

Computational Skills for Biostatistics I: Lecture 5

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Theme for today: Computation time

“Programmers waste enormous amounts of time thinking about, or worrying about, the speed of noncritical parts of their programs, and these attempts at efficiency actually have a strong negative impact when debugging and maintenance are considered.”

— Donald Knuth

Theme for today: Computation time

“R was purposely designed to make data analysis and statistics easier for you to do. It was not designed to make life easier for your computer. While R is slow compared to other programming languages, for most purposes, it's fast enough.”

— (BFF) Hadley Wickham

Theme for today: Computation time

A language that was built for data analysis: R

- ▶ Of course you can do scientific computing in R

Languages that were built for scientific computing: Python, MatLab, C++

- ▶ Of course you can do data analysis in these languages

Theme for today: Computation time

However, with some understanding of how R works, and some understanding of how to use your computer effectively, you can significantly speed up runtime.

Evaluating runtime

You can grab the “system time”: how long have you been running your R process

```
proc.time()
```

```
##      user  system elapsed  
##    1.246    0.085    1.334
```

```
t1 <- proc.time()  
Sys.sleep(0.25)  
t2 <- proc.time()  
t2 - t1
```

```
##      user  system elapsed  
##    0.001    0.000    0.255
```

Evaluating runtime

You can also use `system.time()`

```
system.time(rnorm(1e5))
```

```
##      user  system elapsed  
##    0.005    0.001    0.005
```

```
system.time(rnorm(1e7))
```

```
##      user  system elapsed  
##    0.524    0.019    0.582
```

Evaluating runtime

Investigating chunks of code

```
t1 <- Sys.time()
my_inverse <- rnorm(1e6) %>% matrix(nrow = 1e3) %>% solve
t2 <- Sys.time()
t2 - t1
```

```
## Time difference of 0.9289382 secs
```


Microbenchmarking

For very small comparisons, the library `microbenchmark` is great!

```
library(microbenchmark)
x <- runif(100)
mbm <- microbenchmark(
  sqrt(x),
  x ^ 0.5
)
mbm
```

Unit: nanoseconds

##	expr	min	lq	mean	median	uq	max	neval
##	sqrt(x)	255	267.0	351.47	276.5	311.5	6184	100
##	x^0.5	1897	1913.5	1976.04	1923.5	1961.0	5354	100

Microbenchmark

```
mbm <- microbenchmark(  
  sqrt(x),  
  x ^ 0.5  
)  
autoplot(mbm)
```

Coordinate system already present. Adding new coordinate



Microbenchmarking

```
mbm
```

```
## Unit: nanoseconds
```

##	expr	min	lq	mean	median	uq	max	neval
##	sqrt(x)	249	264	344.64	271.5	292	1913	100
##	$x^{0.5}$	1890	1912	2219.53	1935.0	2007	10464	100

Which should you use?

`sqrt()` or `^0.5`: Which should you use?

```
mbm
```

```
## Unit: nanoseconds
##      expr  min   lq   mean median   uq   max neval
##  sqrt(x)  249  264  344.64  271.5  292  1913   100
##    x^0.5 1890 1912 2219.53 1935.0 2007 10464   100
```

It actually doesn't matter – a million square roots will take 0.5 or 2 seconds.

Don't agonise over microbenchmarks: no need to overoptimise

Tools for benchmarking

- ▶ `microbenchmark()`
- ▶ `Sys.time()`

Making larger gains in programming time

Start with the biggest bottleneck, work to speed it up as fast as possible; move onto the next biggest bottleneck, and so on. . .

There are easy and there are hard ways to make code run faster. Start with the easy ways!

