Phosphoproteome analysis

Hazal Koptagel, Julius Lautenbach, Maryia Ropat & Linnea Axelsson

Isoelectric point-based fractionation by HiRIEF coupled to LC-MS allows for in-depth quantitative analysis of the phosphoproteome

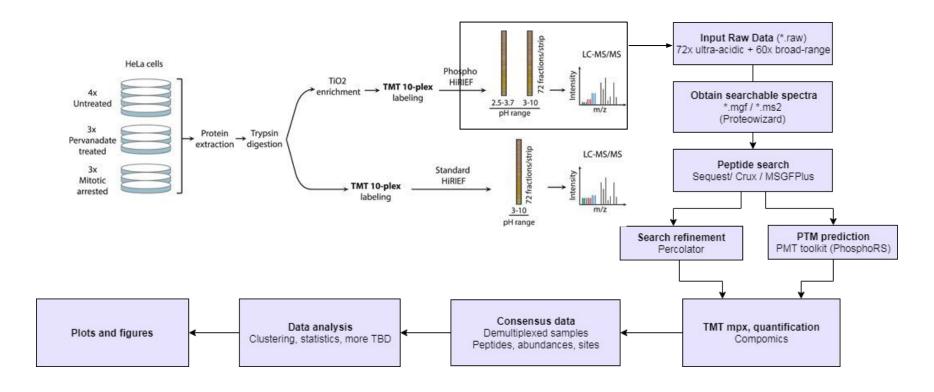
Elena Panizza, Rui M. M. Branca, Peter Oliviusson, Lukas M. Orre & Janne Lehtiö

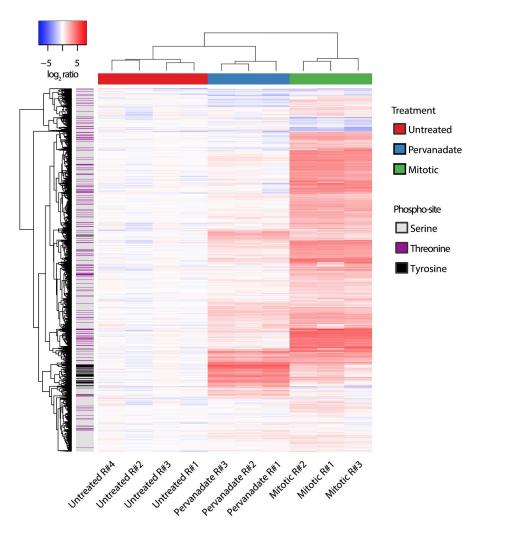
Titanium dioxide phospho-enrichment coupled with isobaric labeling by Tandem Mass Tags (TMT) and high-resolution isoelectric focusing (HiRIEF) before in-depth quantitative phosphoproteomics starting with a low sample quantity

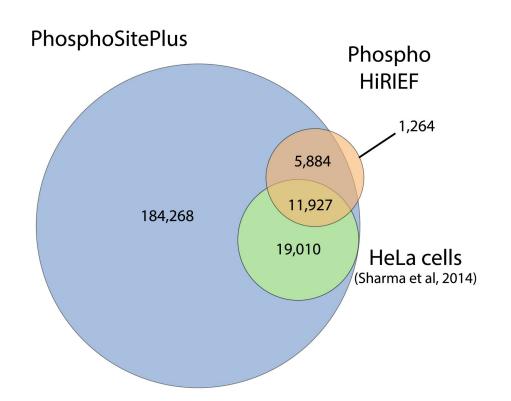
Phosphorylated proteins

- Protein phosphorylation is a fundamental regulatory mechanism in eukaryotic cells
- Deregulated protein phosphorylation is involved in the development of many diseases
- Identification of phosphorylated peptides is challenging due to their low abundance and poor ionization efficiency

