

Genomic Data Analysis using R

Lecture 15

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Overview: Learning Objectives

1. Sequence data

- Databases and online resources for sequence data
- Learn the common sequence data file formats

2. Tools for sequencing data

- Tools to query, inspect, visualize an aligned sequence file
- Learn the contents of sequence data files
- Learn to generate sequencing metrics and to process sequence data
- Learn about Python and R libraries/packages to read sequence data

3. Genome variant analysis (Background; this Lecture)

- Types of genomic variation
- Tools to predict genomic variations
- Learn the common file formats for variation data
- Databases and online resources for human variation data

Genome Variant Analysis Background: Overview

1. Types of genomic variation

2. Visualization using IGV

3. File Formats for Variation Data

Genome Variant Analysis: Types of Genomic Variation

Variant or Mutation or Alteration or Polymorphism

- Changes in the genome sequence of a sample compared to a reference sequence
- Chromosomes: 22 autosomal pairs + 1 sex pair
 - Each set inherited from maternal and paternal germline cells

Germline Variant

- Variant inherited from one or both parental chromosomes
- Source of genetic differences between ancestral populations and individuals
- Polymorphism: >1% frequency in a population

Somatic Variant

- Mutation acquired during individual's lifetime
- Important to identify in sporadic cancers and other non-familial diseases

Genome Variant Analysis: Types of Genomic Variation

a. Single nucleotide base substitutions

- Germline single nucleotide polymorphism (SNP)
- Somatic single nucleotide variant (SNV)

b. Small insertions or deletions

- Germline or somatic insertion or deletion (INDEL)

c. Copy number changes

- Germline copy number variant (CNV) or polymorphism (CNP)
- Somatic copy number variant (CNV) or alterations (CNA)

d. Structural rearrangements

- Germline or Somatic structural variant (SV)

Genome Variant Analysis: Single Nucleotide Polymorphism

- ~1.5 to 2 million **SNPs** per individual
- Identify SNPs from normal peripheral blood mononuclear cells (PBMC)