The easiest way: run code in parallel

Tools for running code in parallel

- ▶ Writing functions: `parallel` and `snow`
- ▶ Data analysis: `multidplyr`
- ▶ Simulations: `simulator`

Quite practical example

An expensive operation to perform repeatedly is matrix inversion:
 $O(n^3)$

- I'm hoping we'll have time to come back to this notation

```
my_matrices <- replicate(n=20,  
                          rnorm(1e2) %>% matrix(nrow = 1e1),  
                          simplify=FALSE)
```

Let's distribute the work multiple cores

parallel

- ▶ The parallel package is an easy way to split computation over multiple cores

```
library(parallel)
detectCores()
```

```
## [1] 8
```

```
my_matrix_inverses <- mclapply(my_matrices,
                                solve,
                                mc.cores=4)
```

parallel

```
microbenchmark(times = 10,  
  mclapply(my_matrices, solve, mc.cores=4),  
  mclapply(my_matrices, solve, mc.cores=2),  
  mclapply(my_matrices, solve, mc.cores=1)  
)
```

Unit: microseconds

##		expr	min
##	mclapply(my_matrices, solve, mc.cores = 4)	8021.691	872
##	mclapply(my_matrices, solve, mc.cores = 2)	4896.039	547
##	mclapply(my_matrices, solve, mc.cores = 1)	269.643	38
##	median	uq	max neval
##	9521.8155	10183.352	18095.226 10
##	6915.5480	7647.442	8174.207 10
##	624.2825	2029.152	2876.631 10

That's weird...

mclapply, mcsapply...

mclapply, mcsapply and friends use “forks”

- ▶ Idea from Unix-based systems
 - ▶ Does not work on Windows
- ▶ Takes a complete copy of the master process, including the workspace and state of the random-number stream
 - ▶ Generally fast but there is overhead

parallel

```
my_matrices <- replicate(n=20,  
                          rnorm(1e2) %>% matrix(nrow = 1e1),  
                          simplify=FALSE)  
more_matrices <- replicate(n=20000,  
                            rnorm(1e2) %>% matrix(nrow = 1e1),  
                            simplify=FALSE)
```

parallel

```
microbenchmark(times = 10,  
  mclapply(more_matrices, solve, mc.cores=4),  
  mclapply(more_matrices, solve, mc.cores=2),  
  mclapply(more_matrices, solve, mc.cores=1)  
)
```

Unit: milliseconds

##		expr	min
##	mclapply(more_matrices, solve, mc.cores = 4)	223.9301	3
##	mclapply(more_matrices, solve, mc.cores = 2)	306.0708	3
##	mclapply(more_matrices, solve, mc.cores = 1)	307.0515	3
##	median	uq	max neval
##	344.7060	368.2690	601.2881 10
##	407.8786	480.7047	610.5487 10
##	373.7049	377.3594	450.6345 10

There is significant overhead involved in splitting work over cores:
check it is justified first!

parallel

- ▶ Argument: `mc.preschedule`
 - ▶ TRUE: short computations or large number of values in `X`
 - ▶ FALSE: high variance of completion time and not too many values of `X` compared to `mc.cores`
- ▶ Be careful of multiple levels of parallelisation
 - ▶ Multiple processes on multiple cores cause chaos (crashes)
 - ▶ Be careful with GUIs and parallelisation (e.g., Shiny)

parallelisation on Windows

Unfortunately it is slightly more work:

```
z <- as.list(1:4)
system.time(lapply(z, function(x) Sys.sleep(1)))
```

```
##      user  system elapsed
##    0.001    0.000    4.005
```

You need to register a cluster, then use it. Don't forget to shut it down!

```
cl <- parallel::makeCluster(4, type="SOCK")
system.time(parallel::clusterApply(cl, z, function(x) Sys.sleep(1)))
```

```
##      user  system elapsed
##    0.000    0.000    1.006
```

```
parallel::stopCluster(cl)
```

Writing for parallelisation

Many well-written R packages will check to see if you have `doParallel` or `doSNOW` available, and then will adapt to your system

- ▶ Check to see if `ncores` (or similar) is an argument to a function that you are using
- ▶ Implementing this is a little advanced; check out `DivNet` if you're interested

Writing for parallelisation: why you should

“If a program takes longer than 8 minutes for me to install, I will never ever use it, no matter how good it is.”

- ▶ (Actual BFF) Chris Quince

An example of parallelisation via parallel

The simulator uses parallel under the hood.

```
generated_model %>%  
  simulate_from_model(nsim = 40,  
                     index = 5:8,  
                     parallel = list(socket_names = 4)) %>%  
  run_method(list(lse),  
             parallel = list(socket_names = 4)) %>%  
  evaluate(list(squared_error))
```

Summary so far

We have now seen

- ▶ How to benchmark options
- ▶ How to run your own functions in parallel
- ▶ How some packages use `parallel` under the hood

What about large-scale data analysis?

