# DDI assessment for examplinib, M1

### Rainer Strotmann

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# Introduction

This assessment of the perpetrator risks for xxx is based on the relevant FDA and EMA guidance documents (FDA 2020 and EMA 2012).

This output was generated using version xx (dd-dd-dddd) of the DDI assessment R script.

# Drug properties

The following physico-chemical, ADME and clinical exposure data were used for the calculation of the maximal gut and portal vein concentrations, and the unbound systemic concentrations.

Table 1: Compound parameters for examplinib

parameter	value	source
$\overline{MW}$ (g/mol)	492.6	
dose  (mg)	450	clinical dose
$C_{max,ss}$ (ng/ml)	3530	study $001$
$f_u$	0.023	study $002$
$f_{u,mic}$	1	default
$R_B$	1	study $003$
$F_a$	0.81	study $003$
$F_g$	1	default
$k_a (1/\min)$	0.00267	unknown

Table 2: Compound parameters for M1

parameter	value	source
MW (g/mol)	506.56	
dose (mg)	NA	
$C_{max,ss}$ (ng/ml)	1038	study 001
$f_u$	0.012	study 002
$f_{u,mic}$	1	default
$R_B$	1	study 002
$F_a$	NA	
$F_g$	NA	
$k_a (1/\min)$	NA	

# Key perpetrator concentrations

The relevant perpetrator concentrations are calculated as outlined in the Appendix:

Table 3: Key perpetrator concentrations for examplinib

parameter	value (ng/ml)	value (µM)
$\overline{I_{gut}}$	1800000.0	3654.080
$I_{max,ss,u}$	81.2	0.165
$I_{max,inlet,u}$	95.0	0.193
$I_{max,intestinal,u}$	74.6	0.151

Table 4: Key perpetrator concentrations for M1

parameter	value (ng/ml)	value (µM)
$\overline{I_{gut}}$	0.0	0.0000
$I_{max,ss,u}$	12.5	0.0246
$I_{max,inlet,u}$	12.5	0.0246
$I_{max,intestinal,u}$	12.5	0.0246

# DDI perpetrator risk assessment

## Basic modeling of CYP inhibition

#### Reversible inhibition

As per the FDA guideline (FDA, 2020), the relevant metric for the basic modeling of the direct CYP inhibition risk is  $R = 1 + [I]/K_{i,u}$  with the relevant inhibitor concentration [I] being  $I_{max,ss,u}$  for hepatic CYP enzymes and  $I_{gut}$  for intestinal CYP enzymes.

Thresholds of R < 1.02 and R < 11 apply for hepatic and intestinal enzymes, respectively. Note that intestinal CYP inhibition is only evaluated for CYP3A4.

Basic modeling as per the FDA guideline results in the following risk assessment for direct CYP inhibition by xxx.

Table 5: Risk for direct CYP inhibition by examplinib (basic model)

CYP	$K_i$	$K_{i,u}$ .	$R_1$	risk (hepatic)	$R_{1,gut}$	risk (intestinal)
CYP1A2	NA	NA I	NA	NA	NA	NA
CYP2B6	NA	NA I	NA	NA	NA	NA
CYP2C8	11.0	11.0	1.015	FALSE	NA	NA
CYP2C9	13.5	13.5 1	1.012	FALSE	NA	NA
CYP2C19	15.0	15.0 1	1.011	FALSE	NA	NA
CYP2D6	NA	NA I	NA	NA	NA	NA
CYP3A4	12.5	12.5 1	1.013	FALSE	293.3	TRUE

Table 6: Risk for direct CYP inhibition by M1 (basic model)

CYP	$K_i$	$K_{i,u}$	$R_1$	risk (hepatic)	$R_{1,gut}$	risk (intestinal)
CYP2C9	4.4	4.4	1.006	FALSE	NA	NA

Note that the ratios used in the EMA guidance correspond to the FDA ratios listed above minus 1.

## Modeling of CYP induction

#### Basic/fold-change method

The basic (EMA) or fold-change (FDA) methods evaluate whether the maximal fold-change in mRNA expression is > 2-fold at the expected unbound hepatic concentration of the drug.