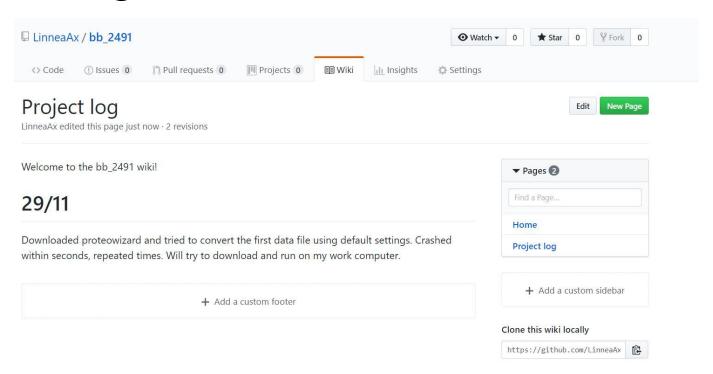
Analysis Workflow



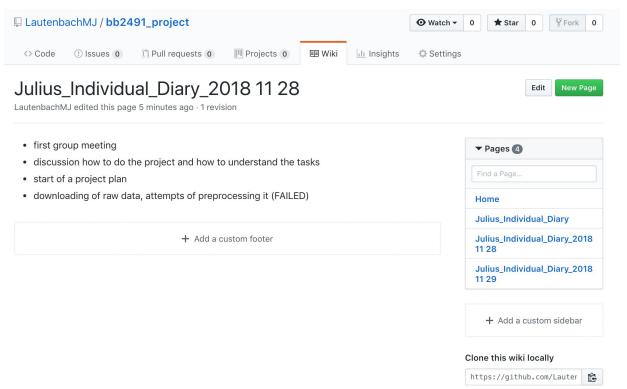




Linnea log



Julius log



https://github.com/LautenbachMJ/bb2491_project/wiki

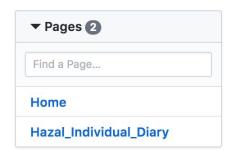
Hazal log

https://github.com/koptagel/bb2490/wiki/Hazal_Individual_Diary

Hazal_Individual_Diary

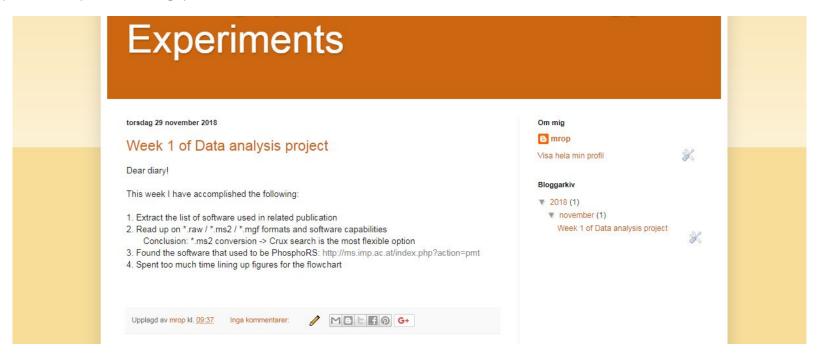
koptagel edited this page 3 hours ago · 3 revisions

- 28-29 Nov: I met with the group to prepare Seminar 1's presentation.
- 29 Nov: I looked at the supplementary data of the article. I started preparing clustering / visualisation scripts.



Maryia log

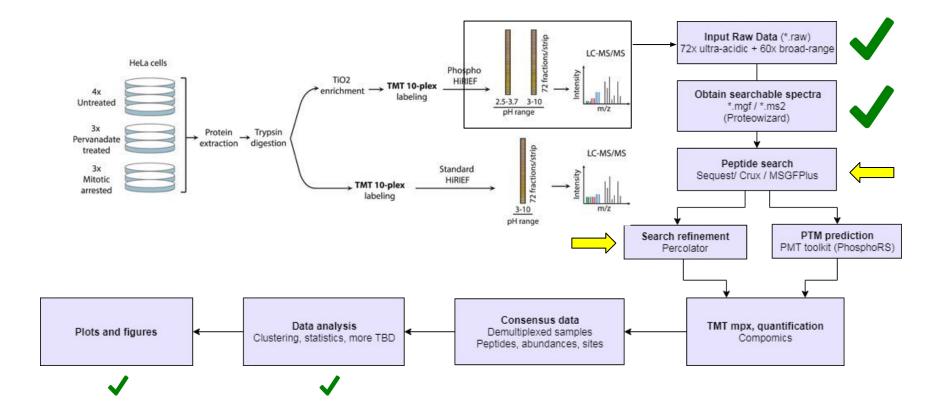
https://maropbb2491.blogspot.com/



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Project Workflow

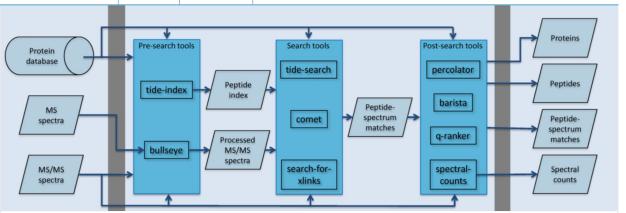


First steps

- preprocessing the raw data into .ms2 format (using Proteowizard msconvert)
 - o focus on the phosphoproteomics runs/files (132 files ~ 32GB)
- running crux/percolator
 - Semi-supervised learning for peptide identification from shotgun proteomics datasets
 - fix-modifications for carbamidomethylation on cysteine (by default)
 - mass modifications for lysine and N-terminus (TMT 10-plex)
- searchGUI with comet
 - good interface
 - possibility to follow Lab4 pipeline
 - output is not compatible percolator

crux







The Crux mass spectrometry analysis toolkit is an open source project that aims to provide users with a cross-platform suite of analysis tools for interpreting protein mass spectrometry data. The toolkit includes several search engines for both standard and cross-linked database search, as well as a variety of pre- and post-processing engines for assigning high-resolution precursor masses to spectra, assigning statistical confidence estimates to spectra, peptides and proteins, and performing label free quantification. Crux comes pre-complied for the Linux, Windows and MacOS operating systems. It is implemented as a single program that offers a wide variety of commands. If you use Crux in your research, please cite:

Christopher Y. Park, Aaron A. Klammer, Lukas Käll, Michael J. MacCoss and William Stafford Noble. "Rapid and accurate peptide identification from tandem mass spectra." *Journal of Proteome Research*, 7(7):3022-3027, 2008.

For a more up-to-date description of Crux, please read:

Sean McIlwain, Kaipo Tamura, Attila Kertesz-Farkas, Charles E. Grant, Benjamin Diament, Barbara Frewen, J. Jeffry Howbert, Michael R. Hoopmann, Lukas Käll, Jimmy K. Eng, Michael J. MacCoss and William Stafford Noble. "Crux: rapid open source protein tandem mass spectrometry analysis." *Journal of Proteome Research.* 13(10):4488-4491, 2014.

- includes several search engines (standard & cross-linked database search)
- pre- and post-processing engines for assigning high-resolution precursor masses to spectra
- estimation of statistical confidence for a spectra, peptides and proteins
- label free quantification

Next steps

- Decide which search engine to use
- Crux/percolator
 - variable modifications of S, T or Y for phosphorylations
- Apply search to all files