Common in genomics, and increasingly common in modern public health

Data analysis of large data frames

```
devtools::install_github("wesm/feather/R")
library(feather)
cakes <- feather::read_feather("iHMP_IBD.MTX.10samples.feather")
cakes
```

```
## # A tibble: 62,248 x 79
##   index SAMN07424551 SAMN07424552 SAMN07424553 SAMN07424554
##   <chr>          <dbl>          <dbl>          <dbl>          <dbl>
## 1 SAMN~          0            0            0.488          0
## 2 SAMN~          0            0            0              0
## 3 SAMN~          0            0            0              0
## 4 SAMN~          0            0.220        2.17           0
## 5 SAMN~        0.535            0            0              8
## 6 SAMN~        1.45            0            0              0
## 7 SAMN~        1.66            0            0.684          0
```

Preparing for data analysis

```
cakes %<>% gather(key="BioSample", "depth", -1)
cakes
```

```
## # A tibble: 4,855,344 x 3
##   index                               BioSample    depth
##   <chr>                               <chr>        <dbl>
## 1 SAMN07424251_OGJIOPKL_100510 SAMN07424551  0
## 2 SAMN07424251_OGJIOPKL_100564 SAMN07424551  0
## 3 SAMN07424251_OGJIOPKL_100587 SAMN07424551  0
## 4 SAMN07424251_OGJIOPKL_100783 SAMN07424551  0
## 5 SAMN07424251_OGJIOPKL_100920 SAMN07424551  0.535
## 6 SAMN07424251_OGJIOPKL_100933 SAMN07424551  1.45
## 7 SAMN07424251_OGJIOPKL_100967 SAMN07424551  1.66
## 8 SAMN07424251_OGJIOPKL_100995 SAMN07424551  0
## 9 SAMN07424251_OGJIOPKL_101094 SAMN07424551  0
## 10 SAMN07424251_OGJIOPKL_101127 SAMN07424551  0
## # ... with 4,855,334 more rows
```

Joining my metadata (mapping) file

```
meta <- read_csv("iHMP_IBD.csv")
meta
```

```
## # A tibble: 378 x 35
```

```
##   Run      host_subject_id host_id timepoint assay_type he
```

```
##   <chr> <chr>                <chr>   <chr>      <chr>      <chr>
```

```
##  1 SRR5~ C3001C1_MGX          C3001    C1         MGX         CF
```

```
##  2 SRR5~ C3001C10_MGX         C3001    C10        MGX         CF
```

```
##  3 SRR5~ C3001C10_MTX         C3001    C10        MTX         CF
```

```
##  4 SRR5~ C3001C2_MGX          C3001    C2         MGX         CF
```

```
##  5 SRR5~ C3001C3_MGX          C3001    C3         MGX         CF
```

```
##  6 SRR5~ C3001C4_MGX          C3001    C4         MGX         CF
```

```
##  7 SRR5~ C3001C5_MGX          C3001    C5         MGX         CF
```

```
##  8 SRR5~ C3001C5_MTX          C3001    C5         MTX         CF
```

```
##  9 SRR5~ C3001C7_MGX          C3001    C7         MGX         CF
```

```
## 10 SRR5~ C3001C8_MGX          C3001    C8         MGX         CF
```

```
## # ... with 368 more rows, and 29 more variables: BioSample
```

```
## #   LibrarySource <chr>, Sample_Name <chr>, Library_Name
```

