

How does snowpack, Nitrogen, and temperature affect the phenology of an alpine grass?

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This script is the final, summary report of a Bayesian hierarchical analysis of flowering time of *Deschampsia cespitosa* as a function of snow, nitrogen, and temperature. More information can be found on each part of the analysis by referring to the associated scripts available on github.

The following libraries are required for this script

```
library(dplyr)
library(tidyr)
library(lme4)
library(ggplot2)
library(rstanarm)
library(rstan)
library(forcats)
options(mc.cores = parallel::detectCores())
theme_set(theme_grey()) #rstanarm overrides default ggplot theme: set it back

source("source/hpdi.R")
```

1. Introduction

The purpose of this analysis is to evaluate whether first flowering date (FFD) changes in response to increases in temperature, snowpack, and/or N. Data was collected for 2007 and 2008, though I only looked at 2007 data. The data are grouped at 2 levels: block and sub-block. There are a total of 3 blocks, each with 2 sub-blocks. Sub-blocks are separated by a snowfence with plots on the leeward side experiencing normal to reduced snowpack, while plots on the windward side experiencing increased snowpack. In each sub-block, 2 plots have warming treatments, 2 plots have increased nitrogen, 2 plots have both warming and increased nitrogen, and 2 plots are left as controls (ambient N and ambient temperature). Figure 1 (below) captures the conceptual layout of the data structure.

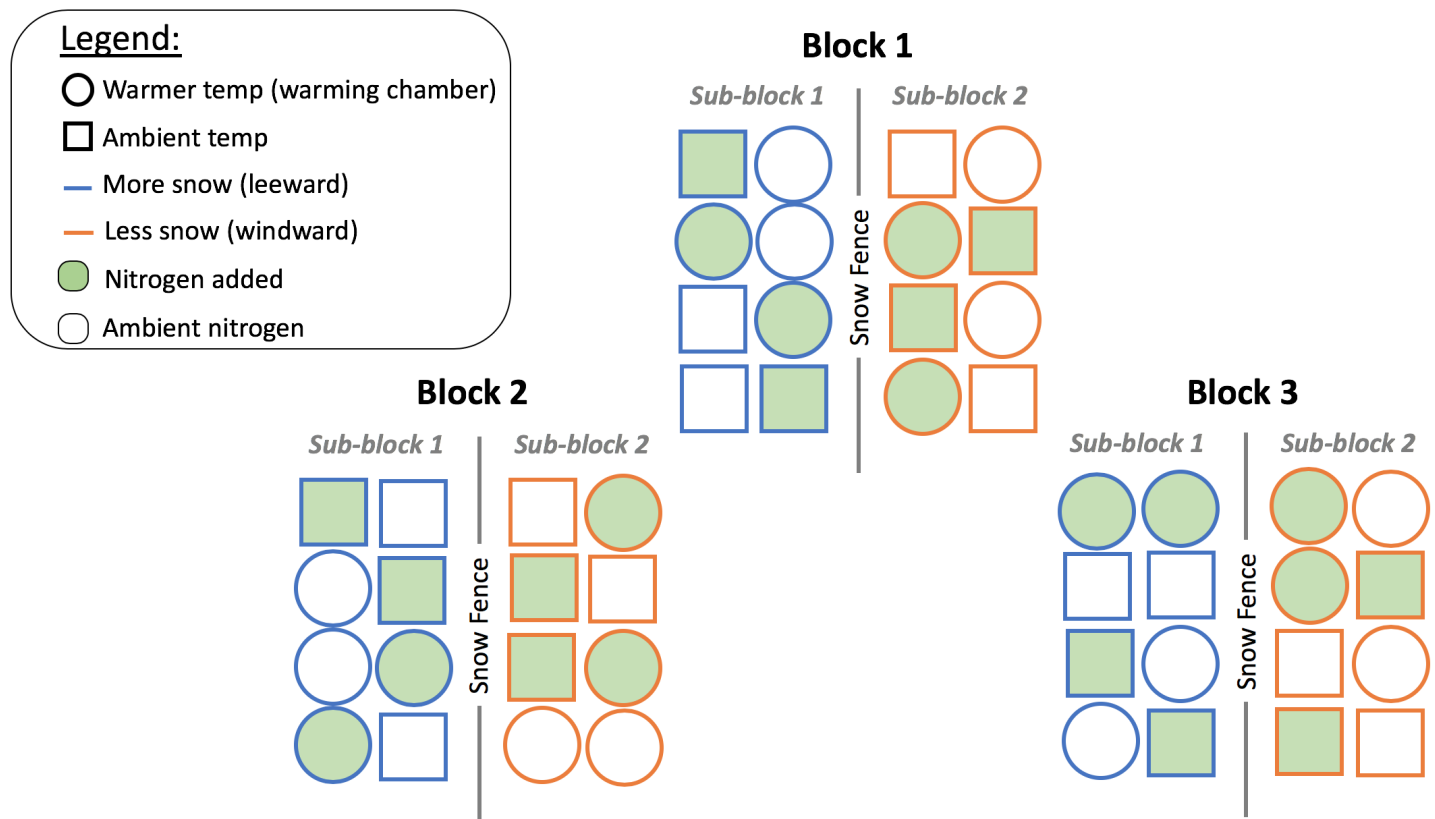


Figure 1. The experimental design shows all 48 plots divided into blocks and sub-blocks. Each plot within a sub-block is 1 meter apart. The blocks are approximately 50 meters apart.

2. Exploring the data

For more, see “[indiv_project_2_eda.md](#)”

Load and clean the data

```
# Load data
nwt_ffd <- read.csv(file = "data/NWT_ITEX_FFD_data_2007_and_2008.csv", na.strings = ".") %>%

# create 0,1 vector for values
mutate(Snow_x = ifelse(Snow == "X", "0", "1")) %>%
mutate(Temp_x = ifelse(Temp == "X", "0", "1")) %>%
mutate(N_x = ifelse(N == "X", "0", "1")) %>%

# rename the values to be more true to their meaning:
mutate(Snow = as.factor(ifelse(Snow == "X", "Reduced_Snowpack", "Increased_Snowpack"))) %>%
mutate(Temp = as.factor(ifelse(Temp == "X", "Normal_Temp", "Higher_Temp"))) %>%
mutate(N = as.factor(ifelse(N == "X", "Ambient_N", "Increased_N"))) %>%

# create a new column for "subblock"
mutate(Subblock = as.factor(ifelse(Snow == "Reduced_Snowpack", 0, 1))) %>%

# define "Block" as a factor
mutate(Block = as.factor(Block)) %>%
```

```

# create a new Block|Subblock column
mutate(Subblock_ID = as.factor(paste0(Block, "_", Subblock))) %>%

# remove 2 unnecessary (and empty) columns
select(-c("X", "X.1"))

# Convert data frame with species as a vector
nwt_ffd2 <- nwt_ffd %>%
  gather(key = Species, value = FFD, 8:35, na.rm = TRUE, factor_key = TRUE) %>%
  mutate(Species = factor(Species)) %>%
  mutate(log_FFD = log(FFD))

# Subset data for 2007 (2008 was incomplete) and centered FFD
nwt_ffd2_2007 <- nwt_ffd2 %>%
  filter(year == "2007") %>%
  mutate(FFD_s = scale(FFD))

# create a new column for FFD based on visit number (rather than Julian day)
days_visited <- as.vector(sort(unique(nwt_ffd2_2007$FFD)))
number_visits <- length(days_visited)
Visit <- as.vector(1:number_visits)
visits_df <- data.frame(days_visited, Visit)
nwt_ffd2_2007 <- merge(nwt_ffd2_2007, visits_df, by.x = "FFD", by.y = "days_visited")

# subset of DESC AE (Deschampsia cespitosa) in 2007, with centered FFD
nwt_descae_2007 <- nwt_ffd2 %>%
  filter(Species == "DESCAE") %>%
  filter(year == "2007") %>%
  merge(visits_df, by.x = "FFD", by.y = "days_visited") %>%
  mutate(FFD_s = scale(FFD))

```

Explore data

```

str(nwt_ffd2_2007)

## 'data.frame':    500 obs. of  17 variables:
## $ FFD          : int  150 150 155 155 155 155 155 155 155 155 ...
## $ year         : int  2007 2007 2007 2007 2007 2007 2007 2007 2007 2007 ...
## $ Block        : Factor w/ 3 levels "1","2","3": 1 3 3 3 3 1 1 3 3 2 ...
## $ Plot         : int   16 42 41 36 44 9 11 43 45 29 ...
## $ Code         : Factor w/ 8 levels "PNW","PNX","PXW",...: 4 1 4 5 3 1 3 2 3 3 ...
## $ Snow         : Factor w/ 2 levels "Increased_Snowpack",...: 1 1 1 2 1 1 1 1 1 1 ...
## $ Temp         : Factor w/ 2 levels "Higher_Temp",...: 2 1 2 1 1 1 1 2 1 1 ...
## $ N            : Factor w/ 2 levels "Ambient_N","Increased_N": 1 2 1 2 1 2 1 2 1 1 ...
## $ Snow_x       : chr   "1" "1" "1" "0" ...
## $ Temp_x       : chr   "0" "1" "0" "1" ...
## $ N_x          : chr   "0" "1" "0" "1" ...
## $ Subblock     : Factor w/ 2 levels "0","1": 2 2 2 1 2 2 2 2 2 2 ...
## $ Subblock_ID : Factor w/ 6 levels "1_0","1_1","2_0",...: 2 6 6 5 6 2 2 6 6 4 ...
## $ Species      : Factor w/ 28 levels "ACOROS","ARTSCO",...: 5 5 5 5 5 5 5 5 18 5 ...
## $ log_FFD      : num   5.01 5.01 5.04 5.04 5.04 ...
## $ FFD_s        : num  [1:500, 1] -2.2 -2.2 -1.86 -1.86 -1.86 ...
## $ Visit        : int   1 1 2 2 2 2 2 2 2 2 ...

