs)                0
Weight (Kg)              0
Height(Cm)               0
BMI                      0
Blood Group              0
Pulse rate(bpm)          0
RR (breaths/min)         0
Hb(g/dl)                 0
Cycle(R/I)               0
Cycle length(days)       0
Marraige Status (Yrs)    1
Pregnant(Y/N)            0
No. of aborptions        0
I   beta-HCG(mIU/mL)     0
II  beta-HCG(mIU/mL)     0
FSH(mIU/mL)              0
LH(mIU/mL)               0
FSH/LH                   0
Hip(inch)                0
Waist(inch)              0
Waist:Hip Ratio          0
```

```

TSH (mIU/L)          0
AMH(ng/mL)           0
PRL(ng/mL)           0
Vit D3 (ng/mL)       0
PRG(ng/mL)           0
RBS(mg/dl)           0
Weight gain(Y/N)     0
hair growth(Y/N)     0
Skin darkening (Y/N) 0
Hair loss(Y/N)       0
Pimples(Y/N)         0
Fast food (Y/N)      1
Reg.Exercise(Y/N)    0
BP _Systolic (mmHg)  0
BP _Diastolic (mmHg) 0
Follicle No. (L)     0
Follicle No. (R)     0
Avg. F size (L) (mm) 0
Avg. F size (R) (mm) 0
Endometrium (mm)     0
Unnamed: 44          539
dtype: int64

```

```

[ ]: # Drop the column containg almost all null and uninterpretable values
df_f = df_f.drop(columns='Unnamed: 44')
# Drop unnecessary columns
df_f = df_f.drop(columns=['Sl. No', 'Patient File No.'])

```

```

[ ]: df_f.head(12).T

```

```

[ ]:

```

	0	1	2	...	9	10	11
PCOS (Y/N)	0	0	1	...	0	0	0
Age (yrs)	28	36	33	...	36	20	26
Weight (Kg)	44.6	65	68.8	...	52	71	49
Height(Cm)	152	161.5	165	...	150	163	160
BMI	19.3	#NAME?	#NAME?	...	#NAME?	#NAME?	#NAME?
Blood Group	15	15	11	...	15	15	13
Pulse rate(bpm)	78	74	72	...	80	80	72
RR (breaths/min)	22	20	18	...	20	20	20
Hb(g/dl)	10.48	11.7	11.8	...	10	10	9.5
Cycle(R/I)	2	2	2	...	4	2	2
Cycle length(days)	5	5	5	...	2	5	5
Marraige Status (Yrs)	7	11	10	...	4	4	3
Pregnant(Y/N)	0	1	1	...	0	1	0
No. of abortions	0	0	0	...	0	2	1
I beta-HCG(mIU/mL)	1.99	60.8	494.08	...	1.99	158.51	1.99
II beta-HCG(mIU/mL)	1.99	1.99	494.08	...	1.99	158.51	1.99



FSH(mIU/mL)	7.95	6.73	5.54	...	2.8	4.89	4.09
LH(mIU/mL)	3.68	1.09	0.88	...	1.51	2.02	1.47
FSH/LH	#NAME?	#NAME?	#NAME?	...	#NAME?	#NAME?	#NAME?
Hip(inch)	36	38	40	...	40	39	39
Waist(inch)	30	32	36	...	38	35	33
Waist:Hip Ratio	#NAME?	#NAME?	#NAME?	...	#NAME?	#NAME?	#NAME?
TSH (mIU/L)	0.68	3.16	2.54	...	6.65	1.56	3.98
AMH(ng/mL)	2.07	1.53	6.63	...	1.61	4.47	1.67
PRL(ng/mL)	45.16	20.09	10.52	...	11.74	13.47	21.1
Vit D3 (ng/mL)	17.1	61.3	49.7	...	27.7	18.1	29.18
PRG(ng/mL)	0.57	0.97	0.36	...	0.25	0.36	0.25
RBS(mg/dl)	92	92	84	...	125	108	100
Weight gain(Y/N)	0	0	0	...	0	0	0
hair growth(Y/N)	0	0	0	...	0	0	0
Skin darkening (Y/N)	0	0	0	...	0	0	0
Hair loss(Y/N)	0	0	1	...	0	0	0
Pimples(Y/N)	0	0	1	...	0	0	0
Fast food (Y/N)	1	0	1	...	0	0	0
Reg.Exercise(Y/N)	0	0	0	...	0	0	0
BP _Systolic (mmHg)	110	120	120	...	110	110	120
BP _Diastolic (mmHg)	80	70	80	...	80	80	80
Follicle No. (L)	3	3	13	...	1	7	4
Follicle No. (R)	3	5	15	...	1	15	2
Avg. F size (L) (mm)	18	15	18	...	14	17	18
Avg. F size (R) (mm)	18	14	20	...	17	20	19
Endometrium (mm)	8.5	3.7	10	...	2.5	6	7.8

[42 rows x 12 columns]

## 6 Handling Missing Values

### 7 Drop Rows with null values

```
[ ]: df_f.dropna(axis=0, inplace=True)
```

```
[ ]: df_f.isnull().sum()
```

```
[ ]: PCOS (Y/N)          0
      Age (yrs)          0
      Weight (Kg)        0
      Height(Cm)         0
      BMI                0
      Blood Group        0
      Pulse rate(bpm)    0
      RR (breaths/min)   0
      Hb(g/dl)           0
```

```

Cycle(R/I)          0
Cycle length(days)  0
Marraige Status (Yrs)  0
Pregnant(Y/N)       0
No. of abortions    0
  I   beta-HCG(mIU/mL)  0
  II  beta-HCG(mIU/mL)  0
FSH(mIU/mL)         0
LH(mIU/mL)          0
FSH/LH              0
Hip(inch)           0
Waist(inch)         0
Waist:Hip Ratio     0
TSH (mIU/L)         0
AMH(ng/mL)          0
PRL(ng/mL)          0
Vit D3 (ng/mL)      0
PRG(ng/mL)          0
RBS(mg/dl)          0
Weight gain(Y/N)    0
hair growth(Y/N)    0
Skin darkening (Y/N) 0
Hair loss(Y/N)      0
Pimples(Y/N)        0
Fast food (Y/N)     0
Reg.Exercise(Y/N)   0
BP _Systolic (mmHg) 0
BP _Diastolic (mmHg) 0
Follicle No. (L)    0
Follicle No. (R)    0
Avg. F size (L) (mm) 0
Avg. F size (R) (mm) 0
Endometrium (mm)    0
dtype: int64

```

```
[ ]: df_f.info()
```

```

<class 'pandas.core.frame.DataFrame'>
Int64Index: 539 entries, 0 to 540
Data columns (total 42 columns):
#   Column                Non-Null Count  Dtype
---  -
0   PCOS (Y/N)            539 non-null   int64
1   Age (yrs)             539 non-null   int64
2   Weight (Kg)           539 non-null   float64
3   Height(Cm)            539 non-null   float64
4   BMI                   539 non-null   object
5   Blood Group           539 non-null   int64

```

```

6 Pulse rate(bpm)          539 non-null    int64
7 RR (breaths/min)        539 non-null    int64
8 Hb(g/dl)                 539 non-null    float64
9 Cycle(R/I)              539 non-null    int64
10 Cycle length(days)      539 non-null    int64
11 Marraige Status (Yrs)   539 non-null    float64
12 Pregnant(Y/N)           539 non-null    int64
13 No. of aborptions       539 non-null    int64
14 I beta-HCG(mIU/mL)     539 non-null    float64
15 II beta-HCG(mIU/mL)    539 non-null    object
16 FSH(mIU/mL)             539 non-null    float64
17 LH(mIU/mL)              539 non-null    float64
18 FSH/LH                  539 non-null    object
19 Hip(inch)               539 non-null    int64
20 Waist(inch)             539 non-null    int64
21 Waist:Hip Ratio         539 non-null    object
22 TSH (mIU/L)            539 non-null    float64
23 AMH(ng/mL)             539 non-null    object
24 PRL(ng/mL)             539 non-null    float64
25 Vit D3 (ng/mL)         539 non-null    float64
26 PRG(ng/mL)             539 non-null    float64
27 RBS(mg/dl)             539 non-null    float64
28 Weight gain(Y/N)       539 non-null    int64
29 hair growth(Y/N)       539 non-null    int64
30 Skin darkening (Y/N)   539 non-null    int64
31 Hair loss(Y/N)         539 non-null    int64
32 Pimples(Y/N)           539 non-null    int64
33 Fast food (Y/N)        539 non-null    float64
34 Reg.Exercise(Y/N)      539 non-null    int64
35 BP _Systolic (mmHg)    539 non-null    int64
36 BP _Diastolic (mmHg)   539 non-null    int64
37 Follicle No. (L)       539 non-null    int64
38 Follicle No. (R)       539 non-null    int64
39 Avg. F size (L) (mm)   539 non-null    float64
40 Avg. F size (R) (mm)   539 non-null    float64
41 Endometrium (mm)       539 non-null    float64
dtypes: float64(16), int64(21), object(5)
memory usage: 181.1+ KB

```

## 8 Solve corrupted values

```
[ ]: df_f
```

```

[ ]:      PCOS (Y/N)  Age (yrs)  ...  Avg. F size (R) (mm)  Endometrium (mm)
0           0        28  ...           18.0           8.5
1           0        36  ...           14.0           3.7
2           1        33  ...           20.0          10.0

```

3	0	37	...	14.0	7.5
4	0	25	...	14.0	7.0
..	...	...	...	...	...
536	0	35	...	10.0	6.7
537	0	30	...	18.0	8.2
538	0	36	...	9.0	7.3
539	0	27	...	16.0	11.5
540	1	23	...	18.0	6.9

[539 rows x 42 columns]

```
[ ]: #calculating BMI
df_f['BMI'] = (df_f['Weight (Kg)']/(df_f['Height(Cm) '])**2)*10000
df_f
```

	PCOS (Y/N)	Age (yrs)	...	Avg. F size (R) (mm)	Endometrium (mm)
0	0	28	...	18.0	8.5
1	0	36	...	14.0	3.7
2	1	33	...	20.0	10.0
3	0	37	...	14.0	7.5
4	0	25	...	14.0	7.0
..	...	...	...	...	...
536	0	35	...	10.0	6.7
537	0	30	...	18.0	8.2
538	0	36	...	9.0	7.3
539	0	27	...	16.0	11.5
540	1	23	...	18.0	6.9

[539 rows x 42 columns]

```
[ ]: df_f.loc[:, "FSH/LH"] = df_f.loc[:, "FSH(mIU/mL)"] / df_f.loc[:, "LH(mIU/mL)"];
df_f.loc[:, "FSH/LH"] = df_f.loc[:, "FSH/LH"].round(2);
```

```
[ ]: df_f.loc[:, "Waist:Hip Ratio"] = df_f.loc[:, "Waist(inch)"] / df_f.loc[:,
↪, "Hip(inch)"]
df_f.loc[:, "Waist:Hip Ratio"] = df_f.loc[:, "Waist:Hip Ratio"].round(2)
```

```
[ ]: df_f.head(12)
```

	PCOS (Y/N)	Age (yrs)	...	Avg. F size (R) (mm)	Endometrium (mm)
0	0	28	...	18.0	8.5
1	0	36	...	14.0	3.7
2	1	33	...	20.0	10.0
3	0	37	...	14.0	7.5
4	0	25	...	14.0	7.0
5	0	36	...	20.0	8.0
6	0	34	...	16.0	6.8

7	0	33	...	18.0	7.1
8	0	32	...	17.0	4.2
9	0	36	...	17.0	2.5
10	0	20	...	20.0	6.0
11	0	26	...	19.0	7.8

[12 rows x 42 columns]