Joining my metadata (mapping) file

```
cakes %<>% left_join(meta, by = "BioSample")
cakes
```

```
## # A tibble: 4,855,344 x 37
```

```
##   index BioSample depth Run   host_subject_id host_id t
```

```
##   <chr> <chr>      <dbl> <chr> <chr>          <chr> <
```

```
## 1 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 2 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 3 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 4 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 5 SAMN~ SAMN0742~ 0.535 SRR5~ M2021C1_MTX    M2021 0
```

```
## 6 SAMN~ SAMN0742~ 1.45  SRR5~ M2021C1_MTX    M2021 0
```

```
## 7 SAMN~ SAMN0742~ 1.66  SRR5~ M2021C1_MTX    M2021 0
```

```
## 8 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 9 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## 10 SAMN~ SAMN0742~ 0      SRR5~ M2021C1_MTX    M2021 0
```

```
## # ... with 4,855,334 more rows, and 29 more variables:
```

```
## #   health_status <chr>, LibrarySource <chr>, Sample_Nam
```

Grab only the necessary data

```
cakes <- cakes %>%  
  select(BioSample, index, depth, health_status) %>%  
  filter(health_status %in% c("CD", "Non-IBD"))
```


Analyzing this data

Goal: Look for significant associations of genes (index) with disease state (health_status)

How long do we think this will take?

```
cakes %>% summarise(n_distinct(index)) %>% unlist
```

```
## n_distinct(index)  
##                62248
```

```
cakes %>% summarise(n_distinct(BioSample)) %>% unlist
```

```
## n_distinct(BioSample)  
##                    57
```

Microbenchmark

```
mbm <- microbenchmark(  
  {y <- rnorm(78); x <- rnorm(78); lm(y ~ x)}  
)  
mbm
```

```
## Unit: microseconds
```

```
##
```

```
## {      y <- rnorm(78)      x <- rnorm(78)      lm(y ~ x) }      expr
```

```
##      mean      median      uq      max neval
```

```
## 467.1768 452.7925 464.8645 1607.225    100
```

```
(mbm %$% time %>% median) * 1e-6 * 62248 / 3600
```

```
## [1] 7.829285
```

Estimated: runtime of 62k regressions on 78 observations: about 8 hours

multidplyr

“multidplyr is a backend for dplyr that partitions a data frame across multiple cores. You tell multidplyr how to split the data up with `partition()` and then the data stays on each node until you explicitly retrieve it with `collect()`. This minimises the amount of time spent moving data around, and maximises parallel performance. . . ”

– BFF Hadley

multidplyr

“Due to the overhead associated with communicating between the nodes, you won’t expect to see much performance improvement on basic dplyr verbs with less than ~10 million observations. . .”

– BFF Hadley

Let's use multtidplyr

Definitely in development but actively maintained

```
devtools::install_github("hadley/multtidplyr")
```

```
## Skipping install of 'multtidplyr' from a github remote, t  
## Use `force = TRUE` to force installation
```

```
library(multtidplyr)
```

multiplyr

Split data up over multiple cores

```
ncores <- 4  
cakes %<>%  
  group_by(index)  
cluster <- create_cluster(cores = ncores)
```

```
## Initialising 4 core cluster.
```

```
by_group <- cakes %>%  
  partition(index, cluster = cluster)
```

```
## Warning: group_indices_.grouped_df ignores extra arguments
```

This ensures that all data in the same group goes in the same node

multidplyr

```
start <- proc.time() # Start clock
processed_in_parallel <- by_group %>%
  summarise(p_val = summary(
    lm(depth ~ health_status)
  )$coef[2, 4],
  coef_est = summary(
    lm(depth ~ health_status)
  )$coef[2, 1]) %>%
  collect() %>% # function to recombine partitions
  as_tibble()
time_elapsed_parallel <- proc.time() - start
```

Always important to check

```
summary(lm(depth ~ health_status,  
  data = cakes %>% filter(  
    index == processed_in_parallel$index[5]  
  )))$coef
```

```
##              Estimate Std. Error  t value Pr(>|t|)  
## (Intercept)    1.0621427   0.8133009   1.305965 0.1970002  
## health_statusNon-IBD -0.9563694   1.8513661  -0.516575 0.6075238
```

```
processed_in_parallel[5, ]
```

```
## # A tibble: 1 x 3  
##   index                p_val coef_est  
##   <chr>              <dbl>    <dbl>  
## 1 SAMN07424251_OGJIOPKL_101467 0.608    -0.956
```

Thank goodness...

Let's see how long it took

```
time_elapsed_parallel
```

```
##      user  system elapsed  
##    0.047    0.029   23.653
```

This was unbelievable to me... and why I am teaching it to you!