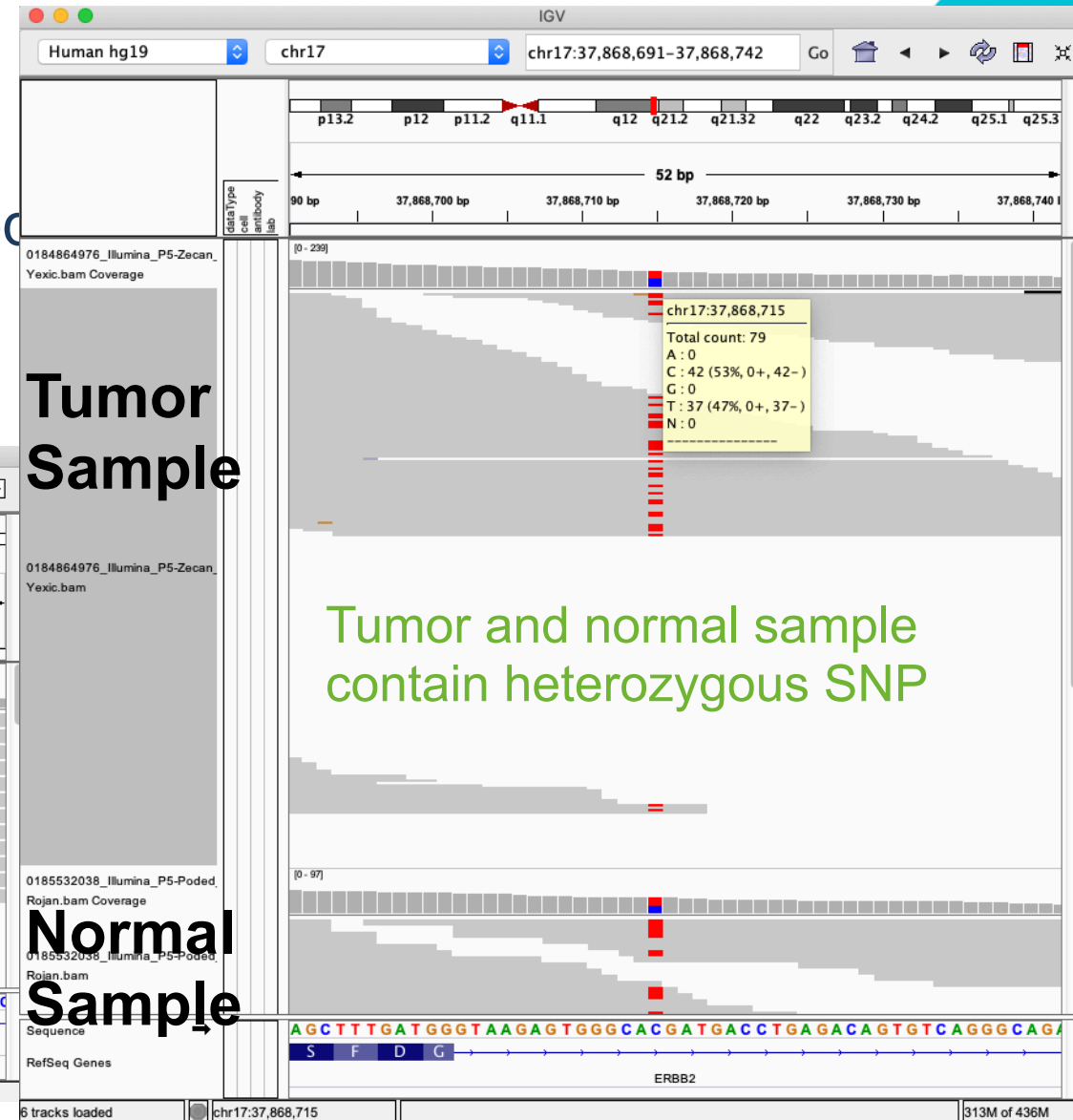


Heterozygous SNP with 37 reads containing the variant and having depth 79 reads