```

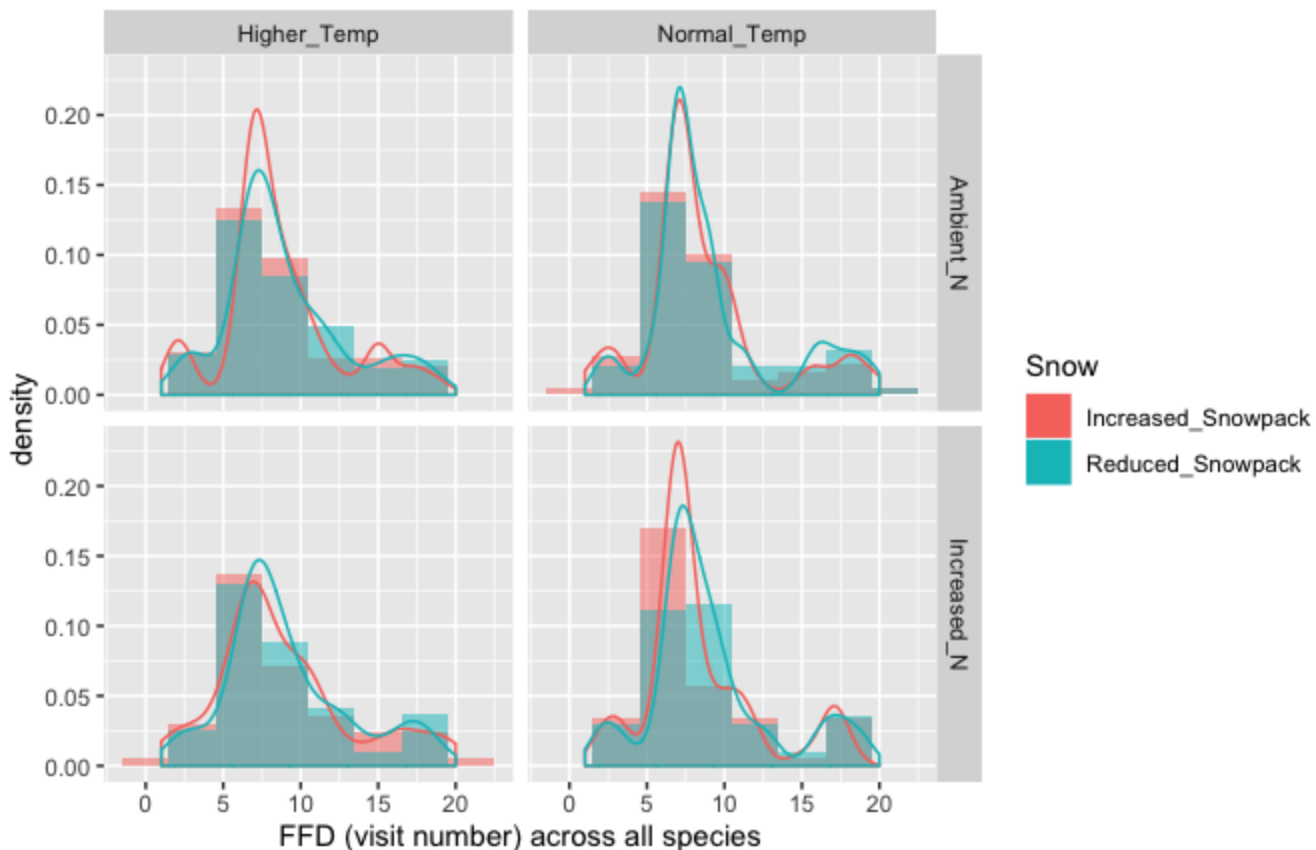
```
## $ visit : int 1 1 2 2 2 2 2 2 2 2 ...
```

```
p2 <- ggplot(data = nwt_ffd2_2007) +
  geom_histogram(mapping = aes(x = Visit, y = stat(density), fill = Snow), position = "identity",
  geom_density(mapping = aes(x = Visit, col = Snow)) +
  facet_grid(N ~ Temp) +
  labs(title = "Figure 2.",
    subtitle = "FFD as a function of Snowpack, Temperature, and Nitrogen",
    x = "FFD (visit number) across all species")
```

p2

Figure 2.

FFD as a function of Snowpack, Temperature, and Nitrogen



Figure

2. A histogram of all species as a function of snowpack, temperature, and nitrogen.

Not all species are equally represented in the dataset:

```
species_list <- unique(nwt_ffd2$Species)
number_obs_df <- data.frame(ncol = 2, nrow = length(species_list))
colnames(number_obs_df) <- c("Species", "NA_count")

number_obs_df <- nwt_ffd2 %>%
  group_by(Species, year) %>%
  summarise(number_obs = n()) %>%
  mutate(number_nas = 48 - number_obs) %>%
  arrange(year, -number_obs)
head(number_obs_df)
```

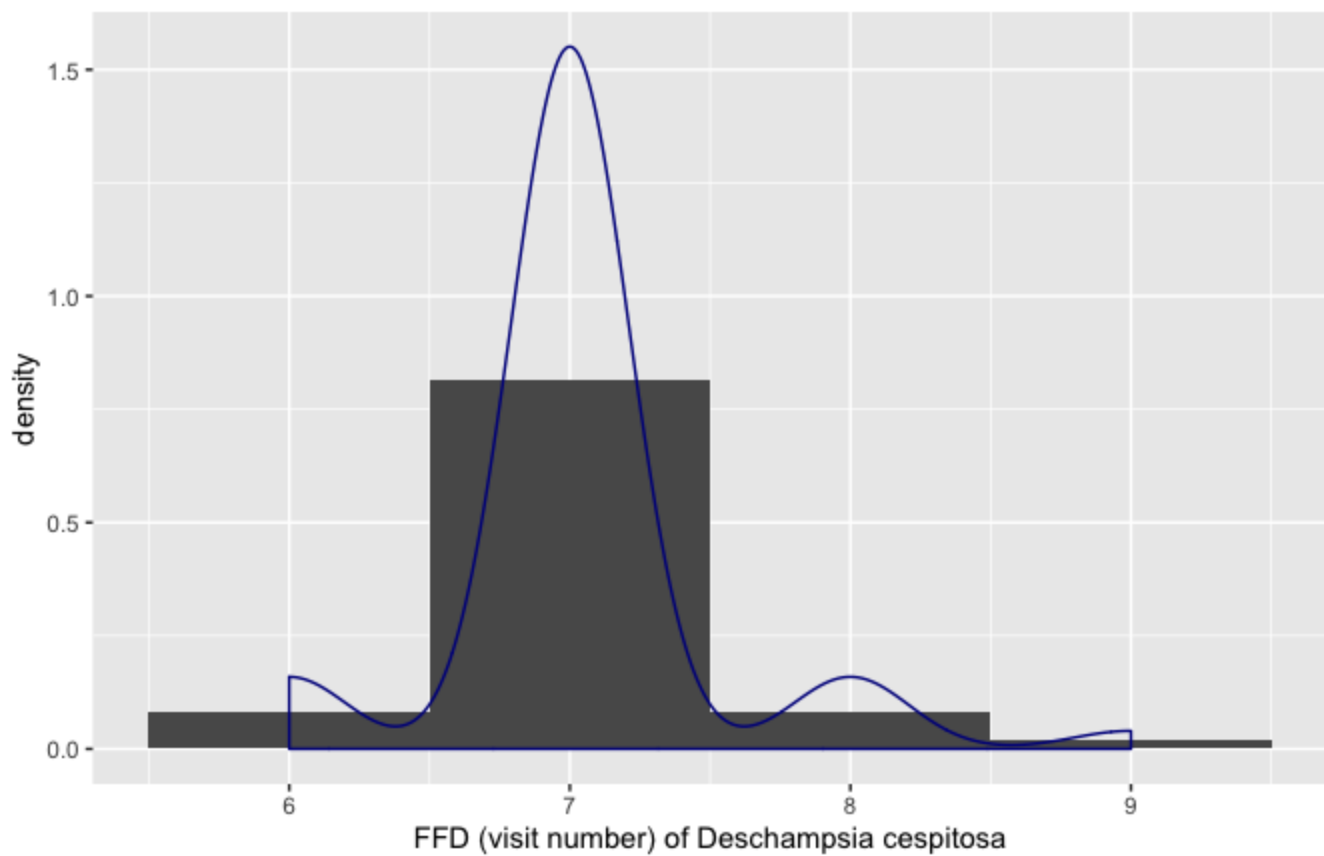
```
## # A tibble: 6 x 4
## # Groups:   Species [6]
##   Species year number_obs number_nas
##   <fct>   <int>     <int>     <dbl>
## 1 DESCAE  2007         48         0
## 2 ACOROS  2007         47         1
## 3 ARTSCO  2007         45         3
## 4 CALLEP  2007         44         4
## 5 GENALG  2007         41         7
## 6 BISBIS  2007         40         8
```

One species, *Deschampsia cespitosa* (DESCAE), is present in all the plots in the 2007 data. For simplification, let's just use *Deschampsia cespitosa* for the hierarchical analysis.

```
p3 <- ggplot() +
  geom_histogram(data = nwt_descae_2007,
    mapping = aes(x = Visit, y = stat(density)),
    binwidth = 1) +
  geom_density(data = nwt_descae_2007,
    mapping = aes(x = Visit),
    col = "darkblue") +
  labs(title = "Figure 3.",
    subtitle = "FFD of Deschampsia cespitosa",
    x = "FFD (visit number) of Deschampsia cespitosa")
p3
```

Figure 3.

FFD of *Deschampsia cespitosa*



Figure

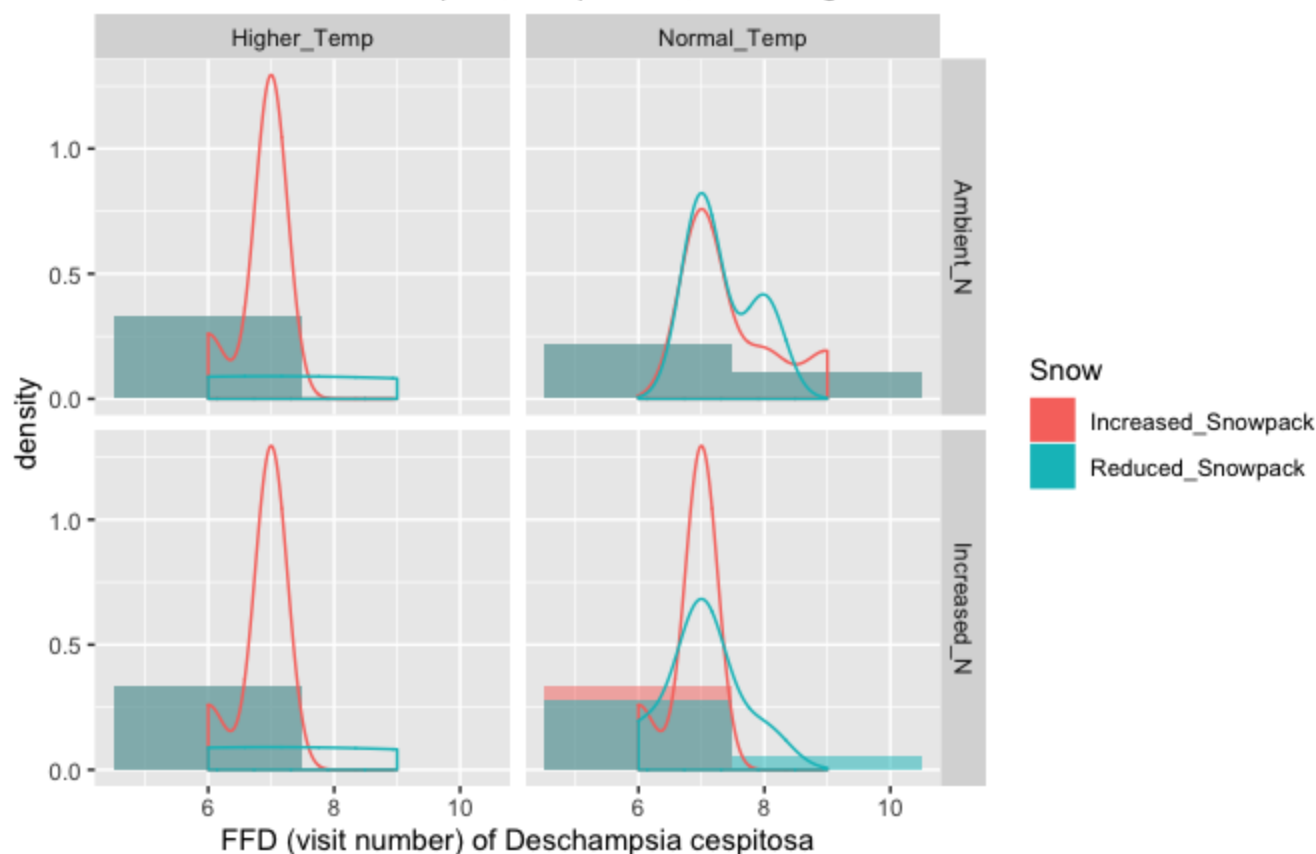
3. A histogram of FFD of *Deschampsia cespitosa* across all plots.

```
p4 <- ggplot(data = nwt_descae_2007) +  
  geom_histogram(mapping = aes(x = Visit, y = stat(density), fill = Snow,), position = "ident"  
  geom_density(mapping = aes(x = Visit, col = Snow)) +  
  facet_grid(N ~ Temp) +  
  labs(title = "Figure 4.",  
        subtitle = "FFD as a function of Snowpack, Temperature, and Nitrogen",  
        x = "FFD (visit number) of Deschampsia cespitosa")
```