A quick look at the data

```
processed_in_parallel %>%  
  arrange(desc(coef_est))
```

```
## # A tibble: 62,248 x 3  
##       index                p_val coef_est  
##       <chr>                <dbl>    <dbl>  
## 1 SAMN07424559_ANOFLIEK_24652 0.237    1514.  
## 2 SAMN07424306_PBIHEEJJ_19786 0.0783   1224.  
## 3 SAMN07424567_LNNPIEOP_08899 0.0803    833.  
## 4 SAMN07424568_JKFDFLGO_06732 0.159     769.  
## 5 SAMN07424284_ALEGCIBH_40364 0.746     661.  
## 6 SAMN07424514_CJJNBMBF_16787 0.0440    619.  
## 7 SAMN07424501_DNNKMFDB_22371 0.0742    597.  
## 8 SAMN07424566_MGKBKFLG_12111 0.111     569.  
## 9 SAMN07424536_CCBBMNP_23079 0.0120    456.  
## 10 SAMN07424614_AOEHBBOC_04646 0.220     408.  
## # ... with 62,238 more rows
```

A quick look at the data

```
processed_in_parallel %>%  
  arrange(p_val)
```

```
## # A tibble: 62,248 x 3
```

##	index	p_val	coef_est
##	<chr>	<dbl>	<dbl>
##	1 SAMN07424564_NPKIDKIG_01029	0.00000889	2.07
##	2 SAMN07424395_GBDDKALD_52613	0.0000168	0.935
##	3 SAMN07424369_ODEMOCKO_76309	0.0000214	0.768
##	4 SAMN07424520_IMOKJCLF_52792	0.0000305	0.848
##	5 SAMN07424413_AKPBBDGAN_35653	0.0000362	5.62
##	6 SAMN07424369_ODEMOCKO_81434	0.0000365	0.559
##	7 SAMN07424522_HDNAFFAI_105792	0.0000450	2.55
##	8 SAMN07424334_LOEEILJP_47089	0.0000494	5.29
##	9 SAMN07424289_MODOLPOI_89206	0.000115	1.10
##	10 SAMN07424310_ONNGJNML_51693	0.000123	1.77
##	... with 62,238 more rows		

Save to file

```
processed_in_parallel %>%  
  arrange(p_val) %>%  
  write_csv(path="cd_vs_not_50pct.csv")
```

Multiple ways to make your code run faster

- ▶ Not intelligently
- ▶ Intelligently

It's not a bad thing to have your code run faster in a not-intelligent way!

Intelligent ways of coding

In methods development (and many job interviews), you only get points for the intelligent ways

- ▶ Thoughtfully using algorithms, or developing new ones
 - ▶ There is a somewhat standard toolkit for developing statistical computing algorithms, and it varies by field
 - ▶ Think about if you want to learn more about algorithms and tell me in your feedback sheet

A common problem

```
list.files("tricky_example/")
```

```
## [1] "file1.csv" "file10.csv" "file11.csv" "file12.csv"  
## [6] "file14.csv" "file15.csv" "file16.csv" "file17.csv"  
## [11] "file19.csv" "file2.csv" "file20.csv" "file21.csv"  
## [16] "file23.csv" "file24.csv" "file25.csv" "file26.csv"  
## [21] "file28.csv" "file29.csv" "file3.csv" "file30.csv"  
## [26] "file32.csv" "file33.csv" "file34.csv" "file35.csv"  
## [31] "file37.csv" "file38.csv" "file39.csv" "file4.csv"  
## [36] "file5.csv" "file6.csv" "file7.csv" "file8.csv"
```

Reading data from multiple files

I want to read in all of these files and append them to make one long data frame

```
read_csv("tricky_example/file1.csv")
```

```
## # A tibble: 2 x 4
##   V1      V2      V3      V4
##   <chr> <chr> <dbl> <dbl>
## 1 D      M      0.960  1.80
## 2 T      G     -1.33  -1.34
```

```
read_csv("tricky_example/file40.csv")
```

```
## # A tibble: 4 x 4
##   V1      V2      V3      V4
##   <chr> <chr> <dbl> <dbl>
## 1 G      S     -0.127  0.945
## 2 U      H     -0.385 -0.139
## 3 Y      N      0.378  1.42
## 4 O      L      1.70  -0.326
```


