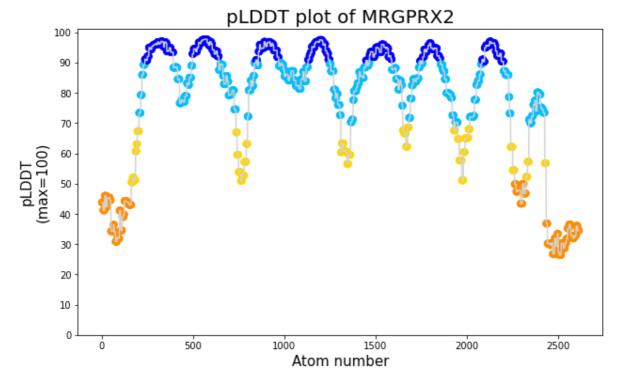
```
##Citation
In [15]:
          #Jumper, J., Evans, R., Pritzel, A. et al. Highly accurate protein structure predict
          #Mihaly Varadi, Stephen Anyango, Mandar Deshpande, Sreenath Nair, Cindy Natassia, Ga
          #https://alphafold.ebi.ac.uk/entry/A0A6J3EQU8
          #https://biopython.org/docs/1.75/api/Bio.PDB.Residue.html
          #https://biopython.org/docs/1.75/api/Bio.PDB.Atom.html
In [16]: import os #Bring the os to check working directory
          os.getcwd()
           'C:₩WUsers₩Wpanky₩WDesktop₩WpLDDT'
Out[16]:
In [17]:
           from Bio.PDB import * #Bring the Bio.PDB for ananlysis, Bio.PDB is included in Biok
          ##Set up the PDB file
In [18]:
          par = PDBParser() #Set PDBParser() to par for convenient
          protein = par.get_structure("MRGPRX2", "AF-AOA6J3EQU8-F1-model_v3.pdb") #Load the PD
          protein #Check the file
In [19]:
          <Structure id=MRGPRX2>
Out[19]:
In [20]: print(dir(protein)) #Check the module
          ['__class__', '__contains__', '__delattr__', '__delitem__', '__dict__', '__dir__
'__doc__', '__eq__', '__format__', '__ge__', '__getattribute__', '__getitem__',
                 '__lt__', '__module__', '__ne__', '__new__', '__reduce__', '__reduce_ex__', '__repr_
_', '__setattr__', '__sizeof__', '__str__', '__subclasshook__', '__weakref__', '_gen
erate_full_id', '_id', '_reset_full_id', 'add', 'atom_to_internal_coordinates', 'cen
          ter_of_mass', 'child_dict', 'child_list', 'copy', 'detach_child', 'detach_parent',
          'full_id', 'get_atoms', 'get_chains', 'get_full_id', 'get_id', 'get_iterator', 'get_
          level', 'get_list', 'get_models', 'get_parent', 'get_residues', 'has_id', 'header',
          'id', 'insert', 'internal_to_atom_coordinates', 'level', 'parent', 'set_parent', 'tr
          ansform', 'xtra']
In [21]: print(protein.header["name"]) #Check the header name
          print(protein.header["release_date"]) #Check the release date
          alphafold monomer v2.0 prediction for mas-related g-protein coupled receptor member
          x2 (a0a6j3equ8)
          1909-01-08
          model = protein.get_models() #Get model to use PDB file easily
In [22]:
          <generator object Structure.get_models at 0x000002847D5E67A0>
Out[22]:
          models = list(model) #Make model as list
In [23]:
          models
In [24]:
          [<Model id=0>]
Out[24]:
          chains = list(models[0].get_chains()) #Check the numer of chain
In [25]:
          chains #In this case, the protein has one chain
          [<Chain id=A>]
Out[25]:
```

```
In [26]: residue = list(chains[0].get_residues()) #Set the chain number and get residue data
