



Biology of the *Macrocystis* resource in North America

- [1. IDENTITY](#)
 - [2. DISTRIBUTION, ECOLOGY AND METABOLISM](#)
 - [3. LIFE HISTORY](#)
 - [4. POPULATION STRUCTURE AND MORTALITY](#)
 - [5. PRODUCTIVITY OF THE RESOURCE](#)
 - [6. METHOD OF HARVESTING AND HARVESTING CYCLE](#)
 - [7. EQUIPMENT USED FOR HARVESTING](#)
 - [8. PROTECTION AND MANAGEMENT OF THE RESOURCE](#)
 - [9. UTILIZATION](#)
 - [10. REFERENCES](#)
-

By
Wheeler J. North
W.M. Keck Engineering Laboratories
California Institute of Technology
Pasadena; California 91125 USA

ABSTRACT

The major North American source of alginate is *Macrocystis* from California. The area, standing crop and yield in California have varied greatly, e.g., from a maximum of 358,425 wet tons from 13,171 ha in 1918 to a minimum of 236 T from 13,171 ha in 1931 to recent tonnages of about 100,000 to, in 1984, 47,320 T from 4,765 hectares. To some extent variations in the southern sea level index and appearance of the El Nino current off the west coast of South America are related: a complex of grazer and other phenomena have been implicated.. A large amount of experimental physiological and population work is reviewed.

1. IDENTITY

- [1.1 Nomenclature](#)
 - [1.2 Taxonomy](#)
 - [1.3 Morphology and anatomy](#)
-

1.1 Nomenclature

The genus *Macrocystis* (Macro = large, Kystis = bladder) is presently recognized as containing three species¹ *M. pyrifera*, (refers to the pear-shaped or pyriform bladders), *M. angustifolia* (narrow leaves), and *M. integrifolia* (complete leaves). There were formerly many more species recognized, based on frond morphology. Further study revealed that frond morphology could be profoundly affected by environmental factors.

Later workers consequently chose holdfast morphology as a basis for species separation (Womersley, 1954). The holdfast of *M. pyrifera* is terete with an upright axis usually conical for plants a year or more old. Holdfasts of *M. integrifolia* are more or less prostrate with fronds developing from a creeping rhizome with a flattened axis. Holdfasts of *M. angustifolia* are intermediate between *M. pyrifera* and *M. integrifolia*. Neushul and coworkers have obtained viable crosses between all three species.

¹Note added in proof, a fourth species, *M. laevis*, has just been proposed by C.H. Hay (Phycologia, Vol. 25, 1986, pp. 241-252).

1.2 Taxonomy

Macrocystis belongs to the Order Laminariales, which includes those Phaeophyta growing by means of an intercalary meristem as well as a superficial meristoderm and has oogamous reproduction by microscopic dioecious gametophytes. *Macrocystis* above the holdfast is differentiated into a blade and stipe, with the blade being split forming several intercalary meristems during early sporophytic development. This places it in the Family Lessoniaceae.

1.3 Morphology and anatomy

Microscopic stages in the *Macrocystis* life cycle (i.e. gametophytes and embryonic sporophytes) resemble those of other members of the Lessoniaceae (see below) and they cannot be distinguished from one another. The morphologies of the macroscopic adult sporophytes differ widely in this family and constitute the basis for separation into genera and species. The typical *Macrocystis* adult sporophyte (**Figure 1A**) consists of numerous blade-bearing fronds that arise from a basal branching system of stipes. Lower portions of the basal stipe system also produce root-like haptera that grow downwards, wrapping around and forming attachment to irregularities in solid substrata. Uniquely the haptera of *M. angustifolia* can also attach to sedimentary bottoms (See Section 2. 3). The mass of haptera constitutes the holdfast.

Figure 1A. Diagram of the *Macrocystis* life cycle showing (left side) development of the young diploid sporophyte, increasing frond numbers through production of basal and apical meristematic blades; (.right side) growth habit of an adult diploid sporophyte ca two years old, standing in 10 m of water depth, and liberating haploid zoospores; (below center) development of haploid gametophytes from settled zoospores, proceeding to gametogenesis, and fertilization yielding the zygote and, thence, a diploid embryonic sporophyte.

New young fronds arise (**Figure 1A**) from longitudinal splits in meristematic blades located on the basal branches just above the holdfast (the basal meristems, **Figure 1A**). The outer blade of the two produced by the splitting becomes an apical frond initial while the inner portion remains as a basal meristem that will give rise to yet further new frond and basal meristem initials.

Splits appear near the base of the outer (apical) frond initial of a young frond, and developing longitudinally toward the blade edge, subdivide the original lamina into lanceolate strips each of which becomes one blade on the young frond. Elongation occurs in that part of the blade base where the splits arise, yielding the stipe of the young frond and causing separation of the newly forming blades. Further stipe growth and splitting produces an apical scimitar-shaped blade distinct from the remaining

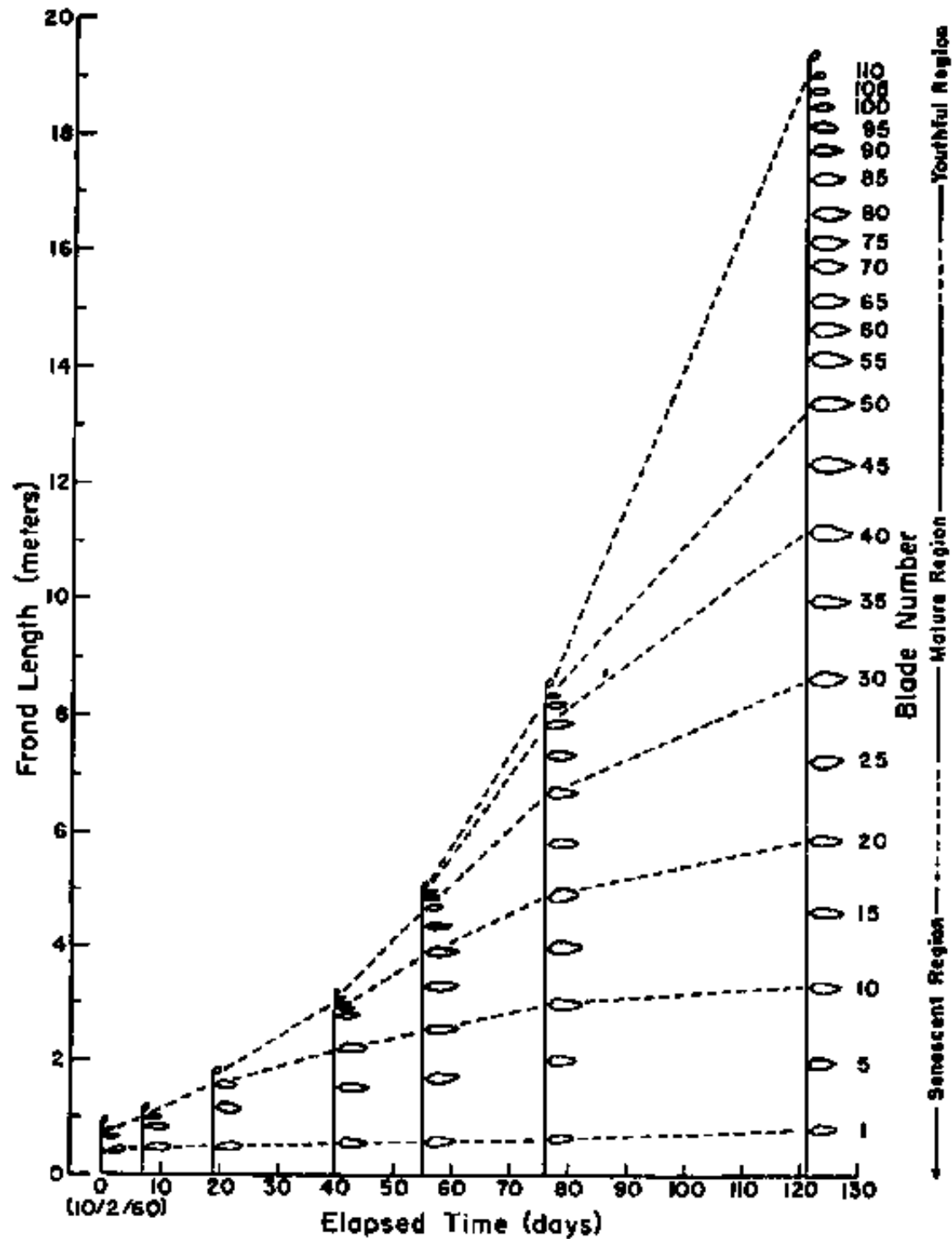
blades. This is the apical meristem which generates additional blades and stipes as the frond continues to develop. The lowermost 2 or 3 blades lack pneumatocysts (blisters) but thereafter all upper blades carry these pyriform flotation devices joining the laminae to the stipe. Continued elongation in the stipe plus production of new blades by the apical meristem results (**Figure 1B**) in elongation of the frond and increase in the numbers of blades. Eventually the apical meristem reaches the water surface and then is pushed horizontally in the canopy by continued growth throughout the stipe length. Finally, meristematic activity ceases in the apical blade and a terminal blade is formed. A fully grown frond may display (**Figure 1B**) 100 to 200 or more blades.

A mature frond thus ultimately consists (**Figure 1B**) of a basal senescent, a central mature and a terminal youthful region as sections of differing ages. The oldest blades occur near the base with a gradient extending upwards to the youngest blades at the apex. Senile deterioration often occurs among the lowest blades of a mature frond even while new youthful blades are developing in the upper regions. The lifespan of an entire frond is about six months but may be somewhat longer or shorter, depending on environmental conditions in the surrounding water. Senile, deteriorating fronds are replaced by young new fronds constantly arising from the basal meristems just above the holdfast. If an apical meristematic blade is severed from a growing frond, further blade production ceases. The remaining basal portion of this frond survives until it is overcome by senility near the end of its normal lifespan.

Harvesting a *Macrocystis* canopy removes all apical meristems lying above the cutting depth. Canopy regeneration is accomplished by growth from fronds whose apical meristems all lie beneath the cutting depth. The lowermost two to six laminae (called sporophylls [**Figure 1A**]) at the base of a frond develop reproductive sporangia. These sporangia are densely packed in certain areas, called sori, on the sporophyll blade surfaces. Each saclike sporangium at maturity releases numerous biflagellated zoospores which develop into gametophytes if they attach to a suitable substratum (see below). The chromosome numbers for a haploid *M. integrifolia* cell are 14 to 16 (Cole, 1968).

Adult *Macrocystis* sporophytes generally display increased numbers of fronds as they age. Very large plants in protected waters may bear several hundred fronds and may be many years old. In exposed locations, however, storms rip holdfasts loose from their attachments and such plants do not usually survive longer than 2 to 3 years. The largest plants under such circumstances may bear 40 to 60 fronds. Fronds growing from holdfasts in deep water do not produce substantially more blades than are found on comparable fronds growing in shallow water. Fronds compensate for greater water depth by increasing internodal distances (lengths of the stipe sections between adjacent pneumatocysts), not by increasing the total blade numbers.

Figure 1B. Time scale frond development illustrated from consecutive observations of one frond of an adult *Macrocystis* plant standing in a water depth of 20 m off La Jolla California. The senescent; mature, and youthful regions of the frond are shown but only some of the blades (from North, 1979a).



2. DISTRIBUTION, ECOLOGY AND METABOLISM

[2.1 Geographical extent](#)

[2.2 Local vertical and horizontal distribution](#)

[2.3 Effects of ecological determinants](#)

[2.4 Nutrition and growth](#)

2.1 Geographical extent

The genus *Macrocystis* occurs naturally around every major land mass in the Southern Hemisphere except for Antarctica (i.e. South America, South Africa, Australia, New Zealand), and also along the eastern Pacific coast in the Northern Hemisphere. Presumably a cold water bridge that formed across the Gulf of Panama on the Pacific side of Latin America permitted transport of *Macrocystis* from South America northward to Mexico, United States and Canada. *Macrocystis pyrifera* was introduced to Asiatic waters in 1978 by scientists from the People's Republic of China (Liu *et al.*, 1984). The species has also been successfully cultured in North Atlantic waters on at least three separate occasions. Thus it appears that *Macrocystis* will survive and grow in any of the temperate oceanic waters of our planet. It has not been able to colonize in either tropical or high arctic climates. *M. pyrifera* and *M. integrifolia* occur in both Hemispheres. *M. angustifolia* was described from Australia (Womersley, 1954). Neushul (1971) identified the *Macrocystis* growing near Santa Barbara, California, as *M. angustifolia*, based on Womersley's description.

