

Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Change “Student Name” on line 3 (above) with your name.
2. Work through the steps, **creating code and output** that fulfill each instruction.
3. Be sure to **answer the questions** in this assignment document.
4. When you have completed the assignment, **Knit** the text and code into a single PDF file.
5. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai. Add your last name into the file name (e.g., “Salk_A03_DataExploration.Rmd”) prior to submission.

The completed exercise is due on <>.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively.

```
# Checking my working directory
getwd()
```

```
## [1] "/Users/kaitlynkrejsa/Desktop/Duke/Classes/ENVIRON 872 - Environmental Data Analytics/Environment"
```

```
# loading packages
library(tidyverse)
```

```
# 3. Importing datasets
Neonics <- read.csv("./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv")
Litter <- read.csv("./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv")
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency’s ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: We might be interested in the ecotoxicology of neonicotinoids on insects because different insecticides are more or less effective depending on a number of variables such as the species of insect, the life stage of the insect, the age of the insect, application time of year, number of chemical applications, and concentrations of chemical applications, among others.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: We might be interested in studying litter and woody debris that falls on the ground in forests because it can provide information on decomposition rate and soil development. It can also provide information on how much carbon is being store in those sinks.

4. How is litter and woody debris sampled as part of the NEON network? Read the [NEON_Litterfall_UserGuide.pdf](#) document to learn more. List three pieces of salient information about the sampling methods here:

Answer: * “Mass data for each collection event are measured separately for the following functional groups, to an accuracy of 0.01 grams.” * “One litter trap pair (one elevated trap and one ground trap) is deployed for every 400 m2 plot area, resulting in 1-4 trap pairs per plot.” * “Ground traps are sampled once per year. Target sampling frequency for elevated traps varies by vegetation present at the site, with frequent sampling (1x every 2weeks) in deciduous forest sites during senescence, and in- frequent year-round sampling (1x every 1-2 months) at evergreen sites.”

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset? 4623 rows and 30 columns

```
dim(Neonics)
```

```
## [1] 4623 30
```

6. Using the `summary` function on the “Effects” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

Effect is a character, so cannot run a summary on it. Need to convert to a factor first

```
Neonics$Effect <- as.factor(Neonics$Effect)
summary(Neonics$Effect)
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##             12             102             360             11
##      Cell(s)      Development      Enzyme(s)      Feeding behavior
##             9             136             62             255
##      Genetics      Growth      Histology      Hormone(s)
##            82             38             5             1
##      Immunological      Intoxication      Morphology      Mortality
##            16             12             22             1493
##      Physiology      Population      Reproduction
##             7             1803             197
```

Answer: The most common effects studied are population, mortality, and behavior. These effects are specifically of interest to determine how effective the insecticide is on reducing the population size of the target insect (population), on killing the target insect (mortality), and on behavioral changes of the target insect, such as feeding on the target crop (behavior). These effects are also of interest regarding non-target insects, such as pollinator species, because it may be important to make sure the insecticide is not reducing their population, not killing them, and not changing their pollinating behaviors.

- Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

species.common.name is a character, so cannot run a summary on it. Need to convert to a factor first