```
[ ]: df_f.info()
```

```
<class 'pandas.core.frame.DataFrame'>
Int64Index: 539 entries, 0 to 540
Data columns (total 42 columns):
#   Column                                Non-Null Count  Dtype
---  -
0   PCOS (Y/N)                            539 non-null    int64
1   Age (yrs)                             539 non-null    int64
2   Weight (Kg)                           539 non-null    float64
3   Height(Cm)                            539 non-null    float64
4   BMI                                   539 non-null    float64
5   Blood Group                           539 non-null    int64
6   Pulse rate(bpm)                       539 non-null    int64
7   RR (breaths/min)                      539 non-null    int64
8   Hb(g/dl)                              539 non-null    float64
9   Cycle(R/I)                            539 non-null    int64
10  Cycle length(days)                    539 non-null    int64
11  Marraige Status (Yrs)                  539 non-null    float64
12  Pregnant(Y/N)                          539 non-null    int64
13  No. of absorptions                     539 non-null    int64
14  I    beta-HCG(mIU/mL)                  539 non-null    float64
15  II   beta-HCG(mIU/mL)                  539 non-null    object
16  FSH(mIU/mL)                           539 non-null    float64
17  LH(mIU/mL)                            539 non-null    float64
18  FSH/LH                                539 non-null    float64
19  Hip(inch)                             539 non-null    int64
20  Waist(inch)                           539 non-null    int64
21  Waist:Hip Ratio                        539 non-null    float64
22  TSH (mIU/L)                           539 non-null    float64
23  AMH(ng/mL)                            539 non-null    object
24  PRL(ng/mL)                            539 non-null    float64
25  Vit D3 (ng/mL)                        539 non-null    float64
26  PRG(ng/mL)                            539 non-null    float64
27  RBS(mg/dl)                            539 non-null    float64
28  Weight gain(Y/N)                       539 non-null    int64
29  hair growth(Y/N)                       539 non-null    int64
30  Skin darkening (Y/N)                   539 non-null    int64
31  Hair loss(Y/N)                         539 non-null    int64
32  Pimples(Y/N)                           539 non-null    int64
```

```

33 Fast food (Y/N)          539 non-null    float64
34 Reg.Exercise(Y/N)       539 non-null    int64
35 BP _Systolic (mmHg)     539 non-null    int64
36 BP _Diastolic (mmHg)    539 non-null    int64
37 Follicle No. (L)        539 non-null    int64
38 Follicle No. (R)        539 non-null    int64
39 Avg. F size (L) (mm)    539 non-null    float64
40 Avg. F size (R) (mm)    539 non-null    float64
41 Endometrium (mm)        539 non-null    float64
dtypes: float64(19), int64(21), object(2)
memory usage: 181.1+ KB

```

## 9 Replace Irrelevant Values

```

[ ]: # df[df["Cycle(R/I)"] == 5]
df_f["Cycle(R/I)"].replace({5: 4}, inplace=True)
df_f["Cycle(R/I)"].replace({2: 0, 4: 1}, inplace=True)

```

```

[ ]: df_f["II    beta-HCG(mIU/mL)"].replace({"1.99.": 1.99}, inplace=True)

```

```

[ ]: df_f["II    beta-HCG(mIU/mL)"] = df_f["II    beta-HCG(mIU/mL)"].astype(float)

```

```

[ ]: df_f[df_f["AMH(ng/mL)"] == "a"].T

```

```

[ ]:
PCOS (Y/N)          0
Age (yrs)           37
Weight (Kg)         56
Height(Cm)          152
BMI                 24.2382
Blood Group         13
Pulse rate(bpm)     74
RR (breaths/min)    20
Hb(g/dl)            11.7
Cycle(R/I)          0
Cycle length(days)  5
Marraige Status (Yrs) 9
Pregnant(Y/N)       0
No. of abortions    0
I    beta-HCG(mIU/mL) 42
II   beta-HCG(mIU/mL) 1.99
FSH(mIU/mL)         2.91
LH(mIU/mL)          0.35
FSH/LH              8.31
Hip(inch)           35
Waist(inch)         33
Waist:Hip Ratio     0.94

```

TSH (mIU/L)	16
AMH(ng/mL)	a
PRL(ng/mL)	2.22
Vit D3 (ng/mL)	38.6
PRG(ng/mL)	0.3
RBS(mg/dl)	100
Weight gain(Y/N)	0
hair growth(Y/N)	0
Skin darkening (Y/N)	0
Hair loss(Y/N)	0
Pimples(Y/N)	1
Fast food (Y/N)	0
Reg.Exercise(Y/N)	1
BP _Systolic (mmHg)	120
BP _Diastolic (mmHg)	70
Follicle No. (L)	4
Follicle No. (R)	5
Avg. F size (L) (mm)	17
Avg. F size (R) (mm)	16
Endometrium (mm)	5.6

```
[ ]: # df_f.drop(df_f["AMH(ng/mL)"]== "a", inplace=True)
df_f.drop(df_f.loc[df_f["AMH(ng/mL)"]== "a"].index, inplace=True);
```

```
[ ]: df_f[df_f["AMH(ng/mL)"]== "a"]
```

```
[ ]: Empty DataFrame
Columns: [PCOS (Y/N), Age (yrs), Weight (Kg), Height(Cm) , BMI, Blood Group,
Pulse rate(bpm) , RR (breaths/min), Hb(g/dl), Cycle(R/I), Cycle length(days),
Marraige Status (Yrs), Pregnant(Y/N), No. of absorptions, I beta-HCG(mIU/mL),
II beta-HCG(mIU/mL), FSH(mIU/mL), LH(mIU/mL), FSH/LH, Hip(inch), Waist(inch),
Waist:Hip Ratio, TSH (mIU/L), AMH(ng/mL), PRL(ng/mL), Vit D3 (ng/mL),
PRG(ng/mL), RBS(mg/dl), Weight gain(Y/N), hair growth(Y/N), Skin darkening
(Y/N), Hair loss(Y/N), Pimples(Y/N), Fast food (Y/N), Reg.Exercise(Y/N), BP
_Systolic (mmHg), BP _Diastolic (mmHg), Follicle No. (L), Follicle No. (R), Avg.
F size (L) (mm), Avg. F size (R) (mm), Endometrium (mm)]
Index: []
```

```
[ ]: df_f["AMH(ng/mL)"] = df_f["AMH(ng/mL)"].astype(float)
```

```
[ ]: df_f.info();
```

```
<class 'pandas.core.frame.DataFrame'>
Int64Index: 538 entries, 0 to 540
Data columns (total 42 columns):
#   Column                                Non-Null Count  Dtype
---  -
0   PCOS (Y/N)                            538 non-null    int64
```



1	Age (yrs)	538 non-null	int64
2	Weight (Kg)	538 non-null	float64
3	Height(Cm)	538 non-null	float64
4	BMI	538 non-null	float64
5	Blood Group	538 non-null	int64
6	Pulse rate(bpm)	538 non-null	int64
7	RR (breaths/min)	538 non-null	int64
8	Hb(g/dl)	538 non-null	float64
9	Cycle(R/I)	538 non-null	int64
10	Cycle length(days)	538 non-null	int64
11	Marraige Status (Yrs)	538 non-null	float64
12	Pregnant(Y/N)	538 non-null	int64
13	No. of absorptions	538 non-null	int64
14	I beta-HCG(mIU/mL)	538 non-null	float64
15	II beta-HCG(mIU/mL)	538 non-null	float64
16	FSH(mIU/mL)	538 non-null	float64
17	LH(mIU/mL)	538 non-null	float64
18	FSH/LH	538 non-null	float64
19	Hip(inch)	538 non-null	int64
20	Waist(inch)	538 non-null	int64
21	Waist:Hip Ratio	538 non-null	float64
22	TSH (mIU/L)	538 non-null	float64
23	AMH(ng/mL)	538 non-null	float64
24	PRL(ng/mL)	538 non-null	float64
25	Vit D3 (ng/mL)	538 non-null	float64
26	PRG(ng/mL)	538 non-null	float64
27	RBS(mg/dl)	538 non-null	float64
28	Weight gain(Y/N)	538 non-null	int64
29	hair growth(Y/N)	538 non-null	int64
30	Skin darkening (Y/N)	538 non-null	int64
31	Hair loss(Y/N)	538 non-null	int64
32	Pimples(Y/N)	538 non-null	int64
33	Fast food (Y/N)	538 non-null	float64
34	Reg.Exercise(Y/N)	538 non-null	int64
35	BP _Systolic (mmHg)	538 non-null	int64
36	BP _Diastolic (mmHg)	538 non-null	int64
37	Follicle No. (L)	538 non-null	int64
38	Follicle No. (R)	538 non-null	int64
39	Avg. F size (L) (mm)	538 non-null	float64
40	Avg. F size (R) (mm)	538 non-null	float64
41	Endometrium (mm)	538 non-null	float64

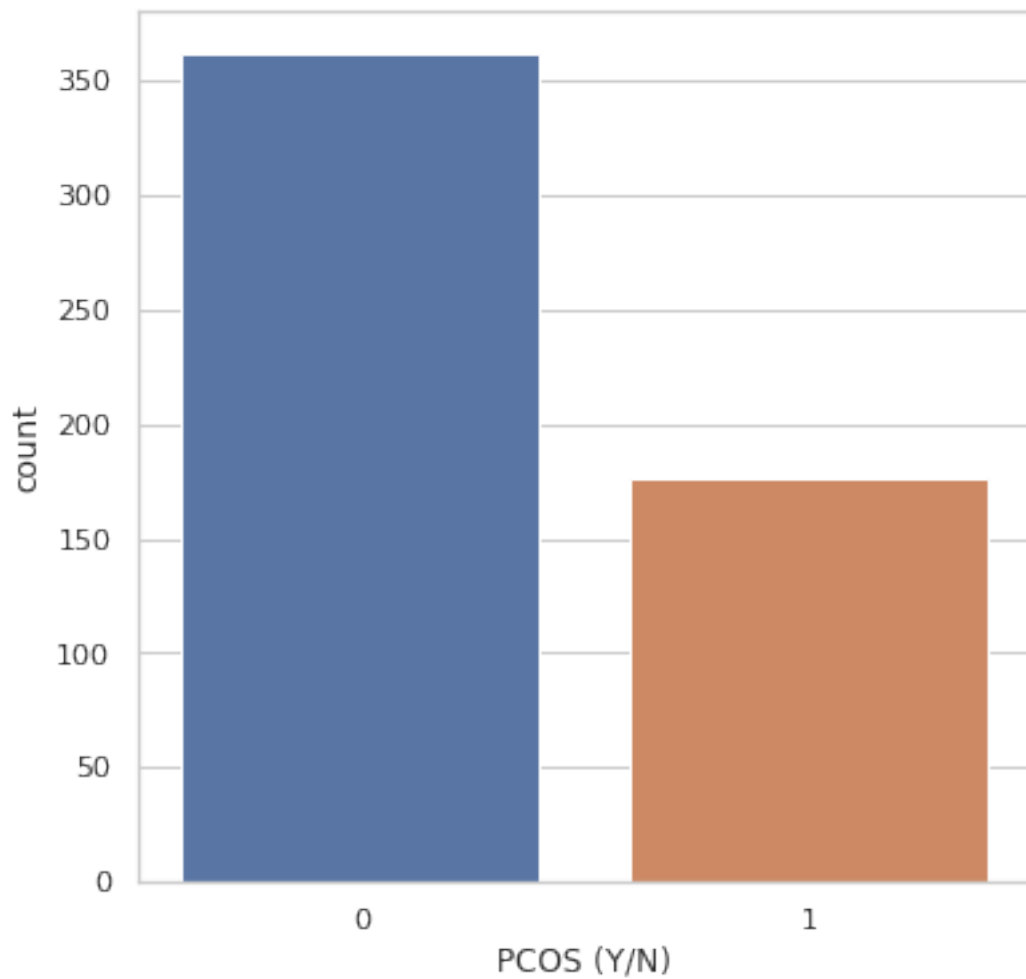
dtypes: float64(21), int64(21)

memory usage: 180.7 KB

## 9.1 Count