2.2 Local vertical and horizontal distribution

Vertical distribution of *Macrocystis* is apparently affected more by local conditions than by preferences or a specific range of depths. For example, given year-round suitably low water temperatures, ample nutrients, and protection from wave action, both *M. pyrifera* and *M. integrifolia* can colonize low intertidal pools. On exposed coasts, however, the inner limit of large *Macrocystis* beds is usually at 7 to 10 m depths and is apparently controlled by wave action. The outer limit of *Macrocystis* beds in California coastal waters usually occurs at depths of 10 - 20 m. The depth range may increase to 25 - 30 m in very clear waters around offshore islands. Presumably the lower depth limit is governed by the bottom illumination required to sustain development of the microscopic gametophytes and small sporophytes. The deepest adult plant ever observed by the author was 40.2 m beneath the surface at Punta Banda, Mexico, an area of extremely clear water.

Local horizontal distributions are most frequently controlled by the availability of rocky substrata. Holdfasts of *M. pyrifera* or *M. integrifolia* require solid substrata for firm attachment. *Macrocystis angustifolia* does occur on sedimentary bottoms. (See Section below.)

2.3 Effects of ecological determinants

The primary ecological factors affecting success and survival of *Macrocystis* populations in the United States and Mexico (and probably elsewhere in the world) are water motion, nutrient availability, water temperature, competition, grazing organisms, bottom illumination and sedimentation, more or less in that order of importance. We discuss nutrients and grazing below, so primary emphasis here will be on wave action, water temperature and illumination.

Importance of wave action as a disruptive factor is demonstrated by the distribution of *Macrocystis* in southern California, comparing the well-protected Santa Barbara coastline with the remaining exposed habitat. Harger (1983) reported that the coastline from Santa Barbara to Point Conception (including San Miguel Island) contained 65 percent of all kelp bed areas in southern California along only 18 percent of the entire shoreline. Variation of canopy coverage was smallest for the Santa Barbara region.

These statistics suggest that some degree of protection against wave action is very beneficial for increasing area of coverage and stability of kelp beds.

Studies of plant mortality in exposed kelp beds clearly indicate that maximum life span of adult plants is short, usually around two to three years for most of the population (North, 1971a; Rosenthal et al., 1974; Dayton et al., 1984). Juvenile recruitment thus is necessary to assure continuation of the population. While wave action can be destructive, water motion is also necessary for nutrient renewal. Back and forth motion such as caused by waves creates a scrubbing action across blade surfaces enhancing nutrient exchange (W.N. Wheeler, 1982; Gerard, 1982a). Longer period motion resulting from tidal cycles, internal waves, and alongshore current replenishes dissolved nutrients within beds and transports food and waste products for the associated fauna. Resistance to flow reduces current velocities within the kelp beds and causes deviation of flow around kelp stands (Jackson and Winant, 1983).

Nutrient availability is negatively correlated with water temperature for values greater than 15.5°C (Zimmerman, 1983). Consequently, the many reported observations of summertime deterioration by *Macrocystis* attributed to high temperature, probably represent combined effects from elevated water temperatures and low nutrients (Jackson, 1977; Zimmerman and Kremer, 1984). North and Zimmerman (1984) showed that fertilizing an experimental group of kelp plants resulted in substantial canopy preservation during summertime compared to an unfertilized group of control plants. In natural beds, stratification in the water column during summer results in a warm, nutrient-poor environment above the thermocline and a cool, nutrient-enhanced milieu below. Consequently, kelp deterioration is greatest in the surface layers while basal portions may survive with little, if any, damage. When sea surface temperatures fall during autumn, basal portions may regenerate the canopies in a few weeks or months.

Summertime deterioration is typically accompanied by a number of symptoms or signs which may conveniently be grouped into a single term we shall call "summer syndrome". Most of these symptoms can be seen within kelp beds at other times of the year as well but less frequently. Prevalence may simply increase sharply during warm summers. Summer syndrome includes loss of coloration in kelp tissues, lesions in the blades, tissues may become brittle and break easily or they may become soft and slough away, coverage by encrusting animals may increase, bladders become perforated or may simply fill with liquid so that fronds lose buoyancy and sink, apical blades become scarce and roost fronds display terminal blades. At a physiological level, photosynthetic capacity declines (Gerard, 1984a). Some of these symptoms undoubtedly arise from nutrient insufficiency (i.e. loss of color). Others may be indirect effects of increased temperature (e.g. lesions and sloughing, caused by enhanced microbial activity). Clendenning (1971) reported that light-saturated photosynthesis was always highest over the range of 20° to 25°C among all kelp tissues he studied (except for kelp from central Baja California which peaked in the range 25° to 30°C) during short term exposures. Summer syndrome, however, usually develops at around 20°C. Appearance of summer syndrome apparently involves relationships between duration of a period of elevated temperature and the level to which temperature rises. Thus a temperature of 20° may be tolerated for 2 or 3 months while 23°C or more for a month will result in rapid deterioration.

The microscopic stages of *Macrocystis* may be more sensitive than adult plants to elevated water temperatures. Dean and Deysher (1983) reported that the upper limit for sporophyte production from gametophytes they outplanted to a natural kelp bed was 16.3°C.

Competition from other algal species takes several forms including shading of juvenile *Macrocystis* (i.e. competition for light), competition for space, substratum, and possibly competition for nutrients. There may also be poorly understood competitive mechanisms. For example, small *Macrocystis* and other laminarian sporophytes appear to recruit poorly on substrata thickly covered by encrustations of coralline algae and animals when compared to non-encrusted surfaces. On the other hand, remains of old decaying *Macrocystis* holdfasts sometimes support numbers of young sporophytes while none occurs on the surrounding bottom. These observations suggest that chemical mechanisms may exist which affect suitability of a substratum for colonization by kelp spores. Barren rocky bottoms severely and constantly grazed by sea urchins are usually excellent substratum for colonization by *Macrocystis* and other brown algae after the urchins are eliminated; provided, of course, that fertile adult plants are close by enough to serve as spore sources. When territory dominated by *Macrocystis* is lost due to some catastrophic effect, other seaweed species may colonize the area and prevent *Macrocystis* from reestablishing. Persistence, resistance to invasion, and resilience or recoverability after loss of territory are discussed by Dayton and Tegner (1984) and by Dayton *et al.* (1984). Physical abrasion from nearby blades of tough species such as *Laminaria* and *Agarum* may damage small *Macrocystis* sporophytes, (R. McPeak, pers. comm.).

Significant grazing damage can be inflicted on *Macrocystis* plants by several herbivores including sea urchins, crustaceans, mollusks and fishes. Probably the most damaging grazers in California waters are the sea urchins, *Strongylocentrotus franciscanus*, *S. purpuratus* and *Lytechinus anamesus*. These echinoids can occur in enormous populations at concentrations well above 10/m². The animals migrate slowly across the sea floor, consuming all plant life except for crustose corallines, as well as many sessile invertebrates such as sponges, tunicates, and bryozoans. Under some conditions, urchin populations persist indefinitely at high concentrations, preventing vegetation from reestablishing and creating "urchin barrens" (Dean *et al.*, 1984). Urchin barrens have been observed in many shallow benthic areas other than California. Disappearances or significant reductions of predators that control urchin numbers permit development of huge urchin populations. Urchin predators include lobster, fishes, sea otters, and asteroids (Tegner and Levin, 1983; Dayton *et al.*, 1980; Estes and Palmisano, 1974). Human exploitation of urchins as food may also act as a significant control, (Wilson and North, 1983). Very severe storms may even remove urchins from kelp bed depths (Ebling *et al.*, 1984). Techniques for artificially controlling urchin populations are described below.

Herbivorous fishes are not usually sources of significant damage to *Macrocystis* populations in dense healthy kelp forests because the amounts of plant tissue consumed are small in relation to total productivity. Fish grazing may assume importance when productivity declines as, for example, in a kelp bed under stressful conditions or where small plants colonize the sea floor after the adult *Macrocystis* plants were destroyed by some catastrophe. Two herbivorous fishes in California waters, *Girella nigricans* and *Medialuna californiensis*, preferentially feed on youthful kelp tissues such as the apical meristems and the blades of very young sporophytes (Harris *et al.*, 1984). A third fish species, *Oxyjulis californiensis* adventitiously removes chunks of kelp tissue when feeding on encrusting bryozoans (Berstein, 1977), sometimes damaging blades significantly. For grazing of small sporophytes or apical meristems, the damage inflicted may be far out of proportion to actual amounts of tissue consumed. Effects from fish grazing may be especially significant when small young *Macrocystis* plants are first reestablished in an area where a catastrophe has eliminated all the adults.

Leighton (1971) studied food preferences of eleven invertebrate herbivores common in California kelp beds. All eleven showed preferences for *Macrocystis* tissues. He did not include herbivorous fishes in his studies but our field observations suggest that *Macrocystis* is strongly preferred by *Girella* and *Medialuna*. We frequently record instances where extensive damage from fish grazing occurs on blades of small *Macrocystis* plants but none is seen on other recruiting kelp species such as *Eisenia*, *Egregia*, *Pterygophora*, and *Laminaria*. Fish grazing undoubtedly enhances competition from other kelp species on such occasions. Grazing fishes can be a serious problem when we undertake to develop *Macrocystis* stands by transplantation techniques (see below).

Grazing by crustaceans and gastropods infrequently may cause significant damage to *Macrocystis* populations. Numbers of these herbivorous species are usually so small that their activities only result in trivial losses of kelp tissue. Very occasionally, however, dense concentrations of a species may appear so that grazing effects reach destructive proportions. We have observed significantly destructive grazing on *Macrocystis* by large numbers of kelp crabs (*Pugettia*), abalone (*Haliotis*), dove snails (*Mitrella*), and microcrustaceans (primarily *Ampithoe* and *Idothea*).

Adequate sunlight is, of course, a requirement for all plants, but effects of cloudy days or turbid water are less for adult *Macrocystis* than for short-statured algae because of the extensive canopies which extend just beneath the surface where submarine illumination is maximal (cloudy days may affect kelp productivity but not survival of adult plants). The primary influence on *Macrocystis* populations by attenuation of submarine light is the effect on microscopic plants and recruitment of small plants. Dean and Deysher (1983) outplanted gametophytes to a natural kelp bed at San Onofre, California, and after six weeks recorded densities of resulting sporophytes on their artificial substrata. They found that no sporophytes were produced at irradiances below 0.4 E/m²/d. They also noted an interactive effect between water temperature and irradiance. As temperatures approached 16°C, about 1E/m²/d was required for moderate spore-phyte production. These authors also measured daily irradiance on the bottom in clear areas within the San Onofre kelp bed for 3 1/2 years (July, 1978 to January, 1982). Favorable irradiances over 2-week intervals combined with suitable bottom temperatures occurred only eight times during the 3 1/2 year interval and each recruitment was traceable to a period of favorable light and temperature as defined by Dean and Deysher's measurements. These authors postulated that "windows" of environmental conditions favoring recruitment occurred only occasionally and were critical to persistence of this kelp bed. Photosynthesis by *Macrocystis* gametophytes and embryonic sporophytes became light saturated at 70 m E/m²/s (Fain and Murray, 1982). Compensation irradiance for embryonic sporophytes was 2.8 m E/m²/s. Luning and Neushul (1978) estimated that 20 m E/m²/s was adequate for saturation of vegetative growth in *M. pyrifera* and *M. integrifolia* gametophytes but 2 to 3 times this irradiance was necessary to induce gametogenesis.

Sedimentary deposits can affect *M. pyrifera* in several ways. Even very light sedimentary films may interfere with spore settling under laboratory conditions (Devinny and Volsse, 1978). An approximately tenfold increase in sedimentary load caused burial of already settled spores and interfered with their survival. Presumably due to sediment scouring, water motion in the presence of fine sediment further reduced survival. Dean and Deysher (1983) noted a substantial influence from sedimentation on sporophyte production by their gametophyte outplants to the San Onofre kelp beds (in their regression analysis, sedimentation accounted for 11% of the variance observed, temperature 25%, light 10%, nitrogen 0%). The thin sedimentary deposits that may bury

microscopic stages of *Macrocystis* are, of course, ineffective against larger plants. Major sedimentary shifts have been noted, however, damaging young and even adult sporophytes (Neu-shul, 1959; North, numerous observations, unpub.).

Partial burial of holdfasts does not appear to affect haptera adversely, e.g., the haptera of *M. angustifolia* actively growing downward into the sedimentary bottom. The haptera penetrate deeply into the sediments, and interstices within that part of the holdfast above the bottom become filled with sand or mud. This plant produces holdfasts that may be 3 to 5 m in diameter but less than one meter tall (i.e., low mounds). By trapping and penetrating the sediment, it creates a heavy mass that withstands displacement by wave action.