          len(residue) #Check the number of residues
Out[26]:
         a=residue[1].get_resname() #Use .get_resname() to extract residue name
In [27]:
          print(a)
         ASP
          ##Generate the sequence of protein
In [28]:
          #Create the dictionary to make shorter amioacids sequence
          d= {'CYS': 'C', 'ASP': 'D', 'SER': 'S', 'GLN': 'Q', 'LYS': 'K',
               'ILE': 'I', 'PRO': 'P', 'THR': 'T', 'PHE': 'F', 'ASN': 'N',
               'GLY': 'G', 'HIS': 'H', 'LEU': 'L', 'ARG': 'R', 'TRP': 'W', 'ALA': 'A', 'VAL':'V', 'GLU': 'E', 'TYR': 'Y', 'MET': 'M'}
          #len(residue)
          nr_0=range(0,len(residue)) #Select the residue range, be aware of that python starts
          R=[] #Create the empty list
          for i in nr_0: #Repeat the for loop
              rr=residue[i].get_resname() #Get the name of each residue
              rrs=str(rr) #Convert residue name object to string
              nrr=d[rrs] #Bring the simple code from dictionary above
              R.append(nrr) #Add the code to list
          seq="".join(R) #Use "".join() to make continuous sequence with no space
          print(seq) #View the result sequence
          len(seq) #Check the length of sequence
         MDPT|PAWGTKSTTMNGDDQALPLLCGKETL|PVLL|LF|GLVGLVGNAVVLWFLGFHMRRNAFSVYVLSLAGADFLCLCFQ||
         DCLAYLSDFYHSLYTYFPSFLTAMITCAYLAGLNILSAISAERCLSVLCPIWYRCRRPRHLSTVMCALLWAVSLLLSILEGKFC
          GFLFTDGDSGWCQTFDFITAAWLIFLFVVLCGSSLALLVRILCGSRKMPLTRLYVTILLTVLVFLLCGLPFGIQWFLILWIWKN
         FDDFLCH1HPVSLVLSSLNSSANP11YFFVGSFRQQWRLRQPTLKLALQRALQDTAEVDHSEGSFRQDTLEMSGSSLV
Out[29]:
          ##Draw the pLDDT plot
In [30]:
          atoms = list(residue[1].get_atoms()) #Check atoms in each residue
          atoms
          [<Atom N>,
Out[30]:
          <Atom CA>.
           <Atom C>,
           <Atom CB>.
           <Atom 0>,
           <Atom CG>.
           <Atom OD1>,
           <Atom 0D2>1
          atoms[1].get_bfactor() #Get the data in b factor column, in alphfold PDB files, pLDD
In [31]:
          41.16
Out[31]:
In [32]:
          #Prepare the x-value and y-value for plot
          X=[] #Create the empty list
          Y=[] #Create the empty list
          nr=range(0,len(residue)) #Select the residue range, remember the len(residue)
          N=nr[0] #Set the base value for for loop
          for i in nr: #Repeat the for loop
              atoms_1=list(residue[i].get_atoms()) #Extract the atom list of each residue
              for k in atoms_1: #Additional for loop for atoms list
                  bb=k.get_bfactor() #Get pLDDT value for each atom
                  Y.append(bb) #Add pLDDT to y-value list
```