```
[ ]: print(df_f['PCOS (Y/N)'].value_counts())  
plt.figure(figsize=(6, 6))  
sns.countplot(  
    x='PCOS (Y/N)',  
    data=df_f  
);
```

```
0    362  
1    176  
Name: PCOS (Y/N), dtype: int64
```

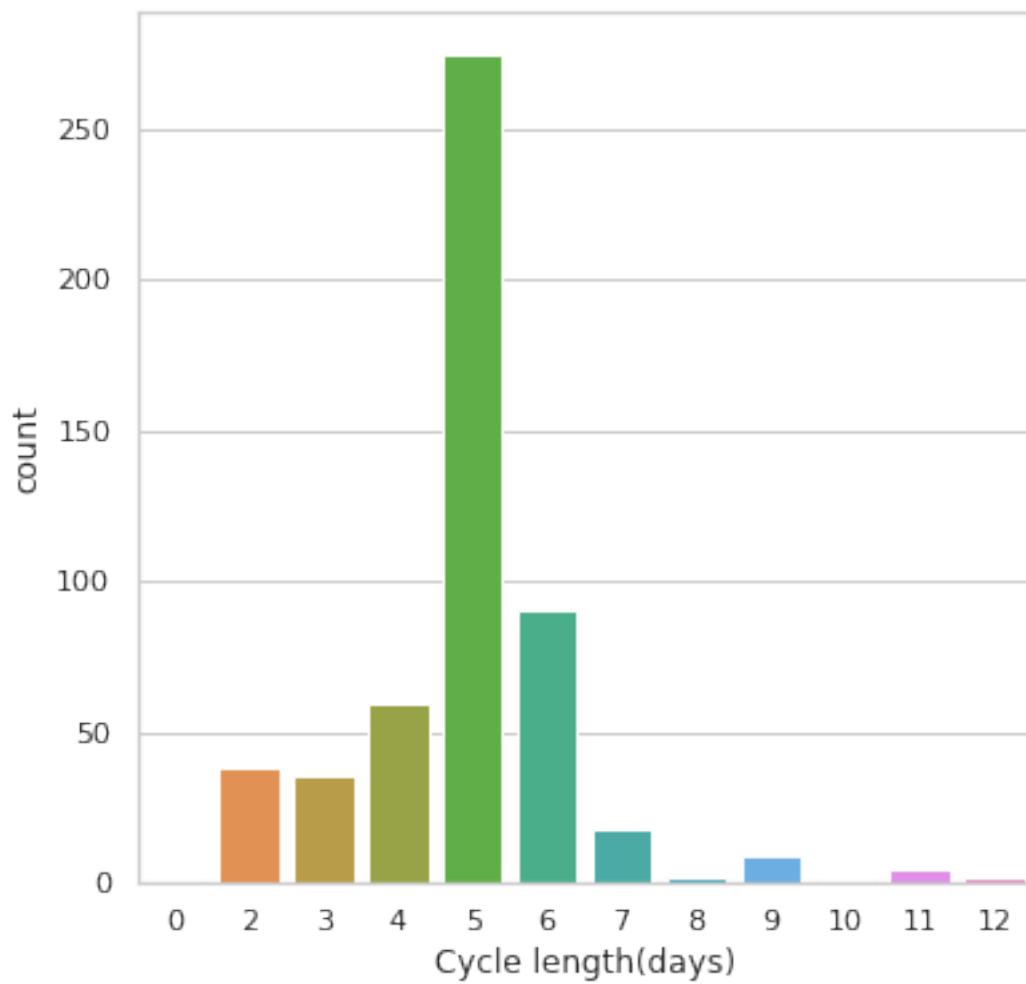


```
[ ]: print(df_f['Cycle length(days)'].value_counts())  
plt.figure(figsize=(6, 6))  
sns.countplot(  
    x='Cycle length(days)',
```

```
data=df_f  
)
```

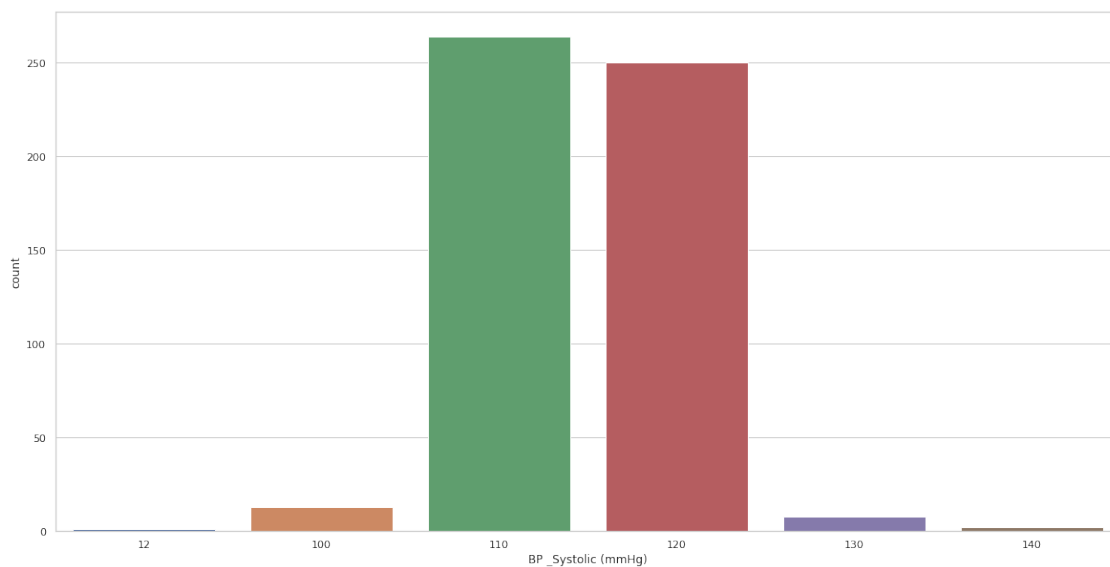
```
5    275  
6     91  
4     60  
2     38  
3     36  
7     18  
9      9  
11     5  
12     2  
8      2  
10     1  
0      1
```

Name: Cycle length(days), dtype: int64



```
[ ]: print(df_f['BP _Systolic (mmHg)'].value_counts())
plt.figure(figsize=(20, 10))
sns.countplot(
    x='BP _Systolic (mmHg)',
    data=df_f
);
```

```
110    264
120    250
100     13
130      8
140      2
12       1
Name: BP _Systolic (mmHg), dtype: int64
```



```
[ ]: df_f["BP _Systolic (mmHg)"].replace({12: 120}, inplace=True)
df_f["BP _Diastolic (mmHg)"].replace({8: 80}, inplace=True)
```

## 10 Save Cleaned Dataset

```
[ ]: #df_f.to_csv('../datasets/PCOS_clean_data_without_infertility.csv',
    ↪ index=False)
```

# **FEATURE SELECTION**

# PCOS\_ML\_Part\_2

October 29, 2021

## 1 Feature Selection For The Dataset

### 1.1 Importing Libraries

```
[ ]: import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
```

```
[ ]: df=pd.read_csv("/content/drive/MyDrive/PCOS/PCOS_clean_data_without_infertility.
↪csv")
```

```
[ ]: df.describe().T
```

```
[ ]:
```

	count	mean	...	75%	max
PCOS (Y/N)	538.0	0.327138	...	1.0000	1.00
Age (yrs)	538.0	31.420074	...	35.0000	48.00
Weight (Kg)	538.0	59.644052	...	65.0000	108.00
Height(Cm)	538.0	156.480104	...	160.0000	180.00
BMI	538.0	24.323625	...	26.6625	38.90
Blood Group	538.0	13.802974	...	15.0000	18.00
Pulse rate(bpm)	538.0	73.250929	...	74.0000	82.00
RR (breaths/min)	538.0	19.236059	...	20.0000	28.00
Hb(g/dl)	538.0	11.160558	...	11.7750	14.80
Cycle(R/I)	538.0	0.276952	...	1.0000	1.00
Cycle length(days)	538.0	4.938662	...	5.0000	12.00
Marraige Status (Yrs)	538.0	7.683457	...	10.0000	30.00
Pregnant(Y/N)	538.0	0.382900	...	1.0000	1.00
No. of abortions	538.0	0.289963	...	0.0000	5.00
I beta-HCG(mIU/mL)	538.0	667.215271	...	297.0500	32460.97
II beta-HCG(mIU/mL)	538.0	239.550333	...	99.1750	25000.00
FSH(mIU/mL)	538.0	14.669054	...	6.4175	5052.00
LH(mIU/mL)	538.0	6.503933	...	3.6800	2018.00
FSH/LH	538.0	6.900279	...	3.8700	1372.83
Hip(inch)	538.0	37.998141	...	40.0000	48.00
Waist(inch)	538.0	33.840149	...	36.0000	47.00
Waist:Hip Ratio	538.0	0.891245	...	0.9300	0.98
TSH (mIU/L)	538.0	2.960935	...	3.5700	65.00
AMH(ng/mL)	538.0	5.623035	...	6.9750	66.00

PRL(ng/mL)	538.0	24.393216	...	29.9500	128.24
Vit D3 (ng/mL)	538.0	50.036781	...	34.4750	6014.66
PRG(ng/mL)	538.0	0.612660	...	0.4575	85.00
RBS(mg/dl)	538.0	99.864684	...	107.0000	350.00
Weight gain(Y/N)	538.0	0.379182	...	1.0000	1.00
hair growth(Y/N)	538.0	0.275093	...	1.0000	1.00
Skin darkening (Y/N)	538.0	0.306691	...	1.0000	1.00
Hair loss(Y/N)	538.0	0.453532	...	1.0000	1.00
Pimples(Y/N)	538.0	0.490706	...	1.0000	1.00
Fast food (Y/N)	538.0	0.516729	...	1.0000	1.00
Reg.Exercise(Y/N)	538.0	0.245353	...	0.0000	1.00
BP _Systolic (mmHg)	538.0	114.832714	...	120.0000	140.00
BP _Diastolic (mmHg)	538.0	77.081784	...	80.0000	100.00
Follicle No. (L)	538.0	6.120818	...	9.0000	22.00
Follicle No. (R)	538.0	6.646840	...	10.0000	20.00
Avg. F size (L) (mm)	538.0	15.014498	...	18.0000	24.00
Avg. F size (R) (mm)	538.0	15.448643	...	18.0000	24.00
Endometrium (mm)	538.0	8.477454	...	9.8000	18.00

[42 rows x 8 columns]