```
Neonics$Species.Common.Name <- as.factor(Neonics$Species.Common.Name)
summary(Neonics$Species.Common.Name)
```

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113
##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27

##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee
##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17
##	Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
##	16	16
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
##	13	13
##	Yellow Fever Mosquito	Braconid Parasitoid
##	13	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12

##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Spotless Ladybird Beetle	Glasshouse Potato Wasp
##	11	10
##	Lacewing	Southern House Mosquito
##	10	10
##	Two Spotted Lady Beetle	Ant Family
##	10	9
##	Apple Maggot	(Other)
##	9	670

Answer: The six most commonly studied species in the dataset are: Other, Honey Bee, Parasitic Wasp, Buff Tailed Bumblebee, Carniolan Honey Bee, and Bumble Bee. All of these species are pollinator species. Pollinator species are critical for maintaining high crop yields. It is important to study the effects of insecticides on pollinators to make sure the insecticides are not having a deleterious effect on these species.

8. Concentrations are always a numeric value. What is the class of Conc.1..Author. in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

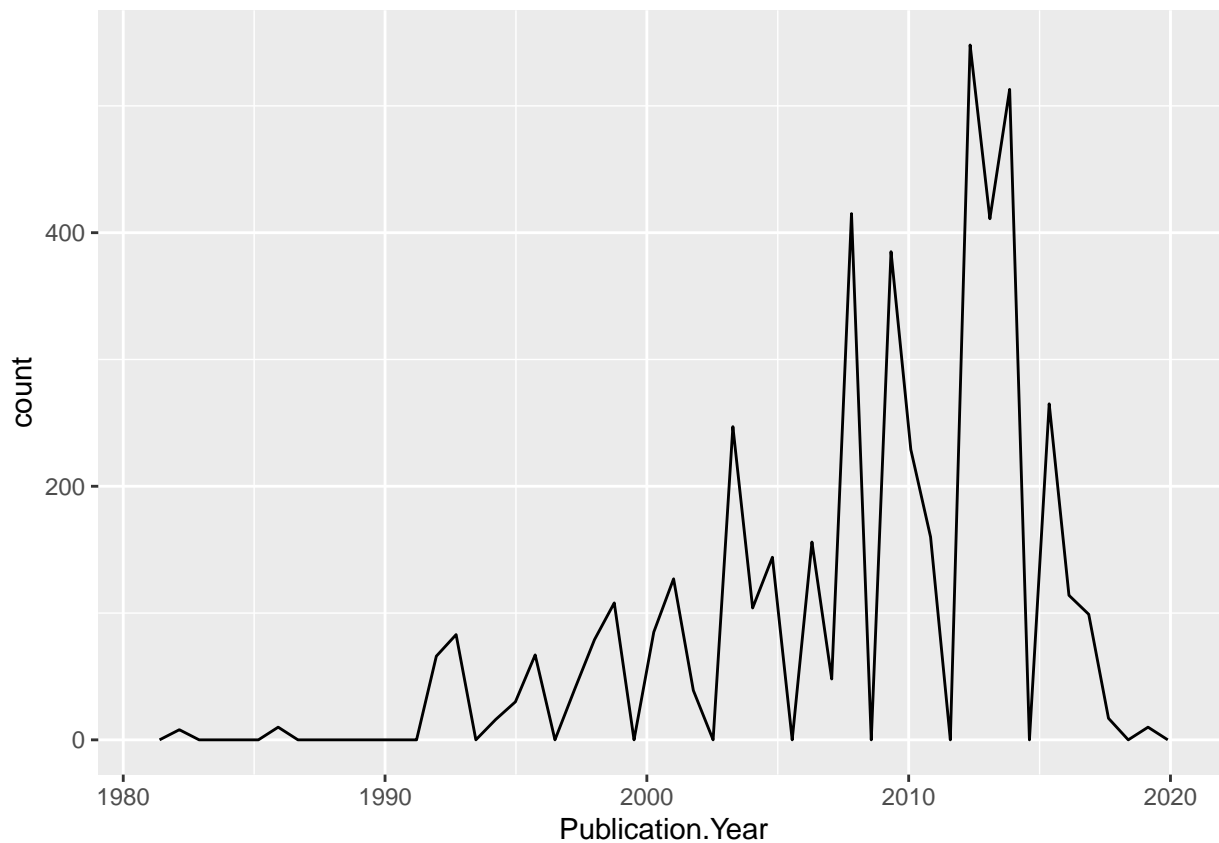
```
## [1] "character"
```

Answer: It is a character. It is not numeric because it has some non-numeric values like “NR” and “~10”, so the class of the entire column defaults to character.

Explore your data graphically (Neonics)

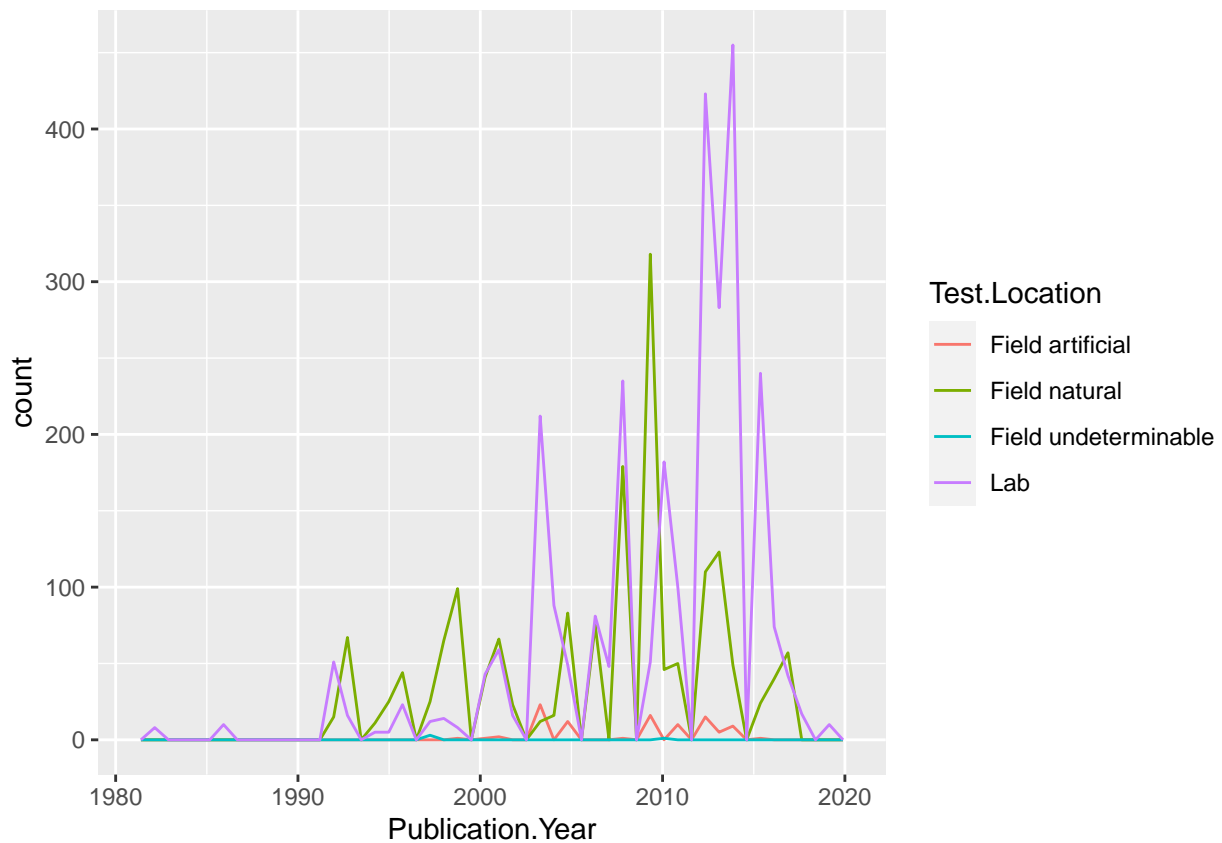
9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year), bins = 50)
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 50)
```