p4

Figure 4.

FFD as a function of Snowpack, Temperature, and Nitrogen



Figure

4. A density histogram of FFD as a function of Snowpack, Temperature, and Nitrogen with a density smoother overlaid.

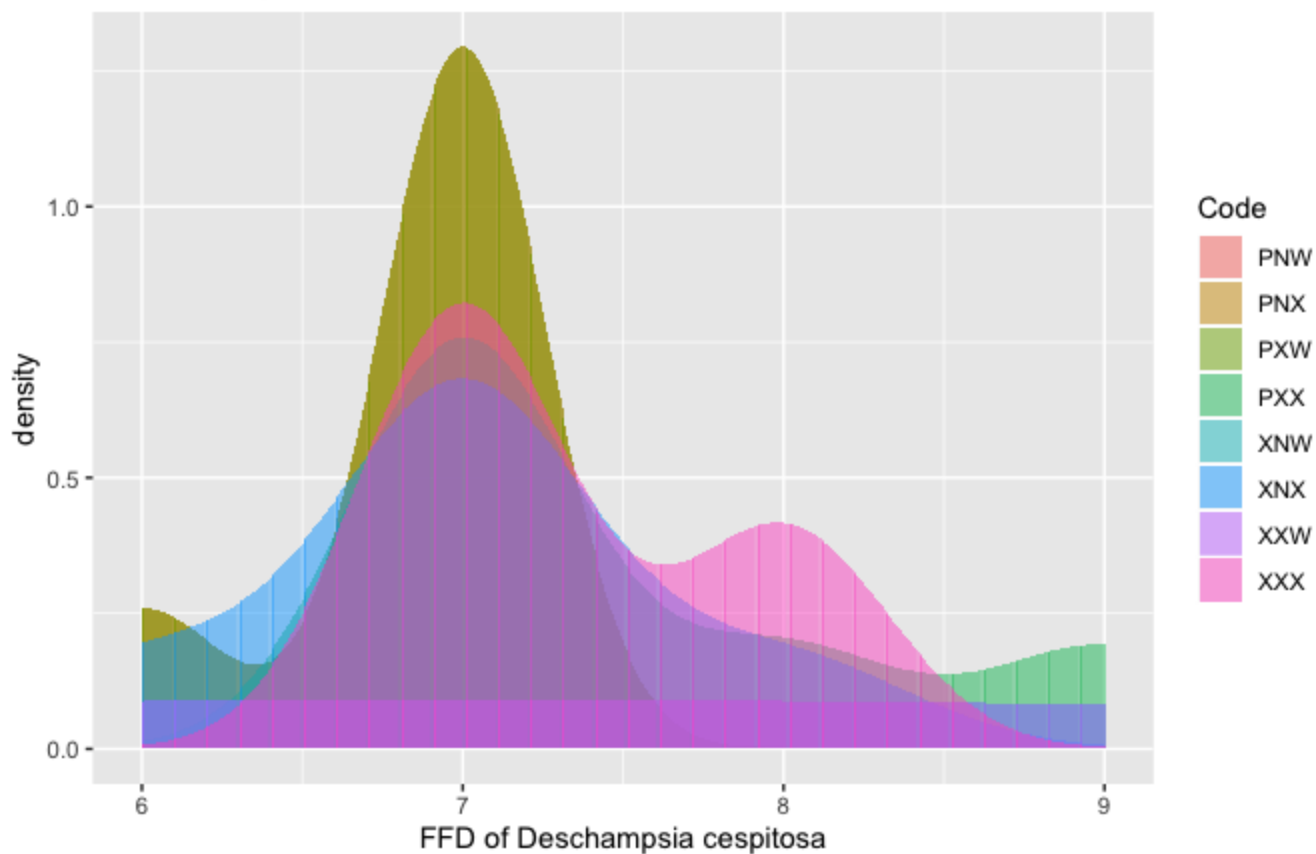
```
p5 <- ggplot(data = nwt_descae_2007) +
  geom_histogram(mapping = aes(x = Visit, y = stat(density), fill = Code), position = "identity",
  labs(title = "Figure 5",
    subtitle = "FFD of all 8 types of plots",
    x = "FFD of Deschampsia cespitosa")
```

Warning: Ignoring unknown parameters: binwidth, bins, pad

p5

Figure 5

FFD of all 8 types of plots



Figure

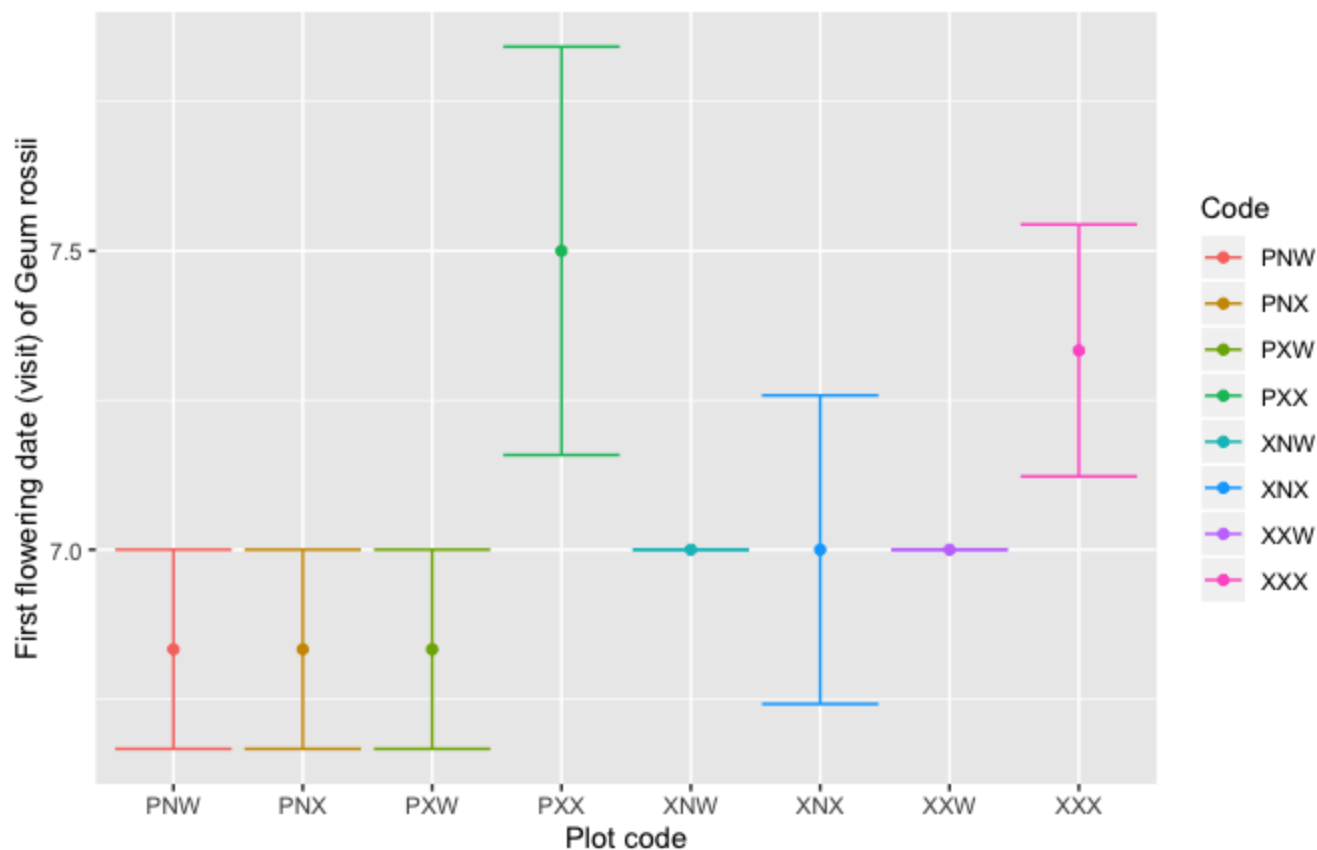
5 A density plot of FFD for *Deschampsia cespitosa* in each of the 8 different types of plots (combinations of snow, nitrogen, and temperature).

```
p6 <- ggplot(data = nwt_descae_2007, aes(x = Code, y = Visit, fill = Code, col = Code)) +  
  stat_summary(geom = "point", fun.y = mean, position = "dodge") +  
  stat_summary(geom = "errorbar", fun.data = mean_se, position = "dodge") +  
  labs(title = "Figure 6",  
        subtitle = "First flowering data of each plot type",  
        x = "Plot code",  
        y = "First flowering date (visit) of Geum rossii")
```

p6

Figure 6

First flowering data of each plot type



Figure

6. Mean and SE of FFD for *Deschampsia cespitosa* for all plot types.

3. Define conceptual (biological) model

The purpose of the model is to evaluate how snowpack, temperature, and nitrogen impact the phenology of alpine plants. As discussed above, I will only look at one species (*Deschampsia cespitosa*) in one year (2007) for the sake of simplicity.

The structure of the experimental design has **2 grouping variables**: blocks (3 replicates separated in space) and 2 sub-blocks per block (separated by a snowfence).

There are **3 explanatory variables**: snowpack (*Snow*), temperature (*Temp*), and nitrogen (*N*).

The **response variable** is the first flowering date of *Deschampsia cespitosa*, which is indicated by the visit number (*visit*).

