## Package 'ouch'

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```
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     'ouchtree.R'
     'ape2ouch.R'
     'bimac.R'
     'rmvnorm.R'
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## Description

The **ouch** package provides facilities for phylogenetic comparative analysis based on Ornstein-Uhlenbeck models of trait evolution along a phylogeny. Multivariate data and complex adaptive hypotheses are supported.

## Classes

The basic class, ouchtree, is provided to encode a phylogenetic tree. Plot and print methods are provided.

The class browntree derives from class ouchtree and encodes the results of fitting a Brownian Motion model to data.

The class hansentree derives from class ouchtree and encodes the results of fitting a Hansen model to data.

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## **Detailed Documentation**

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#### Author(s)

Aaron A. King

#### References

Butler, M.A. and A.A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.

Cressler, C. E., Butler, M. A., and King, A. A. 2015. Detecting adaptive evolution in phylogenetic comparative analysis using the Ornstein-Uhlenbeck model. Syst. Biol., 64:953–968.

anolis.ssd

Greater Antillean anolis lizard sexual size dimorphism data

## **Description**

The dataset consists of sexual size-dimorphism data for 38 species of anoles from Cuba, Hispaniola, Jamaica, and Puerto Rico (Butler, Schoener, and Losos 2000). Each of these species belongs to one of six microhabitat types, or "ecomorphs" (sensu Williams, 1972): trunk-ground, grass-bush, trunk, trunk-crown, twig, and crown-giant. The data were used to demonstrate an evolutionary association between habitat type and degree of sexual size dimorphism.

## **Format**

A data frame with 38 observations on the following 6 variables.

**node** Labels for the nodes.

species Names of extant species.

log.SSD Log sexual size dimorphism of extant species.

ancestor Ancestor node.

time Time of node.

OU.1 a factor with levels ns

**OU.7** a factor with levels corresponding to ecomorph (tg tc gb cg tw tr anc)

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## **Details**

Size dimorphism was calcuated as the log-ratio of male snout-to-vent length to female snout-to-vent length (males are larger).

In this example, we tested three models of evolution: Brownian motion, Ornstein-Uhlenbeck with one global optimum, and Ornstein-Uhlenbeck with 7 optima (one for each ecomorph type plus an additional one for an "unknown" type).

For the 7-optima model, we assigned each terminal branch to an optimum according to the ecomorph type of the extant species. Because we had no information to help guide hypotheses about internal branches, we assigned internal branches to the "unknown" selective regime. The phylogeny of these species is consistent with and adaptive radiation, with a burst of speciation events early in the evolutionary history of this clade (see phylogeny in Butler & King (2004) or example below).

## Author(s)

Marguerite A. Butler, Aaron A. King

#### Source

Butler, M.A. and A.A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.

#### References

Butler, M. A., T. W. Schoener, and J. B. Losos. 2000. The relationship between sexual size dimorphism and habitat use in Greater Antillean Anolis lizards. Evolution, 54:259–272.

Williams, E. E. 1972. The origin of faunas. Evolution of lizard congeners in a complex island fauna: a trial analysis. Evol. Biol., 6:47–89.

#### **Examples**

```
tree <- with(anolis.ssd,ouchtree(node,ancestor,time/max(time),species))
plot(tree,node.names=TRUE)

h1 <- brown(anolis.ssd['log.SSD'],tree)
h1
plot(h1)

h2 <- hansen(anolis.ssd['log.SSD'],tree,anolis.ssd['OU.1'],sqrt.alpha=1,sigma=1)
h2
plot(h2)

h3 <- hansen(anolis.ssd['log.SSD'],tree,anolis.ssd['OU.7'],sqrt.alpha=1,sigma=1)
h3
plot(h3)</pre>
```

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ape2ouch	Convert an "ape" tree to an "ouch" tree.

#### **Description**

ape2ouch translates **ape**'s phylo representation of a phylogenetic tree into **ouch**'s ouchtree representation. The user can change the branch lengths while preserving the topology.