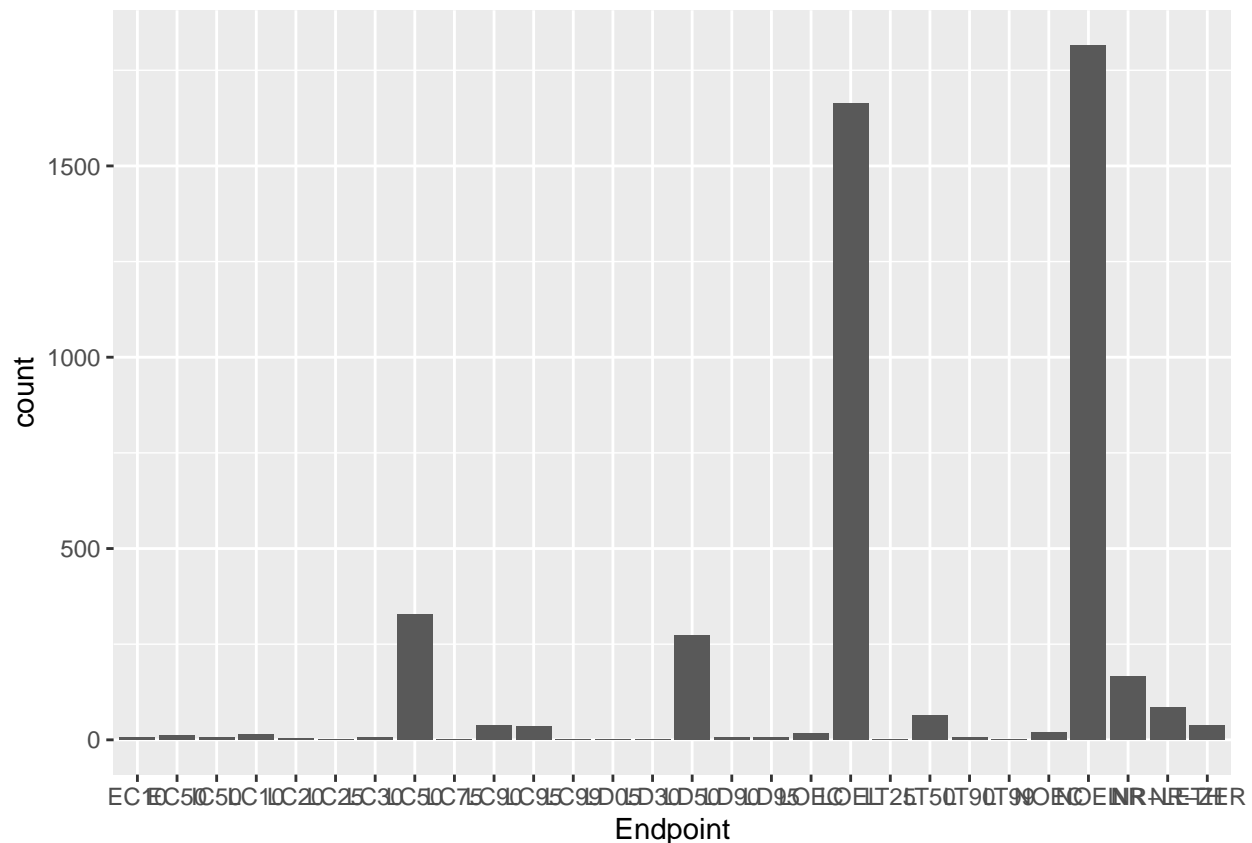


Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are lab and field natural. Lab and field natural locations differ quite a bit overtime, but overall these two test locations are more common than the other two locations over time.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

```
ggplot(Neonics, aes(x = Endpoint)) +  
  geom_bar()
```



```
Neonics$Endpoint <- as.factor(Neonics$Endpoint)
summary(Neonics$Endpoint)
```

```
##      EC10      EC50      IC50      LC10      LC20      LC25      LC30      LC50      LC75      LC90
##         6        11         6        15         5         1         6       327         1        37
##      LC95      LC99      LD05      LD30      LD50      LD90      LD95      LOEC      LOEL      LT25
##       36         2         1         1       274         6         7        17     1664         1
##      LT50      LT90      LT99      NOEC      NOEL      NR NR-LETH NR-ZERO
##       65         7         2        19     1816     167        86        37
```

Answer: The two most common endpoints are NOEL and LOEL. NOEL is defined as “No-observable-effect-level: highest dose (concentration) producing effects not significantly different from responses of controls according to author’s reported statistical test.” LOEL is defined as “Lowest-observable-effect-level: lowest dose (concentration) producing effects that were significantly different (as reported by authors) from responses of controls.”