```
[ ]: df.corr()
```

```
[ ]:
```

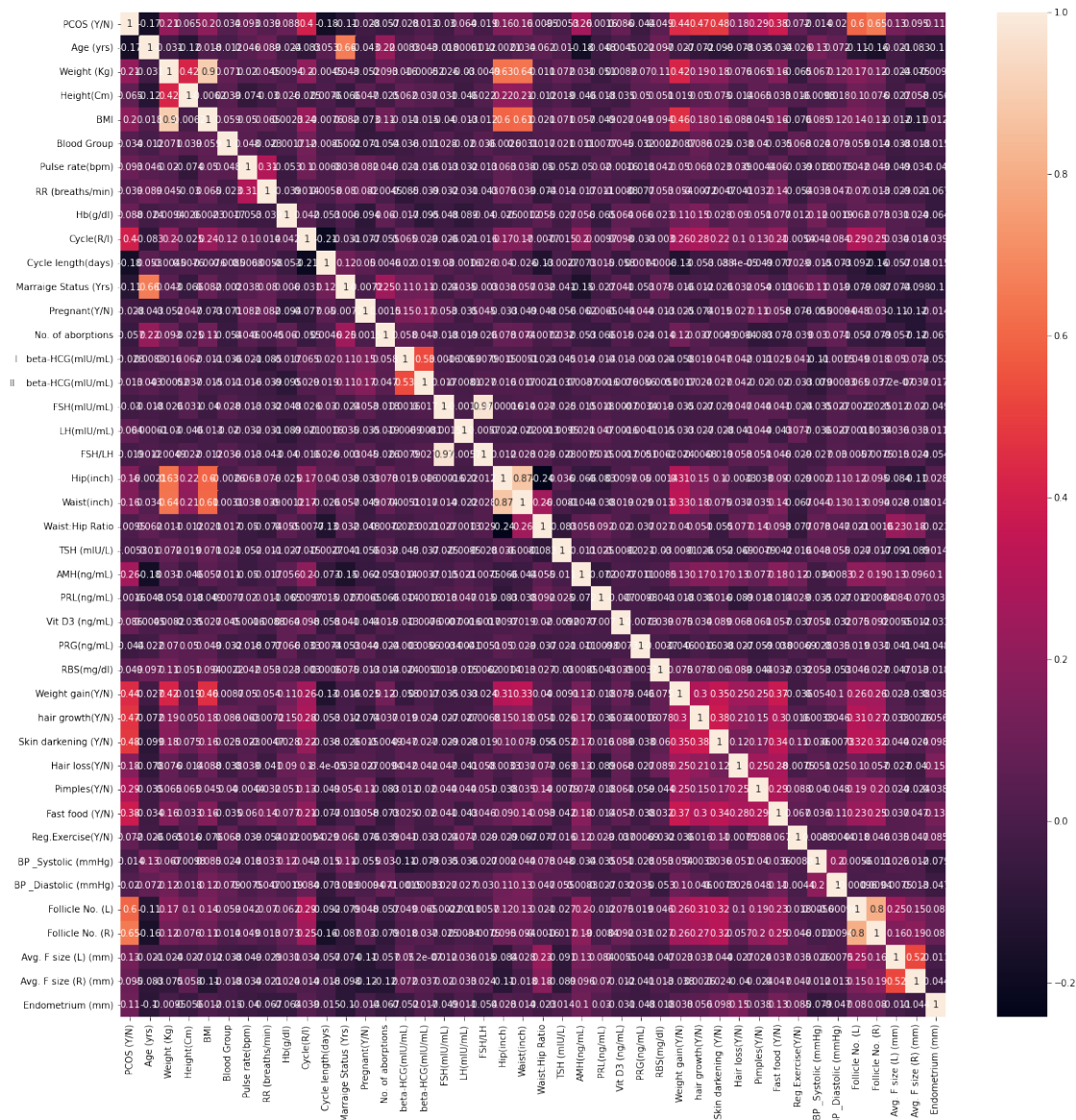
	PCOS (Y/N)	...	Endometrium (mm)
PCOS (Y/N)	1.000000	...	0.107639
Age (yrs)	-0.171349	...	-0.100115
Weight (Kg)	0.209969	...	-0.009452
Height(Cm)	0.065465	...	-0.056273
BMI	0.199302	...	0.012331
Blood Group	0.033701	...	-0.015257
Pulse rate(bpm)	0.092699	...	-0.040456
RR (breaths/min)	0.038641	...	-0.066551
Hb(g/dl)	0.087809	...	-0.063592
Cycle(R/I)	0.400668	...	0.039000
Cycle length(days)	-0.183811	...	-0.014630
Marraige Status (Yrs)	-0.113406	...	-0.104838
Pregnant(Y/N)	-0.027632	...	-0.013675
No. of abortions	-0.057316	...	-0.067136
I beta-HCG(mIU/mL)	-0.028074	...	-0.051580
II beta-HCG(mIU/mL)	0.012808	...	0.017295
FSH(mIU/mL)	-0.030384	...	-0.049132
LH(mIU/mL)	0.064074	...	0.010917
FSH/LH	-0.018512	...	-0.053760
Hip(inch)	0.160882	...	0.028465
Waist(inch)	0.161922	...	0.014308
Waist:Hip Ratio	0.009505	...	-0.023082
TSH (mIU/L)	-0.005277	...	0.014302



AMH(ng/mL)	0.263974 ...	0.104496
PRL(ng/mL)	0.001649 ...	0.030021
Vit D3 (ng/mL)	0.085825 ...	-0.031307
PRG(ng/mL)	-0.043897 ...	-0.047946
RBS(mg/dl)	0.049452 ...	-0.017851
Weight gain(Y/N)	0.443093 ...	0.038255
hair growth(Y/N)	0.466508 ...	0.055972
Skin darkening (Y/N)	0.481323 ...	0.097781
Hair loss(Y/N)	0.176496 ...	0.151038
Pimples(Y/N)	0.290335 ...	0.038164
Fast food (Y/N)	0.380985 ...	0.130807
Reg.Exercise(Y/N)	0.071979 ...	0.084927
BP _Systolic (mmHg)	-0.013765 ...	-0.079425
BP _Diastolic (mmHg)	0.019870 ...	-0.047305
Follicle No. (L)	0.601208 ...	0.080313
Follicle No. (R)	0.650929 ...	0.079607
Avg. F size (L) (mm)	0.129997 ...	-0.011326
Avg. F size (R) (mm)	0.094528 ...	-0.043720
Endometrium (mm)	0.107639 ...	1.000000

[42 rows x 42 columns]

```
[ ]: plt.figure(figsize=(20,20))
      sns.heatmap(df.corr(), annot=True)
      plt.show()
```



## 1.2 Feature Selction Based On Correlation

The Pearson Method is used to choose the features for model building. The method implemented includes choosing of only those features whose correlation i.e corr\_features is greater than 0.4 while discarding all the others.

This gave us the following features for final model building in subsequent notebooks. - Cycle(R/I) - Weight gain(Y/N) - hair growth(Y/N) - Skin darkening (Y/N) - Follicle No. (L) - Follicle No. (R) - PCOS (Y/N)—> Dependent Variable

```
[ ]: corr_features=df.corrwith(df["PCOS (Y/N)"],method='pearson').abs().
    ↪sort_values(ascending=True)
```

```
corr_features=corr_features[corr_features>0.4].index
corr_features
```

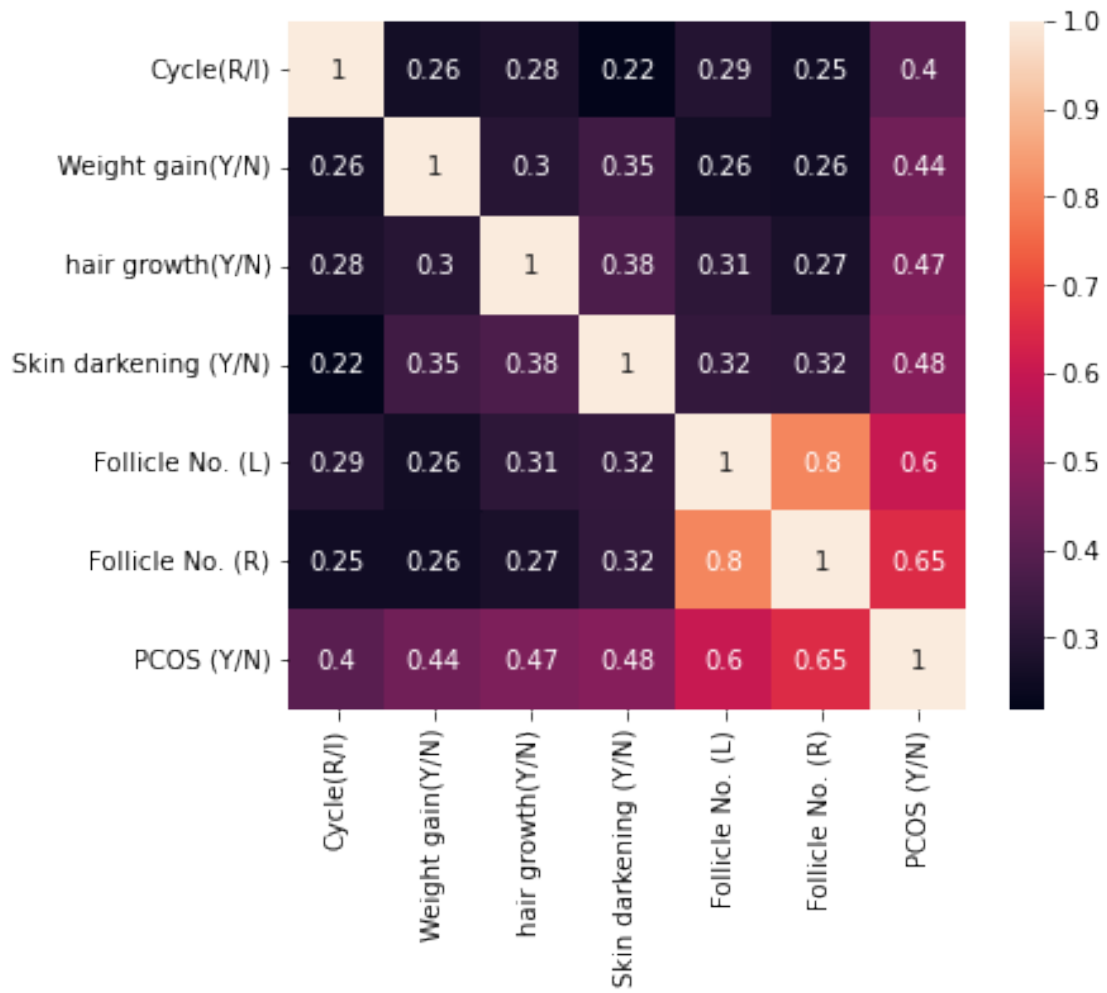
```
[ ]: Index(['Cycle(R/I)', 'Weight gain(Y/N)', 'hair growth(Y/N)',
          'Skin darkening (Y/N)', 'Follicle No. (L)', 'Follicle No. (R)',
          'PCOS (Y/N)'],
          dtype='object')
```

```
[ ]: df_f=df[corr_features]
df_f.head()
```

```
[ ]:   Cycle(R/I)  Weight gain(Y/N)  ...  Follicle No. (R)  PCOS (Y/N)
0           0           0  ...           3           0
1           0           0  ...           5           0
2           0           0  ...          15           1
3           0           0  ...           2           0
4           0           0  ...           4           0
```

[5 rows x 7 columns]

```
[ ]: plt.figure(figsize=(6,5))
sns.heatmap(df_f.corr(), annot=True)
plt.show()
```



**LOGISTIC  
REGRESSION  
&  
NAÏVE BAYES**

# PCOS\_ML\_Part\_3

October 29, 2021

## 1 Algorithm: Logistic Regression & Naive Bayes

### 1.1 Importing Libraries

```
[ ]: import pandas as pd
import matplotlib.pyplot as plt

from sklearn.model_selection import train_test_split
from sklearn.linear_model import LogisticRegression
from sklearn import metrics

from sklearn.metrics import confusion_matrix
from sklearn.metrics import accuracy_score
from sklearn.metrics import classification_report

import pickle
```

```
[ ]: from google.colab import drive
drive.mount('/content/drive')
```

Mounted at /content/drive

```
[ ]: df = pd.read_csv("/content/drive/MyDrive/PCOS/clean_data.csv")
```

```
[ ]: df.shape
```

```
[ ]: (534, 7)
```

```
[ ]: X = df.drop(columns=["PCOS (Y/N)"])
X.head()
```

```
[ ]: 
```

	Follicle No. (R)	Follicle No. (L)	...	Weight gain(Y/N)	Cycle(R/I)
0	3	3	...	0	2
1	5	3	...	0	2
2	15	13	...	0	2
3	2	2	...	0	2
4	4	3	...	0	2

[5 rows x 6 columns]

```
[ ]: y = df["PCOS (Y/N)"].values
```

## 1.2 Splitting Dataset

```
[ ]: X_train, X_test, y_train, y_test = train_test_split(X,y,test_size=0.2,
↳random_state=12)
```

## 2 Logistic Regression Implementation