Burial of the primary or secondary stipes at the holdfast apex is not tolerated well. Stipe tissues rot in a few days if they are not freely exposed to surrounding water. Blades also are intolerant of even light sedimentary deposits and may disintegrate after a few weeks of coverage. This type of damage is common among *Macrocystis* plants growing in quiet waters of bays and inlets where scrubbing action by wave surge is lacking. The Santa Barbara region, however, is protected from long period waves by offshore islands and orientation of the mainland.

Ecological determinants of lesser importance affect *Macrocystis* from time to time. Aggregations of encrusting animals may appear on *Macrocystis* blades so densely as to cause loss of buoyancy and sinking of fronds. Encrustation of kelp by the Bryozoan, *Membranipora*, enhances blade loss by reducing flexibility and by encouraging fish grazing on the Bryozoan colonies (Dixon *et al.*, 1981; Kirkwood, 1977). Compensating light intensity was about 50% higher for *Membranipora* encrusted than for clean *Macrocystis* blades (Wing and Clendenning, 1971). Isolated plants and those near the edges of kelp beds are more susceptible to colonization by *Membranipora* (Bernstein and Jung, 1979).

Salinity remains sufficiently constant along open coastlines so that distribution of *Macrocystis* populations is rarely, if ever, affected in California. Lowering of salinity in bays and estuaries may regulate *Macrocystis* distribution in these habitats, especially north of southern California where rainfall increases. According to Druehl (1979), beds of *M. integrifolia* are able to grow at Nootka, Vancouver Island, where the yearly salinity range was from about 23 ‰ to 30 ‰. Druehl indicated that high temperatures (i.e. 14° - 18°C) during periods of low salinity were associated with absence of *Macrocystis* and that the season of low salinity at Nootka coincided with a time of low temperatures (8° - 12°C). North (unpub.) grew gametophytes of *M. pyrifera* in serial dilutions of open ocean seawater and distilled water. Gametophytes did not survive at dilutions greater than 3 to 1 (i.e. just below 25 ‰ salinity if we assume the salinity of the undiluted seawater was 33 ‰. This conclusion was in rough agreement with Druehl's (1979) observations.

2.4 Nutrition and growth

Kelp growth is probably rarely, if ever, limited by availability of micronutrients in surface seawater. Kuwabara and North (1980) demonstrated nine elemental requirements by *Macrocystis* including the macronutrients nitrogen and phosphorus as well as the micronutrients copper, zinc, cobalt, manganese, iron, molybdenum and iodine. Kuwabara (1982) found indications that sea-water from 300 m depths in southern California was somewhat deficient in manganese and cobalt for growth by *Macrocystis* gametophytes.

Nitrogen may at times be a limiting element in surface water and very occasionally phosphorus (Manley and North, 1984). Luxury uptake of nitrogen and phosphorus occurs in *Macrocystis* during times when these elements are plentiful and reserves are utilized when external supplies are inadequate (Gerard, 1982b; Manley and North, 1984). These authors indicated that values of critical tissue levels (where the element is neither in excess nor in short supply) for *Macrocystis* were 0.2% dry wt. for Phosphorus and 1.0% dry wt. for Nitrogen in blade tissues. P.A. Wheeler and North (1980) found that blade N-contents of juvenile sporophytes reached a maximal value of 3% dry weight when external nitrate concentration in a flowing water culturing system was at 13 m M. Gerard (1982b) transplanted large adult sporophytes with laminar tissues initially containing 2.2 to 3.5% dry wt. Nitrogen from a coastal kelp forest to an offshore island site where ambient external Nitrogen was well below m M (as indicated by continual temperature measurements). Growth rates stayed at relatively high levels for two weeks, then declined sharply. Ambient Nitrogen rose during the fourth week and growth rates recovered somewhat. The Nitrogen contents of laminae declined steadily during the initial 3 weeks of starvation, then remained constant or rose slightly thereafter. Holdfast Nitrogen remained constant throughout. Gerard (1982c) further proposed a mathematical model for nitrate uptake by whole plants. Zimmerman and Kremer (1984) demonstrated a relationship between estimated nitrate concentration and frond growth rate. Growth rates fell for nitrate concentrations below 1 m g-at/1.

Haines and P.A. Wheeler (1978) listed values of $K_S=13.1$ m q-at N/1, $V_{max}=3.05$ m g-at N/wet g/h for NO_3 uptake by juvenile *Macrocystis* and $K_S=5.3$ m g-at N/1, $V_{max}=2.38$ m g-at N/wet g/h for NH_4 for concentrations below 22 m g-at/1. W.N. Wheeler (1978) reported maximum uptake rates by tissue discs cut from blades on an adult frond as 75 m m/cm²/h for NO_3 , 275 nM/cm²/h for NH_4 , and 3.8 nM/cm²/h for PO_4 under saturating water motion. All workers reported that nitrate and ammonium were taken up simultaneously without any effects on uptake rates by presence of another form of nitrogen. Manley (1985) found values of $K_S=3.51$ m M and $V_{max}=5.29$ nM/cm²/h for PO_4 uptake by blade tissue discs from adult sporophytes. Manley suggested that differences between his results and those of W.N. Wheeler might arise from differences in tissue Phosphorus levels. Gerard (1982c), using whole blades attached to adult plants, showed that NO_3 uptake rates differed substantially for tissues from different parts of the plant and similar findings have been shown for PO_4 uptake (Manley, 1985; Schmitz and Srivastava, 1979) and for NO_3 uptake by *M. integrifolia* (W.N. Wheeler and Srivastava, 1984). Uptake of NO_3 and PO_4 was more rapid in light than in darkness or under shaded conditions (W.N. Wheeler, 1982; Gerard, 1982a; Manley, 1985). Culturing tissues in water high in NO_3 or PO_4 reduces uptake of these nutrients and Gerard (1982b) found an inverse relation between depth and NO_3 uptake. W.N. Wheeler (1978, 1980a, 1982) found that water motion affected carbon and nutrient uptake rates with uptake saturating in the range of 3 to 5 cm/sec current velocity. Gerard (1982c) concluded that NO_3 uptake regardless of concentration, became saturated at a water flow of 3 cm/sec. She found that water motion under field conditions exceeds this critical value even inside dense canopies during calm seas. W.N. Wheeler (1980a) reported that transport rate of inorganic carbon through the boundary layer on *Macrocystis* blades saturated at flow velocities of 4 cm/sec.

Uptake rates of the micronutrients I, Fe, Zn, Co, and Mn as a function of external concentration were presented by Manley and North (1981), using an artificial seawater. Manley (1984) showed that these elements as well as Mo and Ni were concentrated in

sieve tube sap. He calculated that adult plants sometimes might not be able to fulfill their requirements for Mn and Co, resulting in growth limitation by availability of these elements. Manley (1981) showed that Fe uptake is light independent and energy dependent. Manley (1.c.) and Anderson (1984) both showed that iron is reduced from Fe^{3+} to Fe^{2+} during uptake.

Ability to translocate inorganic and organic ions and compounds is an important feature of nutrition in the Laminariales. Structural elements of the translocation system are maximally developed in the tree-like kelps such as *Macrocystis*. Translocation processes nourish portions of adult plants growing under low illumination near the bottom by photosynthate created in the upper well-lit parts of the water column. The translocation capability allows *Macrocystis* plants to grow in dense aggregates with over-lapping canopies which effectively shade out competitors on the bottom, yet supports rapid growth by young fronds, sporophylls, haptera and other tissues near bases of the plants. The sieve tubes occurring around the periphery of medullary tissue centrally within blades and stipes, are the conducting pathways supporting translocation. The sieve tubes are enclosed by cellular membranes. Osmotic pressure is believed to be the driving force causing movement of fluid through the sieve tubes (Schmitz, 1982). Synthesis of organics and rapid uptake of inorganics in the "source" region of a frond creates high concentrations of solutes there, creating a tendency for intake of water from outside and movement of water plus solutes through the sieve tubes towards more diluted solutions in the "sink" regions. Accommodation of the flow is accomplished by loss of water in the sink region. Flow can be bidirectional within a single mature frond (i.e. from the center toward both ends). Thus, both apical meristems in the upper parts of the water columns and newly developing fronds at the plant base receive photosynthate and nutrients from mature blades located in the central part of a long frond.

Lobban (1978a) showed that newly developing fronds of *M. pyrifera* act as sinks until they are approximately three meters long. Loss or disappearance of an apical meristem caused all translocation to occur toward the base. Translocation in *M. integrifolia* differed from that in *M. pyrifera* in terms of seasonal changes in direction of translocation and in distances from the apices where changes occur (Lobban, 1978b). Manley (1984) postulated that young short fronds probably depend on translocation of Mn, Co and Fe to meet growth requirements because their small total blade areas provided insufficient uptake capacities.

Translocation velocities of 25 to 45 cm/hr were measured using radioactive ^{32}P compounds as tracers in *M. integrifolia* (Schmitz and Srivastava, 1979). These velocities were comparable to velocities shown for ^{14}C compounds. Translocation velocities in *M. pyrifera* blades were only 6 to 22 cm/hr., possibly because of small diameter pores in sieve plates forming the interconnections between adjacent sieve tube cells (Buggeln et al., 1985).

Phosphorous is assimilated from the exterior in inorganic form but is primarily incorporated into organic compounds during translocation in the sieve tube sap. Similarly, organic forms of nitrogen are taken up very slowly or not at all by *M. pyrifera* (P.A. Wheeler, 1979), yet nitrogen in sieve tube sap occurs almost entirely as amino acids (Jackson, 1976). Jackson (1977) calculated that the flux of nitrogen due to translocation in *Macrocystis* stipes amounted to 0.2mM/hr, while that of carbon was 2mM/hr. The principal amino acid in *M. pyrifera* was alanine.

The initial product of photosynthesis in *Macrocystis* is mannitol (Vaughn, 1959) and the primary low molecular weight carbohydrate was mannitol in both *M. pyrifera* and *M.*

integrifolia (Jackson, 1.c., Schmitz and Srivastava, 1979).

The basis of growth and productivity in *Macrocystis*, as in all plants, is photosynthesis. Photosynthesis in an adult *Macrocystis* sporophyte is affected by a number of factors, resulting in an extremely complex situation. Different tissue types (i.e. blades, stipes, sporophylls, haptera) display different photo-synthetic capacities (Arnold and Manley, 1985). Gradients in photosynthetic capacities exist from base to tip within blades and between blades along the frond length (Arnold and Manley, 1.c.; W.N. Wheeler, 1980b; Clendenning, 1963). The blades account for nearly all the photosynthetic activity of *Macrocystis* sporophytes in nature). The submarine light field that drives photosynthesis is extremely variable and complex in kelp beds as a consequence of canopy absorption, refraction by ripples and waves, and constant movement by blades resulting in flashing light patterns in the upper part of the water column (Gerard, 1984b). Vertical distribution of blade area is not uniform (Jackson et al., 1985) with 50% or more of the total area occurring near the surface in adult plants displaying full canopies. Very substantial seasonal changes in photosynthetic capacity may occur for both *M. pyrifera* (Clendenning 1971) and *M. integrifolia* (Smith et al., 1983) due to changes in water temperature, availability of micronutrients and general healthiness of the blades. Jackson (personal communication) is developing a computer-based mathematical model of kelp bed photosynthesis and productivity that takes account of the many factors affecting photosynthesis in *Macrocystis*. The model is still in a preliminary state of development, but in its present state has yielded interesting predictions as to effects of planting density, changes in latitude, and seasonal changes in plant size (the seasonal changes in plant size predictions were confirmed by measurements conducted by Kirkwood, 1977, in a *Macrocystis* bed at Palos Verdes, California).

Growth in *Macrocystis*, on a short term basis, consists of development by individual fronds and haptera and, over the longer term, increase in holdfast size and total numbers of fronds per plant. The short term growth parameters can be easily determined through in situ measurements of hapteral and frond lengths by divers. Long term results of growth can also be estimated in situ (measuring holdfast dimensions, counting numbers of fronds), or by removing plants from the water and weighing them (Neushul and Harger, 1983; the latter technique requires correction for added weights from extraneous factors such as entrapped water and sediment as well as encrusting organisms). Gerard (personal communication) developed a technique for estimating changes in frond biomass based on in situ measurements. First, a relationship is determined for frond length vs frond weight by sampling fronds from a target population. Then the frond size distribution pattern is determined by measuring overall lengths of a large number of randomly selected fronds from the population in situ. Concurrently, frond density is estimated in situ by counting all fronds within randomly positioned quadrats of known areas. Biomass per unit area is then estimated as the sum of the mean weights of the various size classes of fronds, adjusted for the frequency of occurrence of each size class in the total frond population, multiplied by the density of fronds per unit area. The determinations are repeated at a later date and the difference between the two sets of measurements provides an estimate of biomass change during the interval. The productivity of each frond size class can be assessed by measuring elongations of fronds in the population over a given time period and calculating the resulting weight increment from the length-weight relationship. Summing productivities of each frond size class (adjusted for frequency of each size class) yields productivity by the entire frond population.

Methods for assessing frond growth have been variously proposed (North, 1971b; Kain, 1982; Gerard, 1982b; Grua, 1964; Manley, 1984). Changes in mean frond growth rates

are useful for assessing healthiness of *Macrocystis* populations. Frond growth rates, measured either as length or weight changes, are not uniform with time, but change as the frond ages. It is thus necessary that measurements either be made on a sample of fronds all of similar ages or that corrections for the age factor be introduced if the sample includes fronds of various ages.

3. LIFE HISTORY

[3.1 Life cycle and reproduction](#)

3.1 Life cycle and reproduction

The *Macrocystis* life cycle (**Figure 1A**) consists of a dimorphic alternation of generations between a macroscopic diploid sporophyte and microscopic haploid gametophytes. Biflagellated zoospore production begins after the sporophytes are 6 to 12 months old. Spores are transported away from the parent plant primarily by natural water movements and secondarily by their own swimming activity. Those settling on suitable substrata produce germ tubes in a few hours. The spore content migrates down the germ tube in producing the first cell of the gametophyte (a few gametophytes apparently do grow directly from the spore without intervention of a germ tube). The initial gametophytic cell grows and divides into one or two cells (females) or several cells (males) in a week or two.

Male and female gametophytes can be differentiated after about a week in laboratory cultures by color and shape. Males are pale or bluish, the cells are small, and the overall shape is often filamentous. Females are yellow-green, gold, or brown with round or ovoid shapes. Gametogenesis may begin in 7 to 10 days. Antheridia on males liberate biflagellated sperm. The entire gametophyte in single celled females may form the deeply pigmented ovum, or only one cell may become an ovum in multicellular females. All cells, however, may eventually develop into ova.

The ovum is extruded from the oogonium and becomes fertilized. Extrusion may or may not rupture attachment of the ovum to the oogonium. If detached, the ovum or zygote is capable of settling elsewhere and forming a new attachment. Rapid cell division in the zygote produces the embryonic sporophyte. Within a few weeks, growth yields tiny blades just visible to the naked eye.

The single blade stage in *Macrocystis* closely resembles that of other Laminariales. Positive identification as *Macrocystis* can be made when traces of the first or primary split occurs immediately at the junction of blade and stipe which eventually creates two separate blades or basal meristems (**See Figure 1A**). Further splitting and appearance of frond initials leads to development of the adult morphology in the young plant.

Although spore production is enormous (Neushul, 1959, estimated there were 10,000 sporangia/cm² in *Macrocystis* sporo-phylls), probably only a small fraction ever settle on suitable substrata and vastly smaller numbers survive through the microscopic phases of the life cycle (Anderson and North, 1966; Deysher, 1984).

Young sporophytes normally appear at only distances of a few meters to ca 100 m from attached plants. Drifting adult sporophytes, however, may disperse spores in areas lacking attached *Macrocystis*, possibly providing opportunities for establishing the

species.

4. POPULATION STRUCTURE AND MORTALITY

[4.1 Age, weight or size composition](#)

[4.2 Sporophyte - gametophyte sex composition](#)

[4.3 Mortality, morbidity](#)

4.1 Age, weight or size composition

Natural *Macrocystis* populations vary widely in their population structures. Sometimes nearly all plants will be of similar sizes and ages while in other cases plants of varying ages and sizes occur intermixed. The former type of age-size structure sometimes arises after a major catastrophe such as an exceptional storm destroys almost all *Macrocystis* throughout a large area. Normally, the bottom in such an area would be well seeded with microscopic spores and, thus, gametophytes. Removal of most adult plants would increase bottom illumination dramatically, stimulating gametophytic growth and production of embryonic sporophytes. Assuming good survival, a population of adult sporophytes of uniform age would eventually develop. Once a canopy is formed, further recruitment of juvenile plants ceases and only the oldest plants with fronds in the canopy receive enough sunlight to survive. The shorter plants perish for lack of sunlight, leaving a population of uniform age and size. If a few of these survivors perish more or less randomly as time passes, canopies of the remaining plants will still prevent recruitment of juveniles. Random attrition and continued growth by the survivors produces a population of scattered old plants with large holdfasts, each supporting many fronds.

Populations of variously sized plants occur where forces of attrition are high so that mortalities occur frequently. Patches of open water can appear between adult plants, allowing enough sunlight into the water column to support areas of juvenile recruitment during periods of clear water. Thus, plants of all sizes and ages coexist (for harvesting effects, cf. Kimura, 1980).

One of the major agents of attrition in *Macrocystis* forests is dislodgement of plants by storm-generated waves. Storm-attrition is greater the shallower the water. Consequently, the offshore edges of kelp beds experience less storm attrition than inshore edges. We frequently observe uniformly large and widely spaced old plants along the offshore borders of *Macrocystis* beds, while inshore portions of the beds support mixed sizes of fairly young plants, often in dense aggregates.

A unit commonly employed for assessing size among adult *Macrocystis* plants is the number of fronds. The typical adult *Macrocystis* is composed of fronds of varying lengths ranging from initials a few cm long to fully mature fronds with 5 to 10 m or more of their length stretching horizontally in the canopy (the longest frond I ever measured was 44.8 m and probably 3 m was missing from the distal end). Although frond lengths on a given plant vary, frond size distribution per plant stays within narrower limits (North, 1968), particularly for populations growing at a given depth. The average weight of all fronds in a population thus tends to be similar, increasing slightly with depth. Thus the mean wet weight of all fronds from plants growing at depths of 10-20 m is slightly more than 1 kg, while for plants growing at 20-25 m depths, the mean wet weight is ca 1.5 kg (North, 1958). Total biomass in a population can be estimated by multiplying frond density

(obtained by counting all fronds at the plant bases occurring within several randomly-located quadrats in the bed) by the mean wet weight of a frond (i.e. ca 1 kg). For natural kelp beds with coherent canopies, a range of biomasses of 3 to 22 kg/m² was reported by North (1971a).

4.2 Sporophyte - gametophyte sex composition

Nothing is known about proportions of *Macrocystis* sporophytes to gametophytes in natural populations, or of the sex composition of gametophytes. The microscopic dimensions of gametophytes make them difficult to detect on natural substrates such as rock or hard organic material. Fates of gametophytes and sporophytes have been followed on outplanted artificial substrates (Dean and Deysher, 1983), and recruitment has been monitored on concrete blocks (Foster 1975a, 1975b). Outplanted *Macrocystis* gametophytes survived only 40 to 45 days off San Onofre, California (probably a fairly hostile environment). In laboratory cultures, approximately equal numbers of male vs female gametophytes develop from settled spores.

4.3 Mortality, morbidity

Rosenthal *et al.*, (1974) monitored tagged adult *Macrocystis* in a bed off Del Mar, California for 5.7 years and reported that the primary causes of mortality were: storms, entanglement between drifting and attached plants, and kelp harvesting (holdfasts were presumably pulled loose during cutting). North (1971a) studied *Macrocystis* mortality by tagging plants and following their disappearances over a few months, computing half-lives by assuming that mortality remained fairly constant (a half-life is the time required for 50% of a tagged population to disappear). Half-lives ranged from two months for very exposed populations during winter, to several years for well-protected plants off Santa Barbara, California. Similar results were reported by Dayton *et al.* (1984).

Gametophyte mortalities are probably much greater. Deysher (1984) found that *Macrocystis* gametophytes outplanted on rope substrata, survived only 40 to 45 days off San Onofre (as judged by cessation of sporophyte production on the ropes). San Onofre is believed to be a somewhat hostile environment for gametophytes because of sedimentation (Devinney and Volse, 1978, showed that even light sedimentary deposits were very harmful towards *Macrocystis* gametophytes).

5. PRODUCTIVITY OF THE RESOURCE

[5.1 Productivity and harvesting yield](#)

[5.2 Principal factors affecting productivity](#)

[5.3 Genetic studies and breeding studies](#)

[5.4 Relative contributions from vegetative & sexual reproduction to the harvest](#)

[5.5 Environmental enhancement](#)

5.1 Productivity and harvesting yield

Biomass production by plants depends on availability of sunlight and nutrients. Most of the photon flux through the water surface is presumably utilized when plant biomass is

distributed so as to completely cover the surface, as in dense *Macrocystis* canopies. Nutrient availability is influenced not only by concentrations of nutrients in the surrounding waters, but also, by water motion and amounts of plant surfaces exposed for taking up nutrients. Ryther and coworkers have shown that approximately parabolic relationships exist between production and density of biomass when aquatic plants and seaweeds are grown under light limiting conditions (i.e. water motion and nutrients are not growth-limiting). That is, there is an optimum density of plant tissue at which biomass production and yield are maximal. The optimum tissue density is different for differing kinds of plants (e.g. 0.02 kg dry/m² for *Lemna minor*, 0.5 to 1 dry kg/m² for *Eichornia crassipes*, Ryther *et al.*, 1979) and is affected somewhat by season (Ryther *et al.*, 1977).

Neushul and coworkers attempted to determine the optimal biomass density for maximizing yield from *Macrocystis angustifolia* by growing adult transplants at three biomass densities (initial values 1.7, 6.4 and 24.3 fronds/m², representing 0.0625 to 0.25 and 1 plant/m²) on test plots. Half of the plots were fertilized with ammonium sulfate, half were unfertilized. Harvested yields varied seasonally and were highest for the period May-July for all three planting densities (Neushul and Harger, 1983). Projected yields (i.e. assumes that no frond was lost and that all fronds produced were eventually harvested) from five harvests the yields averaged 5.6, 13.9 and 24.8 DAFMT/ha/yr (dry, ash-free metric tons per hectare per year; presumes that one DAFMT = 1.786 dry ton = 14.286 wet ton) from the three densities respectively. Total yields from the fertilized plots were slightly higher than from the unfertilized areas for the two highest densities, but yields from the fertilized plots for all three densities were substantially higher for the July-September period, a time when ambient nutrients were low (Neushul and Harger, 1983). The study thus indicated that the optimal biomass density for *Macrocystis* lies above 24.3 fronds/m². Although I have measured a density of 30 fronds/m² in a natural *Macrocystis* bed, it seems likely that such very high densities cannot be sustained for long. During the course of this 15 month study, Neushul and Harger (1.c.) reported that plants in their high density plots decreased in size so that the overall frond density declined by a half during the study. North (1971a) found a range of 2 to 15/fronds/m² for large size samples from *Macrocystis* beds in California and Mexico. Most populations fall in the range of 3 to 6 fronds/m².

Gerard and North (1984) measured production and harvested yield on a group of 63 adult *M. pyrifera* held for 2 1/2 months in an enclosure with intermittent fertilizing (9-13 uM NO₃ and 1 uM PO₄). Deterioration in the basal portions of the plants occurred, probably from lack of water motion due to the enclosure. In spite of these difficulties, they estimated that 50% of the total productivity was recovered as harvested yield.

Considerable data are gathered by the California Department of Fish & Game as to fresh weights of *Macrocystis* harvested from beds within the State. In most cases, kelp bed areas involved in the harvesting were not distinguished. Neushul *et al.*, (1982) conducted a special study in cooperation with a kelp harvesting concern in which both the weight of kelp and the area harvested were determined for a kelp bed off Goleta, California. They obtained a yield of 8.3 DAFMT/ha/yr. A similar one year study by North *et al.*, (1982) for a fertilized bed off Leucadia, California gave a yield of 12.3 to 14.8 DAFMT/ha/yr. Barilotti (pers. comm.) reported that yields from the kelp bed off Pt. Loma, California, ranged from 1.2 to 4.9 DAFMT/ha/yr.

The total annual harvest of kelp (primarily *Macrocystis*) from California waters shows considerable year to year variation resulting from interactions of both environmental, economic, and other factors (**Table 1**). Data from Neushul (1981a) suggest that areas of canopies have varied substantially, probably accounting for some of the variations in

total harvest. (Examples of canopy variation are shown in **Figures 2 and 3** below). *Macrocystis* was harvested maximally from 1916 to 1918 and from at least 1955 onward because of favorable economics. Kelp was used as a source of potash during World War I, but the demand collapsed after the war terminated (Scofield, 1959). Interest in kelp for animal feed additives and chemicals led to renewed harvesting activity in the late 1920's. Demand for alginate products caused steady increases in harvests in the latter 1930's, the 1940's, and has sustained the industry ever since. High water temperatures, unusual storms and reduced availability of nutrients associated with major El Nino oceanographic events, were responsible for reduced harvests in certain years (1957, 1959, 1983).

Table 1. Estimated southern California kelp areas in hectares (modified from Neushul, 1981a) and annual *Macrocystis* harvest from California waters in wet metric tons (Pinkas, 1977 through 1976; subsequent figures courtesy of K. Wilson and E. Smith, California fish and Game). The area of kelp in hectares is underscored at the head of the respective list of paired years and tonnages.

Total area		Total area		Total area	
Year	Tonnage	Year	Tonnage	Year	Tonnage
	13,171	1949	75,610		<u>4,753</u>
1911		1950	91,265	1967	119,290
1916	122,050	1951	104,109	1968	122,337
1917	358,314	1952	99,935	1969	119,058
1918	358,425	1953	114,894	1970	115,248
1919	15,125	1954	96,358		
1920	23,101				<u>5,971</u>
1931	236		<u>7,110</u>	1971	141,121
1932	9,358	1955	112,548	1972	147,428
1933	19,614	1956	106,881	1973	138,872
1934	14,406	1957	84,961	1974	154,386
1935	27,762	1958	103,475	1975	155,671
1936	44,740			1976	143,672
1937	39,874		<u>6,510</u>		
1938	43,270	1959	81,283		<u>6,202</u>
1939	51,470	1960	109,	1977	143,918
1940	53,528	1961	117,259	1978	186,270
1941	50,546	1962	127,217		
1942	56,153				<u>4765</u>
1943	43,507		<u>2,818</u>	1979	188,464
1944	48,108	1963	109,798	1980	162,693
1945	53,688	1964	115,443	1981	83,822
1946	82,616	1965	122,587	1982	95,326
1947	67,347	1966	108,376	1983	5,334
1948	71,342			1984	47,320

5.2 Principal factors affecting productivity

5.2.1 Physical factors

As noted above, wave exposure is reckoned as the primary environmental factor causing losses of kelp tissues and entire plants on exposed coasts (Rosenthal *et al.*, 1974; Gerard 1976). High water temperatures and lack of nutrients lead to losses of canopy tissues during warm summers (Jackson, 1977; North, 1983). A combination of record storms, warm water and low nutrients associated with an El Nino event in 1983, destroyed more than 90 percent of the kelp canopies in southern California, leading to the poorest annual harvest yield in recent times (see **Table 1**). Factors affecting recruitment of juvenile plants have a delayed influence on the adult population. Increased sedimentation from construction projects and discharged waste materials may have temporary or long-lasting effects.

Figure 2. Historical charts showing changes in canopy size for a large *Macrocystis pyrifera* bed off Point Loma California. Kelp is shown as black. Kelp restoration activities (primarily sea urchin control) began near the south end of Point Loma in 1963.

Figure 3. Historical charts showing changes in canopy size for beds of *Macrocystis angustifolia* (before 1974) and *M. pyrifera* (after 1974) off the Palos Verdes coastline in southern California. Kelp is shown as black. Kelp disappeared completely from the area in 1968 but was restored by urchin control and kelp transplantation efforts.

5.2.2 Biological factors

Grazing by sea urchins has led to declines and, in some cases, complete disappearances of major kelp areas in California and Baja California during the period 1940 to 1965 (Leighton *et al.*, 1965). Probably declines in major urchin predators such as sea otters, lobster, and certain fishes eliminated some of the natural factors controlling size of urchin populations (Tegner and Levin, 1983; Leighton *et al.* 1.c.). Programs of artificial control of urchins were associated with reappearances of *Macrocystis* beds at Pt. Loma and Palos Verdes, California, in the 1960's and 1970's respectively (**Figures 2 and 3**).

Grazing by herbivorous fishes does not usually reach destructive proportions in large established *Macrocystis* beds. Fish grazing may cause significant difficulties, as noted above, when attempts are made to introduce transplants of adult or juvenile kelp into areas where no *Macrocystis* exists. Presence of the plants probably attracts herbivorous fishes to the transplant sites and a large amount of grazing is focused on their rather small biomass.

Competitive algae can be a serious impediment to either natural recruitment of juvenile *Macrocystis* or to reestablishment of this species in an area by transplanting or "seeding" with microscopic stages. Removal of competitors from 1 to 3 m of bottom provides an opportunity for a small transplant to grow to a size where it can overcome competitive effects.

Several kelp diseases have been described (North, 1979b; Scotten, 1971). Most are associated with summertime deterioration of canopies during periods of elevated water temperature and low nutrients. Some localized rotting of stipes near plant bases on

Macrocystis holdfasts has been observed in areas of organic enrichment by sewage (North, 1979b).

5.2.3 Possibilities of control measures

Several techniques have been utilized in southern California for controlling overgrazing by sea urchins. Urchins have been destroyed by quicklime, crushed with hammers, and sucked up by dredging machinery (Wilson & McPeak, 1983; Wilson & North, 1983; Leighton *et al.*, 1965). Attempts to control herbivorous fishes by gill-netting, spearing and dispersals of poisons were only partially successful.

As noted above, removal by cutting or plucking of competitive algae from the immediate vicinity of a young Macrocystis (either transplanted or naturally attached) will assist survival until the young plant attains a height of 2 to 3m and begins to eliminate surrounding competitors by shading them. There are no known methods for controlling diseases of Macrocystis plants. Since most of the diseases occur during summer when surface waters are warm and depleted of nutrients, fertilizing might be expected to improve plant healthiness. Research is needed, however, because excessive fertilizing might benefit the disease organisms.

5.3 Genetic studies and breeding studies

Neushul and coworkers have undertaken breeding studies involving crosses between the three species of Macrocystis, between Macrocystis and closely related kelps such as Pelagophycus and development of clones derived from highly productive adult Macrocystis raised on their test farm at Ellwood, California (Sanbonsuga and Neushul, 1978a, 1978b; Neushul, 1981b; Neushul and Harger, 1985). Work is still in progress and it is not yet established whether enhanced productivity will be possible. Stock cultures of gametophytes of the various species and strains are maintained by this research group. One possibility for enhancing Macrocystis productivity would involve development of a strain which would have tolerance for elevated water temperatures and could be cultivated at low latitudes where wintertime reduction in available sunlight is not as pronounced as in California. Clendenning (1971) reported that Macrocystis from Turtle Bay, Baja California (near the southern limit of the *Macrocystis* range in North America) exhibited a photosynthetic capacity per unit area that was about 50% higher than the capacity found for plants from California. The temperature optimum for photosynthesis from Turtle Bay kelp was between 25°C and 30°C, about 5°C higher than for kelp from southern California.

Another interesting possibility for improving harvest yield might be utilization of a Macrocystis strain occurring only in Paterson/inlet, New Zealand (Gerard and Kirkman, 1984). This plant can generate several apical meristems on a single frond through production of side branchings along the frond length. Use of this characteristic in a harvested crop might substantially reduce the period required for canopy regeneration after cutting.

5.4 Relative contributions from vegetative & sexual reproduction to the harvest

Both sexual and vegetative reproduction are important to maintaining the Macrocystis crops available for harvest. Effects of sexual reproduction, however, are indirect and serve only to replace adult plants lost to storm or other adversities. Vegetative growth by

young fronds with intact apical blades is entirely responsible for regenerating kelp canopies following harvesting.

5.5 Environmental enhancement

Macrocystis populations in southern California are sometimes adversely affected by nutrient starvation. As noted above, depletion of nutrients in surface waters above the thermocline is probably a major factor causing canopy deterioration during summer. Nutrient depletion probably also occurs in association with major El Nino events (a complex sequence of oceanographic events beginning near the equator, spreading to higher latitudes, sometimes lasting from a few months to 2-3 years; Dayton and Tegner, 1984; Gerard, 1984a). Fertilizing *Macrocystis* canopies prior to deterioration might serve to preserve for harvesting much organic material that might otherwise be lost. Neushul and Harger (1983) observed greater yield during summer from fertilized than from unfertilized plots on their test farm. Fertilizing a kelp bed by crop dusting techniques using helicopters was described by North *et al.*, (1982).

Fertilizing might also be helpful if a *Macrocystis* crop growing under low-nutrient conditions were to be utilized in preparation of animal feeds. Contents of N and C can be raised substantially in a few days by addition of nitrogen as nitrate or as ammonium to the medium (North, 1978). P-contents of kelp tissues also rise as external concentration of inorganic phosphate is raised (Manley *et al.*, 1982b).

Sizes of *Macrocystis* beds can be increased by provision of artificial substrate for transplanting (**Figure 4**). All that is required is an anchoring mechanism. The adult plant does not need to be attached to the bottom (Wilson and North, 1983). Small plants can, however, easily be affixed to solid substrates if so desired (McPeak *et al.*, 1973).

6. METHOD OF HARVESTING AND HARVESTING CYCLE

[6.1 Annual operations](#)

[6.2 Manpower productivity](#)

[6.3 Alternate employment](#)

6.1 Annual operations

Given appropriate environmental conditions favoring *Macrocystis* growth, canopies can be available throughout the year. Kelp beds occur in a variety of environments in southern California, so at least some canopy is available somewhere for harvesting at all times of the year. Winter storms and warm waters during summer can create seasonal scarcities of plant material available for harvesting.

6.2 Manpower productivity

Operational and economic statistics from the California kelp harvesting industry are closely guarded and not available to the public. It is believed, however, that manpower requirement for operation of even large harvesting vessels is modest, amounting to four to six individuals. Manpower for unloading and processing the cut kelp is unknown and undoubtedly depends to some extent on intended uses of the kelp and the degree of

automation available in the processing factory.

Figure 4. Technique for (A) threading a rope in a *Macrocystis* holdfast preparatory to transplanting, (B) moving several kelp transplants to a new location. (Threading the Holdfast)

Figure 4. Technique for (B) moving several kelp transplants to a new location. (Transplanting Operation)

Two economic analyses of kelp harvesting operations and costs were prepared for marine biomass studies undertaken by the U.S. Naval Ocean Systems Center (NOSC) and the Gas Research Institute (GRI). The NOSC study envisioned a *Macrocystis* farm of about 40,000 ha with an annual productivity of 946 wet metric tons per ha (Integrated Sciences, Inc., 1976). The personnel estimate specified 220 individuals required for harvesting the kelp and 403 shore-based personnel for processing the raw material into the ultimate product (methane gas). These figures indicate that 172,000 t/yr would be produced per individual involved in the harvesting. If we assume that the annual California kelp harvest of about 180,000 t (**Table 1**) can be obtained by three vessels or 15 individuals, a figure of 12,000 t/man-yr is calculated, considerably below the productivity estimate from NOSC. The discrepancy arises partially from the circumstance that productivity of the wild crop in California at about 35 t/ha/yr, is much lower than that assumed in the NOSC study. The comparison is much closer if it is drawn on the basis of areas harvested, 182 ha/yr/individual for the NOSC study and 333 ha/yr/individual for the California kelp industry.

The GRI-sponsored study envisioned a *Macrocystis* farm of 16/300 ha yielding an estimated 674 wet t/ha/yr (R.M. Parsons Co., 1983). Their scheme depicted two individuals working 12 hr. shifts in operating the harvesting vessel. This is equivalent to six individuals per day working 8 hr. shifts, giving 1,830,000 t/man-yr. or 2,700 ha/man-yr. These higher figures assume a much greater degree of automation in the harvesting machinery than actually exists in the California kelp harvesting industry.

6.3 Alternate employment

There is no need for alternate employment in the California kelp harvesting industry, as far as is known.

7. EQUIPMENT USED FOR HARVESTING

[7.1 Wild resources](#)

[7.2 *Macrocystis* culturing](#)

[7.3 Sources of credit and insurance](#)

7.1 Wild resources

As stated above, detailed operating characteristics and economics of harvesting are not available to the public from the California kelp harvesting industry. The harvesting barges carry the cutting apparatus at the sterns of the vessels and proceed in reverse when operating in a kelp canopy. Cutting is accomplished by reciprocating blades attached to a horizontal bar that is lowered to a fixed depth of 1.2 m below the waterline.

Cut ends of the kelp fronds are carried against a conveyor belt just behind the cutting bar. The cut kelp is carried up on the belt over the gunwale, and deposited in an open hold that extends for most of the barge's length. A mechanically driven steel claw or grapnel is used to distribute the cut kelp evenly throughout the hold. The bridge crew's quarters and other necessary spaces are on the bow, forward of the hold. Typical harvesting vessels may load several hundred wet tons of kelp per trip. Cameron (1915) described a harvesting vessel of his day in considerable detail (**Figures 5 to 7**). The essential features of a kelp harvester, as described above, were fully developed even then.

Harger (1979) studied harvesting activity in Goleta Bay for four years, seeking correlations between amount of kelp obtained and ten environmental variables which he monitored. The most important were surface irradiance, water temperature, swell height, and nitrate concentrations (surface & bottom). He developed a model wherein environmental conditions accounted for 46% of the variability in harvest. Harger found that the harvest rate could vary from about 13 to 110 wet t/hr/with a majority of values lying between 40 and 80 wet t/hr.

Gerard and North (1984) calculated that their simulated harvesting operations removed about 50% of the plant biomass. *Macrocystis* beds are generally harvested only when dense canopies justify such activity, otherwise a bed is left unharvested until an adequate canopy develops. A bed may thus be left uncut for several years. Productive beds, however, will usually support three harvests per year (C. Martin, personal communication). Brandt (1923) reported that some beds were harvested continuously for five or more months per year, but yields fell significantly following such intense usage.

Capital and maintenance costs of harvesting vessels, docking and unloading facilities, etc., for the California industry are not public information. The NOSC economic analysis budgeted \$133,800,000 for start-up harvesting costs with annual operating expenses of \$17,400,000 (Integrated Sciences, Inc., 1976). Ralph M. Parsons Co. (1983) listed the cost of a 12x27 m harvesting vessel as \$1,677,000 plus \$805,000 for a 12x45 m barge to receive the cut kelp. Their operating expenses were \$4,830,200 which included all activities, not just harvesting. Thus the NOSC estimates seem extremely high, but the number of harvesting vessels involved was not specified. Also, they planned to operate a kelp farm 160 km offshore, which would increase operating costs considerably since the processing plant was onshore.

The manpower requirement, as stated above appears to be in the range of 200 to 300 ha of kelp bed that can be harvested per man-year using mechanical harvesters as they presently exist. Automation might be able to increase these values.

7.2 *Macrocystis* culturing

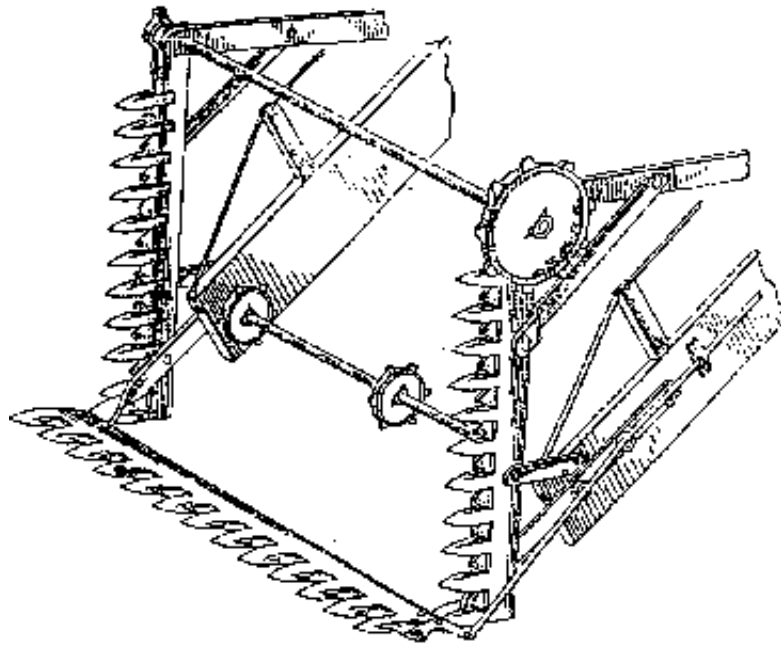
Presently no commercial-scale culturing of *Macrocystis* is conducted in California. North (1976, 1972) described experimental facilities for mass-culturing *Macrocystis* gametophytes and embryonic sporophytes for dispersal in kelp areas to enhance the recruitment processes. Wilson and North (1983) reviewed techniques for transplanting young and adult *Macrocystis* plants to initiate new populations in depleted areas. McPeak (1975) described a four-year transplantation effort that involved installation of about 28,000 young *Macrocystis* at the north end of the La Jolla kelp bed.

Figure 5. Elevation view, showing bow section of a *Macrocystis* harvesting vessel as used in the Palos Verdes area of southern California during World War I (from

[Cameron, 1915\). Kelp cutting apparatus and conveyor belt arrangement are similar in principle to equipment in present-day usage.](#)

[Figure 6. Plan view of *Macrocystis* harvesting vessel as shown in Figure 4 \(from Cameron, 1915\). The cut kelp was loaded onto a barge on the starboard side, while propulsion through the water was supplied from a separate launch. Modern harvesters are larger vessels, loading the kelp onboard in a bin aft from the front conveyor, and are self-propelled.](#)

Figure 7. Perspective view showing details of the cutting apparatus located at the bow of the harvesting vessel of Figure 4 (from Cameron, 1915). Modern harvesters have the cutting and loading equipment at the vessel's stern and the ship moves in reverse while cutting kelp. The prow is V-shaped/providing for maximum efficiency in travelling to and from port.



7.3 Sources of credit and insurance

Industrial loans from commercial banking sources presumably have been available and used by kelp harvesting concerns in California. The major remaining such organization at the present time is Kelco Company of San Diego, a division of Merck & Co., Inc. During periods of crop shortages in California waters, Kelco imports raw material from foreign sources. Dependence on imports, rather than insurance, appears to be the preferred method for sustaining operations in case of natural disasters.

8. PROTECTION AND MANAGEMENT OF THE RESOURCE

[8.1 Management of *Macrocystis* resources](#)

[8.2 Regulation](#)

8.1 Management of *Macrocystis* resources

Commercial harvesting of *Macrocystis* in the USA does not occur north of Monterey Bay in central California. *Macrocystis* is also harvested in northern Baja California, Mexico, and transported to the processing plant of Kelco.

The kelp harvesting concerns in California have all operated as private entrepreneurs, independent of linkages to governmental agencies. Interest in kelp resources led to formation of several kelp harvesting enterprises in the early 1900's (Scofield, 1959; P. Neushul in Neushul and Harger, 1985). Some of the organizations utilized harvesting techniques that may have damaged the resource and contributed unnecessarily to beach litter. Since all the kelp beds lie within three miles of the shoreline, their ownership resides in the State of California. The responsible State agency for management of living marine resources is the Department of Fish and Game with policy direction from the Fish and Game Commission. Concern about welfare of the kelp resource around 1916 led to a joint effort by the Commission, conservationists, and the larger kelp harvesting concerns, to control harvesting methods and practices. Results were the first set of regulations covering kelp harvesting activities, enacted July 26, 1917. The regulations have been expanded and amended since then on several occasions.

Regulations now divide the California kelp beds into two categories: open beds (available for harvest by anyone) and closed beds (restricted for harvesting by a lessee only). A third of the beds are required to be held in the open category. Harvesters must purchase a license and pay a privilege fee of 10 cents for each wet ton harvested and a second assessment of \$100 for each square mile of kelp bed harvested. Since 1950, leases are granted by negotiation (before, leases were awarded on the basis of sealed bids). Leases are granted for periods of fifteen years or less. Lease rights confer only harvesting privileges and have no control over fishing or navigational activities.

8.2 Regulation

In 1956, the Fish and Game Commission decreed that depth of kelp cutting should be no greater than four feet (1.22 m) below the surface and all material cut must be fully utilized (i.e. no escapement of cut fronds is allowed. This latter is to prevent increasing beach litter as a result of harvesting). Violations may result in fines or suspensions of the license to harvest. Enforcement is accomplished by wardens operating from patrol vessels.

9. UTILIZATION

[9.1 Chemical and nutritional content](#)

[9.2 Human Food](#)

[9.3 Animal fodder and feed additives](#)

[9.4 Manure and fertilizer](#)

[9.5 Industrial products and processes](#)

9.1 Chemical and nutritional content

Paine and Vadas (1969) reported the caloric content of *M. pyrifera* as 2.85 kcal/g dry wt, or 4.3 kcal/g dry minus ash. Similar values were obtained for a closely related kelp, *Nereocystis leutkeana*. Because of the high ash content, the above caloric values were among the lowest of seventeen seaweed species assayed by these workers. With ash

corrections most species were very similar. Effects of various light intensities and nitrate concentrations on caloric value of cultured juvenile *Macrocystis* sporophytes were determined by Shivji (1985).

Extensive analyses of organic and salt composition of the Pacific coast kelps, including *Macrocystis*, were reported by Balch and by Turrentine from work in the early 1900's (cf. Cameron, 1912, 1915). The most recent general studies were conducted in connection with the Marine Biomass Program (**Tables 2 and 3**). As a result of these studies, it became known that concentrations of some of the organic and inorganic constituents vary substantially according to environmental conditions and the recent history of the tissue under analysis. Thus the ranges of mannitol contents of *M. pyrifera* fronds increased from 5 to 15% dry wt to 14 to 33% dry wt in a plant transferred for three weeks into a low-nutrient environment (Gerardy 1982b). Tissue contents of nitrogen and phosphorus in young *M. pyrifera* sporophytes reflected concentrations of the inorganic salts of these elements in the culturing media (P.A. Wheeler and North, 1980; Manley et al., 1982). Seasonal variations in dry weight, ash, N, P, S, Cu, Zn, Fe, ether solubles, algin, mannitol, laminarin, and fucoidin were studied for 2 years in *M. integrifolia* by Wort (1955).

Table 2. Gross composition of *Macrocystis*, combining data from Point Loma and San Clemente Island (Lindner et al. 1977) and from Monterey Bay, Santa Cruz, and Soquel Point (Hart et al., 1976). Standard deviations are shown for ash, solids, and alginic acid for all samples combined.

Substance and % Wet or Dry Wt.	Percent Composition		
	All Samples	So. Calif.	Cen. Calif.
% WET WT			
Water	88.2		
Solids	11.8 ± 1.0 ^a	12.2	11.3
% DRY (550°C)			
Ash	38.96 ± 6.24 ^b	37.29	40.49
KCl	29		
NaCl	7		
Na ₂ SO ₄	4		
Other	4		
Volatiles	66-53		
Mannitol	6-22		
Laminarin		1-2	
Fucoidan		0.5-2	
Alginic Acid	15.71 ± 2.56 ^c	18.6	11.4
Cellulosics	3-8		
Protein	5-14		
Fat	0.5-2		

^a 42 samples

^b 44 samples

°35 samples

Table 3. Concentrations of selected elements in wet *Macrocystis* tissues (modified from North, 1980) and in seawater according to Brewer (1975). The ratio of these two sets of numbers was taken as the concentration factor for each element. The tissue and seawater values are in micromoles per given unit.

Element	Concentration		
	Tissue per gm wet wt.	Seawater per liter	Concentration Factor
Phosphorus	11.6	0.2	58,000
Beryllium	0.025	0.00063	40,000
Nitrogen	10.5	0.5	21,000
Antimony	0.0125	0.002	6,000
Iron	0.175	0.05	5,000
Cobalt	0.00375	0.0008	4,700
Arsenic	0.175	0.05	3,500
Manganese	0.0125	0.0036	3,500
Aluminum	0.120	0.074	1,600
Copper	0.01	0.008	1,300
Iodine	0.588	0.5	1,200
Zinc	0.0688	0.076	900
Vanadium	0.0375	0.05	750
Nickel	0.00625	0.028	220
Molybdenm	0.00125	0.01	130
Boron	3.13	410	8
Lithium	0.19	26	7
Strontium	0.475	91	5
Silicon	0.35	71	5
Potassium	15	10,200	1.5
Uncertain			
Selenium	0.00125	0.0025	500
Chromium	0.00125	0.0057	220

Bromine, Rubidium, and Fluorine are not concentrated.

9.2 Human Food

Usage of *Macrocystis* tissues as human food is minor in North America. *Macrocystis* and *Nereocystis* slices are occasionally prepared as pickles by individuals, but there is no known commercial product of this kind. Dried powdered kelp (probably *M. pyrifera*) was formerly marketed by Philip R. Park, Inc. A fraction of the product entered specialty items in health food stores (P. Neushul *in* Neushul and Harger, 1985).

9.3 Animal fodder and feed additives

Dried kelp (probably *Macrocystis pyrifera*) was formerly produced by Philip R. Park Inc. and Kelp Organic Products Co. of Los Angeles. Most of the material was incorporated into animal feeds. Neither of these concerns exist at this time (P. Neushul, 1.c.). The value of this *Macrocystis* as fodder might be enhanced by fertilizing shortly before harvesting to increase nitrogen and phosphorus contents of the tissues via (cf. Mateus et al., 1976) its amino acid composition.