The following conceptual model considers all three explanatory variables and their interactions. I predicted that interactions might be important. I will evaluate this assumption when I build and compare models later in the script.

Plot level:

$$y_i \sim \text{Normal}(\mu_i, \sigma_y^2)$$

$$\mu_i = \alpha_{j[i]} + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_1 x_2$$

where...

- y_i represents the observed value of FFD for a given plot i
- μ_i represents the expected FFD for plot i
- $\alpha_{j[i]}$ represents the expected value for a plot i given it is within sub-block j
- x_1 represents N (2 levels: increased and ambient) and
- x_2 represents temperature (2 levels: increased and ambient)
- x_3 represents snow (2 levels: increased or decreased)

Subblock level:

$$\alpha_j \sim Normal(\gamma_{k[i]}, \sigma_\mu^2)$$

$$\gamma_k = \omega_k + \beta_4 x_3 + \beta_5 x_1 x_3$$

- $\beta_6 x_2 x_3 + \beta_7 x_1 x_2 x_3$ (https://latex.codecogs.com/png.latex?%5Cgamma_k%20%3D%20%5Comega_%7Bk%5Bi%5D%7D%20%2B%20%5Cbeta_4%20x_3%20%2B%20%5Cbeta_5%20x_1%20x_3%20%2B%20%5Cbeta_6%20x_2%20x_3%20%2B%20%5Cbeta_7%20x_1%20x_2%20x_3 "\gamma_k = \omega_{k[i]} + \beta_4 x_3 + \beta_5 x_1 x_3 + \beta_6 x_2 x_3 + \beta_7 x_1 x_2 x_3")

where....

- x_1 represents N (2 levels: increased and ambient),
- x_2 represents temperature (2 levels: increased and ambient)

Block level:

$$\omega_k \sim Normal(\bar{\omega}, \sigma_\omega^2)$$

where...

- ω_k represents the mean among sub-blocks in blocks,
- $\bar{\omega}$ represents the mean among blocks, and
- σ_ω^2 represents the variance among blocks

Alternative Parameterization:

$$y_i = x_1 * x_2 * x_3 + b_k + s_j + e_i$$

which expands to...

$$y_i = \beta_0 + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_1 x_2 + \beta_4 x_3 + \beta_5 x_1 x_3 + \beta_6 x_2 x_3 + \beta_7 x_1 x_2 x_3 + b_k + s_j + e_i$$

where...

- x_1 represents N (2 levels: increased and ambient),
- x_2 represents temperature (2 levels: increased and ambient),
- x_3 represents snow (2 levels: increased or decreased),
- b_k represents variance of a specific block k from the mean of all blocks such that $b_k \sim Normal(0, \sigma_\omega^2)$,
- s_j represents variance of a specific sub-block j from the mean of sub-blocks such that

$s_j \sim \text{Normal}(0, \sigma_\mu^2)$, and

- e_i represents variance of a specific plot i from the mean of plots such that $e_i \sim \text{Normal}(\sigma_y^2)$

Build and run Bayesian hierarchical models

A selection of models are presented here for comparison. The following models were chosen based on a much more thorough model selection comparison in “indiv_project_5_model_selection”.

To start, let’s look at the proposed conceptual model, which includes snow, temperature, nitrogen, and all their interactions as explanatory variables. I will use a normal, “gaussian” distribution for now, but I will return to this assumption later. Let’s call this “**Model 1**”.

Model 1: N x Temp x Snow

```
# Model 1: Visit, gaussian
bayes_NxTxS <- stan_glmer(Visit ~ N*Temp*Snow + (1|Block / Subblock),
                          family = gaussian,
                          data = nwt_descae_2007,
                          adapt_delta = 0.999999)
```

I used default priors for the model, which mean that all parameters (including the intercept) have a default prior mean of 0, with variance of 10 for the intercept and 2.5 for the other parameters (default for normal distributions). The default priors are appropriate for this model because they are of the appropriate magnitude for the data, are weakly informative, and I don’t have a good reason to use any other prior.

```
prior_summary(bayes_NxTxS)

## Priors for model 'bayes_NxTxS'
## -----
## Intercept (after predictors centered)
## ~ normal(location = 0, scale = 10)
##      **adjusted scale = 5.04
##
## Coefficients
## ~ normal(location = [0,0,0,...], scale = [2.5,2.5,2.5,...])
##      **adjusted scale = [1.26,1.26,1.26,...]
##
## Auxiliary (sigma)
## ~ exponential(rate = 1)
##      **adjusted scale = 0.50 (adjusted rate = 1/adjusted scale)
##
## Covariance
```

```
## ~ decov(reg. = 1, conc. = 1, shape = 1, scale = 1)
## -----
## See help('prior_summary.stanreg') for more details
```

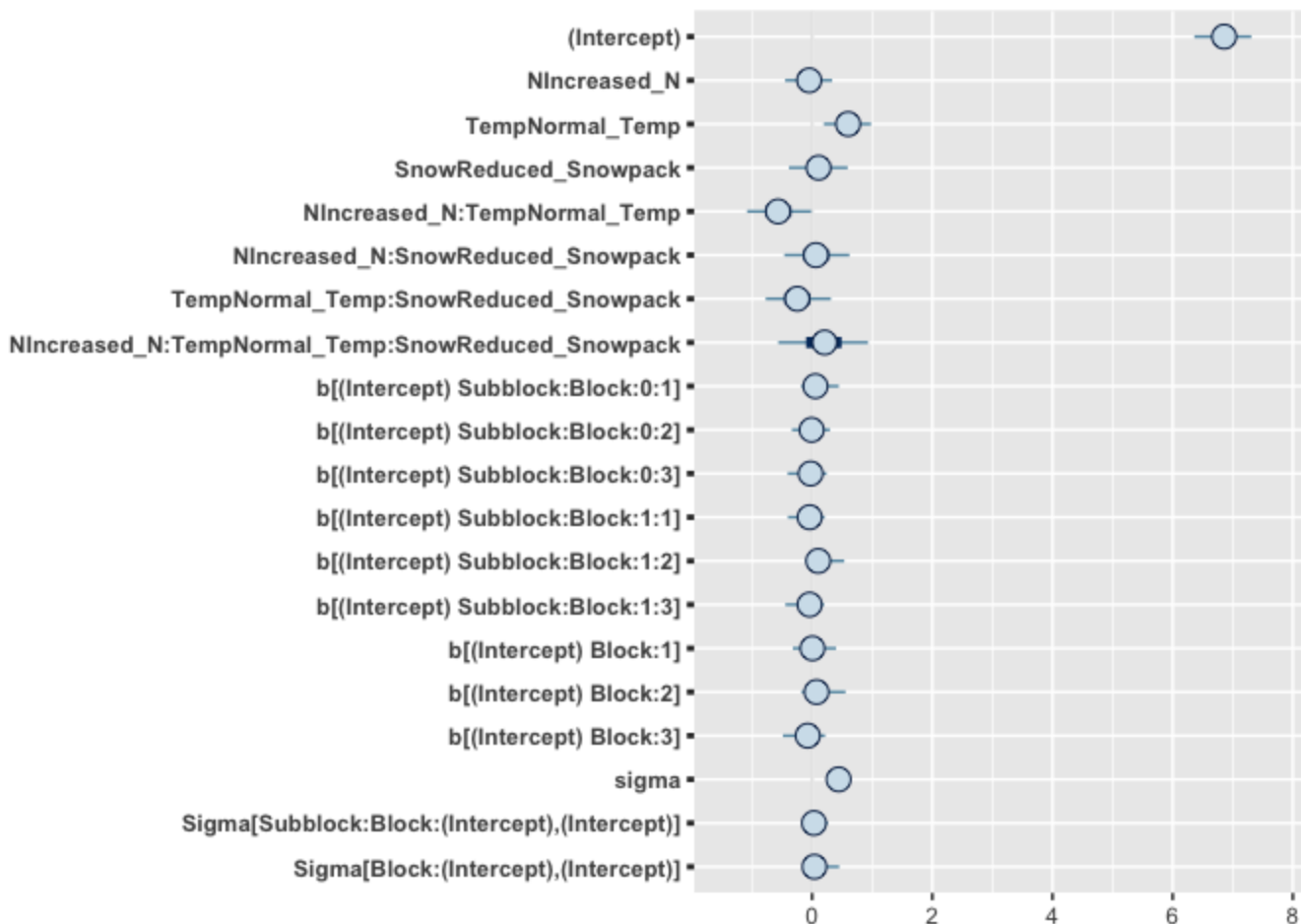
The covariance plot looks good. Values are below +/- 0.7:

```
vcov(bayes_NxTxS, correlation=TRUE)
```

The diagnostics look fine too:

```
launch_shinystan(bayes_NxTxS)
```

```
plot(bayes_NxTxS)
```



Figure

7. Looking at the posterior distribution, it appears that only nitrogen and temperature had a non-zero slope. Increased nitrogen or increased temperature appear to cause earlier flowering in *Deschampsia cespitosa*. The interaction between N and temperature is on the cusp, but interestingly, shows that the combination of increased N and increased temperature can delay flowering.

Based on the results of the conceptual model, it appears that only N and temperature (and maybe their interaction) affect the flowering time of *Deschampsia cespitosa*. To explore this, here is “**Model**

2", which only includes N, temperature, and N:temperature.

Model 2: N x Temp

```
# Model 2: Visit, gaussian
bayes_NxT <- stan_glmer(Visit ~ N*Temp + (1|Block / Subblock),
  family = gaussian,
  data = nwt_descae_2007,
  adapt_delta = 0.999999)
```

I chose to use default priors because they are weakly informative and I don't have a good reason to use any other prior.

```
prior_summary(bayes_NxT)

## Priors for model 'bayes_NxT'
## -----
## Intercept (after predictors centered)
## ~ normal(location = 0, scale = 10)
##      **adjusted scale = 5.04
##
## Coefficients
## ~ normal(location = [0,0,0], scale = [2.5,2.5,2.5])
##      **adjusted scale = [1.26,1.26,1.26]
##
## Auxiliary (sigma)
## ~ exponential(rate = 1)
##      **adjusted scale = 0.50 (adjusted rate = 1/adjusted scale)
##
## Covariance
## ~ decov(reg. = 1, conc. = 1, shape = 1, scale = 1)
## -----
## See help('prior_summary.stanreg') for more details
```