37/79 (47%) variant allele fraction (VAF)

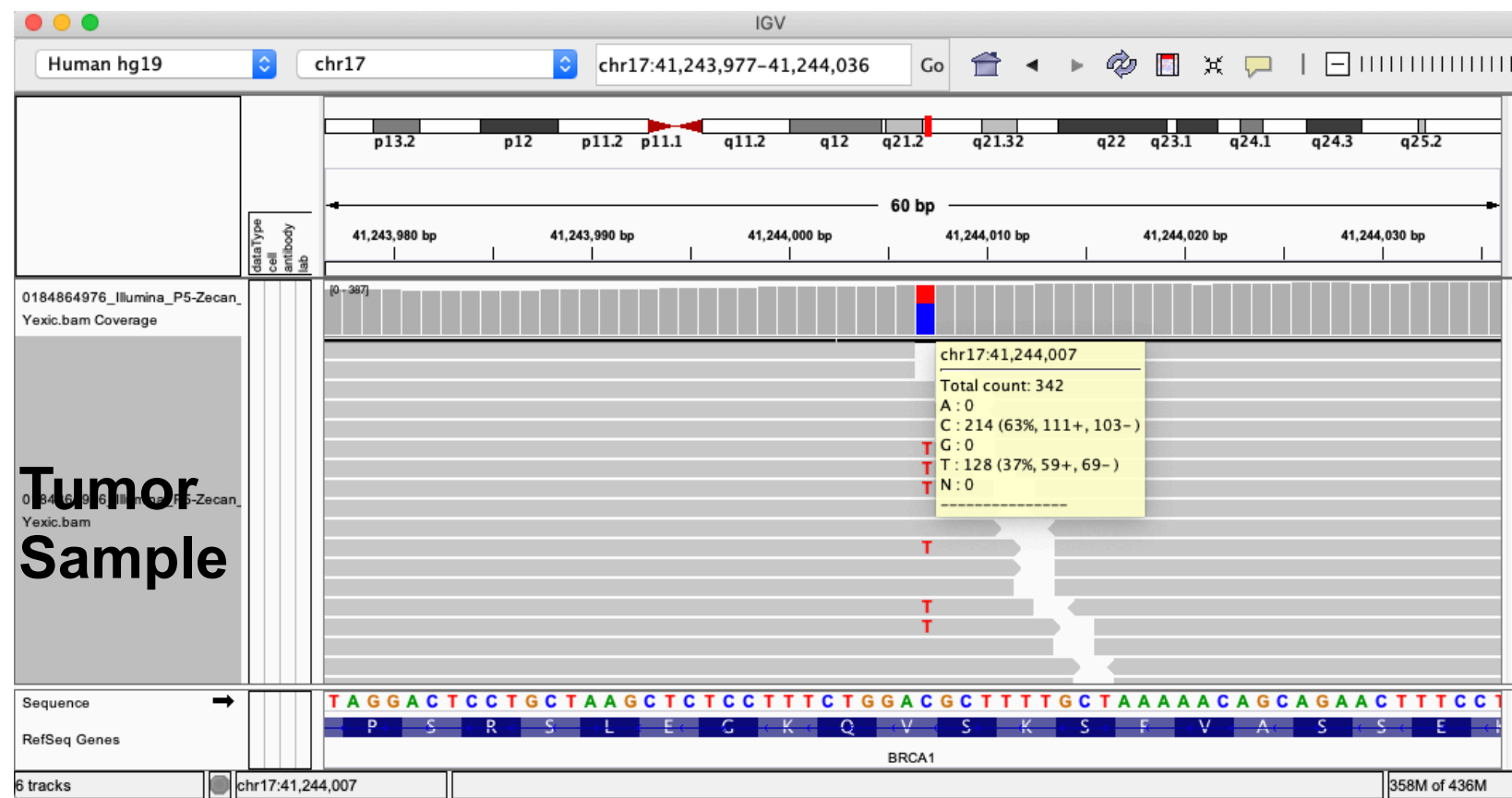
Genome Variant Analysis: Single Nucleotide Polymorphism