N=N+1 #Create x-value which means atom number X.append(N) #Add atom numer to x-value list

```
#Draw the plot
In [33]:
          import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot
          import matplotlib as mpl
          ids=str(protein) #Make the string of protein name
         pn=ids[14:-1] #Extract the name
         #plot
         fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
         cmaps = mpl.colors.ListedColormap(['darkorange', 'gold', 'deepskyblue', 'b']) #Set the
         bounds = [0,50,70,90,100] #Set the bound for colormap
         norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap
         ax.plot(X,Y,color='lightgrey') #Set the color of line plot
         ax.scatter(X,Y,s=50,c=Y,cmap=cmaps,norm=norms) #Draw the scatter plot and set the co
         ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize
         ax.set_xlabel("Atom number", fontsize=15) #Set the x label and fontsize
         ax.set_ylabel("pLDDT\(max=100\)",fontsize=15) #Set the y label and fontsize, remeber
         plt.yticks(range(0, 109,10)) #Custom the y sticks
         plt.show() #Show the plot
```

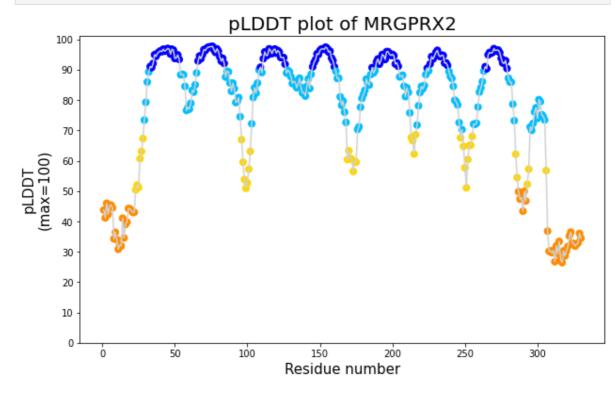


```
#Use the residue number to view the pLDDT value

X1=[] #Create the empty list
Y1=[] #Create the empty list
nr_1=range(0,len(residue)) #Select the residue range, remember the len(residue)
N1=nr_1[0] #Set the base value for for loop
for i in nr_1: #Repeat the for loop
    atoms1=list(residue[i].get_atoms()) #Extract the atom list of each residue
    bb=atoms1[0].get_bfactor() #All pLDDT value is same in on residue, just use the
    Y1.append(bb) #Add pLDDT to y-value list
    N1=N1+1 #Create x-value which means residue number
    X1.append(N1) #Add residue numer to x-value list
```

```
In [35]: #Draw the plot
    import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot
    import matplotlib as mpl
    ids=str(protein) #Make the string of protein name
    pn=ids[14:-1] #Extract the name
```

```
#plot
fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
cmaps = mpl.colors.ListedColormap(['darkorange', 'gold', 'deepskyblue', 'b']) #Set the
bounds = [0,50,70,90,100] #Set the bound for colormap
norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap
ax.plot(X1,Y1,color='lightgrey') #Set the color of line plot
ax.scatter(X1,Y1,s=50,c=Y1,cmap=cmaps,norm=norms) #Draw the scatter plot and set the
ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize
ax.set_xlabel("Residue number",fontsize=15) #Set the x label and fontsize
ax.set_ylabel("pLDDT\m(max=100)",fontsize=15) #Set the y label and fontsize, remeber
plt.yticks(range(0, 109,10)) #Custom the y sticks
plt.show() #Show the plot
```



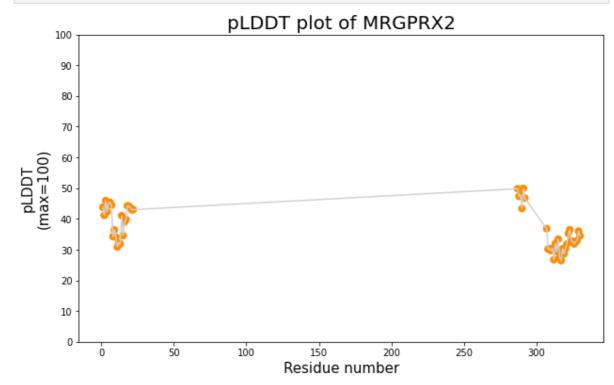
```
In [36]: ##Extract the residue number, pLDDT<50
X2=[] #Create the empty list
Y2=[] #Create the empty list
nr_2=range(0,len(residue)) #Select the residue range, remember the len(residue)
cr_s2=0 ##Set the criteria of pLDDT
cr_e2=50 ##Set the criteria of pLDDT
for i in nr_2: #Repeat the for loop
    atoms1=list(residue[i].get_atoms()) #Extract the atom list of each residue
    bb=atoms1[0].get_bfactor() #All pLDDT value is same in on residue, just use the
    if bb >= cr_s2 and bb < cr_e2:
        Y2.append(bb) #Add pLDDT to y-value list
        N2=i+1 #Create x-value which means residue number
        X2.append(N2) #Add residue numer to x-value list</pre>
```

```
import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot
import matplotlib as mpl
ids=str(protein) #Make the string of protein name
pn=ids[14:-1] #Extract the name

#plot
fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
cmaps = mpl.colors.ListedColormap(['darkorange','gold','deepskyblue','b']) #Set the
bounds = [0,50,70,90,100] #Set the bound for colormap
norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap
ax.plot(X2,Y2,color='lightgrey') #Set the color of line plot
```

```
ax.scatter(X2,Y2,s=50,c=Y2,cmap=cmaps,norm=norms) #Draw the scatter plot and set the ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize ax.set_xlabel("Residue number",fontsize=15) #Set the x label and fontsize ax.set_ylabel("pLDDT\n(max=100)",fontsize=15) #Set the y label and fontsize, remeber plt.yticks(range(0, 109,10)) #Custom the y sticks plt.show() #Show the plot

#View the residue number print('0<pLDDT<50') print('Total:',len(X2)) print(X2)
```



0<pLDDT<50
Total: 52
[1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 287, 288, 289, 290, 291, 292, 307, 308, 309, 310, 311, 312, 313, 314, 315, 316, 317, 318, 319, 320, 321, 322, 323, 324, 325, 326, 327, 328, 329, 330]</pre>