```
[ ]: lr = LogisticRegression()
lr.fit(X_train, y_train)
```

```
[ ]: LogisticRegression(C=1.0, class_weight=None, dual=False, fit_intercept=True,
intercept_scaling=1, l1_ratio=None, max_iter=100,
multi_class='auto', n_jobs=None, penalty='l2',
random_state=None, solver='lbfgs', tol=0.0001, verbose=0,
warm_start=False)
```

```
[ ]: print(f"Score in Train Data : {lr.score(X_train,y_train)}")
```

Score in Train Data : 0.9016393442622951

```
[ ]: y_pred=lr.predict(X_test)
```

```
[ ]: print("Logistic Regression model accuracy(in %):", metrics.
↳accuracy_score(y_test, y_pred)*100)
```

Logistic Regression model accuracy(in %): 90.65420560747664

```
[ ]: print(classification_report(y_test, y_pred))
```

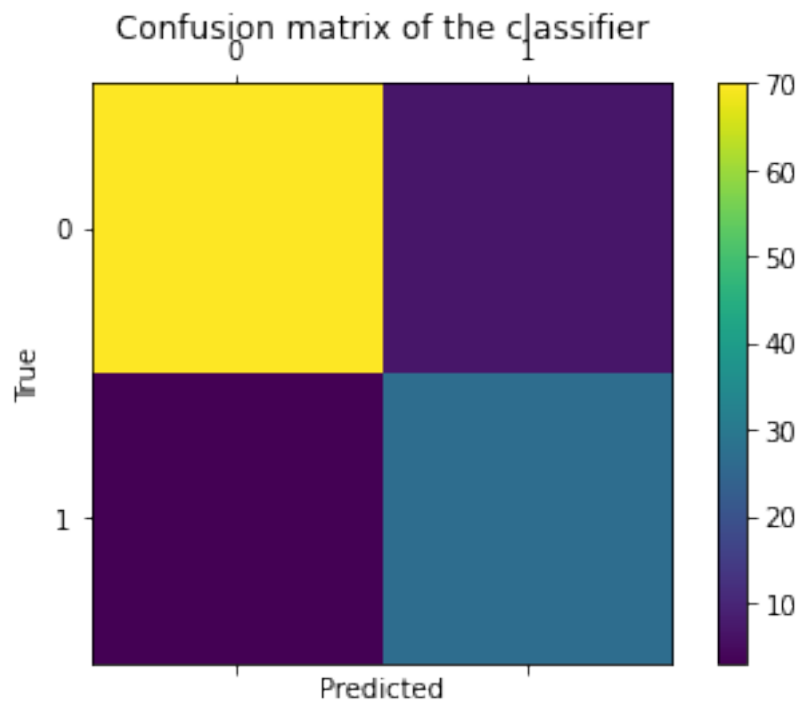
	precision	recall	f1-score	support
0	0.96	0.91	0.93	77
1	0.79	0.90	0.84	30
accuracy			0.91	107
macro avg	0.88	0.90	0.89	107
weighted avg	0.91	0.91	0.91	107

```
[ ]: cm = confusion_matrix(y_test, y_pred)
fig = plt.figure()

ax = fig.add_subplot(111)
cax = ax.matshow(cm)
plt.title('Confusion matrix of the classifier')
fig.colorbar(cax)
```



```
plt.xlabel('Predicted')
plt.ylabel('True')
plt.show()
```



```
[ ]: tn, fp, fn, tp = confusion_matrix(y_test, y_pred).ravel()
print("True Negatives: ",tn)
print("False Positives: ",fp)
print("False Negatives: ",fn)
print("True Positives: ",tp)
specificity = tn / (tn+fp)
print(specificity)
```

```
True Negatives: 70
False Positives: 7
False Negatives: 3
True Positives: 27
0.9090909090909091
```

```
[ ]: sensitivity=tp/(fn+tp)
print(sensitivity)
```

```
0.9
```

## 2.1 Naive Bayes Implementation

```
[ ]: from sklearn.naive_bayes import GaussianNB
```

```
[ ]: gnb = GaussianNB()  
gnb.fit(X_train, y_train)
```

```
[ ]: GaussianNB(priors=None, var_smoothing=1e-09)
```

```
[ ]: y_pred = gnb.predict(X_test)  
print("Gaussian Naive Bayes model accuracy(in %):", metrics.  
      ↳accuracy_score(y_test, y_pred)*100)
```

Gaussian Naive Bayes model accuracy(in %): 87.85046728971963

```
[ ]: tn, fp, fn, tp = confusion_matrix(y_test, y_pred).ravel()  
print("True Negatives: ",tn)  
print("False Positives: ",fp)  
print("False Negatives: ",fn)  
print("True Positives: ",tp)  
specificity = tn / (tn+fp)  
print(specificity)
```

True Negatives: 66  
False Positives: 11  
False Negatives: 2  
True Positives: 28  
0.8571428571428571

```
[ ]: sensitivity=tp/(fn+tp)  
print(sensitivity)
```

0.9333333333333333

```
[ ]: print(classification_report(y_test, y_pred))
```

	precision	recall	f1-score	support
0	0.97	0.86	0.91	77
1	0.72	0.93	0.81	30
accuracy			0.88	107
macro avg	0.84	0.90	0.86	107
weighted avg	0.90	0.88	0.88	107

# **RANDOM FOREST CLASSIFIER (RFC)**

# RFC Final

October 29, 2021

## 1 Algorithm: Random Forest Classifier(RFC)

```
[19]: import pandas as pd
```

```
[20]: df=pd.read_csv("/content/drive/MyDrive/PCOS/clean_data.csv")
```

```
[21]: X = df.drop(['PCOS (Y/N)'], axis=1)

y = df['PCOS (Y/N)']
```

## 2 Splitting Dataset

```
[22]: from sklearn.model_selection import train_test_split

X_train, X_test, y_train, y_test = train_test_split(X, y, test_size = 0.2,
↳random_state = 0)
```

```
[23]: X_train.shape, X_test.shape
```

```
[23]: ((427, 6), (107, 6))
```

```
[24]: cols = X_train.columns
```

### 2.1 Importing Necessary Libraries

```
[33]: from sklearn import metrics
from sklearn.model_selection import train_test_split
from sklearn import model_selection
from sklearn.ensemble import RandomForestClassifier

from sklearn.metrics import confusion_matrix
from sklearn.metrics import accuracy_score
from sklearn.metrics import classification_report
```

```
[34]: X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.33,
↳random_state=23)
```

## 2.2 RFC Implementation

```
[39]: rfc = RandomForestClassifier(n_estimators=1000)
```

```
rfc.fit(X_train,y_train)
# predictions
rfc_predict = rfc.predict(X_test)
```

```
[40]: print("Random forest model accuracy(in %):", metrics.accuracy_score(y_test,
↪rfc_predict)*100)
```

Random forest model accuracy(in %): 89.83050847457628

```
[43]: tn, fp, fn, tp = confusion_matrix(y_test, rfc_predict).ravel()
print("True Negatives: ",tn)
print("False Positives: ",fp)
print("False Negatives: ",fn)
print("True Positives: ",tp)
specificity = tn / (tn+fp)
print(specificity)
```

True Negatives: 112  
False Positives: 5  
False Negatives: 13  
True Positives: 47  
0.9572649572649573

```
[44]: sensitivity=tp/(fn+tp)
print(sensitivity)
```

0.7833333333333333

```
[41]: print(classification_report(y_test, rfc_predict))
```

	precision	recall	f1-score	support
0	0.90	0.96	0.93	117
1	0.90	0.78	0.84	60
accuracy			0.90	177
macro avg	0.90	0.87	0.88	177
weighted avg	0.90	0.90	0.90	177

# **K-NEAREST NEIGHBORS (KNN)**

# KNN

October 29, 2021

## 1 Algorithm: K Nearest Neighbour (KNN)

### 1.1 Importing Libraries

```
[1]: import numpy as np
import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
%matplotlib inline

from sklearn.preprocessing import StandardScaler
from sklearn.model_selection import train_test_split

from sklearn.neighbors import KNeighborsClassifier

from sklearn import metrics
from sklearn.metrics import confusion_matrix
from sklearn.metrics import accuracy_score
from sklearn.metrics import classification_report
```

```
[2]: df = pd.read_csv('/content/drive/MyDrive/clean_data.csv')
```

```
[3]: X = df.drop(columns=["PCOS (Y/N)"])
y = df["PCOS (Y/N)"].values
```

### 1.2 Feature Scaling

```
[4]: scaler = StandardScaler()
scaler.fit(df.drop('PCOS (Y/N)', axis=1))
scaled_features = scaler.transform(df.drop('PCOS (Y/N)', axis=1))
scaled_data = pd.DataFrame(scaled_features, columns = df.drop('PCOS (Y/N)', axis=1).columns)
```

```
[5]: x = scaled_data
y = df['PCOS (Y/N)']
```

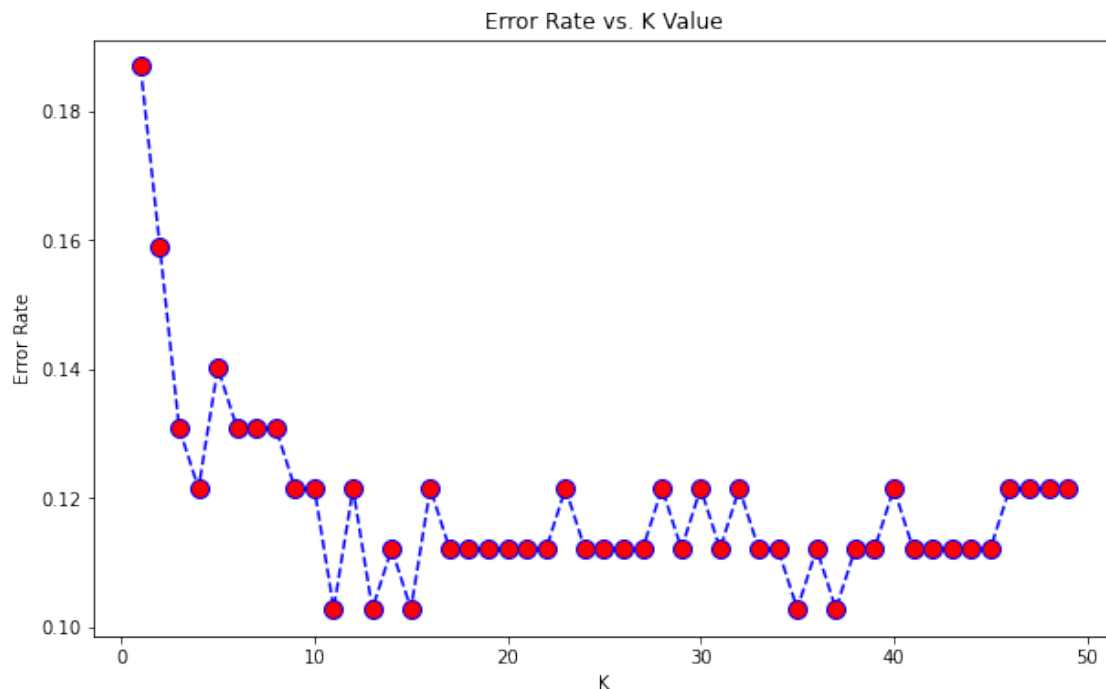
### 1.3 Splitting Dataset

```
[6]: x_training_data, x_test_data, y_training_data, y_test_data =  
      ↪train_test_split(x, y, test_size = 0.2, random_state=42)
```

### 1.4 KNN Implementation