- ~1.5 to 2 million **SNPs** per individual
- Identify SNPs from normal peripheral blood



Genome Variant Analysis: Single Nucleotide Variant (SNV)

- Somatic **SNV** requires comparing case (tumor) with control (PBMC)

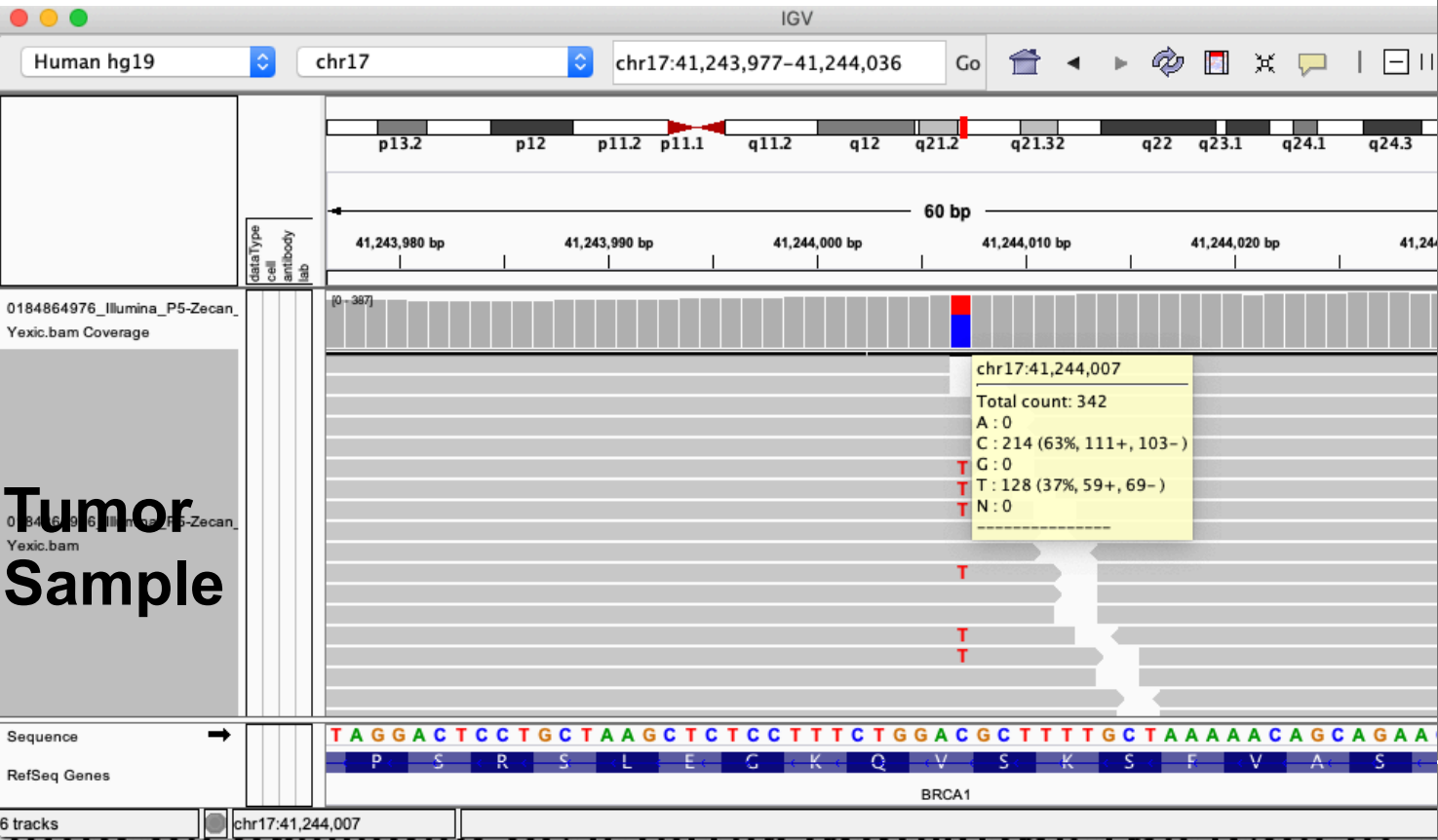


Potential SNV with
128/342 (37%) VAF

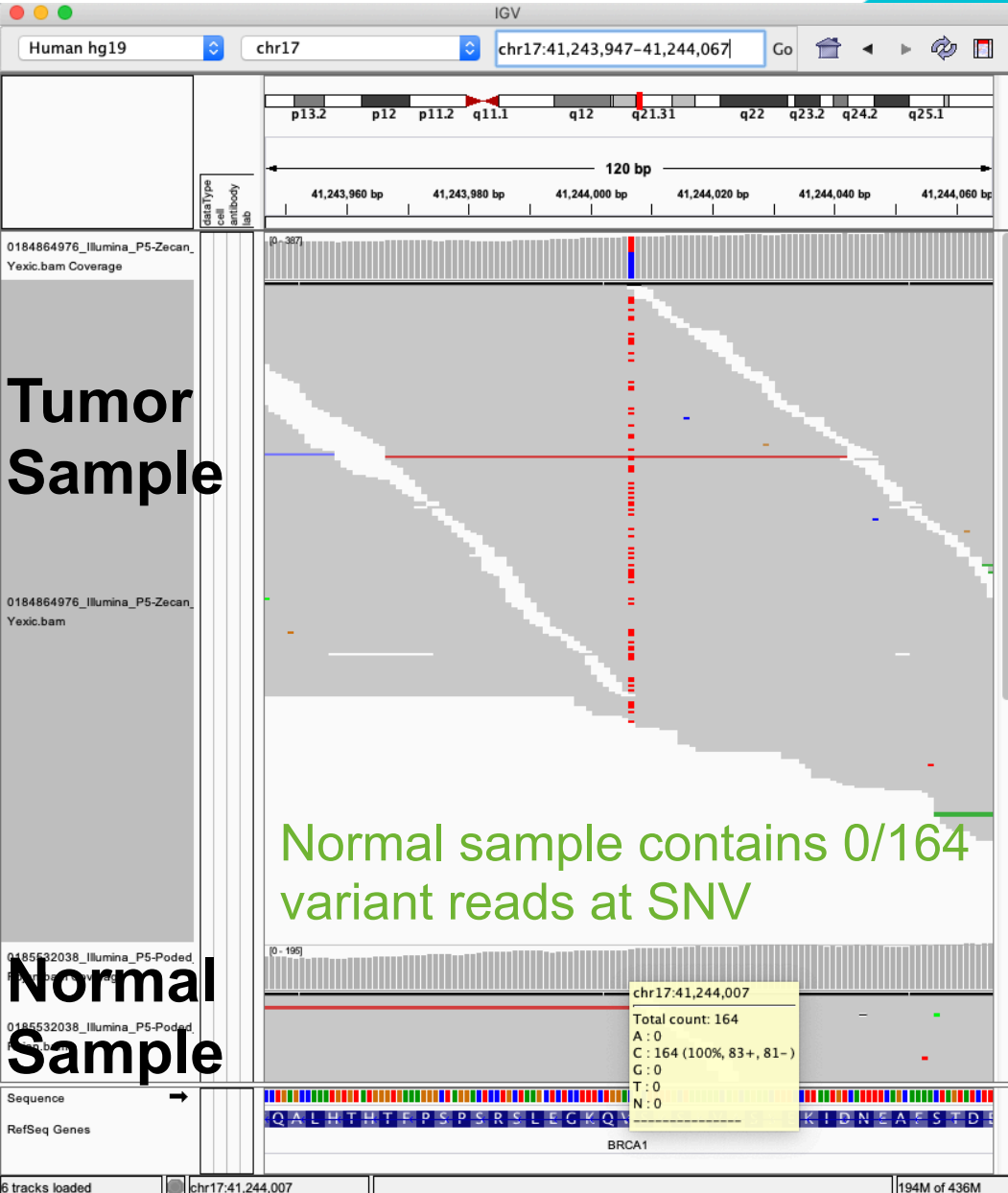
p.V1181I

Genome Variant Analysis: Single Nucleotide Variant (SNV)

– Somatic **SNV** requires comparing case (tumor)