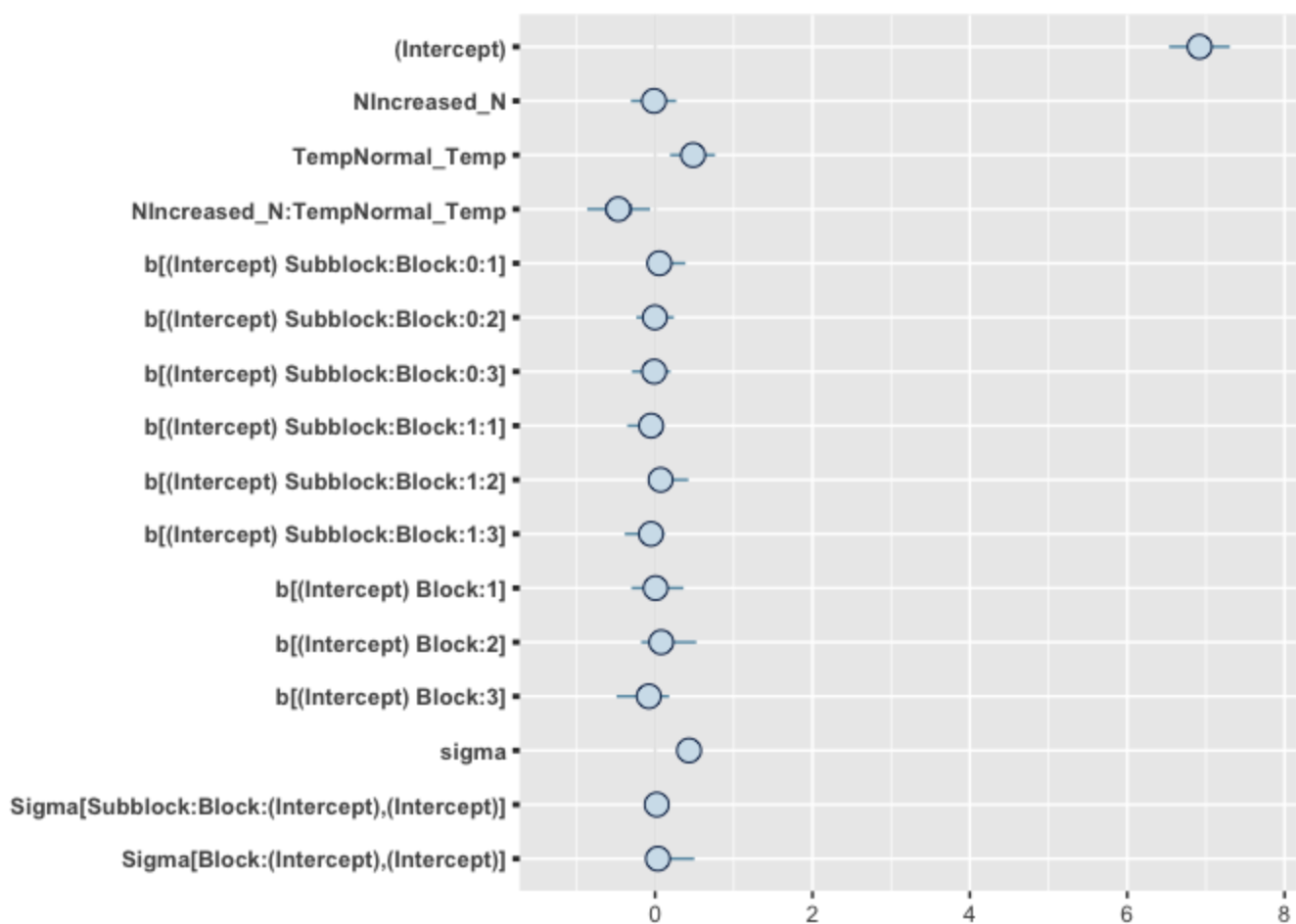
The covariance plot looks good. Values are below +/- 0.7:

```
vcov(bayes_NxT, correlation=TRUE)
```

The diagnostics look fine too:

```
launch_shinystan(bayes_NxT)
```

```
plot(bayes_NxT)
```



Figure

8. The posterior distribution for model 2 looks similar to model 1, with only nitrogen and temperature having a non-zero slope. The interaction between N and temperature is still on the cusp.

For exploration sake, let's build __“model 3”, which excludes the interaction term.

****Model 3: N + Temp ****

```
# Model 1: Visit, gaussian
bayes_NT <- stan_glm(Visit ~ N + Temp + (1|Block / Subblock),
  family = gaussian,
  data = nwt_descae_2007,
  adapt_delta = 0.99999999)
```

```
## Warning: There were 1 divergent transitions after warmup. Increasing adapt_delta above 0.999
## http://mc-stan.org/misc/warnings.html#divergent-transitions-after-warmup
```

```
## Warning: Examine the pairs() plot to diagnose sampling problems
```

I chose to use default priors because they are weakly informative and I don't have a good reason to use any other prior.

```
prior_summary(bayes_NT)
```

```

## Priors for model bayes_NT'
## -----
## Intercept (after predictors centered)
## ~ normal(location = 0, scale = 10)
## **adjusted scale = 5.04
##
## Coefficients
## ~ normal(location = [0,0], scale = [2.5,2.5])
## **adjusted scale = [1.26,1.26]
##
## Auxiliary (sigma)
## ~ exponential(rate = 1)
## **adjusted scale = 0.50 (adjusted rate = 1/adjusted scale)
##
## Covariance
## ~ decov(reg. = 1, conc. = 1, shape = 1, scale = 1)
## -----
## See help('prior_summary.stanreg') for more details

```

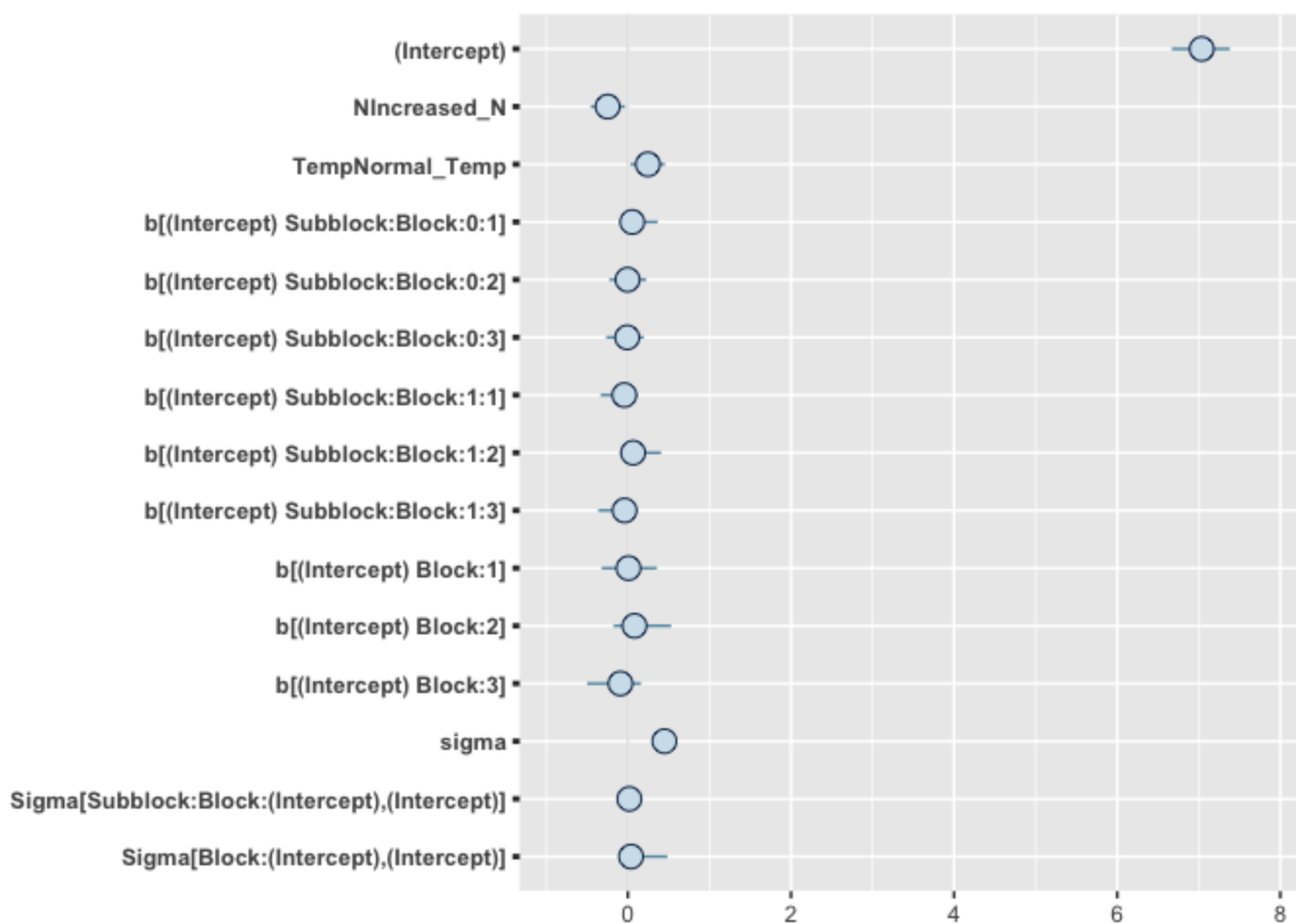
The covariance plot looks good. Values are below +/- 0.4:

```
vcov(bayes_NT, correlation=TRUE)
```

The diagnostics look fine too:

```
launch_shinystan(bayes_NT)
```

```
plot(bayes_NT)
```



Figure

9. *The results of the posterior distribution for model 3 match the results from model 1 and model 2. The posterior shows that increased N or increased temperature lead to earlier flowering.*

To compare the models, I used LOOIC. The LOOIC values for each model are reported below and show that the model with nitrogen, temperature, and nitrogen:temperature is likely the preferred model. The SE for the LOOIC values is relatively large, however, so all three models could be used. Fewer parameters is better, so either model 2 (N x temperature) or model 3 (N + temperature) are good contenders. I chose to explore model 2 (N x temperature) for the rest of the analysis because (1) it has the lowest LOOIC and (2) the interaction term has the opposite effect than each of the individual terms (the interaction delays flowering, while the individual terms cause earlier flowering), which could be biologically important.

```
loo_NxTxS <- loo(bayes_NxTxS, k_threshold = 0.7)

## 2 problematic observation(s) found.
## Model will be refit 2 times.

##
## Fitting model 1 out of 2 (leaving out observation 1)

##
## Fitting model 2 out of 2 (leaving out observation 48)
```



```

loo_NxT <- loo(bayes_NxT, k_threshold = 0.7)

## All pareto_k estimates below user-specified threshold of 0.7.
## Returning loo object.

loo_NT <- loo(bayes_NT, k_threshold = 0.7)

## 1 problematic observation(s) found.
## Model will be refit 1 times.

##
## Fitting model 1 out of 1 (leaving out observation 48)

compare_models(loo_NxTxS, loo_NxT, loo_NT)

##
## Model comparison:
## (ordered by highest ELPD)
##
##               elpd_diff se_diff
## bayes_NxT      0.0         0.0
## bayes_NT     -1.4         2.0
## bayes_NxTxS  -3.2         1.1

```

Simulate data to test model

Before continuing exploration of the model, I want to test the model algorithm using simulated data on a null model.