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)
```

```
## [1] "character"
```



```
# collectDate is a character, so I will change it to a date.
```

```
Litter$collectDate <- as.Date(Litter$collectDate, format = "%Y-%m-%d")  
class(Litter$collectDate)
```

```
## [1] "Date"
```

```
unique(Litter$collectDate)
```

```
## [1] "2018-08-02" "2018-08-30"
```

```
# Litter was sampled in on August 2 and August 30, 2018
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
unique(Litter$plotID)
```

```
## [1] "NIWO_061" "NIWO_064" "NIWO_067" "NIWO_040" "NIWO_041" "NIWO_063"  
## [7] "NIWO_047" "NIWO_051" "NIWO_058" "NIWO_046" "NIWO_062" "NIWO_057"
```

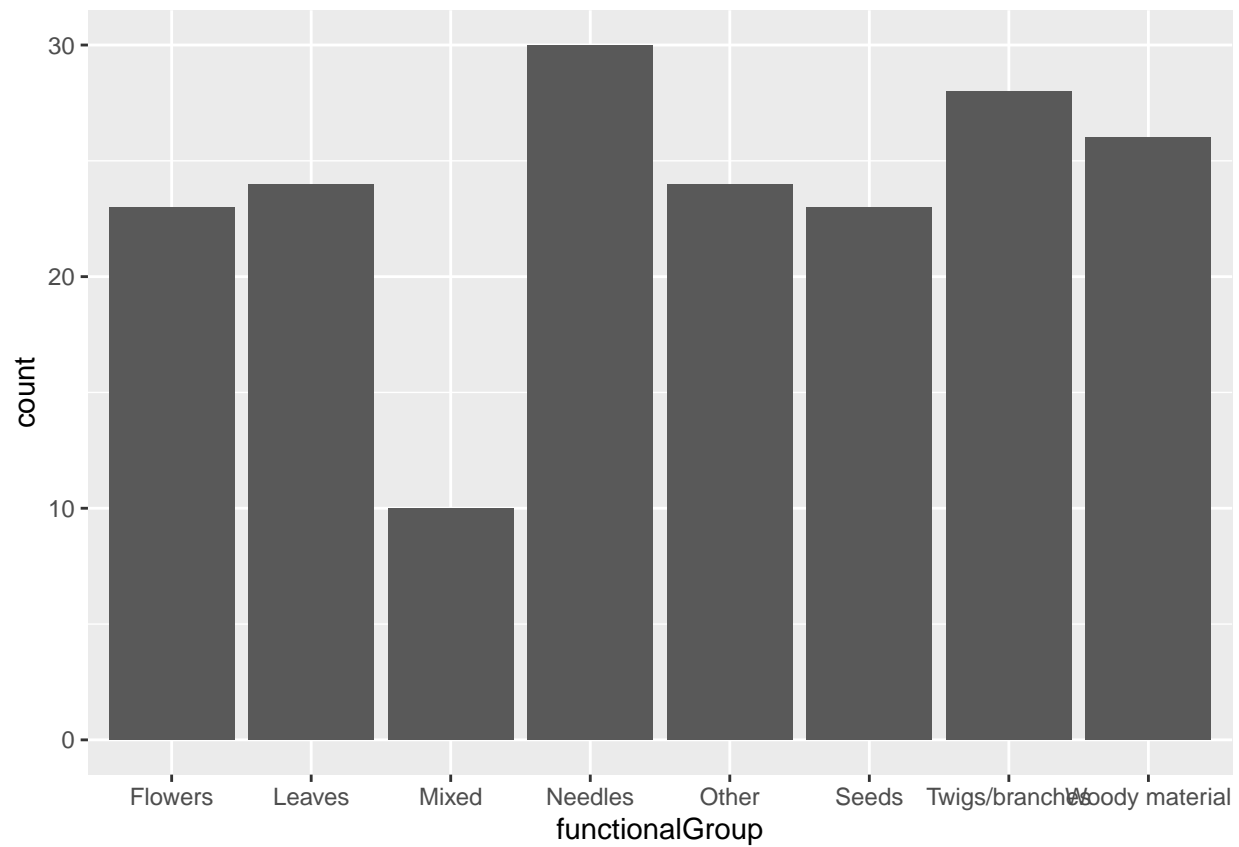
```
summary(Litter$plotID)
```