Tumor Sample

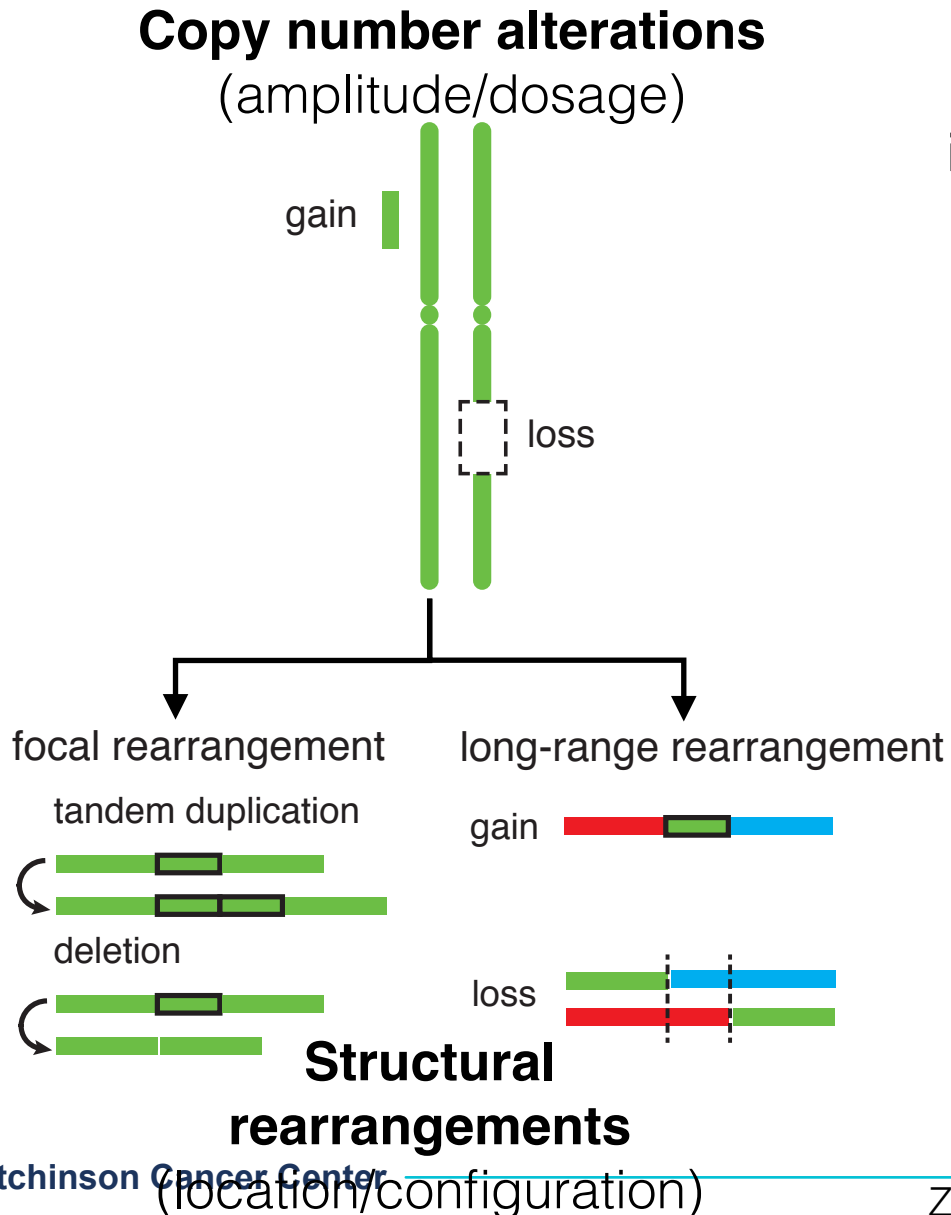


Tumor Sample

Normal Sample

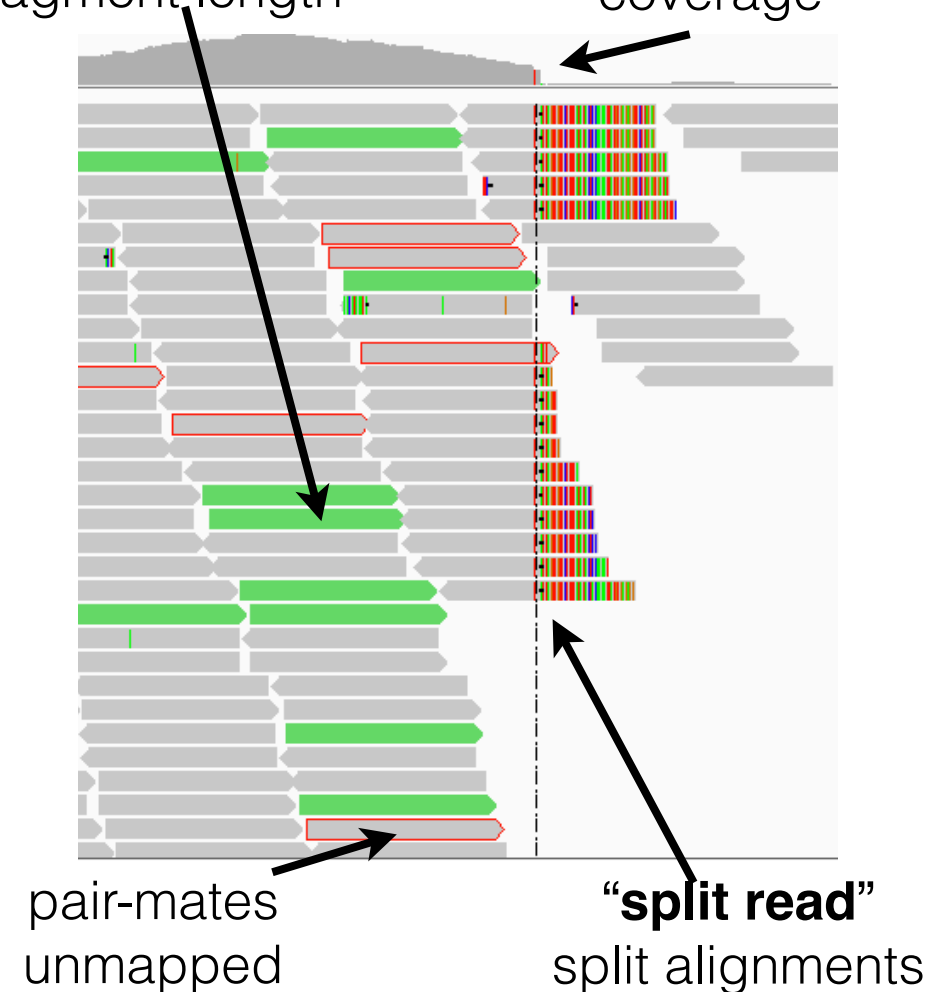
Normal sample contains 0/164 variant reads at SNV