Define parameters

```

# Parameters for block level
meanblock <- 5.32 # also omega_bar (since data is centered, the value should be 0: 0 = average,
vblock <- 0.72 # variation between block 1, 2, 3: close to 1 day
nblock <- 3

#simulate block level
block_sim <- rnorm(nblock, meanblock, sqrt(vblock)) # the output would be the omega_k (see equa

# parameters for subblock level
vsub <- 1.37 # bigger than between block variance (snow might be different?)
nsub <- 2 # 2 for each block

```

```

# simulate subblock level
blockmean_vector <- rep(block_sim, each = nsub) # repeat simulated mean of each block 2x (since
subblock_sim <- rnorm(nsub*nblock, blockmean_vector, sqrt(vsub))

# parameters for data level
vy <- 2.32 # variance in FFD
n <- 32 # number of data points within a sub-block (32 = 4x as many as reality)

# Simulate data level (vectorized)
subblockmean_vector <- rep(subblock_sim, each = n) # repeat simulated mean of each subblock n t
y <- rnorm(n*nsub*nblock, subblockmean_vector, sqrt(vy)) #Simulate y depending on group means

# Compile into a dataframe
dat <- data.frame(Visit = y,
                  Block = factor(rep(1:nblock, each = nsub)),
                  Subblock = factor(rep(1:nsub, each = n)))
dat <- dat[order(dat$Block, dat$Subblock),]
head(dat)

```

```

##      Visit Block Subblock
## 1  1.107648     1         1
## 2  6.763521     1         1
## 7  6.002802     1         1
## 8  7.108182     1         1
## 13 4.429291     1         1
## 14 4.378326     1         1

```

```

# Add a random factor for species
dat2 <- data.frame(Visit = y,
                  Block = factor(rep(1:nblock, each = nsub)),
                  Subblock = factor(rep(1:nsub, each = n)),
                  Species = sample(1:20, n*nsub*nblock, replace = TRUE))
head(dat2)

```

```

##      Visit Block Subblock Species
## 1  1.107648     1         1         7
## 2  6.763521     1         1        15
## 3  4.130240     2         1        20
## 4  4.063392     2         1        13
## 5  2.504303     3         1        15
## 6  5.915298     3         1        13

```

Null model

```

# Null model (no explanatory variables)
bayes_null_sim <- stan_glmer(Visit ~ 1 + (1 | Block / Subblock),

```

```

      data = dat,
      family = gaussian,
      adapt_delta = 0.99999)
summary(bayes_null_sim)[,c(1,3,10,9)] #a sample of the full output

```

```

##                               mean      sd
## (Intercept)                   5.53952093 0.5113716
## b[(Intercept) Subblock:Block:1:1] 0.01128872 0.3287982
## b[(Intercept) Subblock:Block:1:2] 0.27413037 0.3933258
## b[(Intercept) Subblock:Block:1:3] 0.09269091 0.3265139
## b[(Intercept) Subblock:Block:2:1] -0.14624495 0.3469849
## b[(Intercept) Subblock:Block:2:2] -0.10643360 0.3515429
## b[(Intercept) Subblock:Block:2:3] -0.14646444 0.3361924
## b[(Intercept) Block:1]           -0.14816496 0.5102817
## b[(Intercept) Block:2]           0.17802224 0.5203611
## b[(Intercept) Block:3]           -0.04753336 0.5060269
## sigma                           2.08200813 0.1081697
## Sigma[Subblock:Block:(Intercept),(Intercept)] 0.25472921 0.5265494
## Sigma[Block:(Intercept),(Intercept)] 0.75434549 2.3964752
## mean_PPD                        5.52843455 0.2092060
## log-posterior                   -438.15149944 3.4216629
##                               Rhat n_eff
## (Intercept)                   1.0050652 1162
## b[(Intercept) Subblock:Block:1:1] 0.9998293 3172
## b[(Intercept) Subblock:Block:1:2] 1.0003011 2388
## b[(Intercept) Subblock:Block:1:3] 1.0012033 2917
## b[(Intercept) Subblock:Block:2:1] 0.9998183 3130
## b[(Intercept) Subblock:Block:2:2] 1.0005388 2684
## b[(Intercept) Subblock:Block:2:3] 1.0009352 3032
## b[(Intercept) Block:1]           1.0035180 1280
## b[(Intercept) Block:2]           1.0028532 1283
## b[(Intercept) Block:3]           1.0035473 1536
## sigma                           1.0004380 3661
## Sigma[Subblock:Block:(Intercept),(Intercept)] 1.0005745 1937
## Sigma[Block:(Intercept),(Intercept)] 1.0009899 1634
## mean_PPD                        1.0006726 4220
## log-posterior                   1.0037124 996

```

```

# Explore the number of columns here so that I can adjust ncol in keep dataframe (when setting
results <- c(fixef(bayes_null_sim), as.data.frame(VarCorr(bayes_null_sim))[,4])

loo_bayes_null_sim <- loo(bayes_null_sim)

```

Repeat simulation numerous times (5000 or so is best, but I'll do 10)

```

# Setup
reps <- 10 #Number of replicate simulations
keep <- matrix(NA, nrow = reps, ncol = 8) # it was 6, but I think it will be 8 for my data

# Useful to add a timer. You could set reps to say 10 at first, then calculate
# how long it will take to run a larger simulation.
system.time(
for ( i in 1:reps ) {

# Simulate block level
block_sim <- rnorm(nblock, meanblock, sqrt(vblock))

# simulate subblock level
blockmean_vector <- rep(block_sim, each = nsub) # repeat simulated mean of each block 2x (s
subblock_sim <- rnorm(nsub*nblock, blockmean_vector, sqrt(vsub))

# Simulate data level (vectorized)
subblockmean_vector <- rep(subblock_sim, each = n) # repeat simulated mean of each subblock
y <- rnorm(n*nsub*nblock, subblockmean_vector, sqrt(vy)) #Simulate y depending on group mea

# Fit model to simulated data
dat <- data.frame(Visit = y,
                  Block = factor(rep(1:nblock, each = nsub)),
                  Subblock = factor(rep(1:nsub, each = n)))
mlfit <- lmer(Visit ~ 1 + (1|Block/Subblock),
             data = dat,
             REML = FALSE)
baysfit <- stan_lmer(Visit ~ 1 + (1|Block/Subblock),
                   data = dat,
                   adapt_delta = 0.9999999999999999,
                   chains = 3)

# Record results (VarCorr extracts the variance estimates)
keep[i,] <- c(fixef(mlfit), as.data.frame(VarCorr(mlfit))[,4],
             fixef(baysfit), as.data.frame(VarCorr(baysfit))[,4] )

# Tidy up
rm(block_sim, blockmean_vector, subblock_sim, subblockmean_vector, y, dat, mlfit, baysfit)

# Monitoring progress
if ( i %% 10 == 0 ) {
  print(i)
}
}
)
save(keep, file="indiv_project_simulation_kelsey-e.RData") #Save the simulation for future use

```

The model had trouble converging on 3 of the 10 simulation, despite an adapt_delta very close to 1 and reducing the number of chains to 3. This may be an indication of a bigger problem with the model structure.

```
load(file="indiv_project_simulation_kelsey-e.RData")
head(keep)
```

```
##           [,1]           [,2]           [,3]           [,4]           [,5]           [,6]
## [1,] 5.702924 3.085854e-15 0.008320889 3.002527 5.691862 0.12220676
## [2,] 4.597481 0.000000e+00 0.000000000 4.655488 4.610469 0.24137425
## [3,] 5.623230 0.000000e+00 0.000000000 7.450803 5.593068 0.18836378
## [4,] 5.301740 0.000000e+00 0.000000000 5.593922 5.304842 0.19020921
## [5,] 5.452605 0.000000e+00 0.000000000 2.709984 5.444654 0.06920163
## [6,] 5.746652 0.000000e+00 0.000000000 6.843679 5.735840 0.19106128
##           [,7]           [,8]
## [1,] 0.4748647 2.997167
## [2,] 0.5598741 4.652053
## [3,] 1.2221168 7.487201
## [4,] 0.7221918 5.615731
## [5,] 0.3589208 2.736293
## [6,] 0.8188178 6.910245
```

```
# Expected values of the algorithm
```

```
ml_blockmean <- mean(keep[,1])
ml_vblock <- mean(keep[,2])
ml_vsub <- mean(keep[,3])
ml_vy <- mean(keep[,4])
b_blockmean <- mean(keep[,5])
b_vblock <- mean(keep[,6])
b_vsub <- mean(keep[,7])
b_vy <- mean(keep[,8])
```

Compare to the true values (as determined in defining the simulation data)

```
# block-level mean
cbind(meanblock,ml_blockmean,b_blockmean)
```

```
##      meanblock ml_blockmean b_blockmean
## [1,]      5.32      5.371151      5.36386
```

```
# block-level variance
cbind(vblock,ml_vblock,b_vblock)
```

```
##      vblock ml_vblock b_vblock
## [1,]    0.72 0.01156604 0.1903296
```

```
# sub-block level variance
cbind(vsub,ml_vsub,b_vsub)

##          vsub      ml_vsub      b_vsub
## [1,] 1.37 0.008716926 0.6550091

# data-level variance
cbind(vy,ml_vy,b_vy)

##          vy      ml_vy      b_vy
## [1,] 2.32 4.504792 4.523215
```

Uh oh! The model did not do a good job recovering the values. The block-level means were pretty close, but the model did not have comparable values for the variance at the block, sub-block, or data-levels. The poor recovery of our initial values may suggest that the data would be better fit to another model structure. After discussions with Brett, the data structure might do better with a binomial, “event-based” approach (see “binomial_test.R” that Brett posted in my github repository). I will explore alternative models in the future, but for the remainder of this script, I will continue to use the normally distributed model.