```
##      Length      Class      Mode  
##      188 character character
```

Answer: 12 plots were sampled at Niwot Ridge. The ‘unique’ function lists all of the unique plot ID values (i.e., the name of each unique plot ID), whereas the ‘summary’ function tells you how many plot ID observations there are (i.e., the length), the class, and the mode.

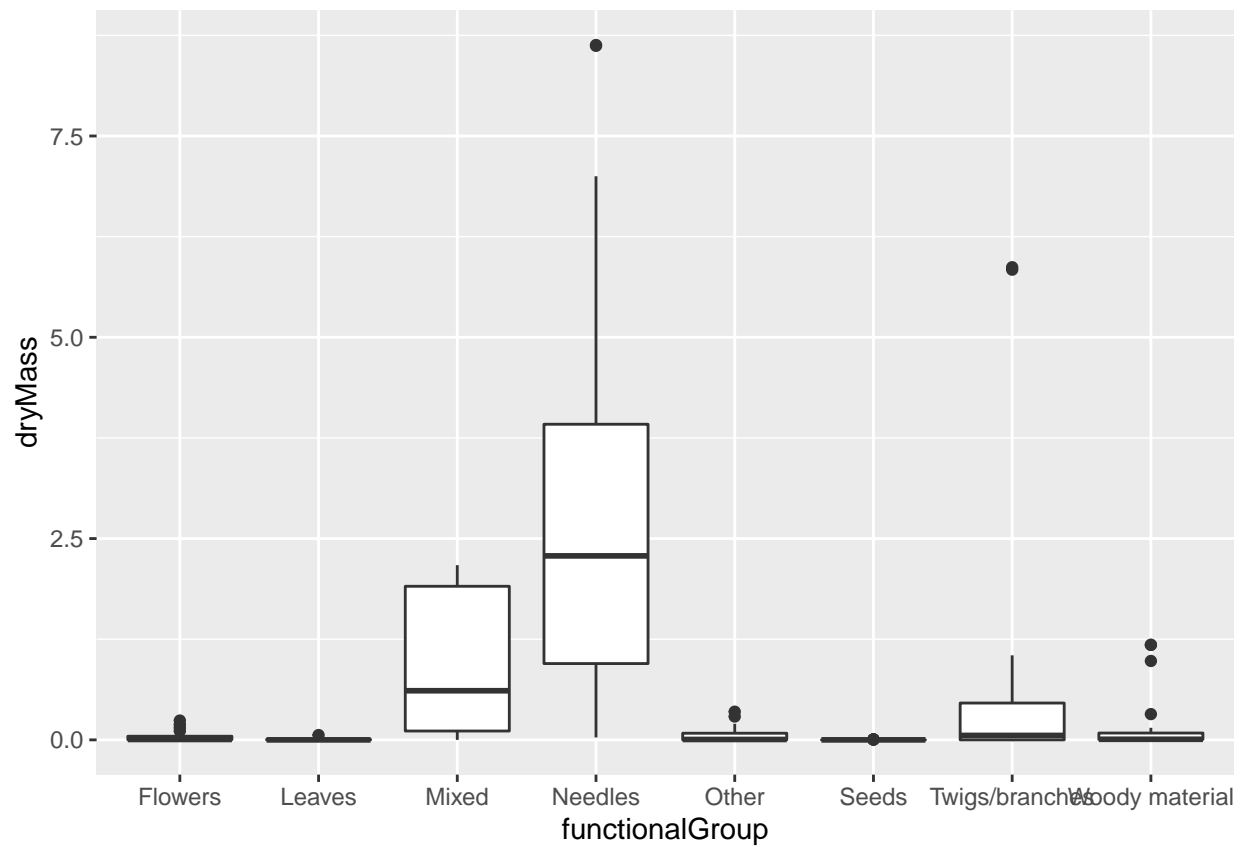
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter, aes(x = functionalGroup)) +  
  geom_bar()
```