For the relevant drug concentrations, the FDA guidance suggests considering  $30 * I_{max,ss,u}$  while the EMA guidance considers  $50 * I_{max,ss,u}$  for hepatic and  $0.1 * I_{gut}$  for intestinal induction. It is expected that the concentrations in the respective in vitro assays cover these concentrations.

Basic modeling as per the FDA guideline results in the following risk assessment:

### Mechanistic static modeling of CYP modulation

As per the FDA guideline (FDA 2020), the relevant metric for mechanistic static modeling of the CYP inhibition potential is the predicted AUC ratio (AUCR) for specific probe substrates. A cut-off of R < 1.25 applies.

In the current implementation of this tool (and listed in the table below), only reversible inhibition and induction (as reflected in the A- and C-terms of the AUCR formula, see Appendix) is included. In general, the FDA guidelines advises to investigate mechanistic static models of inhibition (reversible and mechanism-dependent) and induction separately.

#### CYP inhibition only

Table 7: Mechanistic static modeling of CYP inhibition risk for examplinib

CYP	$K_{i,u}$	substrate	$F_{gut}$	$f_m$	$f_{m,CYP}$	$A_g$	$A_h$	$C_g$	$C_h$	AUC	R risk
CYP2C8	11.0	repaglinide	1.00	1.00	0.61	0.986	0.983	1	1	1.01	FALSE
CYP2C9	13.5	S-	1.00	1.00	0.91	0.989	0.986	1	1	1.01	FALSE
		warfarin									
CYP2C19	15.0	omegrazole	1.00	1.00	0.87	0.990	0.987	1	1	1.01	FALSE
CYP3A4	12.5	midazolam	0.57	0.96	1.00	0.988	0.985	1	1	1.02	FALSE

Table 8: Mechanistic static modeling of CYP inhibition risk for M1

CYP	$K_{i,u}$	substrate	$F_{gut}$	$f_m$	$f_{m,CYP}$	$A_g$	$A_h$	$C_g$	$C_h$	AUCI	R risk
CYP2C9	4.4	S-	1	1	0.91	0.994	0.994	1	1	1.01	FALSE
		warfarin									

#### CYP inhibition and induction

Table 9: Mechanistic static modeling of CYP inhibition risk for examplinib

CYP	$K_{i,u}$	substrate	$F_{gut}$	$f_m$	$f_{m,CYP}$	$A_g$	$A_h$	$C_g$	$C_h$	AUCR	risk
CYP2C8	11.0	repaglinide	1.00	1.00	0.61	0.986	0.983	1.00	1.00	1.011	FALSE
CYP2C9	13.5	S- warfarin	1.00	1.00	0.91	0.989	0.986	1.00	1.00	1.013	FALSE
CYP2C19	15.0	ometrazole	1.00	1.00	0.87	0.990	0.987	1.00	1.00	1.011	FALSE
CYP3A4	12.5	midazolam	0.57	0.96	1.00	0.988	0.985	1.62	1.77	0.463	TRUE

Table 10: Mechanistic static modeling of CYP inhibition risk for M1

CYP	$K_{i,u}$	substrate	$F_{gut}$	$f_m$	$f_{m,CYP}$	$A_g$	$A_h$	$C_g$	$C_h$	AUC	R risk
CYP2C9	4.4	S- warfarin	1	1	0.91	0.994	0.994	1	1	1.01	FALSE

## Basic modeling of UGT inhibition

The relevant metric for basic modeling of the UGT inhibition risk is  $R_1 = I_{max,ss,u}/K_{i,u}$ . For the clinical risk assessment, a cut-off of R < 1.02 applies.

In in vitro UGT inhibition studies, ususally  $IC_{50}$  rather than  $k_i$  values are determined. Assuming that a substrate concentration close to  $K_m$  is used,  $K_i$  is calculated as  $K_i = IC_{50}/2$  (refer to Cheng, Prusoff 1973).

Basic modeling as per the FDA guideline (FDA 2020) results in the following risk assessment:

Table 11: Risk for UGT inhibition by examplinib (basic model)

UGT	$K_{i,u}$	$R_1$	risk
UGT1A1	7.50	1.021976	TRUE
UGT1A3	7.50	1.021976	TRUE
UGT1A4	7.50	1.021976	TRUE
UGT1A6	7.50	1.021976	TRUE
UGT1A9	1.90	1.086747	TRUE
UGT2B7	7.50	1.021976	TRUE
UGT2B15	7.50	1.021976	TRUE
UGT2B17	3.05	1.054039	TRUE

Table 12: Risk for UGT inhibition by M1 (basic model)

UGT	$K_{i,u}$	$R_1$	risk
UGT1A1	0.55	1.044708	TRUE
UGT1A3	2.90	1.008479	FALSE
UGT1A4	3.10	1.007932	FALSE
UGT1A6	7.50	1.003279	FALSE
UGT1A9	1.80	1.013661	FALSE
UGT2B7	7.50	1.003279	FALSE
UGT2B15	4.80	1.005123	FALSE

# Inhibition of drug transporters

The relevant metric for the assessment of transporter interactions is  $R = [I]/K_i$ . In in vitro transporter inhibition studies,  $IC_{50}$  values are experimentally determined. Since the transporter substrate concentration is ususally kept very low in relation to  $K_m$  in order to minimze passive permeation,  $K_i = IC_{50}$  can be assumed.

The relevant perpetrator concentrations [I] are:  $I_{gut}$  for intestinal P-gp and BRCR,  $I_{max,inlet,u}$  for the hepatic basolateral transporters OCT1, OATP1B1 and OATP1B3, and  $I_{max,ss,u}$  for the renal basolateral transporters OAT1, OAT3 and OCT2, as well as the apical transporters outside the intestinal mucosa, i.e., hepatic P-gp and BCRP, and MATE1, MATE2-k.

The FDA and EMA guidelines differ in their threshold definitions. The risk assessments according to both guidelines are presented below:

Table 13: Risk for drug transporter inhibition by examplinib

transporter	$IC_{50}$	source	R	thld FDA	risk FDA	thld EMA	risk EMA
Pgp_int	0.41	study 005	8912.391	10.0	TRUE	10.00	TRUE
$Pgp\_sys$	0.41	study $005$	0.402	0.1	TRUE	0.02	TRUE
BCRP_int	1.90	study $005$	1923.200	10.0	TRUE	10.00	TRUE
$BCRP\_sys$	1.90	study $005$	0.087	0.1	FALSE	0.02	TRUE
OCT1	2.30	study 006	0.084	NA	NA	0.04	TRUE
OATP1B1	177.00	study $006$	0.001	0.1	FALSE	0.04	FALSE
OATP1B3	35.00	study $006$	0.006	0.1	FALSE	0.04	FALSE
OAT1	271.00		0.001	0.1	FALSE	0.04	FALSE
OAT3	300.00		0.001	0.1	FALSE	0.04	FALSE
BSEP	12.80		0.013	0.1	FALSE	0.02	FALSE
OCT2	67.00	study $006$	0.002	0.1	FALSE	0.02	FALSE
MATE1	3.60	study 006	0.046	0.1	FALSE	0.02	TRUE
MATE2k	1.10	study 006	0.150	0.1	TRUE	0.02	TRUE

# Appendix 1: Calculations and formulae

Gut concentration As per the FDA guideline, the maximal gut concentration  $(I_{gut})$  for the parent compound is to be assumed the administered dose dissolved in 250 ml.

$$I_{gut} = \frac{D}{250}$$

**Systemic concentration** The unbound systemic  $(I_{max,ss,u})$  concentrations of the parent compound and the metabolites that are relevant for the DDI potential are derived from the total maximal plasma concentration and the respective unbound fractions:

$$I_{max,ss,u} = I_{max,ss} * f_u$$

**Hepatic inlet concentration** For the parent compound, the portal contribution to the hepatic inlet concentration is calculated as:

$$portal\ term = D*\frac{F_a*F_g*k_a}{Q_h*R_B}*1000\ ng/ml$$

With D the administered dose in mg,  $F_a$  the fraction absorbed after oral administration,  $F_g$  the fraction available after gut metabolism,  $k_a$  the absorption rate,  $Q_h$  the hepatic blood flow and  $R_B$  the blood-to-plasma ratio.

The standard hepatic blood flow is 97 l/h/70 kg or 1.61 l/min/70 kg.

The relevant hepatic inlet  $(I_{max,inlet,u}, \text{ also called } I_h \text{ in the mechanistic static modeling equations})$  concentration is the sum of the maximal systemic plasma concentration and the portal contribution:

$$I_{max,inlet,u} = (I_{max,ss} + portal\ term) * f_u$$

Enteric concentration For the parent compound, the villous concentration in the gut ( $I_{enteric}$ , also called  $I_g$  in the mechanistic static modeling equations) is calculated as:

$$I_{enteric,u} = D*\frac{F_a*k_a}{Q_{ent}}*1000~ng/ml$$

with  $F_a$  the fraction absorbed after oral administration,  $k_a$  the absorption rate,  $Q_{ent}$  the enteric villous blood flow and  $R_B$  the blood-to-plasma ratio.

Note that as per the FDA guideline (refer to FDA, 2020, Fig. 7, and Rostami-Hodjegan and Tucker, 2004) the blood-to-plasma ratio and the plasma binding of the drug are ignored.

The standard villous blood flow is 18 l/h/70 kg or 0.3 l/min/70 kg.

#### Basic modeling of enzyme inhibition

#### Reversible inhibition

For the basic modeling of direct (reversible) enzyme inhibition, the ratios of the relevant inhibitor concentration to the  $K_i$  are considered (refer to the FDA guidance, FDA 2020, Fig. 1). A cut-off of 1.02 applies.

Liver

$$R_1 = 1 - \frac{I_{max,ss,u}}{K_{i,u}}$$

Gut wall

$$R_{1,gut} = 1 - \frac{I_{gut}}{K_{i,u}}$$

### Basic modeling of CYP induction

$$R_3 = \frac{1}{1 + d * \frac{E_{max} * 10 * I_{max,u}}{EC_{50} + 10 * I_{max,u}}}$$

# Static mechanistic modeling of CYP inhibition/induction

In this approach, AUC ratios for probe substrates are calculated based on their known intestinal and hepatic metabolism. Both direct (competitive) and time-dependent inhibition are considered. The below formula given by the FDA guideline (refer to FDA 2020, Fig. 7) also includes intestinal and hepatic enzyme induction terms ( $C_g$  and  $C_h$ , respectively). At the same time, the guideline states that both inhibition and induction should be considered separately. For the purpose of modeling used here the induction terms are ignored.

$$AUCR = \frac{1}{A_g * B_g * C_g * (1 - F_g) + F_g} * \frac{1}{A_h * B_h * C_h * f_m + (1 - f_m)}$$

The invidual terms are:

Reversible inhibition

$$A_g = \frac{1}{1 + \frac{I_g}{K_i}}$$

$$A_h = \frac{1}{1 + \frac{I_h}{K_i}}$$

Time-dependent inhibition

$$B_g = \frac{k_{deg,g}}{k_{deg,g} + \frac{I_g * k_{inact}}{I_q + K_I}}$$

$$B_h = \frac{k_{deg,h}}{k_{deg,h} + \frac{I_h * k_{inact}}{I_h + K_I}}$$

Induction

$$C_g = 1 + \frac{d * E_{max} * I_g}{I_a + EC_{50}}$$

$$C_h = 1 + \frac{d * E_{max} * I_h}{I_h + EC_{50}}$$

with the hepatic inlet concentration  $I_h = I_{max,inlet,u}$  and the intestinal concentration  $I_g = I_{enteric,u}$  (see above). d is an induction scaling factor (assumed to be 1 but can be adjusted).

# Appendix 2: R Session Info

This document was created using R version 4.3.1 (2023-06-16) and the following packages:

name	version
ddir	0.2.0
knitrdata	0.6.1
knitr	1.43
lubridate	1.9.2
forcats	1.0.0
stringr	1.5.0
dplyr	1.1.3
purrr	1.0.2
readr	2.1.4
tidyr	1.3.0
tibble	3.2.1
ggplot2	3.4.2
tidyverse	2.0.0