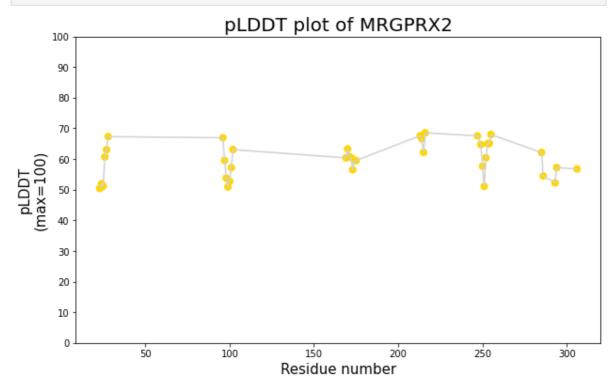
```
In [38]: ##Extract the residue number, 50<pLDDT<70
X3=[] #Create the empty list
Y3=[] #Create the empty list
nr_3=range(0,len(residue)) #Select the residue range, remember the len(residue)
cr_s3=50 ##Set the criteria of pLDDT
cr_e3=70 ##Set the criteria of pLDDT
for i in nr_3: #Repeat the for loop
    atoms1=list(residue[i].get_atoms()) #Extract the atom list of each residue
    bb=atoms1[0].get_bfactor() #All pLDDT value is same in on residue, just use the
    if bb >= cr_s3 and bb < cr_e3:
        Y3.append(bb) #Add pLDDT to y-value list
        N2=i+1 #Create x-value which means residue number
        X3.append(N2) #Add residue numer to x-value list</pre>
```

```
In [39]: #Draw the plot
    import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot
    import matplotlib as mpl
    ids=str(protein) #Make the string of protein name
    pn=ids[14:-1] #Extract the name

#plot
    fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
    cmaps = mpl.colors.ListedColormap(['darkorange','gold','deepskyblue','b']) #Set the
```

```
bounds = [0,50,70,90,100] #Set the bound for colormap norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap ax.plot(X3,Y3,color='lightgrey') #Set the color of line plot ax.scatter(X3,Y3,s=50,c=Y3,cmap=cmaps,norm=norms) #Draw the scatter plot and set the ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize ax.set_xlabel("Residue number",fontsize=15) #Set the x label and fontsize ax.set_ylabel("pLDDTWn(max=100)",fontsize=15) #Set the y label and fontsize, remeber plt.yticks(range(0, 109,10)) #Custom the y sticks plt.show() #Show the plot

#View the residue number
print('50<pLDDT<70')
print('Total:',len(X3))
print(X3)
```

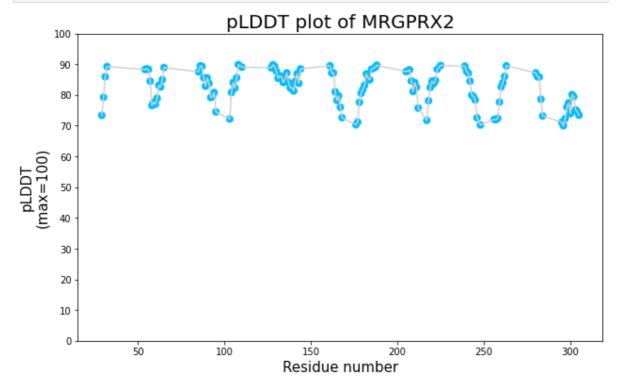


50<pLDDT<70
Total: 37
[23, 24, 25, 26, 27, 28, 96, 97, 98, 99, 100, 101, 102, 169, 170, 171, 172, 173, 17
4, 175, 213, 214, 215, 216, 247, 249, 250, 251, 252, 253, 254, 255, 285, 286, 293, 2
94, 306]

```
In [40]: ##Extract the residue number, 70<pLDDT<90
X4=[] #Create the empty list
Y4=[] #Create the empty list
nr_4=range(0,len(residue)) #Select the residue range, remember the len(residue)
cr_s4=70 ##Set the criteria of pLDDT
cr_e4=90 ##Set the criteria of pLDDT
for i in nr_4: #Repeat the for loop
    atoms1=list(residue[i].get_atoms()) #Extract the atom list of each residue
    bb=atoms1[0].get_bfactor() #All pLDDT value is same in on residue, just use the
    if bb >= cr_s4 and bb < cr_e4:
        Y4.append(bb) #Add pLDDT to y-value list
        N2=i+1 #Create x-value which means residue number
        X4.append(N2) #Add residue numer to x-value list</pre>
```

```
In [41]: #Draw the plot import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot import matplotlib as mpl ids=str(protein) #Make the string of protein name pn=ids[14:-1] #Extract the name
```

