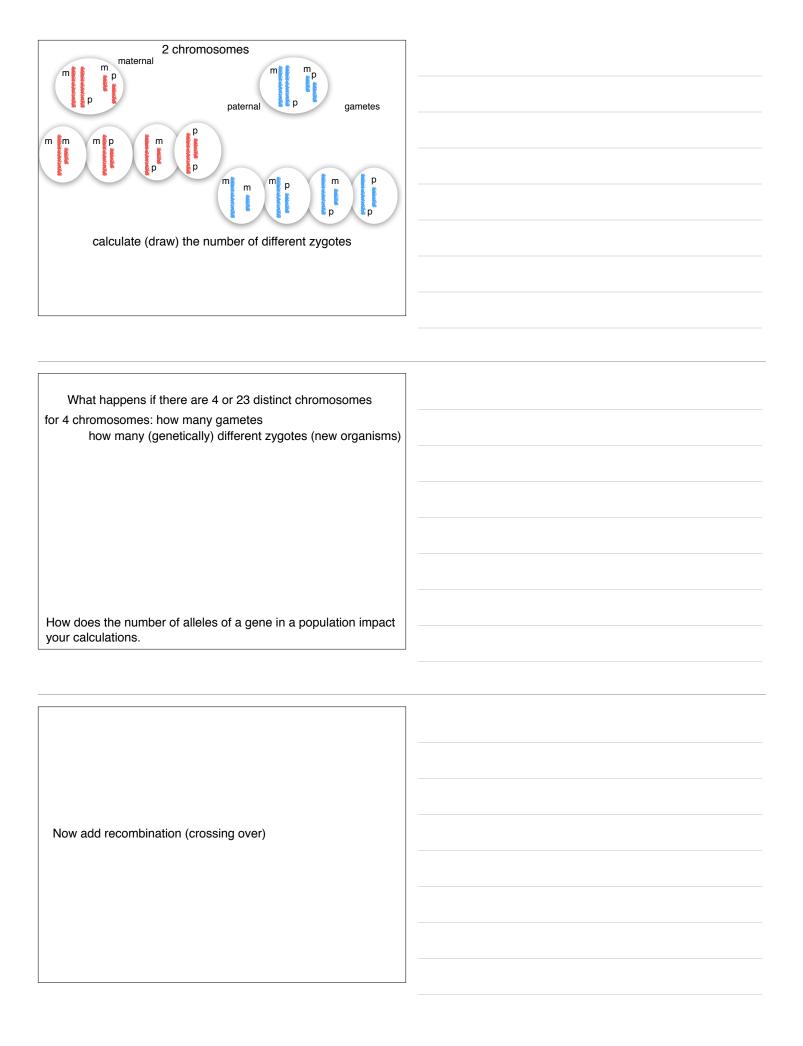
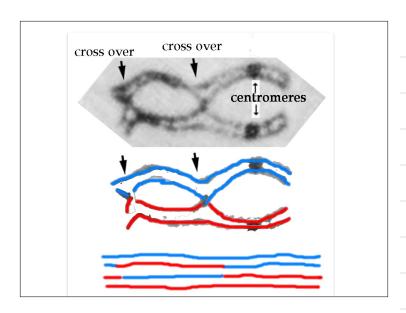
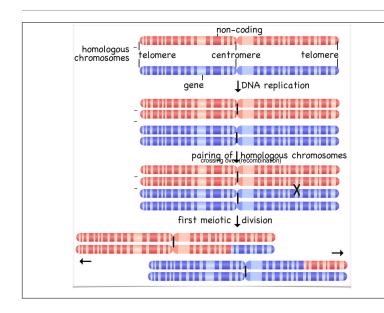
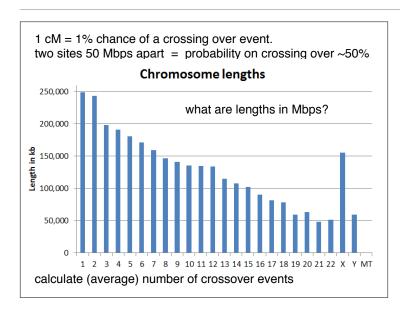


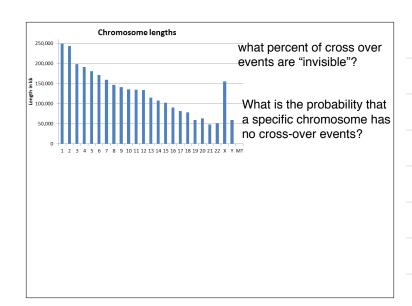
mechanism of synaptonemal complex formation (veronica's question from last time) Modeling meiotic chromosome pairing: nuclear envelope attachment, telomere-led active random motion, and anomalous diffusion Wallace F Marshall and Jennifer C Fung² Department of Biochemistry and Biophysics, University of California San Francisco, USA Department of Obstetrics and Gynecology, Center for Reproductive Science, University of California San Francisco, USA Output Department of Obstetrics and Gynecology, Center for Reproductive Science, University of California San Francisco, USA Keywords: diffusion, homologous pairing, computer simulation, meiosis, telomere, homologous chromosome, nuclear envelope Abstract The recognition and pairing of homologous chromosomes during meiosis is a complex physical and $molecular\ process\ involving\ a\ combination\ of\ polymer\ dynamics\ and\ molecular\ recognition\ events.$ $Two\ highly\ conserved\ features\ of\ meiotic\ chromosome\ behavior\ are\ the\ attachment\ of\ telomeres\ to$ the nuclear envelope and the active random motion of telomeres driven by their interaction with cytoskeletal motor proteins. Both of these features have been proposed to facilitate the process of homolog pairing, but exactly what role these features play in meiosis remains poorly understood. Here we investigate the roles of active motion and nuclear envelope tethering using a Brownian dynamics $simulation\ in\ which\ meiotic\ chromosomes\ are\ represented\ by\ a\ Rouse\ polymer\ model\ subjected\ to$ tethering and active forces at the telomeres. We find that tethering telomeres to the nuclear envelope slows down pairing relative to the rates achieved by unattached chromosomes, but that randomly directed active forces applied to the telomeres speed up pairing dramatically in a manner that depe mechanism of synaptonemal complex formation (veronica's question) nuclear envelope