#### Usage

```
ape2ouch(tree, scale = TRUE, branch.lengths = tree$edge.length)
```

## **Arguments**

tree a tree of class phylo created in package **ape**.

scale if scale=TRUE, the tree's depth will be scaled to 1. If scale is a number, then

the branch lengths will be scaled by this number.

branch.lengths optional vector of branch lengths.

## Author(s)

A. A. King, D. Ackerly

bimac

Anolis bimaculatus lizard size data.

## **Description**

This is the *Anolis bimaculatus* dataset used in Butler & King (2004). It is used to test a hypothesis of character displacement using an interspecific dataset of body sizes and current data on sympatry/allopatry. The data frame has the following columns: species which are species names, size which is the phenotypic data, and the variables ancestor and time which specify the topology of the phylogeny and the location of the nodes in time, respectively. The columns 0U.1, 0U.3, 0U.4, and 0U.LP specify four hypothetical arrangements of selective regimes. Explanations of the data follow:

**Body size.** We use the phenotypic data and phylogeny of Losos (1990), which employed the head lengths (of males) as a proxy for body size. In this group of lizards, head length correlates very strongly with snout-to-vent length and the cube root of mass, which are standard measures of body size. The data are head lengths in mm; note that we use the log of this value in analyses.

**Tree topology** The tree topology is encoded via two vectors: ancestor and time. Each node of the' phylogenetic tree has a corresponding row in the data frame, numbered from 1 to 45. The columns ancestor and time specify the phylogeny. The ancestor variable specifies the topology: it is a list indicating the ancestor of each node. The root node has ancestor 0. The variable time specifies the temporal location of each node, with the root node being at time 0.

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Specifications of selective regimes. (Columns 0U.1, 0U.3, 0U.4, 0U.LP). These columns are factors, the levels of which correspond to the "paintings" of the respective adaptive regime hypotheses onto the phylogeny. Each selective regime is named (small, medium, large, etc.). Each column corresponds to a different painting of the selective regimes, and thus to a different hypothesis. In this example, there are 3 alternative models (see Butler & King 2004): 0U.4 is 4-regime model, 0U.3 is 3-regime model (all ancestors are medium), 0U.LP is the linear parsimony model.

#### **Format**

A data frame with 45 observations on the following 8 variables.

**node** Labels for the nodes.

species Species names for extant species.

size Body size (head length in mm) of extant species.

ancestor Ancestral node.

time Time of node.

OU.1 a factor with levels ns

OU.3 a factor with levels small, medium, large

**OU.4** a factor with levels small, medium, large, and

OU.LP a factor with levels small, medium, large

#### Author(s)

Marguerite A. Butler and Aaron A. King

#### **Source**

Butler, M.A. and A.A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.

#### References

Lazell, J. D. 1972. The anoles (Sauria: Iguanidae) of the Lesser Antilles. Bull. Mus. Comp. Zool., 143:1–115.

Losos, J. B. 1990. A phylogenetic analysis of character displacement in Caribbean Anolis lizards. Evolution, 44:558–569.

## **Examples**

```
tree <- with(bimac,ouchtree(node,ancestor,time/max(time),species))
plot(tree,node.names=TRUE)

h1 <- brown(log(bimac['size']),tree)
h1
plot(h1)

h2 <- hansen(log(bimac['size']),tree,bimac['OU.1'],sqrt.alpha=1,sigma=1)</pre>
```

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```
h2
plot(h2)

h3 <- hansen(log(bimac['size']),tree,bimac['OU.3'],sqrt.alpha=1,sigma=1)
h3
plot(h3)

h4 <- hansen(log(bimac['size']),tree,bimac['OU.4'],sqrt.alpha=1,sigma=1)
h4
plot(h4)

h5 <- hansen(log(bimac['size']),tree,bimac['OU.LP'],sqrt.alpha=1,sigma=1,reltol=1e-5)
h5 <- update(h5,method='subplex',reltol=1e-11,parscale=c(0.1,0.1),hessian=TRUE)
h5

simdat <- simulate(h5,nsim=10)
hsim <- update(h5,data=simdat[[1]])
summary(hsim)
bsim <- update(h1,data=simdat[[1]])
summary(bsim)
```

bootstrap

Bootstrapping for uncertainty quantification

#### **Description**

Generic bootstrapping for **ouch** models.

## Usage