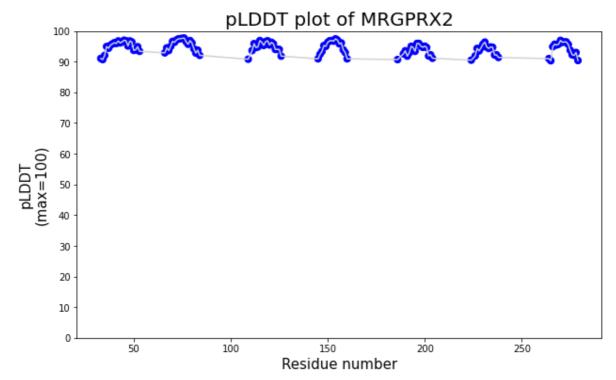
```
#plot
fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
cmaps = mpl.colors.ListedColormap(['darkorange','gold','deepskyblue','b']) #Set the
bounds = [0,50,70,90,100] #Set the bound for colormap
norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap
ax.plot(X4,Y4,color='lightgrey') #Set the color of line plot
ax.scatter(X4,Y4,s=50,c=Y4,cmap=cmaps,norm=norms) #Draw the scatter plot and set the
ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize
ax.set_xlabel("Residue number", fontsize=15) #Set the x label and fontsize
ax.set_ylabel("pLDDT\(max=100\)",fontsize=15) #Set the y label and fontsize, remeber
plt.yticks(range(0, 109,10)) #Custom the y sticks
plt.show() #Show the plot
#View the residue number
print('70<pLDDT<90')</pre>
print('Total:',len(X4))
print(X4)
```



70<pLDDT<90
Total: 121
[29, 30, 31, 32, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 103, 104, 105, 106, 107, 108, 110, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 161, 162, 163, 164, 165, 166, 167, 168, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 187, 188, 205, 206, 207, 208, 209, 210, 211, 212, 217, 218, 219, 220, 221, 222, 223, 225, 239, 240, 241, 242, 243, 244, 245, 246, 248, 256, 257, 258, 259, 260, 261, 262, 263, 280, 281, 282, 283, 284, 295, 296, 297, 298, 299, 300, 301, 302, 303, 304, 305]

```
In [42]: ##Extract the residue number, 90<pLDDT<100
X5=[] #Create the empty list
Y5=[] #Create the empty list
nr_5=range(0,len(residue)) #Select the residue range, remember the len(residue)
cr_s5=90 ##Set the criteria of pLDDT
cr_e5=100 ##Set the criteria of pLDDT
for i in nr_5: #Repeat the for loop
    atoms1=list(residue[i].get_atoms()) #Extract the atom list of each residue
    bb=atoms1[0].get_bfactor() #AII pLDDT value is same in on residue, just use the
    if bb >= cr_s5 and bb < cr_e5:
        Y5.append(bb) #Add pLDDT to y-value list
        N2=i+1 #Create x-value which means residue number
        X5.append(N2) #Add residue numer to x-value list</pre>
```

```
#Draw the plot
In [43]:
          import matplotlib.pyplot as plt #Bring the matplotlib to draw the plot
          import matplotlib as mpl
          ids=str(protein) #Make the string of protein name
         pn=ids[14:-1] #Extract the name
         #plot
         fig, ax = plt.subplots(figsize=(10, 6)) #Set the size of figure
         cmaps = mpl.colors.ListedColormap(['darkorange','gold','deepskyblue','b']) #Set the
         bounds = [0,50,70,90,100] #Set the bound for colormap
         norms = mpl.colors.BoundaryNorm(bounds,cmaps.N) #Set the norm for colormap
         ax.plot(X5,Y5,color='lightgrey') #Set the color of line plot
         ax.scatter(X5,Y5,s=50,c=Y5,cmap=cmaps,norm=norms) #Draw the scatter plot and set the
         ax.set_title("pLDDT plot of "+pn, fontsize=20) #Set the title and fontsize
         ax.set_xlabel("Residue number",fontsize=15) #Set the x label and fontsize
         ax.set_ylabel("pLDDT\(max=100\)", fontsize=15) #Set the y label and fontsize, remeber
         plt.yticks(range(0, 109,10)) #Custom the y sticks
         plt.show() #Show the plot
         #View the residue number
         print('90<pLDDT<100')</pre>
         print('Total:',len(X5))
         print(X5)
```



909090909090100171718191919101819101819101910191019101910191019191010191011111111111111111111111111111112</p