9.4 Manure and fertilizer

No present usage of *Macrocystis* as a manure exists in California. A large and successful effort was mounted during the first World War to secure kelp as a source of potash for manufacture of fertilizer and explosives (Cameron, 1915; Scofield, 1959; P. Neushul, 1.c.). Presently, all *Macrocystis* harvested in California and Baja California is processed for alginate production and the resource is fully utilized. An economic study of the feasibility of using *Macrocystis* for methane production proposed dispersal of sludge remaining from digestion of the kelp, into their kelp farm as a fertilizer (Ralph M. Parsons, 1983).

9.5 Industrial products and processes

As noted above, alginates are the only products currently manufactured from harvested *Macrocystis* in California. Various compounds of the complex polysaccharides, alginic acid, serve as stabilizers, thickeners, emulsifiers, water-holders, and film-forming or suspending agents. Most of the usages are in foods, pharmaceuticals, beverages, dairy products, and in the manufacture of latex, textiles, as well as paper and cardboard, plus other applications (Kelco, 1961; P. Neushul, 1.c.). Two methods for alginate manufacture are the Le Gloahec-Herter and Green's cold processes, both described with flow charts by Chapman (1970). Salts are first leached from cut or slurried kelp in weak acid. The alginate is then solubilized or induced to form a gel by soaking the kelp in a solution of sodium carbonate or other weak alkali. Reduction of pH by dilution with water or addition of acid causes precipitation and the recovered material can be sold as crude alginate or refined further by repeated solubilization and precipitation (Tseng, 1945).

In the past, a number of other industrial products have been derived from *Macrocystis* in California, particularly during the period of intense utilization during World War I. These included potash and acetone for explosives manufacturing (potash was also employed in manufacture of matches), iodine, and char (P. Neushul, 1.c.). The early kelp industry was unable to compete with German sources of potash which once more became available to American industry after termination of the war in 1919. The California kelp industry collapsed for about a decade until interests in alginate production and the uses of kelp in animal feeds revived harvesting activities in 1927-28.

A possible future usage of *Macrocystis* is as alternative sources of energy in the form of gaseous or liquid fuels. The feasibility of liquid fuel production has already been demonstrated on a commercial scale by production of acetone in World War I by Hercules Powder Company (P. Neushul, 1.c.). Extensive investigations of the potential for methane production from kelp were recently completed (Tompkins and Bryce, 1984). This study, the Marine Biomass Program, also yielded considerable information relating to *Macrocystis* biology, and many of the resulting publications were cited above. We also referred to two of the economic analyses that were performed (Integrated Sciences, Inc., 1976; Ralph M. Parsons, 1983). *Macrocystis* tissue proved to be an excellent substrate for methane production by bacterial digestion processes. Yields of 4.5 SCF/1bVS (0.28

liters of combustible gas per gram of volatile solids) were reported by Chenoweth et al. (1978). Maximal methane yields were recovered from those kelp samples highest in mannitol.

In closing, we should mention that the *Macrocystis* forests in nature provide an extensive habitat for fishes and shellfish utilized by humans (North and Hubbs, 1968; Leighton, 1971). As such, *Macrocystis* and other large kelps are an important sustaining element in recreational and commercial fisheries, both of which are well-established activities in California.

10. REFERENCES

- Anderson, L.M., 1984 Iron reduction by juvenile *Macrocystis pyrifera* (L.) C. Agardh. *Hydrobiologia*, 116/117:493-97
- Anderson, E.K., and W.J. North, 1966 In situ studies of spore production and dispersal in the giant kelp, *Macrocystis*. *Proc.Int.Seaweed Symp.*, 5:73-86
- Arnold, K.E. and S.L. Manley, 1985 Carbon allocation in *Macrocystis pyrifera* (Phaeophyta): intrinsic variability in photosynthesis and respiration. *J. Phycol.*, 21:154-67
- Bernstein, B., 1977 Selective pressures and coevolution in a kelp canopy community in southern California. Ph.D. Thesis. University of California, San Diego, 135 p.
- Bernstein, B., and N. Jung, 1979 Selective pressures and coevolution in a kelp canopy community in southern California. *Ecol.Monogr.*, 49:335-55
- Brandt, R.P., 1923 Potash from kelp: early development and growth of the giant kelp, *Macrocystis pyrifera*. *Bull.U.S. Dept-Agric.*, (1191):40 p.
- Brewer, G., 1975 Minor elements in seawater. In *Chemical oceanography*, edited by J.P. Riley and G. Skirrow. London, Academic Press, Vol.1:415-96 2nd ed.
- Buggeln, R.G., D.S. Fensom, and C.J. Emerson, 1985 Translocation of ¹⁴C-photoassimilates in the blade of *Macrocystis pyrifera* (Phaeophyceae). *J. Phycol.*, 21:35-40
- Cameron, F.K., 1912 A preliminary report on the fertilizer resources of the United States (with special reference to seaweeds). Washington, D.C., U.S. Senate Doc. (100): 40-290
- Cameron, F.K., 1915 Potash from kelp. I. Pacific kelp as a source of potassium salts. *Rep.U.S.Dep.Agric.*, (100):9-32
- Chapman, V.J., 1970 Seaweeds and their uses. London, Methuen and Co. Ltd., 304 p. 2nd ed.
- Chenoweth, D.P., L. Klass, and S. Ghosh, 1978 Biomethanation of giant brown kelp *Macrocystis pyrifera*. In *Energy from biomass and wastes*. Chicago Institute of Gas Technology, pp. 229-51
- Clendenning, K.A., 1963 Photosynthesis and growth in *Macrocystis pyrifera*. *Proc.Int.Seaweed Symp.*, 4:55-65
- Clendenning, K.A., 1971 Photosynthesis and general development in *Macrocystis*. In *The*

biology of Giant Kelp beds (*Macrocystis*) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 169-90

Cole, K., 1968 Gametophytic development and fertilization in *Macrocystis integrifolia*. Can.J.Bot., 46:777-82

Dayton, P.K., B.D. Keller, and D.A. Ven Tresca, 1980 Studies of a nearshore community inhabited by sea otters. Report to U.S. Marine Mammal Commission. Washington D.C., Rep.No.(MMC-78/14):91 p. Available from National Technical Information Service, Springfield, Virginia, PB81-109860)

Dayton, P.K., and M.J. Tegner, 1984 Catastrophic storms and patch stability in a southern California kelp community. Science, Mash., 224:283-5

Dayton, P.K., et al., 1984 Patch dynamics and stability of some California kelp communities. Eco L. Monogr., 54:253-89

Dean, T.A., and L.E. Deysher, 1983 The effects of suspended solids and thermal discharges on kelp. In The effects of waste disposal on kelp communities edited by W. Bascom, Long Beach, Southern California Coastal Water Research Project, pp. 114-35

Dean, T.A., S.C. Schroeter, and J. Dixon, 1984 Effects of grazing by two species of sea urchins (*Strongylocentrotus franciscanus* and *Lytechinus anamesus*) on recruitment and survival of two species of kelp (*Macrocystis pyrifera* and *Pterygophora californica*). Mar.Biol., 78:301-13

Devinny, J.S., and L.A. Volse, 1978 Effects of sediments on the development of *Macrocystis pyrifera* gametophytes. Mar.Biol., 48:343-8

Deysher, L.E., 1984 Recruitment processes in benthic marine algae. Ph.D. Thesis, University of California, San Diego, 324 p.

Dixon, J., S.C. Schroeter, and J. Kastendiek, 1981 Effects of the encrusting bryozoan, *Membranipora membranacea*, on the loss of blades and fronds by the giant kelp, *Macrocystis pyrifera* (Laminariales). J.Phycol., 17:341-5

Druehl, L.D., 1979 The distribution of *Macrocystis integrifolia* in British Columbia as related to environmental parameters. Can.J.Bot., 56:69-59

Ebeling, A.W., D.R. Lauer, and R.J. Rowley, 1984 Severe storm disturbances and reversal of community structure in a southern California kelp forest. Mar.Biol., 75:1-8

Estes, J.A., and J.F. Palmisano, 1974 Sea otters: their role in structuring nearshore communities. Science, Wash., 185:1058-60

Fain, S.R., and S.N. Murray, 1982 Effects of Light and temperature on net photosynthesis and dark respiration of gametophytes and embryonic sporophytes of *Macrocystis pyrifera*. J.Phycol., 18:92-8

Foster, M.S., 1975 Algal succession in a *Macrocystis pyrifera* forest. Mar.Biol., 32:313-29

Foster, M.S., 1975a Regulation of algal community development in a *Macrocystis pyrifera* forest. Mar.Biol., 32:331-42

Gerard, V.A., 1976 Some aspects of material, dynamics and energy flow in a kelp forest in Monterey Bay, California. Ph.D. Thesis, University of California Santa Cruz, 173 p.

Gerard, V.A., 1982 In situ rates of nitrate uptake by giant kelp, *Macrocystis pyrifera* (L.) C. Agardh: tissue differences, environmental effects, and predictions of nitrogen-limited growth. J.Exp.Mar.Biol.Ecol., 62:211-24

Gerard, V.A., 1982a Growth and utilization of internal nitrogen reserves by the giant kelp *Macrocystis pyrifera* in a Low-nitrogen environment. Mar. Biol., 66:27-35

Gerard, V.A., 1982b In situ water motion and nutrient uptake by the giant kelp *Macrocystis pyrifera*. Mar.Biol., 69:51-4

Gerard, V.A., 1984 Physiological effects of EL Nino on giant kelp in southern California. Mar.Biol. Lett., 5:317-22

Gerard, V.A., 1984a The Light environment in a giant kelp forest: influence of *Macrocystis pyrifera* on spatial and temporal variability. Mar.Biol., 84:189-95

Gerard, V.A., and H. Kirkman, 1984 Ecological observations on a branched, Loose-Lying form of *Macrocystis pyrifera* (L.) C. Agardh in New Zealand. Bot.Mar., 27:105-9

Gerard, V.A., and J. North, 1984 Measuring growth, production, and yield of the giant kelp, *Macrocystis pyrifera*. Hydrobiologia, 116/117:321-4

Grua, P., 1964 Premières données sur le biomasses de herbier a *Macrocystis pyrifera* de la baie du Morbihan (Archi-pel Kerguelen). Terre Vie, 2:215-20

Haines, K.C., and P.A. wheeler, 1978 Ammonium and nitrate uptake by the marine macrophytes *Hypnea musciformis* (Rhodophyta) and *Macrocystis pyrifera* (Phaeophyta). J.Phycol., 14: 319-24

Harger, B.W.W., 1979 Coastal oceanography and hard substrate ecology in a California kelp bed. Ph.D. Thesis, University of California, Santa Barbara, 437 p.

Harger, B. W., 1983 A historical overview of kelp in southern California. In The effects of waste disposal on kelp communities, edited by W. Bascom. Long Beach Southern California Coastal water Research Project, pp. 70-83

Harris, G., et al., 1984 Community recovery after storm damage: a case of facilitation in primary succession. Science, Wash., 224:1336-8

Hart, M.R., et al., 1976 Ocean food and energy farm kelp pretreatment and separation processes. Berkeley, California, Western Regional Research Center, U.S. Department of Agriculture Report for Agreement No. 12-14-5001-6402:125 p.

Integrated Sciences Corp., 1976 Systems analysis of the ocean food and energy farm (DFEF). Vol. 1, overall economic analysis of the base line design. Santa Monica, California, Integrated Sciences Corporation, 69 p.

Jackson, G.A., 1976 Nutrients and productivity of the giant kelp, *Macrocystis pyrifera* in the nearshore. Ph.D. Thesis, California Institute of Technology, Pasadena, 134 p.

Jackson, G.A., 1977 Nutrients and production of giant kelp, *Macrocystis pyrifera*, off

southern California. Limnol.Oceanogr., 22:979-95

Jackson, G.A., D.E. James, and W.J. North submitted, Morphological relationships among fronds of giant kelp, Macrocystis pyrifera, off La Jolla, California, Mar.Ecol.(Prog. Ser.), 26(3):261-70

Jackson, G.A., and C.D. Winant, 1983 Effect of a kelp forest on coastal currents. Cont.Shelf Res., 2:75-80

Kain, J.M., 1982 Morphology and growth of the giant kelp Macrocystis pyrifera in New Zealand and California. Mar.Biol., 67:143-57

Kelco Company, 1961 Algin. San Diego, Kelco Co., 12 p.

Kimura, S., 1980 The effects of harvesting Macrocystis pyrifera on understory algae in Carmel Bay, California. M.S. Thesis, California State University, Fresno, 108 p.