```
[7]: #Selecting an optimal K value  
  
error_rates = []  
for i in np.arange(1, 50):  
    new_model = KNeighborsClassifier(n_neighbors = i)  
    new_model.fit(x_training_data, y_training_data)  
    new_predictions = new_model.predict(x_test_data)  
    error_rates.append(np.mean(new_predictions != y_test_data))  
  
plt.figure(figsize=(10,6))  
plt.plot(range(1,50),error_rates,color='blue', linestyle='dashed',  
         marker='o',markerfacecolor='red', markersize=10)  
plt.title('Error Rate vs. K Value')  
plt.xlabel('K')  
plt.ylabel('Error Rate')  
print("Minimum error:-",min(error_rates),"at K =",error_rates.  
      ↪index(min(error_rates)))
```

Minimum error:- 0.102803738317757 at K = 10



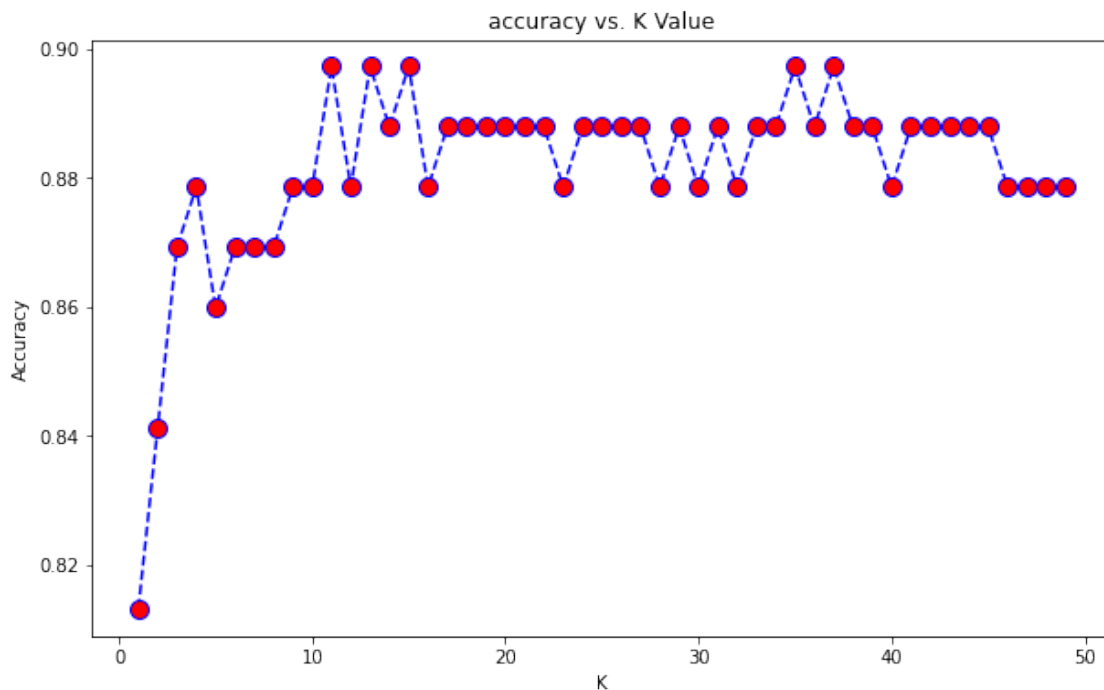


```
[19]: acc = []

for i in range(1,50):
    neigh = KNeighborsClassifier(n_neighbors = i).fit(x_training_data,
    ↪y_training_data)
    yhat = neigh.predict(x_test_data)
    acc.append(metrics.accuracy_score(y_test_data, yhat))

plt.figure(figsize=(10,6))
plt.plot(range(1,50),acc,color = 'blue',linestyle='dashed',
        marker='o',markerfacecolor='red', markersize=10)
plt.title('accuracy vs. K Value')
plt.xlabel('K')
plt.ylabel('Accuracy')
print("Maximum training accuracy:-",max(acc),"at K =",acc.index(max(acc)))
```

Maximum training accuracy:- 0.897196261682243 at K = 10



```
[13]: neigh = KNeighborsClassifier(n_neighbors = 10).fit(x_training_data,
    ↪y_training_data)
predictions = neigh.predict(x_test_data)
```

```
[20]: print("K Nearest Neighbour model for K = 10 testing accuracy(in %):", metrics.  
      ↪accuracy_score(y_test_data, predictions)*100)
```

K Nearest Neighbour model for K = 10 testing accuracy(in %): 87.85046728971963

```
[15]: print(classification_report(y_test_data, predictions))
```

	precision	recall	f1-score	support
0	0.88	0.96	0.92	75
1	0.88	0.69	0.77	32
accuracy			0.88	107
macro avg	0.88	0.82	0.84	107
weighted avg	0.88	0.88	0.87	107

```
[16]: tn, fp, fn, tp = confusion_matrix(y_test_data, predictions).ravel()  
      print("True Negatives: ",tn)  
      print("False Positives: ",fp)  
      print("False Negatives: ",fn)  
      print("True Positives: ",tp)
```

True Negatives: 72  
False Positives: 3  
False Negatives: 10  
True Positives: 22

```
[17]: specificity = tn / (tn+fp)  
      print(specificity)
```

0.96

```
[18]: sensitivity=tp/(fn+tp)  
      print(sensitivity)
```

0.6875

# **SUPPORT VECTOR MACHINES (SVM)**

# SVM\_PCOS (1)

October 29, 2021

## 1 Algorithm: Support Vector Machines

### 1.1 Importing Libraries

```
[1]: import pandas as pd
import matplotlib.pyplot as plt
import seaborn as sns
```

```
[3]: df=pd.read_csv("/content/drive/MyDrive/PCOS/clean_data.csv")
```

```
[4]: X = df.drop(['PCOS (Y/N)'], axis=1)

y = df['PCOS (Y/N)']
```

```
[5]: X.shape
```

```
[5]: (534, 6)
```

```
[6]: y.shape
```

```
[6]: (534,)
```

### 1.2 Splitting Dataset

```
[7]: from sklearn.model_selection import train_test_split

X_train, X_test, y_train, y_test = train_test_split(X, y, test_size = 0.2,
→random_state = 0)
```

```
[8]: # check the shape of X_train and X_test
```

```
X_train.shape, X_test.shape
```

```
[8]: ((427, 6), (107, 6))
```

```
[9]: cols = X_train.columns
```

### 1.3 Feature Scaling

```
[10]: # Feature Scaling
from sklearn.preprocessing import StandardScaler

scaler = StandardScaler()

X_train = scaler.fit_transform(X_train)

X_test = scaler.transform(X_test)
```

```
[11]: X_train = pd.DataFrame(X_train, columns=[cols])
```

```
[12]: X_test = pd.DataFrame(X_test, columns=[cols])
```

### 1.4 SVM Implementation

```
[13]: # 12. Run SVM with default hyperparameters

#Default hyperparameter means C=1.0, kernel=rbf and gamma=auto among other
↳ parameters.

# import SVC classifier
from sklearn.svm import SVC

# import metrics to compute accuracy
from sklearn.metrics import accuracy_score

# instantiate classifier with default hyperparameters
svc=SVC()

# fit classifier to training set
svc.fit(X_train,y_train)

# make predictions on test set
y_pred=svc.predict(X_test)

# compute and print accuracy score
print('Model accuracy score with default hyperparameters: {0:0.4f}'.
↳ format(accuracy_score(y_test, y_pred)))
```

Model accuracy score with default hyperparameters: 0.8879

## 1.5 Running SVM with Linear Kernel

Linear kernel is used when the data is linearly separable. It means that data can be separated using a single line. It is one of the most common kernels to be used. It is mostly used when there are large number of features in a dataset. Linear kernel is often used for text classification purposes.

Training with a linear kernel is usually faster, because we only need to optimize the C regularization parameter. When training with other kernels, we also need to optimize the  $\gamma$  parameter. So, performing a grid search will usually take more time.

```
[14]: # Run SVM with linear kernel and C=1.0
      # instantiate classifier with linear kernel and C=1.0
      linear_svc=SVC(kernel='linear', C=1.0)

      # fit classifier to training set
      linear_svc.fit(X_train,y_train)

      # make predictions on test set
      y_pred_test=linear_svc.predict(X_test)

      # compute and print accuracy score
      print('Model accuracy score with linear kernel and C=1.0 : {0:0.4f}'.format(
        accuracy_score(y_test, y_pred_test)))
```

Model accuracy score with linear kernel and C=1.0 : 0.8972

## 1.6 Checking for Overfitting or Underfitting

```
[15]: print('Training set score: {:.4f}'.format(linear_svc.score(X_train, y_train)))

      print('Test set score: {:.4f}'.format(linear_svc.score(X_test, y_test)))
```

Training set score: 0.9157

Test set score: 0.8972

```
[ ]: # The training-set accuracy score is 0.9157 while the test-set accuracy to be 0.8972.
      # These two values are quite comparable. So, there is no question of overfitting.
```

## 1.7 Running SVM with Polynomial Kernel

Polynomial kernel represents the similarity of vectors (training samples) in a feature space over polynomials of the original variables. The polynomial kernel looks not only at the given features of input samples to determine their similarity, but also combinations of the input samples.

```
[16]: # Run SVM with polynomial kernel and C=1.0
# instantiate classifier with polynomial kernel and C=1.0
poly_svc=SVC(kernel='poly', C=1.0)

# fit classifier to training set
poly_svc.fit(X_train,y_train)

# make predictions on test set
y_pred=poly_svc.predict(X_test)

# compute and print accuracy score
print('Model accuracy score with polynomial kernel and C=1.0 : {0:0.4f}'.format(accuracy_score(y_test, y_pred)))
```

Model accuracy score with polynomial kernel and C=1.0 : 0.9065

```
[17]: print('Training set score: {:.4f}'.format(poly_svc.score(X_train, y_train)))

print('Test set score: {:.4f}'.format(poly_svc.score(X_test, y_test)))
```

Training set score: 0.9321

Test set score: 0.9065

```
[19]: from sklearn.metrics import confusion_matrix

cm = confusion_matrix(y_test, y_pred)

print('Confusion matrix\n\n', cm)

print('\nTrue Positives(TP) = ', cm[0,0])

print('\nTrue Negatives(TN) = ', cm[1,1])

print('\nFalse Positives(FP) = ', cm[0,1])

print('\nFalse Negatives(FN) = ', cm[1,0])
```

Confusion matrix

```
[[69  2]
 [ 8 28]]
```

True Positives(TP) = 69

True Negatives(TN) = 28

False Positives(FP) = 2

False Negatives(FN) = 8

```
[21]: from sklearn.metrics import confusion_matrix
      from sklearn.metrics import accuracy_score
      from sklearn.metrics import classification_report
      print(classification_report(y_test, y_pred))
```

	precision	recall	f1-score	support
0	0.90	0.97	0.93	71
1	0.93	0.78	0.85	36
accuracy			0.91	107
macro avg	0.91	0.87	0.89	107
weighted avg	0.91	0.91	0.90	107

```
[23]: sensitivity=cm[0,0]/(cm[1,0]+cm[0,0])
      print(sensitivity)
```

0.8961038961038961

```
[25]: specificity = cm[1,1] / (cm[1,1]+cm[0,1])
      print(specificity)
```

0.9333333333333333

## 1.8 Running SVM with Sigmoid Kernel

Sigmoid kernel has its origin in neural networks. We can use it as the proxy for neural networks.