Read data into list

```
all_dfs_list <- lapply(1:40,  
  function(x) {  
    read_csv(paste("tricky_example/file",  
      x, '.csv', sep=''))  
  })  
all_dfs_list
```

```
## [[1]]  
## # A tibble: 2 x 4  
##   V1      V2      V3      V4  
##   <chr> <chr>   <dbl> <dbl>  
## 1 D      M      0.960  1.80  
## 2 T      G     -1.33  -1.34  
##  
## [[2]]  
## # A tibble: 3 x 4  
##   V1      V2      V3      V4  
##   <chr> <chr>   <dbl> <dbl>  
## 1 R      S      0.0900 0.921  
## 2 A      Q     -1.24   0.945  
## 3 X      I      0.128  0.398  
##
```

Natural approaches that don't work

```
rbind(all_dfs_list)
```

```
##           [,1]  [,2]  [,3]  [,4]  [,5]  [,6]
## all_dfs_list List,4 List,4 List,4 List,4 List,4 List,4 I
##           [,9]  [,10] [,11] [,12] [,13] [,14]
## all_dfs_list List,4 List,4 List,4 List,4 List,4 List,4 I
##           [,17] [,18] [,19] [,20] [,21] [,22]
## all_dfs_list List,4 List,4 List,4 List,4 List,4 List,4 I
##           [,25] [,26] [,27] [,28] [,29] [,30]
## all_dfs_list List,4 List,4 List,4 List,4 List,4 List,4 I
##           [,33] [,34] [,35] [,36] [,37] [,38]
## all_dfs_list List,4 List,4 List,4 List,4 List,4 List,4 I
```

Natural approaches that don't work

```
lapply(all_dfs_list, rbind)
```

```
## [[1]]  
## # A tibble: 2 x 4  
##   V1      V2      V3      V4  
##   <chr> <chr>   <dbl> <dbl>  
## 1 D      M      0.960  1.80  
## 2 T      G     -1.33  -1.34  
##  
## [[2]]  
## # A tibble: 3 x 4  
##   V1      V2      V3      V4  
##   <chr> <chr>   <dbl> <dbl>  
## 1 R      S      0.0900 0.921  
## 2 A      Q     -1.24   0.945  
## 3 X      I      0.128  0.398  
##  
## [[3]]
```

Interesting alternative

`do.call`

- ▶ use a list to hold the arguments of the function

`sapply`

- ▶ use a vector to hold the arguments of the function

`do.call` is just like `sapply`, but better

do.call

```
all_dfs_list <- lapply(1:40,  
  function(x) {  
    read_csv(paste(  
      "tricky_example/file", x,  
      '.csv', sep=''))  
  })  
answer <- do.call(rbind, all_dfs_list)  
answer %>% dim
```

```
## [1] 224  4
```

do.call

```
answer
```

```
## # A tibble: 224 x 4
##   V1      V2      V3      V4
##   <chr> <chr>   <dbl> <dbl>
## 1 D      M      0.960  1.80
## 2 T      G     -1.33  -1.34
## 3 R      S      0.0900  0.921
## 4 A      Q     -1.24   0.945
## 5 X      I      0.128   0.398
## 6 N      Y      1.28   -1.03
## 7 G      R      1.08   -0.999
## 8 O      L      0.561   1.27
## 9 A      U      0.159  -0.932
## 10 E     V      0.712   1.30
## # ... with 214 more rows
```

The Max Power way

- ▶ Certain operations are expensive: avoid them
- ▶ A little thought can save a lot of time
- ▶ More on profiling and debugging soon

Coming up

- ▶ Homework 5: due next *Friday* afternoon
 - ▶ Start it after your BIOST 533 final
 - ▶ (Perk of seeing me all the time)
- ▶ Mid-quarter feedback
 - ▶ Specific comments on curriculum/syllabus welcome
 - ▶ Tell me what you want to learn and I will try to accommodate you (and everyone else)!