Genome Variant Analysis: Copy Number and Structural Variation

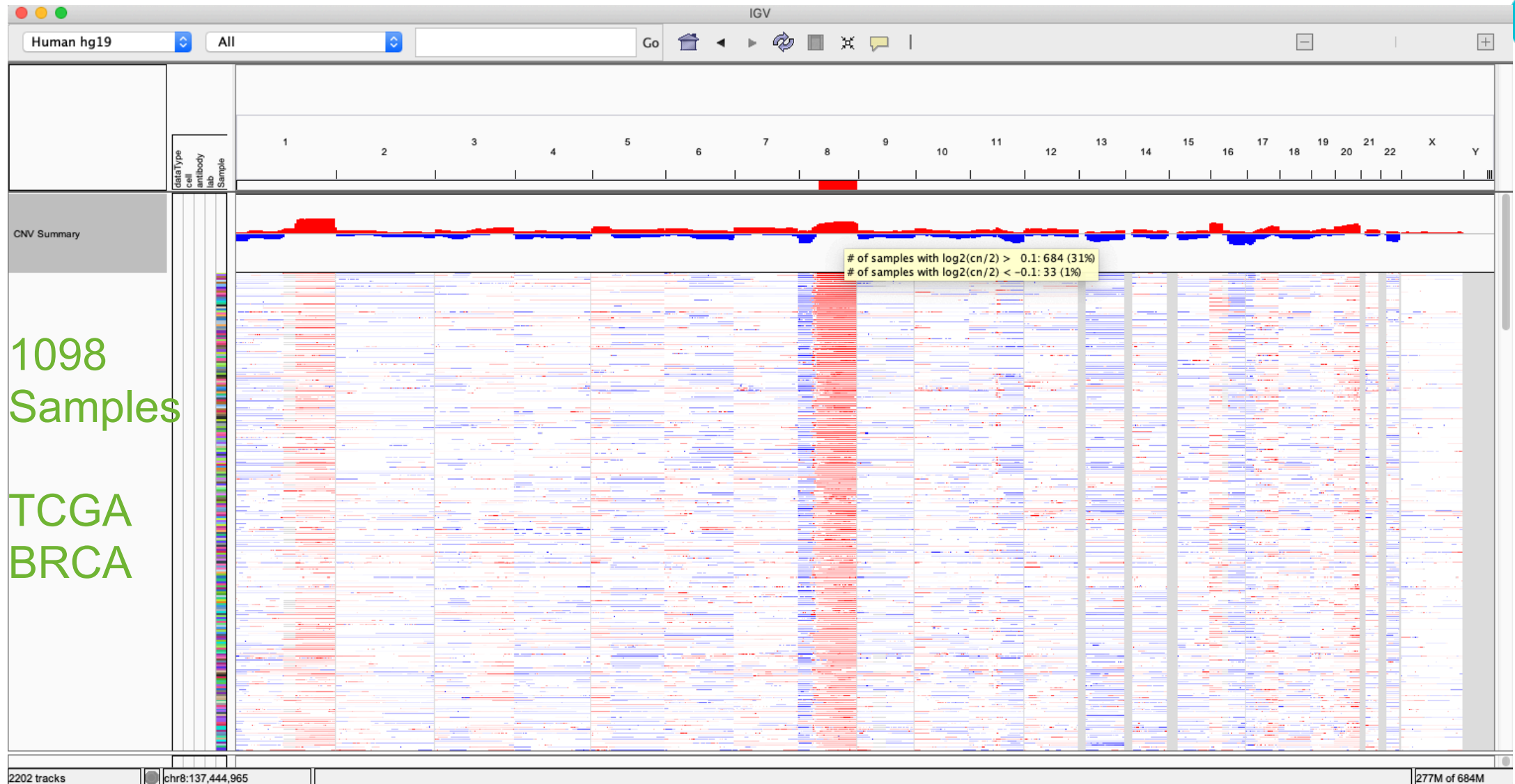


“discordant read pair”
read pairs with aberrant
inferred fragment length

“copy number change”
abrupt change in read
coverage



Genome Variant Analysis: Copy Number Variation



Genome Variant Analysis: Variant Annotation Tools

ANNOVAR (<http://annovar.openbioinformatics.org>)

SnEff (<http://snpeff.sourceforge.net>)

SIFT (<https://sift.bii.a-star.edu.sg/>) - predict amino acid substitution effects on protein function

GATK VariantAnnotator

VariantAnnotation R Package (<https://bioconductor.org/packages/release/bioc/html/VariantAnnotation.html>)

Variant Annotation Integrator (UCSC, <https://genome.ucsc.edu/cgi-bin/hgVai>)

BioMart (<http://www.biomart.org/>)

For your reference.

Genome Variant Analysis: Variant Databases

1000 Genomes Project (<https://www.internationalgenome.org/>)

dbSNP (<https://www.ncbi.nlm.nih.gov/snp/>)

dbVar (<https://www.ncbi.nlm.nih.gov/dbvar/>)

ClinVar (<https://www.ncbi.nlm.nih.gov/clinvar/>)

Exome Aggregation Consortium (ExAC, <http://exac.broadinstitute.org/>)

– Lek et al. Nature, 536, 285-91 (2016)

Genome Aggregation Database (gnomAD, <https://gnomad.broadinstitute.org/>)

– Karczewski et al. bioRxiv (2019)

Genome Data Commons (<https://portal.gdc.cancer.gov/>)

For your reference.