```
## S4 method for signature 'browntree'
bootstrap(object, nboot = 200, seed = NULL, ...)
## S4 method for signature 'hansentree'
bootstrap(object, nboot = 200, seed = NULL, ...)
## S4 method for signature 'missing'
bootstrap(object, ...)
## S4 method for signature 'ANY'
bootstrap(object, ...)
```

## **Arguments**

object A fitted model object.

nboot integer; number of bootstrap replicates.

seed integer; setting seed to a non-NULL value allows one to fix the random seed (see simulate).

Additional arguments are passed to update.

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## **Details**

bootstrap performs a parametric bootstrap for estimation of confidence intervals.

brown

Phylogenetic Brownian motion models

## **Description**

The function brown creates a browntree object by fitting a Brownian-motion model to data.

## Usage

```
brown(data, tree)
```

## **Arguments**

data Phenotypic data for extant species, i.e., at the terminal ends of the phylogenetic

tree. This can either be a numeric vector or a list. If it is a numeric vector, there must be one entry for every node. If it is a list, it must consist entirely of numeric vectors, each of which has one entry per node. A data-frame is coerced to a list.

tree A phylogenetic tree, specified as an ouchtree object.

## Value

brown returns an object of class browntree.

## Author(s)

Aaron A. King

## References

Butler, M.A. and A.A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.

## See Also

ouchtree, hansen, bimac, anolis.ssd

coef,browntree-method 9

coef, browntree-method Model coefficients

## Description

coef extracts the parameters from a fitted model object.

## Usage

```
## S4 method for signature 'browntree'
coef(object, ...)
## S4 method for signature 'hansentree'
coef(object, ...)
```

## **Arguments**

object fitted model object.

... additional arguments, ignored.

#### Value

coef applied to a browntree object extracts a list with three elements:

sigma the coefficients of the sigma matrix.

theta a list of the estimated optima, one per character.

sigma..sq.matrix the sigma-squared matrix itself.

coef applied to a hansentree object returns a named list containing the estimated  $\alpha$  and  $\sigma^2$  matrices(given as the alpha.matrix and sigma.sq.matrix elements, respectively) but also the MLE returned by the optimizer (as sqrt.alpha and sigma, respectively). The latter elements should not be interpreted, but can be used to restart the algorithm, etc.

hansen

Ornstein-Uhlenbeck models of trait evolution

## Description

The function hansen fits an Ornstein-Uhlenbeck model to data. The fitting is done using optim or subplex.

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## Usage