```
[ ]: # Run SVM with sigmoid kernel and C=1.0
      # instantiate classifier with sigmoid kernel and C=1.0
      sigmoid_svc=SVC(kernel='sigmoid', C=1.0)

      # fit classifier to training set
      sigmoid_svc.fit(X_train,y_train)

      # make predictions on test set
      y_pred=sigmoid_svc.predict(X_test)

      # compute and print accuracy score
      print('Model accuracy score with sigmoid kernel and C=1.0 : {0:0.4f}'.format(accuracy_score(y_test, y_pred)))
```



Model accuracy score with sigmoid kernel and C=1.0 : 0.8692

```
[ ]: from sklearn.metrics import confusion_matrix

cm = confusion_matrix(y_test, y_pred_test)

print('Confusion matrix\n\n', cm)

print('\nTrue Positives(TP) = ', cm[0,0])

print('\nTrue Negatives(TN) = ', cm[1,1])

print('\nFalse Positives(FP) = ', cm[0,1])

print('\nFalse Negatives(FN) = ', cm[1,0])
```

Confusion matrix

```
[[69  2]
 [ 9 27]]
```

True Positives(TP) = 69

True Negatives(TN) = 27

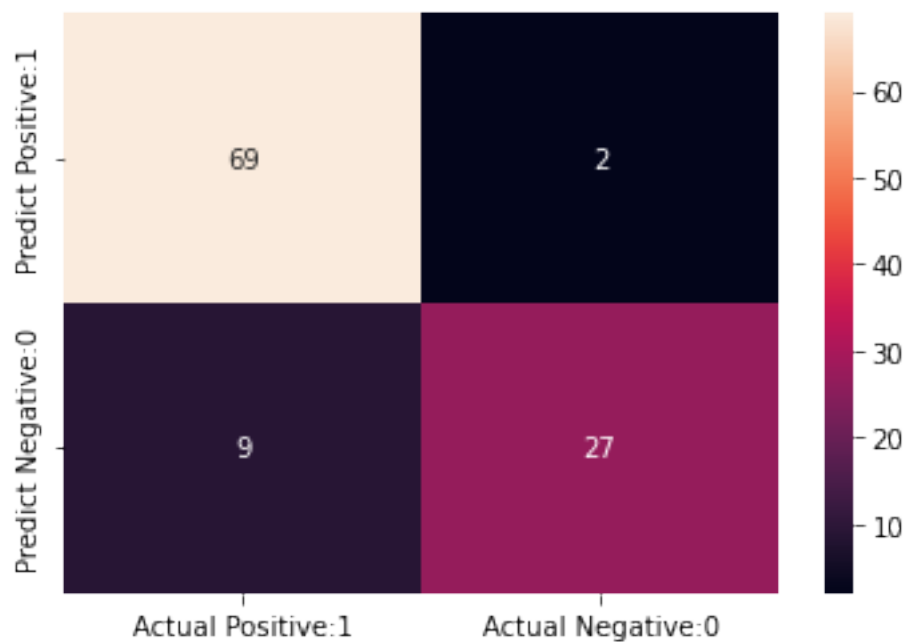
False Positives(FP) = 2

False Negatives(FN) = 9

```
[ ]: cm_matrix = pd.DataFrame(data=cm, columns=['Actual Positive:1', 'Actual_
↪Negative:0'],
                                index=['Predict Positive:1', 'Predict Negative:
↪0'])

sns.heatmap(cm_matrix, annot=True)
```

```
[ ]: <matplotlib.axes._subplots.AxesSubplot at 0x7f7ac552c850>
```



```
[ ]: # Classification report for Poly Kernel
from sklearn.metrics import classification_report
print(classification_report(y_test, y_pred_test))
```

	precision	recall	f1-score	support
0	0.88	0.97	0.93	71
1	0.93	0.75	0.83	36
accuracy			0.90	107
macro avg	0.91	0.86	0.88	107
weighted avg	0.90	0.90	0.89	107

# XGBOOST

# Xgboost\_PCOS

October 29, 2021

## 1 Algorithm: eXtreme Gradient Boosting(XGBoost)

### 1.1 Importing Libraries

```
[1]: import pandas as pd
import matplotlib.pyplot as plt
from numpy import loadtxt
from xgboost import XGBClassifier
from sklearn.model_selection import train_test_split
from sklearn.metrics import accuracy_score
from sklearn.metrics import confusion_matrix
from sklearn.metrics import accuracy_score
from sklearn.metrics import classification_report
```

```
[2]: df = pd.read_csv('/content/drive/MyDrive/PCOS/clean_data.csv')
df.shape
```

```
[2]: (534, 7)
```

```
[3]: X = df.drop(columns=["PCOS (Y/N)"])
y = df["PCOS (Y/N)"].values
```

### 1.2 Splitting Dataset

```
[38]: seed = 13
test_size = 0.2
X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=test_size,
↪random_state=seed)
```

### 1.3 Feature Scaling

```
[39]: # Feature Scaling
from sklearn.preprocessing import StandardScaler

scaler = StandardScaler()

X_train = scaler.fit_transform(X_train)
```

```
X_test = scaler.transform(X_test)
```

## 1.4 XGBoost Implementation

XGBoost (eXtreme Gradient Boosting) is an advanced implementation of gradient boosting algorithm.

The overall parameters have been divided into 3 categories by XGBoost authors:

- General Parameters: Guide the overall functioning
- Booster Parameters: Guide the individual booster (tree/regression) at each step
- Learning Task Parameters: Guide the optimization performed

```
[40]: model = XGBClassifier(learning_rate=0.1, n_estimators=1000, max_depth=5,
min_child_weight=1, gamma=0, subsample=0.8,
colsample_bytree=0.8, objective='binary:logistic',
nthread=4, scale_pos_weight=1, seed=27)
model.fit(X_train, y_train)
```

```
[40]: XGBClassifier(base_score=0.5, booster='gbtree', colsample_bylevel=1,
colsample_bynode=1, colsample_bytree=0.8, gamma=0,
learning_rate=0.1, max_delta_step=0, max_depth=5,
min_child_weight=1, missing=None, n_estimators=1000, n_jobs=1,
nthread=4, objective='binary:logistic', random_state=0,
reg_alpha=0, reg_lambda=1, scale_pos_weight=1, seed=27,
silent=None, subsample=0.8, verbosity=1)
```

### 1. General Parameters:

**booster** [default=gbtree] Select the type of model to run at each iteration. It has 2 options: gbtree: tree-based models gblinear: linear models

**silent** [default=0]: Silent mode is activated is set to 1, i.e. no running messages will be printed. It's generally good to keep it 0 as the messages might help in understanding the model.

**nthread** [default to maximum number of threads available if not set] This is used for parallel processing and number of cores in the system should be entered If you wish to run on all cores, value should not be entered and algorithm will detect automatically

### 2. Booster Parameters

**eta** [default=0.3] Analogous to learning rate in GBM Makes the model more robust by shrinking the weights on each step Typical final values to be used: 0.01-0.2

**min\_child\_weight** [default=1] Defines the minimum sum of weights of all observations required in a child. This is similar to min\_child\_leaf in GBM but not exactly. This refers to min “sum of weights” of observations while GBM has min “number of observations”. Used to control over-fitting. Higher values prevent a model from learning relations which might be highly specific to the particular sample selected for a tree. Too high values can lead to under-fitting hence, it should be tuned using CV.

**max\_depth** [default=6] The maximum depth of a tree, same as GBM. Used to control over-fitting as higher depth will allow model to learn relations very specific to a particular sample. Should be

tuned using CV. Typical values: 3-10 `max_leaf_nodes` The maximum number of terminal nodes or leaves in a tree. Can be defined in place of `max_depth`. Since binary trees are created, a depth of 'n' would produce a maximum of  $2^n$  leaves. If this is defined, GBM will ignore `max_depth`.

`gamma` [default=0] A node is split only when the resulting split gives a positive reduction in the loss function. Gamma specifies the minimum loss reduction required to make a split. Makes the algorithm conservative. The values can vary depending on the loss function and should be tuned.

`max_delta_step` [default=0] In maximum delta step we allow each tree's weight estimation to be. If the value is set to 0, it means there is no constraint. If it is set to a positive value, it can help making the update step more conservative. Usually this parameter is not needed, but it might help in logistic regression when class is extremely imbalanced. This is generally not used but you can explore further if you wish.

`subsample` [default=1] Same as the subsample of GBM. Denotes the fraction of observations to be randomly samples for each tree. Lower values make the algorithm more conservative and prevents overfitting but too small values might lead to under-fitting. Typical values: 0.5-1

`colsample_bytree` [default=1] Similar to `max_features` in GBM. Denotes the fraction of columns to be randomly samples for each tree. Typical values: 0.5-1

`colsample_bylevel` [default=1] Denotes the subsample ratio of columns for each split, in each level. don't use this often because `subsample` and `colsample_bytree` will do the job for you. but you can explore further if you feel so.

`lambda` [default=1] L2 regularization term on weights (analogous to Ridge regression) This used to handle the regularization part of XGBoost. Though many data scientists don't use it often, it should be explored to reduce overfitting.

`alpha` [default=0] L1 regularization term on weight (analogous to Lasso regression) Can be used in case of very high dimensionality so that the algorithm runs faster when implemented  
`scale_pos_weight` [default=1] A value greater than 0 should be used in case of high class imbalance as it helps in faster convergence.

### 3. Learning Task Parameters

`objective` [default=reg:linear] This defines the loss function to be minimized. Mostly used values are: `binary:logistic` –logistic regression for binary classification, returns predicted probability (not class) `multi:softmax` –multiclass classification using the softmax objective, returns predicted class (not probabilities) you also need to set an additional `num_class` (number of classes) parameter defining the number of unique classes `multi:softprob` –same as softmax, but returns predicted probability of each data point belonging to each class.

`eval_metric` [ default according to objective ] The metric to be used for validation data. The default values are rmse for regression and error for classification. Typical values are: rmse – root mean square error mae – mean absolute error logloss – negative log-likelihood error – Binary classification error rate (0.5 threshold) merror – Multiclass classification error rate mlogloss – Multiclass logloss auc: Area under the curve

`seed` [default=0] The random number seed. Can be used for generating reproducible results and also for parameter tuning.

```
[41]: y_pred = model.predict(X_test)
      predictions = [round(value) for value in y_pred]
```

```
[42]: accuracy = accuracy_score(y_test, predictions)
      print("Accuracy: %.2f%%" % (accuracy * 100.0))
```

Accuracy: 90.65%

```
[43]: tn, fp, fn, tp = confusion_matrix(y_test, y_pred).ravel()
      print("True Negatives: ",tn)
      print("False Positives: ",fp)
      print("False Negatives: ",fn)
      print("True Positives: ",tp)
      specificity = tn / (tn+fp)
      print(specificity)
```

True Negatives: 68  
 False Positives: 6  
 False Negatives: 4  
 True Positives: 29  
 0.918918918918919

```
[45]: sensitivity=tp/(fn+tp)
      print(sensitivity)
```

0.8787878787878788

```
[44]: print(classification_report(y_test, y_pred))
```

	precision	recall	f1-score	support
0	0.94	0.92	0.93	74
1	0.83	0.88	0.85	33
accuracy			0.91	107
macro avg	0.89	0.90	0.89	107
weighted avg	0.91	0.91	0.91	107

**MULTI-  
LAYER  
PERCEPTRON  
(MLP  
CLASSIFIER)**



# MLP\_PCOS

October 29, 2021

## 1 Algorithm: Multilayer Perceptron Classifier (MLP)

A multilayer perceptron is a type of feed-forward artificial neural network that generates a set of outputs from a set of inputs

MLPClassifier trains iteratively since at each time step the partial derivatives of the loss function with respect to the model parameters are computed to update the parameters.

It can also have a regularization term added to the loss function that shrinks model parameters to prevent overfitting.

This implementation works with data represented as dense numpy arrays or sparse scipy arrays of floating point values.