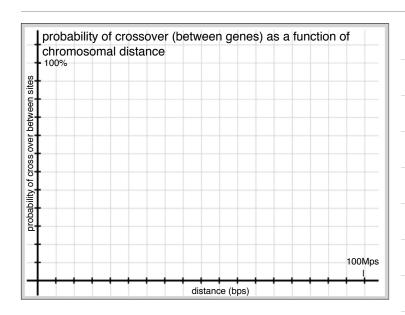


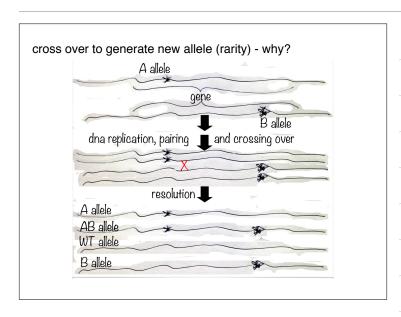




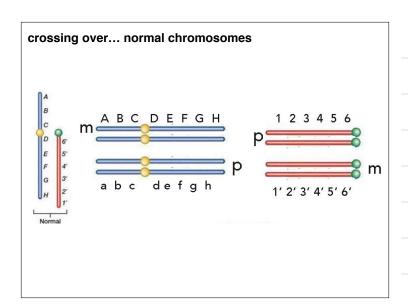


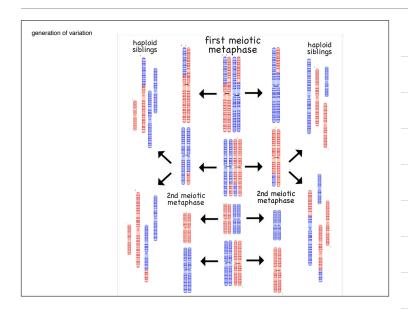


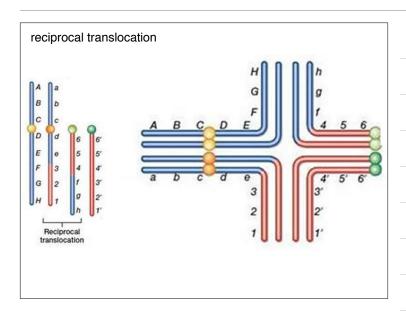


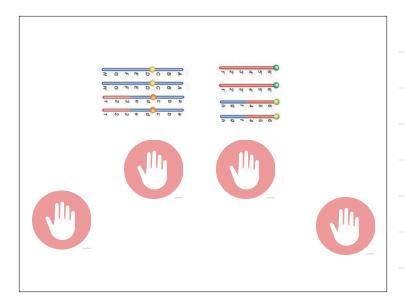


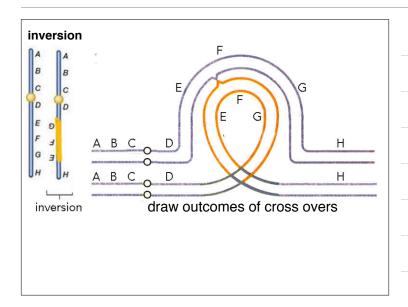
Aidan: why is there crossing over any way?	
 Questions to Consider What is the purpose of chromosome rearmament if genetic information is not gained or lost? When cutting: What are the odds of cutting between section 1 and 6 versus cutting between section 1 and 2 After Trading: What % of same genes do you have after you traded? 	
• 83% chance that you shave a different genome than what you started out with from just crossing over	

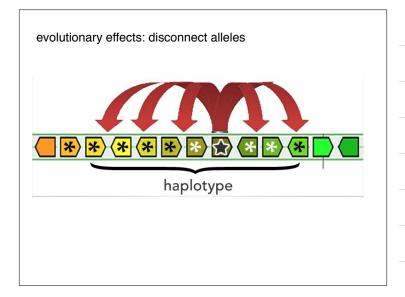












recombination event maternal chromosome paternal chromosome chromo

Now consider mutation rate

Article | OPEN

Differences between germline and somatic mutation rates in humans and mice

Brandon Milholland, Xiao Dong, Lei Zhang, Xiaoxiao Hao, Yousin Suh & Jan Vijg 🖼

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in human: germline mutation rate (males) $\sim 3 \times 10^{-11}$ per bp Somatic mutation rate $\sim 3 \times 10^{-9}$ per bp haploid human genome $\sim 3 \times 10^{9}$ bps

both mutation rates are higher in mouse

Genetics

Fathers pass on four times as many new genetic mutations as mothers – study

The figures mean that a child born to 30-year-old parents would, on average, inherit 11 new mutations from the mother, but 45 from the father.

why, exactly?

Questions (you should be able) toanswer: 212. Graph, as a function of distance, the likelihood that recombination will disconnect a selected (whether positively of negatively) allele from alleles in surrounding genes. 213. Why might a crossing over event inhibit nearby crossing over events? 214. How can you use the size of a conserved genomic region to estimate time of isolation of a population? 215. What are the benefits of recombination in terms of	
environmental adaptation? Questions to ponder: -How does the size of haplotype regions reflect the reproductive history of a population? -How does the presence of a deleterious allele influence the selective pressures on an organism? How might it open up (over generational) time, new evolutionary possibilities?	