```
hansen(
  data,
  tree,
  regimes,
  sqrt.alpha,
  sigma,
  fit = TRUE,
  method = c("Nelder-Mead", "subplex", "BFGS", "L-BFGS-B"),
  hessian = FALSE,
  ...
)
```

## **Arguments**

data

Phenotypic data for extant species, i.e., species at the terminal twigs of the phylogenetic tree. This can either be a single named numeric vector, a list of nchar named vectors, or a data frame containing nchar data variables. There must be an entry per variable for every node in the tree; use NA to represent missing data. If the data are supplied as one or more named vectors, the names attributes are taken to correspond to the node names specified when the ouchtree was constructed (see ouchtree). If the data are supplied as a data-frame, the rownames serve that purpose.

tree

A phylogenetic tree, specified as an ouchtree object.

regimes

A vector of codes, one for each node in the tree, specifying the selective regimes hypothesized to have been operative. Corresponding to each node, enter the code of the regime hypothesized for the branch segment terminating in that node. For the root node, because it has no branch segment terminating on it, the regime specification is irrelevant. If there are nchar quantitative characters, then one can specify a single set of regimes for all characters or a list of nchar regime specifications, one for each character.

sqrt.alpha, sigma

These are used to initialize the optimization algorithm. The selection strength matrix  $\alpha$  and the random drift variance-covariance matrix  $\sigma^2$  are parameterized by their matrix square roots. Specifically, these initial guesses are each packed into lower-triangular matrices (column by column). The product of this matrix with its transpose is the  $\alpha$  or  $\sigma^2$  matrix. See Details.

fit

If fit=TRUE, then the likelihood will be maximized. If fit=FALSE, the likelihood will be evaluated at the specified values of sqrt.alpha and sigma; the optima theta will be returned as well.

method

The method to be used by the optimization algorithm. See subplex and optim for information on the available options.

hessian

If hessian=TRUE, then the Hessian matrix will be computed by optim.

• • •

Additional arguments will be passed as control options to optim or subplex. See optim and subplex for information on the available options.

. . . .

S

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#### **Details**

The Hansen model for the evolution of a multivariate trait X along a lineage can be written as a stochastic differential equation (Ito diffusion)

$$dX = \alpha(\theta(t) - X(t))dt + \sigma dB(t),$$

where t is time along the lineage,  $\theta(t)$  is the optimum trait value, B(t) is a standard Wiener process (Brownian motion), and  $\alpha$  and  $\sigma$  are matrices quantifying, respectively, the strength of selection and random drift. Without loss of generality, one can assume  $\sigma$  is lower-triangular. This is because only the infinitesimal variance-covariance matrix  $\sigma^2 = \sigma \sigma^T$  is identifiable, and for any admissible variance-covariance matrix, we can choose  $\sigma$  to be lower-triangular. Moreover, if we view the basic model as describing evolution on a fitness landscape, then  $\alpha$  will be symmetric. If we further restrict ourselves to the case of stabilizing selection,  $\alpha$  will be positive definite as well. We make these assumptions and therefore can assume that the matrix  $\alpha$  has a lower-triangular square root.

The hansen code uses unconstrained numerical optimization to maximize the likelihood. To do this, it parameterizes the  $\alpha$  and  $\sigma^2$  matrices in a special way: each matrix is parameterized by nchar\*(nchar+1)/2 parameters, where nchar is the number of quantitative characters. Specifically, the parameters initialized by the sqrt.alpha argument of hansen are used to fill the nonzero entries of a lower-triangular matrix (in column-major order), which is then multiplied by its transpose to give the selection-strength matrix. The parameters specified in sigma fill the nonzero entries in the lower triangular  $\sigma$  matrix. When hansen is executed, the numerical optimizer maximizes the likelihood over these parameters.

#### Value

hansen returns an object of class hansentree. For details on the methods of that class, see hansen.

#### Author(s)

Aaron A. King

#### References

Butler, M.A. and A.A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.

Cressler, C. E., Butler, M. A., and King, A. A. 2015. Detecting adaptive evolution in phylogenetic comparative analysis using the Ornstein-Uhlenbeck model. Syst. Biol., 64:953–968.

#### See Also

```
ouchtree, brown, optim, subplex, bimac, anolis.ssd
```

## Examples

```
## Not run:
   if (library(geiger,logical.return=TRUE)) {
### an example data set (Darwin's finches)
   data(geospiza)
```

```
str(geospiza)
    sapply(geospiza,class)
### check the correspondence between data and tree tips:
    print(nc <- with(geospiza,name.check(geospiza.tree,geospiza.data)))</pre>
### looks like one of the terminal twigs has no data associated
### drop that tip:
    tree <- with(geospiza,drop.tip(geospiza.tree,nc$tree_not_data))</pre>
    dat <- as.data.frame(geospiza$dat)</pre>
### make an ouchtree out of the phy-format tree
    ot <- ape2ouch(tree)
### merge data with tree info
    otd <- as(ot, "data.frame")</pre>
### in these data, it so happens that the rownames correspond to node names
### we will exploit this correspondence in the 'merge' operation:
    dat$labels <- rownames(dat)</pre>
    otd <- merge(otd,dat,by="labels",all=TRUE)</pre>
    rownames(otd) <- otd$nodes</pre>
    print(otd)
### this data-frame now contains the data as well as the tree geometry
### now remake the ouch tree
    ot <- with(otd,ouchtree(nodes=nodes,ancestors=ancestors,times=times,labels=labels))</pre>
    b1 <- brown(tree=ot,data=otd[c("tarsusL","beakD")])</pre>
    summary(b1)
### evaluate an OU model with a single, global selective regime
    otd$regimes <- as.factor("global")</pre>
    h1 <- hansen(
      tree=ot,
      data=otd[c("tarsusL", "beakD")],
      regimes=otd["regimes"],
      sqrt.alpha=c(1,0,1),
      sigma=c(1,0,1),
      maxit=10000
    summary(h1)
 }
## End(Not run)
```

logLik, browntree-method

Log likelihood of a fitted model

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## **Description**

logLik extracts the log likelihood from a fitted model object.

## Usage