```
[23]: import pandas as pd
```

```
[24]: df=pd.read_csv("/content/drive/MyDrive/PCOS/clean_data.csv")
```

```
[25]: X = df.drop(['PCOS (Y/N)'], axis=1)

      y = df['PCOS (Y/N)']
```

### 1.1 Splitting Data

```
[26]: from sklearn.model_selection import train_test_split

      X_train, X_test, y_train, y_test = train_test_split(X, y, test_size = 0.2,
      ↪random_state = 0)
```

```
[27]: X_train.shape, X_test.shape
```

```
[27]: ((427, 6), (107, 6))
```

```
[28]: cols = X_train.columns
```

## 1.2 Feature Scaling

```
[29]: # Feature Scaling
      from sklearn.preprocessing import StandardScaler

      scaler = StandardScaler()

      X_train = scaler.fit_transform(X_train)

      X_test = scaler.transform(X_test)

[30]: X_train = pd.DataFrame(X_train, columns=[cols])

[31]: X_test = pd.DataFrame(X_test, columns=[cols])

[38]: def test_results(model, X_test, y_test):
      from sklearn.metrics import confusion_matrix
      y_pred = model.predict(X_test)
      tn, fp, fn, tp = confusion_matrix(y_test, y_pred).ravel()

      accuracy = (tp + tn)/(tp + fp + tn + fn)
      print("Accuracy: ", '{:.2f}'.format(accuracy * 100))
      print("True Negative:", tn)
      print("True Positive:", tp)
      print("False Positive:", fp)
      print("False Negative:", fn)
      print()
      print("-----")
      print("Negative Class Results")
      precision = (tp / (tp + fp))
      recall = (tp / (tp + fn))
      f1_score = (2 * (precision * recall) / (precision + recall))
      print("Precision (N): ", '{:.2f}'.format(precision * 100))
      print("Recall (N): ", '{:.2f}'.format(recall * 100))
      print("F1 Score (N):" , '{:.2f}'.format(f1_score * 100))
      print()
      print("-----")
      print("Positive Class Results")
      precision = (tn / (tn + fn))
      recall = (tn / (tn + fp))
      f1_score = (2 * (precision * recall) / (precision + recall))
      print("Precision (P): ", '{:.2f}'.format(precision * 100))
      print("Recall (P): ", '{:.2f}'.format(recall * 100))
      print("F1 Score (P):" , '{:.2f}'.format(f1_score * 100))
      print("Specificity:")
      specificity = tn / (tn+fp)
      print(specificity)
```

```
print("Sensitivity:")
sensitivity=tp/(fn+tp)
print(sensitivity)
```

### 1.3 Importing MLP Classifier From SKLearn

We can relate this model with the Keras Sequential Model.

```
[33]: from sklearn.neural_network import MLPClassifier
```

```
[34]: mlp = MLPClassifier(hidden_layer_sizes=(60, 60), activation='tanh',
    ↪ solver='adam',
    alpha=0.0001, learning_rate='adaptive',
    ↪ learning_rate_init=0.001,
    power_t=0.5, max_iter=200, shuffle=False, random_state=0,
    tol=0.0001, verbose=False, warm_start=False, momentum=0.9,
    nesterovs_momentum=True, early_stopping=False,
    ↪ validation_fraction=0.1,
    beta_1=0.9, beta_2=0.999, epsilon=1e-08,
    ↪ n_iter_no_change=10, max_fun=15000)
```

Parameters:

**hidden\_layer\_sizes**tuple, length = n\_layers - 2, default=(100,) The ith element represents the number of neurons in the ith hidden layer.

**activation**{‘identity’, ‘logistic’, ‘tanh’, ‘relu’}, default=‘relu’ Activation function for the hidden layer.

‘identity’, no-op activation, useful to implement linear bottleneck, returns  $f(x) = x$

‘logistic’, the logistic sigmoid function, returns  $f(x) = 1 / (1 + \exp(-x))$ .

‘tanh’, the hyperbolic tan function, returns  $f(x) = \tanh(x)$ .

‘relu’, the rectified linear unit function, returns  $f(x) = \max(0, x)$

**solver**{‘lbfgs’, ‘sgd’, ‘adam’}, default=‘adam’ The solver for weight optimization.

‘lbfgs’ is an optimizer in the family of quasi-Newton methods.

‘sgd’ refers to stochastic gradient descent.

‘adam’ refers to a stochastic gradient-based optimizer proposed by Kingma, Diederik, and Jimmy Ba

Note: The default solver ‘adam’ works pretty well on relatively large datasets (with thousands of training samples or more) in terms of both training time and validation score. For small datasets, however, ‘lbfgs’ can converge faster and perform better.

**alpha**float, default=0.0001 L2 penalty (regularization term) parameter.

**learning\_rate**{‘constant’, ‘invscaling’, ‘adaptive’}, default=‘constant’ Learning rate schedule for weight updates.

‘constant’ is a constant learning rate given by ‘learning\_rate\_init’.

‘invscaling’ gradually decreases the learning rate at each time step ‘t’ using an inverse scaling exponent of ‘power\_t’.  $\text{effective\_learning\_rate} = \text{learning\_rate\_init} / \text{pow}(t, \text{power\_t})$

‘adaptive’ keeps the learning rate constant to ‘learning\_rate\_init’ as long as training loss keeps decreasing. Each time two consecutive epochs fail to decrease training loss by at least tol, or fail to increase validation score by at least tol if ‘early\_stopping’ is on, the current learning rate is divided by 5.

Only used when solver=‘sgd’.

**learning\_rate\_init**double, default=0.001 The initial learning rate used. It controls the step-size in updating the weights. Only used when solver=‘sgd’ or ‘adam’.

**power\_t**double, default=0.5 The exponent for inverse scaling learning rate. It is used in updating effective learning rate when the learning\_rate is set to ‘invscaling’. Only used when solver=‘sgd’.

**max\_iter**int, default=200 Maximum number of iterations. The solver iterates until convergence (determined by ‘tol’) or this number of iterations. For stochastic solvers (‘sgd’, ‘adam’), note that this determines the number of epochs (how many times each data point will be used), not the number of gradient steps.

**nesterovs\_momentum**bool, default=True Whether to use Nesterov’s momentum. Only used when solver=‘sgd’ and momentum > 0.

**validation\_fraction**float, default=0.1 The proportion of training data to set aside as validation set for early stopping. Must be between 0 and 1. Only used if early\_stopping is True.

**beta\_1**float, default=0.9 Exponential decay rate for estimates of first moment vector in adam, should be in [0, 1). Only used when solver=‘adam’.

**beta\_2**float, default=0.999 Exponential decay rate for estimates of second moment vector in adam, should be in [0, 1). Only used when solver=‘adam’.

**epsilon**float, default=1e-8 Value for numerical stability in adam. Only used when solver=‘adam’.

**n\_iter\_no\_change**int, default=10 Maximum number of epochs to not meet tol improvement. Only effective when solver=‘sgd’ or ‘adam’.

```
[35]: mlp.fit(X_train, y_train)
```

```
/usr/local/lib/python3.7/dist-  
packages/sklearn/neural_network/_multilayer_perceptron.py:571:  
ConvergenceWarning: Stochastic Optimizer: Maximum iterations (200) reached and  
the optimization hasn't converged yet.  
% self.max_iter, ConvergenceWarning)
```

```
[35]: MLPClassifier(activation='tanh', alpha=0.0001, batch_size='auto', beta_1=0.9,  
                  beta_2=0.999, early_stopping=False, epsilon=1e-08,  
                  hidden_layer_sizes=(60, 60), learning_rate='adaptive',  
                  learning_rate_init=0.001, max_fun=15000, max_iter=200,  
                  momentum=0.9, n_iter_no_change=10, nesterovs_momentum=True,
```

```
power_t=0.5, random_state=0, shuffle=False, solver='adam',
tol=0.0001, validation_fraction=0.1, verbose=False,
warm_start=False)
```

```
[39]: test_results(mlp, X_test, y_test)
```

```
Accuracy: 92.52
True Negative: 69
True Positive: 30
False Positive: 2
False Negative: 6
```

```
-----
Negative Class Results
Precision (N): 93.75
Recall (N): 83.33
F1 Score (N): 88.24
```

```
-----
Positive Class Results
Precision (P): 92.00
Recall (P): 97.18
F1 Score (P): 94.52
Specificity:
0.971830985915493
Sensitivity:
0.8333333333333334
```

## 1.4 Trying Activation Function To Be ReLu

```
[46]: mlp1 = MLPClassifier(hidden_layer_sizes=(60, 60), activation='relu',
    ↪ solver='adam',
    ↪ alpha=0.0001, learning_rate='adaptive',
    ↪ learning_rate_init=0.001,
    ↪ power_t=0.5, max_iter=200, shuffle=False, random_state=0,
    ↪ tol=0.0001, verbose=False, warm_start=False, momentum=0.9,
    ↪ nesterovs_momentum=True, early_stopping=False,
    ↪ validation_fraction=0.1,
    ↪ beta_1=0.9, beta_2=0.999, epsilon=1e-08,
    ↪ n_iter_no_change=10, max_fun=15000)
```

```
[47]: mlp1.fit(X_train, y_train)
```

```
/usr/local/lib/python3.7/dist-
packages/sklearn/neural_network/_multilayer_perceptron.py:571:
ConvergenceWarning: Stochastic Optimizer: Maximum iterations (200) reached and
the optimization hasn't converged yet.
  % self.max_iter, ConvergenceWarning)
```

```
[47]: MLPClassifier(activation='relu', alpha=0.0001, batch_size='auto', beta_1=0.9,
                    beta_2=0.999, early_stopping=False, epsilon=1e-08,
                    hidden_layer_sizes=(60, 60), learning_rate='adaptive',
                    learning_rate_init=0.001, max_fun=15000, max_iter=200,
                    momentum=0.9, n_iter_no_change=10, nesterovs_momentum=True,
                    power_t=0.5, random_state=0, shuffle=False, solver='adam',
                    tol=0.0001, validation_fraction=0.1, verbose=False,
                    warm_start=False)
```

```
[48]: test_results(mlp1, X_test, y_test)
```

```
Accuracy: 87.85
True Negative: 67
True Positive: 27
False Positive: 4
False Negative: 9
```

```
-----
Negative Class Results
Precision (N): 87.10
Recall (N): 75.00
F1 Score (N): 80.60
```

```
-----
Positive Class Results
Precision (P): 88.16
Recall (P): 94.37
F1 Score (P): 91.16
Specificity:
0.9436619718309859
Sensitivity:
0.75
```

# RESULTS

## Author's Results

Algorithm	Accuracy	Sensitivity	Specificity	Precision	F1 Score
Logistic regression	0.8536	0.6451	0.98039	0.952380	0.3845
Gaussian Naive Bayes	0.8414	0.7419	0.90196	0.82142	0.3898
Random Forest Classifier	0.8902	0.7419	0.98039	0.95833	0.4182
K-Nearest neighbors	0.8658	0.8064	0.90196	0.833333	0.4098
Support Vector Machines	0.8292	0.5483	1.0	1.0	0.3541

## Our Results

Algorithm	Accuracy	Sensitivity	Specificity	Precision	F1 Score
Logistic regression	90.6542056	0.9	0.90909	0.91	0.91
Gaussian Naive Bayes	87.850467	0.93333	0.85714	0.90	0.88
Random Forest Classifier	89.83050	0.78333	0.957264	0.90	0.90
K-Nearest neighbors	87.850467	0.6875	0.96	0.88	0.87
Support Vector Machines	90.65	0.896103	0.933333	0.91	0.91

## Our Results- Extra Algorithms Implemented

Algorithm	Accuracy	Sensitivity	Specificity	Precision	F1 Score
XGBoost	90.65	0.878787	0.918918	0.91	0.91
MLP Classifier	92.52	0.833333	0.9718309	93.75	88.24

# Changes and modifications made:

- Our dataset is slightly different as some columns such as 'Diabetes' are missing.
- Where their dataset has a single parameter, ours has the same one split in two. For example, Follicle No. in theirs, Follicle No. (L) and Follicle No. (R) in ours
- The features we have extracted are through EDA by checking the heatmap and correlation matrix. They have extracted features mainly through SPSS software and through discussions with experts in the field.

## Conclusion:

Out of the 43 features from clinical and metabolic test results, 6 potential features were identified. Classification of PCOS with the feature set transformed using standardisation is done using various machine learning techniques such as Naïve Bayes, Logistic Regression, K-Nearest neighbor (KNN), Random Forest Classifier (RFC), Support Vector Machine (SVM), XGBOOST and MLP in Python. Results revealed that the most suitable and accurate method for the PCOS prediction is MLP with an accuracy of almost 92.07%.

This automated system, that we aim to build in the future, can act as an assistive tool for doctors and save considerable time in examining patients and hence reducing delay in diagnosing the risk of PCOS.