Summary of Bayesian Model

```
summary(bayes_NxT)[,c(1,3,10,9)] #a sample of the full output
```

```
##                                     mean      sd
## (Intercept)                      6.918505874 0.25368403
## NIncreased_N                     -0.016084301 0.17460755
## TempNormal_Temp                   0.479989179 0.17509011
## NIncreased_N:TempNormal_Temp     -0.466612874 0.24138665
## b[(Intercept) Subblock:Block:0:1] 0.088268695 0.15660531
## b[(Intercept) Subblock:Block:0:2] -0.004677098 0.14800648
## b[(Intercept) Subblock:Block:0:3] -0.030498544 0.15353729
## b[(Intercept) Subblock:Block:1:1] -0.079549400 0.15158463
## b[(Intercept) Subblock:Block:1:2]  0.108114364 0.16427823
## b[(Intercept) Subblock:Block:1:3] -0.087942334 0.16179618
## b[(Intercept) Block:1]            0.018628394 0.22406524
## b[(Intercept) Block:2]            0.117874091 0.23379745
## b[(Intercept) Block:3]           -0.113047162 0.23403069
## sigma                            0.435122895 0.04887512
## Sigma[Subblock:Block:(Intercept),(Intercept)] 0.046593746 0.07808762
## Sigma[Block:(Intercept),(Intercept)] 0.116277541 0.29209433
## mean_PPD                          7.041013621 0.09065270
```

```
## log-posterior -54.190071384 3.55417721
##
## Rhat n_eff
## (Intercept) 1.0005151 1271
## NIncreased_N 1.0002932 3169
## TempNormal_Temp 0.9993627 2861
## NIncreased_N:TempNormal_Temp 1.0004930 2571
## b[(Intercept) Subblock:Block:0:1] 1.0000449 2765
## b[(Intercept) Subblock:Block:0:2] 0.9997105 2568
## b[(Intercept) Subblock:Block:0:3] 1.0018612 2532
## b[(Intercept) Subblock:Block:1:1] 0.9998627 2848
## b[(Intercept) Subblock:Block:1:2] 1.0015584 2000
## b[(Intercept) Subblock:Block:1:3] 1.0011147 2356
## b[(Intercept) Block:1] 1.0014215 1246
## b[(Intercept) Block:2] 1.0026153 1254
## b[(Intercept) Block:3] 1.0000364 1123
## sigma 1.0005398 3538
## Sigma[Subblock:Block:(Intercept),(Intercept)] 1.0005446 2021
## Sigma[Block:(Intercept),(Intercept)] 1.0024770 1729
## mean_PPD 0.9998270 4095
## log-posterior 1.0014933 1303
```

Note that the Rhat values are all close to, but not quite 1. The number of effective replicates, which would ideally be 4000, is not always that high.

```
posterior_interval(bayes_NxT,prob=0.95)
```

```
##
## 2.5% 97.5%
## (Intercept) 6.392569e+00 7.410384886
## NIncreased_N -3.603532e-01 0.322070552
## TempNormal_Temp 1.256657e-01 0.828611944
## NIncreased_N:TempNormal_Temp -9.374868e-01 -0.004448939
## b[(Intercept) Subblock:Block:0:1] -1.735805e-01 0.455135822
## b[(Intercept) Subblock:Block:0:2] -3.173747e-01 0.320166899
## b[(Intercept) Subblock:Block:0:3] -3.867671e-01 0.267552633
## b[(Intercept) Subblock:Block:1:1] -4.411568e-01 0.179059525
## b[(Intercept) Subblock:Block:1:2] -1.409033e-01 0.501622849
## b[(Intercept) Subblock:Block:1:3] -4.547385e-01 0.174235879
## b[(Intercept) Block:1] -4.389073e-01 0.523240228
## b[(Intercept) Block:2] -2.695108e-01 0.685669579
## b[(Intercept) Block:3] -6.519420e-01 0.304196891
## sigma 3.523896e-01 0.541939189
## Sigma[Subblock:Block:(Intercept),(Intercept)] 4.980471e-05 0.245970867
## Sigma[Block:(Intercept),(Intercept)] 1.261421e-04 0.750479635
```

```
par(mfrow = c(1,1))
bayes_NxT$stanfit
```

```
## Inference for Stan model: continuous.
## 4 chains. each with iter=2000; warmup=1000; thin=1:
```

```

## 1 chain, each with 1000 warmup draws, then 10000
## post-warmup draws per chain=1000, total post-warmup draws=4000.
##
##
##
##
## (Intercept) 6.92 0.01 0.25
## NIncreased_N -0.02 0.00 0.17
## TempNormal_Temp 0.48 0.00 0.18
## NIncreased_N:TempNormal_Temp -0.47 0.00 0.24
## b[(Intercept) Subblock:Block:0:1] 0.09 0.00 0.16
## b[(Intercept) Subblock:Block:0:2] 0.00 0.00 0.15
## b[(Intercept) Subblock:Block:0:3] -0.03 0.00 0.15
## b[(Intercept) Subblock:Block:1:1] -0.08 0.00 0.15
## b[(Intercept) Subblock:Block:1:2] 0.11 0.00 0.16
## b[(Intercept) Subblock:Block:1:3] -0.09 0.00 0.16
## b[(Intercept) Subblock:Block:_NEW_Subblock:Block] 0.00 0.00 0.22
## b[(Intercept) Block:1] 0.02 0.01 0.22
## b[(Intercept) Block:2] 0.12 0.01 0.23
## b[(Intercept) Block:3] -0.11 0.01 0.23
## b[(Intercept) Block:_NEW_Block] 0.00 0.01 0.33
## sigma 0.44 0.00 0.05
## Sigma[Subblock:Block:(Intercept),(Intercept)] 0.05 0.00 0.08
## Sigma[Block:(Intercept),(Intercept)] 0.12 0.01 0.29
## mean_PPD 7.04 0.00 0.09
## log-posterior -54.19 0.10 3.55
##
## 2.5% 25% 50%
## (Intercept) 6.39 6.78 6.92
## NIncreased_N -0.36 -0.13 -0.01
## TempNormal_Temp 0.13 0.36 0.48
## NIncreased_N:TempNormal_Temp -0.94 -0.63 -0.47
## b[(Intercept) Subblock:Block:0:1] -0.17 0.00 0.05
## b[(Intercept) Subblock:Block:0:2] -0.32 -0.07 0.00
## b[(Intercept) Subblock:Block:0:3] -0.39 -0.10 -0.01
## b[(Intercept) Subblock:Block:1:1] -0.44 -0.15 -0.05
## b[(Intercept) Subblock:Block:1:2] -0.14 0.00 0.07
## b[(Intercept) Subblock:Block:1:3] -0.45 -0.17 -0.05
## b[(Intercept) Subblock:Block:_NEW_Subblock:Block] -0.45 -0.08 0.00
## b[(Intercept) Block:1] -0.44 -0.07 0.01
## b[(Intercept) Block:2] -0.27 0.00 0.08
## b[(Intercept) Block:3] -0.65 -0.21 -0.08
## b[(Intercept) Block:_NEW_Block] -0.64 -0.11 0.00
## sigma 0.35 0.40 0.43
## Sigma[Subblock:Block:(Intercept),(Intercept)] 0.00 0.00 0.02
## Sigma[Block:(Intercept),(Intercept)] 0.00 0.01 0.04
## mean_PPD 6.86 6.98 7.04
## log-posterior -61.92 -56.47 -53.89
##
## 75% 97.5% n_eff Rhat
## (Intercept) 7.06 7.41 1271 1
## NIncreased_N 0.10 0.32 3169 1
## TempNormal_Temp 0.59 0.83 2861 1
## NIncreased_N:TempNormal_Temp -0.30 0.00 2571 1
## b[(Intercept) Subblock:Block:0:1] 0.17 0.46 2765 1
## b[(Intercept) Subblock:Block:0:2] 0.06 0.32 2568 1
## b[(Intercept) Subblock:Block:0:3] 0.04 0.27 2532 1
## b[(Intercept) Subblock:Block:1:1] 0.01 0.18 2848 1
## b[(Intercept) Subblock:Block:1:2] 0.19 0.50 2000 1

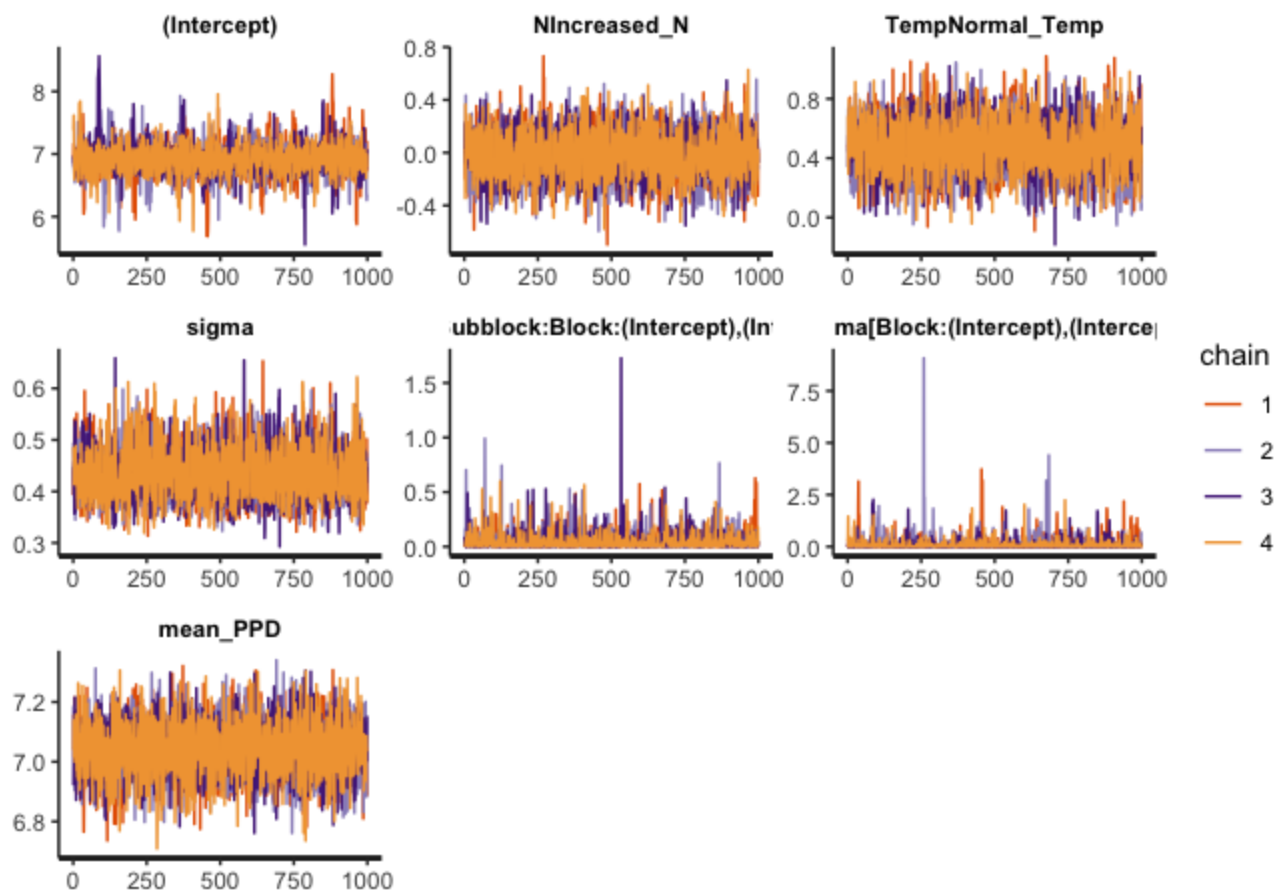
```



```
## b[(Intercept) Subblock:Block:1:3] 0.01 0.17 2356 1
## b[(Intercept) Subblock:Block:_NEW_Subblock:Block] 0.08 0.44 4124 1
## b[(Intercept) Block:1] 0.10 0.52 1246 1
## b[(Intercept) Block:2] 0.22 0.69 1254 1

## b[(Intercept) Block:3] 0.00 0.30 1123 1
## b[(Intercept) Block:_NEW_Block] 0.11 0.67 3939 1
## sigma 0.46 0.54 3538 1
## Sigma[Subblock:Block:(Intercept),(Intercept)] 0.05 0.25 2021 1
## Sigma[Block:(Intercept),(Intercept)] 0.10 0.75 1729 1
## mean_PPD 7.10 7.22 4095 1
## log-posterior -51.64 -48.22 1303 1
##
## Samples were drawn using NUTS(diag_e) at Sat Aug 17 13:54:22 2019.
## For each parameter, n_eff is a crude measure of effective sample size,
## and Rhat is the potential scale reduction factor on split chains (at
## convergence, Rhat=1).
```

```
traceplot(bayes_NxT$stanfit, pars = c("(Intercept)", "NIncreased_N", "TempNormal_Temp", "sigma"
```



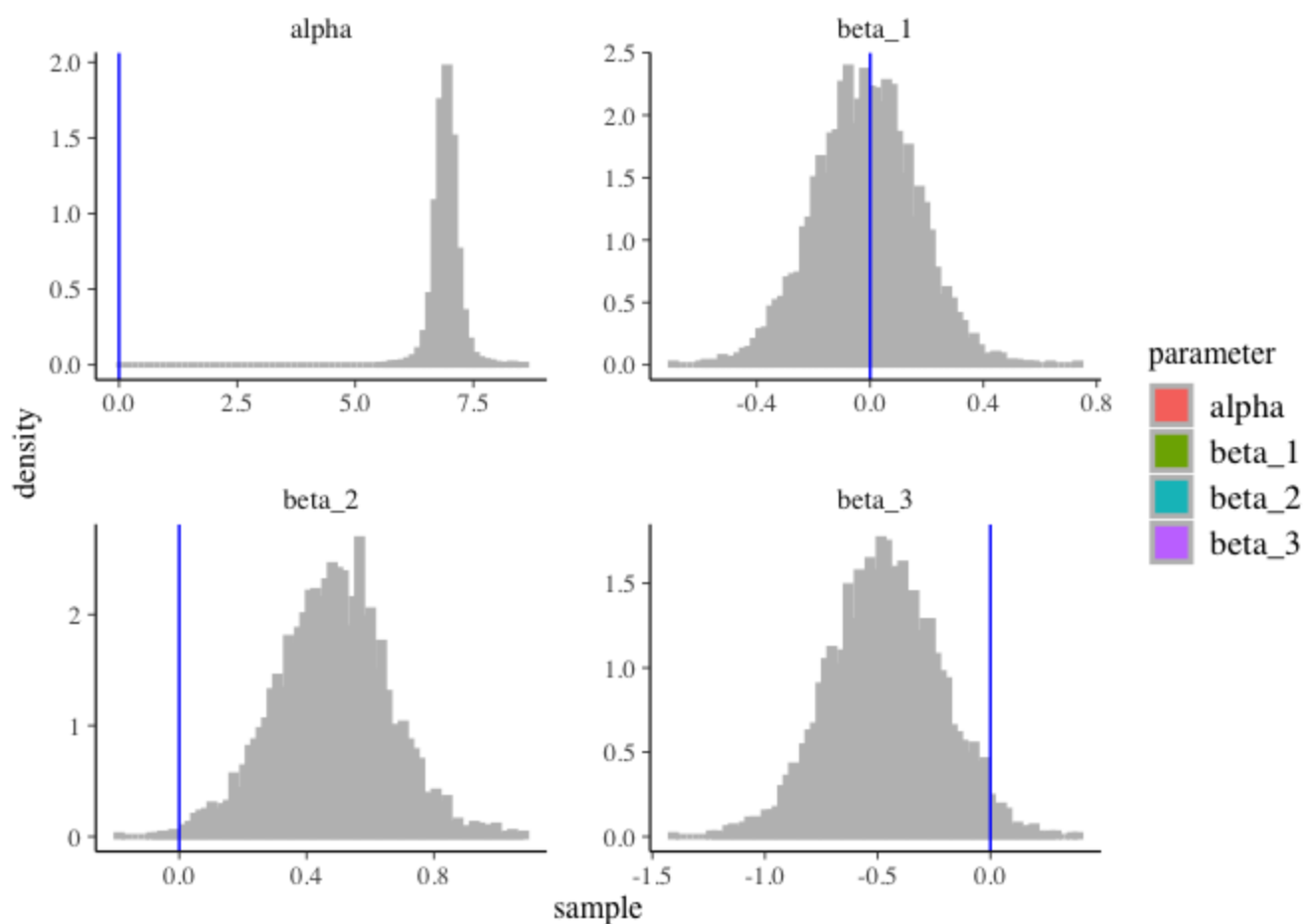
Figure

10. A traceplot of the intercept of Model 2 shows that, although the model converged, there was a lot of variability through the end of the iterations.

Sample from posterior (real data)

```
#' Extract posterior samples:
samples <- extract(bayes_NxT$stanfit)
samplesdf <- data.frame(samples$alpha, samples$beta)
names(samplesdf) <- c("alpha", paste(names(samples[2]),1:3,sep="_"))
```

```
samplesdf %>%
  gather(key = "parameter", value = "sample") %>%
  ggplot() +
  geom_histogram(mapping = aes(x=sample, y=stat(density),fill=parameter),
    bins=75, color="gray",lwd=1) +
  geom_vline(xintercept = 0, color = "blue") +
  facet_wrap(facets = ~ parameter,scales="free")
```



```
# Intercept
hpdi(samples$alpha[,1], prob=0.89)
```

```
##          lower    upper
## [1,] 6.561506 7.294984
```

```
# Nitrogen (Increased N)
hpdi(samples$beta[1,1], prob=0.89)
```

```
##          lower upper
## [1,]      NA     NA
```

```
# Temperature (Warmer Temp)
hpdi(samples$beta[,2], prob=0.89)
```

```
##          lower      upper
## [1,] 0.2126868 0.7656126
```

```
# Nitrogen:Temperature (Increased N, Warmer Temp)
hpdi(samples$beta[,3], prob=0.89)
```

```
##          lower      upper
## [1,] -0.8404694 -0.06739349
```

```
pred <- predictive_interval(bayes_NxT,prob=0.95)
colnames(pred) <- c("pred_l","pred_u")
nwt_descae_2007 <- cbind(nwt_descae_2007, pred)
```

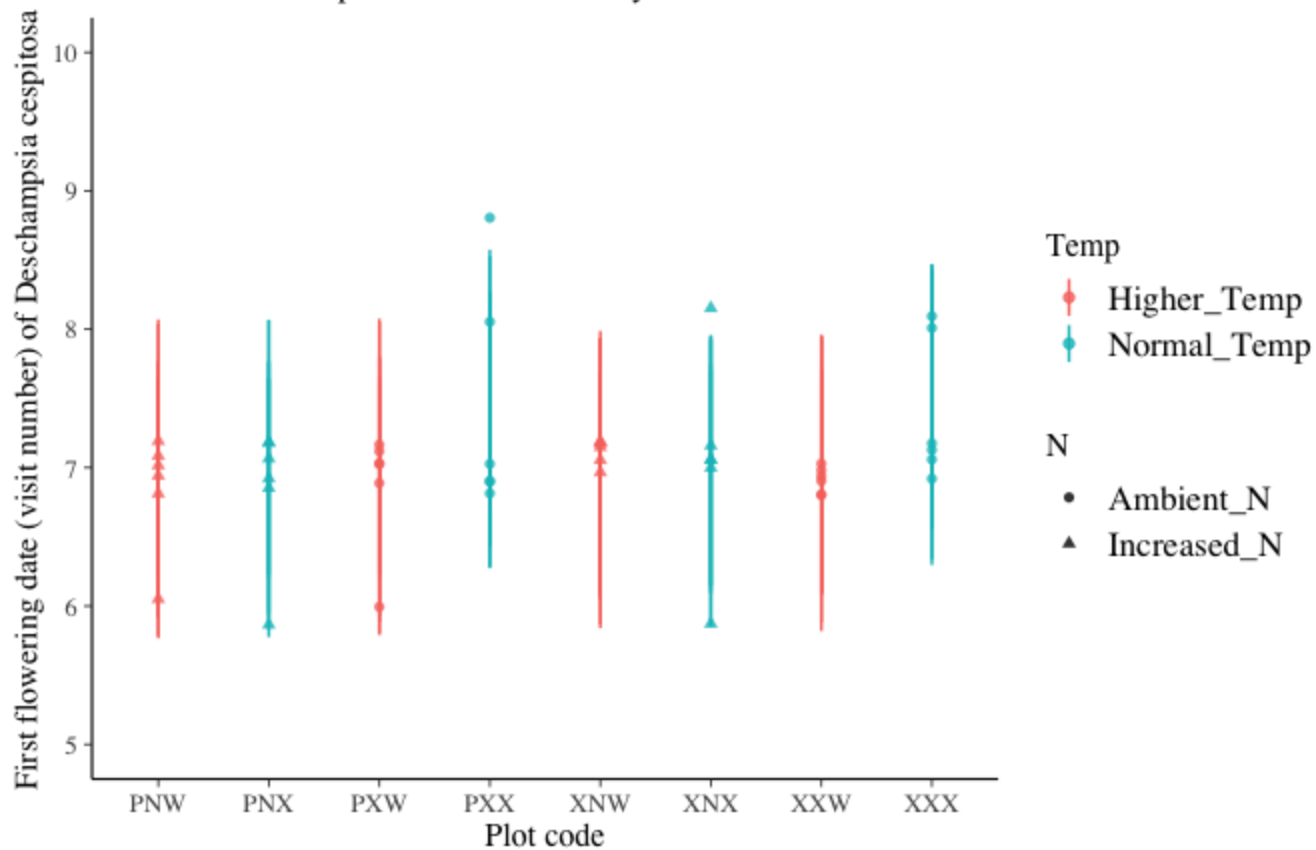
```
p11 <- ggplot(data = nwt_descae_2007, aes(x = Code, fill = Temp, col = Temp, pch = N)) +
  geom_point(aes(y = jitter(Visit)), alpha = 0.8) +
  geom_linerange(aes(y = Visit), ymin = nwt_descae_2007$pred_l, ymax = nwt_descae_2007$pred_u)
labs(title = "Figure 11",
      subtitle = "Observed data and predicted data from Bayesian model",
      x = "Plot code",
      y = "First flowering date (visit number) of Deschampsia cespitosa") +
ylim(5, 10)
```

```
## Warning: Ignoring unknown aesthetics: y
```

```
p11
```

Figure 11

Observed data and predicted data from Bayesian model

**Figure**

11. The observed data (plotted as points) is overlaid with the predicted data from the model (lines). Overall, plots with higher temperatures flowered earlier than plots with normal temperatures. Plots with increased nitrogen, flowered earlier than plots with ambient nitrogen. Plots with both increased temperature and increased nitrogen flowered later.

The preceding analysis showed that there may be some problematic assumptions with using a gaussian distribution for this data. Though the data is distributed relatively normally (see EDA), the number of total observations and the variance in the data are too small to effectively use a gaussian GLMER. Future work should explore using a latent variable model, which was recommended by Brett Melbourne.