```
## S4 method for signature 'browntree'
logLik(object)
## S4 method for signature 'hansentree'
logLik(object)
```

## **Arguments**

object fitted model object

## Value

logLik returns a numeric value.

ouchtree

Phylogenetic tree object in 'ouch' format.

## **Description**

An object containing a phylogenetic tree in a form suitable for using **ouch** methods.

## Usage

```
ouchtree(nodes, ancestors, times, labels = as.character(nodes))
```

## **Arguments**

nodes A character vector giving the name of each node. These are used internally and

must be unique.

ancestors Specification of the topology of the phylogenetic tree. This is in the form of a

character vector specifying the name (as given in the nodes argument) of the immediate ancestor of each node. In particular, the i-th name is that of the ancestor of the i-th node. The root node is distinguished by having no ancestor

(i.e., NA).

times A vector of nonnegative numbers, one per node in the tree, specifying the time

at which each node is located. Time should be increasing from the root node to

the terminal twigs.

labels Optional vector of node labels. These will be used in plots to label nodes. It is

not necessary that these be unique.

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## **Details**

ouchtree creates an ouchtree object. This contains the topology, branch times, and epochs. It also (optionally) holds names of taxa for display purposes.

## Author(s)

```
Aaron A. King
```

#### See Also

```
ouchtree, ape2ouch, brown, hansen
```

## **Examples**

```
tree <- with(
   bimac,
   ouchtree(nodes=node,ancestors=ancestor,times=time,labels=species)
)
tree

plot(tree)
plot(tree,node.names=TRUE)</pre>
```

paint

Painting regimes on a phylogenetic tree

## **Description**

Function to paint selective regimes on a phylogenetic tree.

## Usage

```
paint(tree, subtree, branch, which = 1)
```

## Arguments

tree	An object of class	ouchtree.

subtree An optional named vector specifying the root nodes of subtrees. Each branch

that descends from this node will be painted with the specified regime.

branch An optional named vector specifying the end nodes of branches. The unique

branch that terminates at the named node will be painted with the specified

regime.

which integer; if tree is a hansentree, start not with a blank canvas but with the

regime specifications tree contains for the character indicated by which.

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#### **Details**

The names of subtree and branch must be the names of nodes of tree. The painting proceeds in a particular order: one can overpaint a branch. The subtrees indicated by the elements of subtree are painted first, in order. Then the branches indicated by branch are painted. If tree is a simple ouchtree object, then paint begins with a blank canvas, i.e., a tree painted with the single regime "nonspec". If tree is of class hansentree, then paint begins with the regimes specified in the regimes slot of tree. Note that, if tree is a multivariate hansentree, then there are multiple regime specifications contained in tree. In this case, the argument which lets you pick which one you wish to begin with; by default, the first is used.

## Value

A vector of class 'factor' with names corresponding to the nodes in tree, specifying selective regimes.

#### Author(s)

Aaron A. King

#### See Also

ouchtree, hansen

## **Examples**

```
x <- with(
   bimac,
   ouchtree(nodes=node,times=time/max(time),ancestors=ancestor,labels=species))

r <- paint(x,subtree=c("1"="medium","9"="large","2"="small"),
   branch=c("38"="large","2"="medium"))
plot(x,regimes=r,node.names=TRUE)

## compare to bimac['OU.LP']
h5 <- hansen(data=log(bimac['size']),tree=x,regimes=bimac['OU.LP'],
   sqrt.alpha=1,sigma=1,reltol=1e-5)
r <- paint(h5,branch=c("18"="large"),subtree=c("9"="small"))
plot(x,regimes=r,node.names=TRUE)</pre>
```

plot

Plotting functions.

## Description

Plot phylogenies.

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## Usage

```
## S4 method for signature 'ouchtree'
plot(x, y, regimes = NULL, node.names = FALSE, legend = TRUE, ..., labels)
## S4 method for signature 'hansentree'
plot(x, y, regimes, ...)
```

## **Arguments**

x object to plot.
y ignored.

regimes factor or character; a vector of regime paintings.

node.names logical; should node names be displayed?

legend logical; display a legend?

... additional arguments, passed to text.

labels character; taxon labels.

print, ouch tree-method Print methods

## **Description**

Print methods

## Usage

```
## S4 method for signature 'ouchtree'
print(x, ...)

## S4 method for signature 'ouchtree'
show(object)

## S4 method for signature 'browntree'
show(object)

## S4 method for signature 'browntree'
print(x, ...)

## S4 method for signature 'hansentree'
print(x, ...)

## S4 method for signature 'hansentree'
show(object)
```

simulate, browntree-method

## **Arguments**

```
... additional arguments, ignored. object, x object to display.
```

## Value

print displays the tree as a table, along with the coefficients of the fitted model and diagnostic information.

print displays the tree as a table, along with the coefficients of the fitted model and diagnostic information.

print displays the tree as a table, with (possibly) other information.

```
simulate, browntree-method
```

Simulations of a phylogenetic trait model.

## **Description**

simulate generates random deviates from a fitted model.

## Usage

```
## S4 method for signature 'browntree'
simulate(object, nsim = 1, seed = NULL, ...)
## S4 method for signature 'hansentree'
simulate(object, nsim = 1, seed = NULL, ...)
```

## **Arguments**

object fitted model object to simulate.

nsim integer; number of independent simulations.

seed integer; if non-NULL, the RNG will be initialized with this seed for the simula-

tions. The RNG will be reset to its pre-existing state when simulate returns.

... additional arguments, ignored.

#### Value

simulate returns a list of data-frames, each comparable to the original data.

```
summary, browntree-method
```

Summary methods

## **Description**

Summary methods

## Usage

```
## $4 method for signature 'browntree'
summary(object, ...)
## $4 method for signature 'hansentree'
summary(object, ...)
```

## Arguments

```
object fitted model object.
... additional arguments, ignored.
```

#### Value

summary applied to a browntree object returns information about the fitted model, including parameter estimates and quantities describing the goodness of fit.

summary applied to a hansentree method displays the estimated  $\alpha$  and  $\sigma^2$  matrices as well as various quantities describing the goodness of model fit.

```
update, browntree-method
```

Update and refit a model.

## **Description**

update will update a model and re-fit. This allows one to change the data and/or parameters.

## Usage

```
## S4 method for signature 'browntree'
update(object, data, ...)
## S4 method for signature 'hansentree'
update(object, data, regimes, sqrt.alpha, sigma, ...)
```

## **Arguments**

object fitted model object.

data data that replace those used in the original fit.

... Additional arguments replace the corresponding arguments in the original call.

regimes A vector of codes, one for each node in the tree, specifying the selective regimes

hypothesized to have been operative. Corresponding to each node, enter the code of the regime hypothesized for the branch segment terminating in that node. For the root node, because it has no branch segment terminating on it, the regime specification is irrelevant. If there are nother quantitative characters, then one can specify a single set of regimes for all characters or a list of nother regime

specifications, one for each character.

sqrt.alpha These are used to initialize the optimization algorithm. The selection strength

matrix  $\alpha$  and the random drift variance-covariance matrix  $\sigma^2$  are parameterized by their matrix square roots. Specifically, these initial guesses are each packed into lower-triangular matrices (column by column). The product of this matrix

with its transpose is the  $\alpha$  or  $\sigma^2$  matrix. See Details.

sigma These are used to initialize the optimization algorithm. The selection strength

matrix  $\alpha$  and the random drift variance-covariance matrix  $\sigma^2$  are parameterized by their matrix square roots. Specifically, these initial guesses are each packed into lower-triangular matrices (column by column). The product of this matrix

with its transpose is the  $\alpha$  or  $\sigma^2$  matrix. See Details.

## Value

update returns a new fitted-model object of the same class as object.

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