R/Bioconductor Packages for Genomic Data

Tutorials

1. Analyzing Genomic Data
2. Analyzing and Annotating Variants

Overview: Learning Objectives

R Bioconductor Packages for Genomic Data

- `GenomicRanges`, `plyranges`, `VariantAnnotation`

Tutorials

1. Genomic Data Analysis (`GenomicRanges`, `plyranges`)
 - i. Load, inspect, query a BED/SEG file
 - ii. Genomic regions overlap
2. Genomic Variants and Annotations (`VariantAnnotation`)
 - i. Load, inspect, query a VCF file

Tutorial #1: Genomic Data Analysis

1. Loading and querying BED/SEG text files
 - a. Use packages `GenomicRanges`, `plyranges`
2. Download the VCF and SEG files for this tutorial
 - <https://www.dropbox.com/sh/zoitjnobgp7I7c2/AABBIpTQcNA4IWYOFnV5dIMKa?dl=0>
 - `BRCA.genome_wide_snp_6_broad_Level_3_scna.seg`
3. Files for tutorial on GitHub:
 - R Markdown: `Lecture15_GenomicData.Rmd`
 - Jupyter Notebook: `Lecture15_GenomicData.ipynb`

Genome Variant Analysis: Common Variant File Formats

a. Variant Call Format (VCF)

- <http://samtools.github.io/hts-specs/VCFv4.2.pdf>
- Used mostly for SNV/SNP, INDEL, and SV

b. Mutation Annotation Format (MAF)

- https://docs.gdc.cancer.gov/Data/File_Formats/MAF_Format/
- <http://software.broadinstitute.org/software/igv/MutationData>
- Tab-delimited format containing columns for mutation information and annotations
- Used primarily for SNV/SNP and INDEL data

c. Browser Embedded Data (BED)

a. <https://bedtools.readthedocs.io/>

b. Used for any genomic features/region and annotations, including CNV and SV (BEDPE)

d. Others

a. <http://genome.ucsc.edu/FAQ/FAQformat>

b. GFF, WIG/bigWIG, etc.

Genome Variant Analysis: Variant Call Format (VCF)

<http://samtools.github.io/hts-specs/VCFv4.2.pdf>

a. Header information

```
##fileformat=VCFv4.2
##GATKCommandLine=<ID=HaplotypeCaller,CommandLine="HaplotypeCaller">
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele">
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##INFO=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth; some reads may have been filtered">
##FORMAT=<ID=AD,Number=R,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=PL,Number=G,Type=Integer,Description="Normalized, Phred-scaled likelihoods for genotypes as defined in the VCF specification">
##FORMAT=<ID=PS,Number=1,Type=Integer,Description="ID of Phase Set for Variant">
##FILTER=<ID=PASS,Description="All filters passed">
##FILTER=<ID=LowQual,Description="Low quality">
```

b. Variant record

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	Sample_1
chr1	11542	.	A	T	49.77	PASS	AC=1;AF=0.5;AN=2;DP=4	GT:AD:DP:GQ:PL:PS	0 1:2,2:4:78:78,0,78

Tutorial #2: Variant Call Format (VCF)

1. Loading and querying VCF files in R

a. Use packages `VariantAnnotation`

b. Download the VCF files for this tutorial

- <https://www.dropbox.com/sh/zoitjnobgp7I7c2/AABB1pTQcNA4IWYOFnV5dIMKa?dl=0>
- `GIAB_highconf_v.3.3.2.vcf.gz`
- `GIAB_highconf_v.3.3.2.vcf.gz.tbi`

2. Files for tutorial on GitHub:

- R Markdown: `Lecture15_VariantCalls.Rmd`
- Jupyter Notebook: `Lecture15_VariantCalls.ipynb`

Homework #7: Genomic Data Analysis in R

Problem Sets in R Markdown or Jupyter Notebook

- Contains 3 Problems with some code to prepare you for the questions.
- Please complete the assignment within the Markdown or Jupyter file
- You will be evaluated on
 - i. the results and outputs
 - ii. your code and documentation
 - iii. Partial points awarded for code with correct logic/function even if the final answer may be incorrect