Kirkwood, P.D., 1977 Seasonal patterns in the growth of the giant kelp, Macrocystis pyrifera. Ph.D. Thesis, California Institute of Technology, Pasadena, 139 p.

Kuwabara, J.S., 1982 Micronutrients and kelp cultures: evidence for cobalt and manganese deficiency in southern California deep seawater. Science, wash., 216:1219-21

Kuwabara, J.S., and W.J. North, 1980 Culturing microscopic stages of Macrocystis pyrifera (Phaeophyta) in Aquil, a chemically defined medium. J.Phycol., 16:546-9

Leighton, D.L., 1971 Grazing activities of benthic invertebrates in kelp beds. In The biology of giant kelp beds (*Macrocystis*) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 421-53

Leighton, D.L., G. Jones and J. North, 1966 Ecological relationships between giant kelp and sea urchins in southern California. Proc.Int.Seaweed Symp., 5:141-53

Lindner, E., C.A. Dooley, and R.H. Wade, 1977 Chemical variation of chemical constituents in Macrocystis pyrifera. Final report. San Diego, U.S. Naval Ocean Systems Center, 39 p.

Liu, Tianjing, et al., 1984 Studies on the artificial cultivation and propagation of giant kelp (Macrocystis pyrifera). Hydrobiologia, 116/117:259-62

Lobban, C.S., 1978 Translocation of ¹⁴C in Macrocystis pyrifera (giant kelp). Plant Physiol., 61:585-9

Lobban, C.S., 1978 Translocation of ¹⁴C in Macrocystis integrifolia (Phaeophyceae). J.Phycol., 14:178-82

Luning, K., and M. Neushul, 1978 Light and temperature demands for growth and reproduction of Laminarian gametophytes in southern and central California. Mar. Biol., 45:297-309

Manley, S.L., 1981 Iron uptake and translocation by Macrocystis pyrifera. Plant Physiol., 68:914-8

- Manley, S.L., 1984 Micronutrient uptake and translocation by Macrocystis pyrifera (Phaeophyta). J. Phycol., 20:199-201
- Manley, S.L., Phosphate uptake by blades of Macrocystis pyrifera (Phaeophyta). Sot. Mar. (in press)
- Manley, S.L., and W.J. North, 1981 micronutrient uptake by juvenile Macrocystis pyrifera (L.) C. Agardh. Proc. Int. Seaweed Symp., 10:467-72
- Manley, S.L., and W.J. North, 1984 Phosphorus and the growth of juvenile Macrocystis pyrifera (Phaeophyta) sporophytes. J. Phycol., 20:389-93
- Manley, S.L. et al., 1981 Environmental considerations involved in 1982 farming Macrocystis. In Proceedings of the International Gas Research Conference, Rockville, Maryland, Government Institutions Inc., pp. 693-8
- Mateus, H., J.M. Regenstein, and R.C. Baker, 1976 The amino acid composition of the marine brown alga Macrocystis pyrifera from Baja California. Mexico. Bot. Mar., 19:155-9
- McPeak, H., 1975 Observations and transplantation studies at Point Loma and La Jolla. Annu. Rep. Kelp Habitat Improv. Proj. Calif. Inst. Technol. Pasadena, (1974/5):80-91
- McPeak, R.H., H.C. Fastenau, and D. Bishop, 1973 observations and transplantation studies at La Jolla and Point Loma. Annu. Rep. Kelp Habitat Improv. Proj. Calif. Inst. Technol., Pasadena, (1972/3): 57-73
- Neushul, M., 1959 Studies on the growth and reproduction of the giant kelp, *Macrocystis*. Ph.D. Thesis, University of California, Los Angeles, 134 p.
- Neushul, M., 1971 The species of Macrocystis with particular reference to those of North and south America. In The biology of Giant Kelp beds (Macrocystis) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 211-22
- Neushul, M., 1981 Historical review of kelp beds in the southern California Bight. Goleta, California, Neushul Mariculture Inc., 74 p.
- Neushul, M., 1981 The domestication and cultivation of Californian amacroalgae. Proc. Int. Seaweed Symp., 10:71-96
- Neushul, M. and B.W.W. Harger, 1983 Kelp biomass coastal test farm program. Annual report for 1982. Goleta, California, Neushul Mariculture Inc., 25 p. + 75 p. Appendices.
- Neushul, M. and B.W.W. Harger, 1985 Kelp biomass production yield, genetics, and planting technology. Annual report Jan. 1983-Aug. 1984. Goleta, California, Neushul Mariculture Inc., 85 p. + 39 p. Appendices
- Neushul, M., B.W.W. Harger, and J.W. Woessner, 1982 Laboratory and nearshore field studies of the giant California kelp as an energy crop plant. In Proceedings of the 1981 International Gas Research Conference, Rockville, Maryland, Government Institutions Inc., pp. 699-708
- North, W.J., 1958 Experimental ecology. Kelp Investigations Program, Quarterly progress report, July 1 - Sept. 30. La Jolla, University of California, Institute of Marine

Research, pp. 2-8

North, W.J, 1968 Effects of canopy cutting on kelp growth: comparison of experimentation with theory. In Utilization of kelp-bed resources in Southern California, edited by W.J. North and C.L. Hubbs. Fish Bull.Calif.Dep.Fish Game, (139):223-54

North, W.J, 1971 Growth of individual fronds of the mature giant kelp. Macrocystis. In The biology of Giant kelp beds (Macrocystis) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 123-68

North, W.J, 1972 Mass-cultured Macrocystis as a means of increasing kelp stands in nature. Proc.Int. Seaweed Symp., 7:394-99

North, W.J, 1976 Aquacultural techniques for creating and restoring beds of giant kelp, Macrocystis spp. J.Fish.Res.Board Can, 33(4)Pt2:1015- 23

North, W.J, 1978 Progress in studies of oceanic production of biomass. In Proceedings of the Second Annual Symposium on Fuels from biomass, edited by W.W. Shuster. Washington, D.C., U.S. Department of Energy.(CONF-7806107-P2):99-1040. Available from National Technical Information Service, Springfield, Virginia.

North, W.J, 1979 Adverse factors affecting giant kelp and associated seaweeds. Experientia, 35:445-7

North, W.J., 1980 Trace metals in giant kelp. Macrocystis. Am.J.Bot., 67:1097-101

North, W.J, 1983 Separating the effects of temperature and nutrients. In The effects of waste disposal on kelp communities, edited by W. Bascom. Long Beach, Southern California Coastal Water Research Project, pp. 243-55

North, W.J., V.A. Gerard, and J.S. Kuwabara, 1982 Farming Macrocystis at coastal and oceanic sites. In Proceedings of the Conference on Synthetic and degradative processes in marine macrophytes, edited by L.M. Srivastava. New York, de Gruyter, pp. 247-62

North, W.J., and C.L. Hubbs, (eds). 1968 Utilization of kelp-bed resources in Southern California. Fish Bull.Calif. Dep.Fish Game, (139):264 p.

North, W.J., and R.C. Zimmerman, 1984 Influences of macronutrients and water temperatures on summertime survival of Macrocystis canopies. Hydrobiologia, 116/117:419-24

Paine, R.T., and R.L. Vadas, 1969 Calorific values of benthic marine algae and their postulated relationship to invertebrate food preferences. Mar.Biol., 4:79-86

Parsons Company, Ralph M., 1983 An economic and systems assessment of the concept of nearshore kelp farming for methane production. Final report. Chicago, Gas Research Institute (GRI-82/0067):86 p + 234 p Appendices

Pinkas, L., 1977 California marine fish landings for 1975. Fish.Bull. Calif.Dep.Fish.Game, (168):5 p.

Rosenthal, J., D.Clarke, and P.K. Dayton, 1974 Observations on the ecology and natural history of a stand of giant kelp, Macrocystis pyrifera (Linnaeus) Agardh, off Del Mar,

California. Fish.Bull.NOAA/NMFS, 72:670-84

Ryther, J.H., et al., 1977 Cultivation of seaweeds as a biomass source for energy. In Proceedings Fuel from biomass Symposium. Urbana, University of Illinois, pp. 83-98

Ryther, J.H., et al., 1979 Biomass production by marine and freshwater plants. In Proceedings of the Third Annual Biomass Energy Systems Conference. Golden, Connecticut, SERI, (SERI/TP-33-285):13-23. Available from National Technical Information Service, Springfield, Virginia.

Schmitz, K., 1982 Translocation of organic compounds in Lam Laminariales. In Synthetic and degradative processes in marine macrophytes, edited by L.M. Srivastava, de Gruyter, pp. 16-179

Schmitz, K. and L.M. Srivastava, 1979 Long distance transport in Macrocystis integrifolia. 2. Tracer experiments with ¹⁴C and ³²P. Plant Physiol., 63:1003-9

Scofield, W.L., 1959 History of kelp harvesting in California. Calif. Fish Game Q., 45:135-57

Scotten, H.L., 1971 Microbiological aspects of the kelp bed environment. In The biology of Giant Kelp beds (Macrocystis) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 315-8

Shivji, M., 1985 Interactive effects of Light and nitrogen on growth and chemical composition of juvenile Macrocystis pyrifera (L.) C.Ag. (Phaeophyta) sporophytes. J. E x p. Mar.Biol-Ecol., 89(1):81-96

Smith, R.G., N. Wheeler, and L.M. Srivastava, 1983 Seasonal photo-synthetic performance of Macrocystis integrifolia (Phaeophyceae). J.Phycol., 19:352-9

Tegner, M.J., and L.A. Levin, 1983 Spiny Lobsters and sea urchins: analysis of a predator-prey interaction. J.Exp.Mar. Biol.Ecol., 73:125-50

Tompkins, A.N., and A.J. Bryce, 1984 Marine biomass program, Final report. Chicago, Gas Research Institute, 166 p.

Tseng, Cheng K., 1945 Colloids from kelp. Chem-Metall.Eng., 52:97-100

Vaughn, O.W., 1959 Carbohydrate metabolism in a marine brown alga Macrocystis pyrifera. Ph.D. Thesis, University of California, Berkeley, 65 p.

Wheeler, A., 1979 Uptake of methylamine (an ammonium analogue) by a marine macrophyte Macrocystis pyrifera (Phaeophyta). J.Phycol., 15:12-7

Wheeler, P.A., and W.J. North, 1980 Effect of nitrogen supply on nitrogen content and growth rate of juvenile Macrocystis pyrifera (Phaeophyta) sporophytes. J.Phycol., 16:577-82

Wheeler, W.N., 1978 Ecophysiological studies on the giant kelp, Macrocystis. Ph.D. Thesis, University of California, SantaBarbara, 179 p.

Wheeler, W.N, 1980 Effect of boundary Layer transport on the fixation of carbon by the giant kelp Macrocystis pyrifera. Mar.Biol., 56:103-10

Wheeler, W.N, 1980a Pigment content and photosynthetic rate of the fronds of Macrocystis pyrifera. Mar.Biol., 56:97-102

Wheeler, W.N, 1982 Nitrogen nutrition of *Macrocystis*. In Synthetic and degradative processes in marine macrophytes, edited by L.M. Srivastava. New York, de Gruyter, pp. 121-35

Wheeler, W.N., and L.M. Srivastava, 1984 Seasonal nitrate physiology of Macrocystis integrifolia Bory. J.Exp.Mar.Biol. Ecol., 76:35-50

Wilson, K.C., and R. H. McPeak, 1983 Kelp restoration. In The effects of waste disposal on kelp communities, edited by W. Bascom. Long Beach, Southern California Coastal Water Research Project, pp. 199-216

Wilson, K.C., and W.J. North, 1983 A review of kelp bed management in southern California. J.World Maricult.Soc., 14:347-59

Wing, B.L., and K.A. Clendenning, 1971 Kelp surfaces and associated invertebrates. In The biology of Giant Kelp beds (Macrocystis) in California, edited by W.J. North. Lehre, Germany, J. Cramer, pp. 319-41

Womersley, H.B.S., 1954 The species of Macrocystis with special reference to those on southern Australian coasts. Univ.Calif.Publ.Bot., 27:109-32

Wort, D.J., 1955 The seasonal variation in chemical composition of Macrocystis integrifolia and *Nereocystis leutkeana* in British Columbia coastal waters. Can.J. Bot., 33, pp. 323-40

Zimmerman, R.C., 1983 Seasonal patterns in the productivity of a giant kelp (Macrocystis pyrifera) forest: the effect of nutrient availability. Ph.D. Thesis, University of Southern California Los Angeles, 182 p.

Zimmerman, R.C., and J.N. Kremer, 1984 Episodic nutrient supply to a kelp forest ecosystem in southern California. J.Mar. Res., 42:591-604