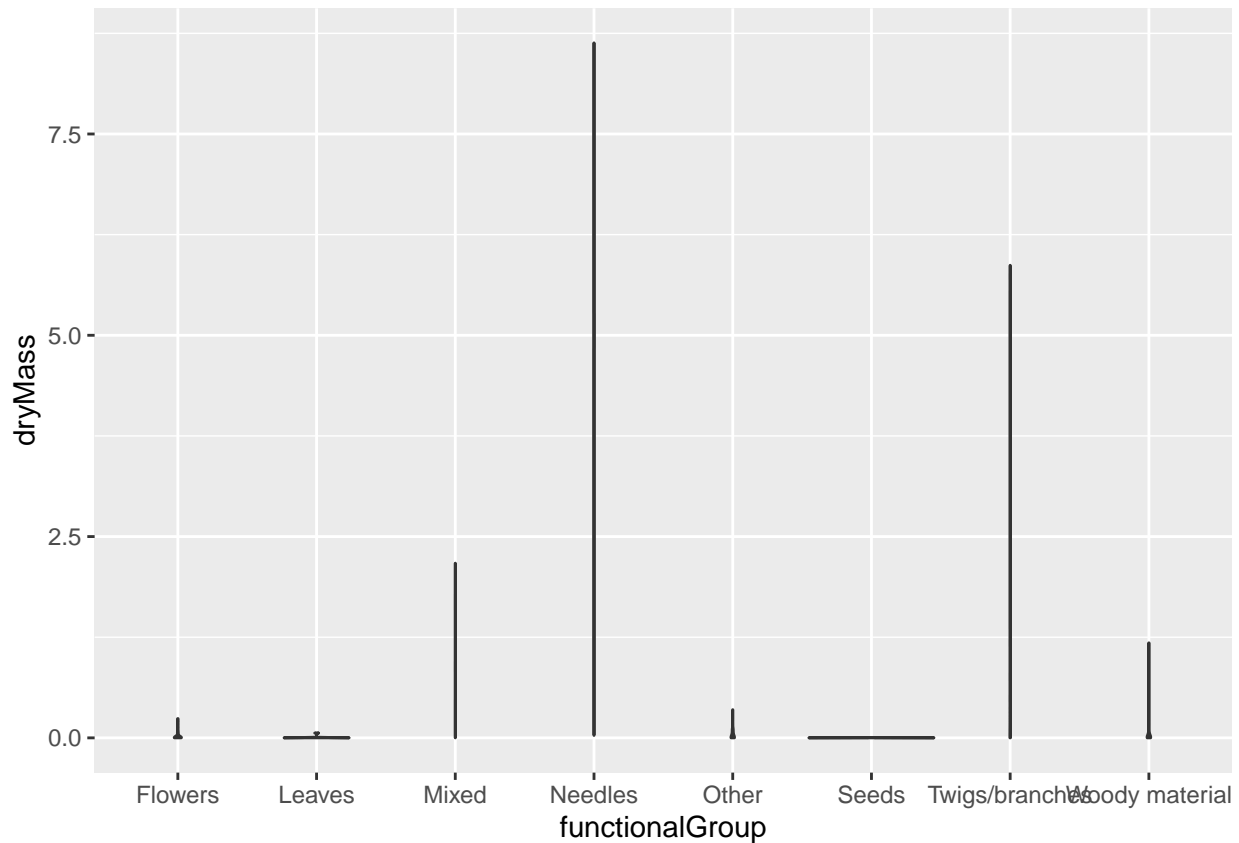


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
# Boxplot
ggplot(Litter) +
  geom_boxplot(aes(x = functionalGroup, y = dryMass))
```



```
# violin plot  
ggplot(Litter) +  
  geom_violin(aes(x = functionalGroup, y = dryMass))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot is a more effective visualization option than the violin plot in this case because, for many of the functional groups, the 'dryMass' data has a tight distribution/little variation. Therefore, a more complex visualization, such as the violin plot, is not as useful.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles