

International Union of Basic and Clinical Pharmacology. LXXXVI. Orexin Receptor Function, Nomenclature and Pharmacology

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Abstract—Orexin signaling is essential for normal regulation of arousal and behavioral state control and represents an attractive target for therapeutics combating insomnia. Alternatively termed hypocretins, these neuropeptides were named to reflect sequence similarity to incretins and their potential to promote feeding. Current nomenclature reflects these molecular and biochemical discovery approaches in which *HCRT*, *HCRT1*, and *HCRT2* genes encode prepro-orexin, the orexin 1 receptor (OX_1) and the orexin 2 receptor (OX_2)—gene names designated by the Human Genome Organization and receptor names designated by the International Union of Basic and Clinical Pharmacology. Orexinergic neurons are most active during wakefulness and fall silent during inactive periods, a prolonged disruption in signaling most profoundly resulting in hypersomnia and narcolepsy. *Hcrtr2* mutations underlie the etiology of canine narcolepsy, defi-

ciencies in orexin-producing neurons are observed in the human disorder, and ablation of mouse orexin neurons or the *Hcrtr* gene results in a narcolepsy-cataplexy phenotype. The development of orexin receptor antagonists and genetic models targeting components of the orexin pathway have elucidated the OX_2 receptor-specific role in histamine-mediated arousal and the contribution of both receptors in brainstem pathways involved in vigilance state gating. Orexin receptor antagonists of varying specificity uncovered additional roles beyond sleep and feeding that include addiction, depression, anxiety, and potential influences on peripheral physiology. Combined genetic and pharmacological approaches indicate that orexin signaling may represent a confluence of sleep, feeding, and reward pathways. Selective orexin receptor antagonism takes advantage of these properties toward the development of novel insomnia therapeutics.

I. Introduction

Orexin/hypocretin signaling has a pre-eminent role in the regulation of arousal and vigilance state. Disruption of the genes encoding the orexin 2 receptor or orexin ligands themselves are associated with canine (Lin et al., 1999) and murine (Chemelli et al., 1999) narcolepsy, and orexin deficiency is associated with the human disorder (Nishino et al., 2000). These initial findings not only demonstrated that orexin governs the normal regulation of arousal but also set off a range of investigations aimed at defining the role of this signaling pathway in arousal and the regulation of sleep. Before the genetic link to mammalian narcolepsy was uncovered, these hypothalamic peptides had been discovered and named based on their similarity to incretins (“hypocretin”) (de Lecea et al., 1998) and their propensity to promote feeding (“orexin” after the Greek word *orexis* for appetite) (Sakurai et al., 1998). Although the designation of hypocretin may be appropriate from a molecular and perhaps a functional standpoint, the term orexin is much more pervasive in the biological, chemical, patent, and popular literature. The present work addresses the nomenclature of these neuropeptides and their cognate receptors as well as the small molecules targeting pathways associated with orexin signaling. Studies using these small-molecule antagonists in concert with genetic manipulation have also been invaluable toward dissecting the function of orexin signaling in arousal, vigilance state, and the mechanisms regulating sleep in general. In addition to its function in arousal, feeding, and en-

ergy homeostasis, remarkable progress has also been made toward understanding the role of orexin in addiction and psychiatric function, as well as peripheral influences on nociception, metabolism and cardiovascular physiology that may or may not be a secondary consequence of its central roles. From this work, the therapeutic potential of modulating orexin signaling for the selective treatment of insomnia and related sleep disorders has become evident, but so too have the possibilities for the treatment of disorders in which sleep/wake dysregulation occurs. This therapeutic potential contrasts with the current standard of care including GABA_A receptor modulators, which have less selectivity for mechanisms controlling sleep/wake regulation.

II. Discovery and Nomenclature of Orexin Signaling Components

A. Orexin-A and -B Are Products of the Hypocretin Gene

Even before the genetic association with narcolepsy was discovered, orexin neuropeptides were simultaneously described by two different groups using distinct molecular and biochemical approaches. From mRNA enriched from rat hypothalamus, de Lecea et al. (1998) identified, cloned, and sequenced a 569-nucleotide transcript encoding a 130-amino acid prepro-peptide based on sequence similarity to secretin, a gut hormone involved in osmoregulation. Because of its CNS expression restricted to large cell bodies of the dorsal lateral hypothalamic area and its sequence similarity to the incretin

TABLE 1
Nomenclature of orexin signaling components

The IUPHAR (International Union of Basic and Clinical Pharmacology) ID was retrieved from <http://www.iuphar-db.org/DATABASE/FamilyMenuForward?familyId=51>. The HGNC gene name is that approved by the Human Genome Nomenclature Committee. Chromosomal location is based on fluorescent in situ hybridization mapping (human) and ISCN (International System for Cytogenetic Nomenclature) lengths (mouse and rat) from UCSC Genome Bioinformatics (<http://genome.ucsc.edu>). ChEMBL is from the MedChem literature data on drug-like molecules and their targets.

IUPHAR ID ^a	HGNC Gene Name ^b	Species	HGNC Symbol	Genomic Locus	Databases				
					IUPHAR Ligand ID	Entrez ID	Unigene	HGNC	ChEMBL
OX-A OX-B	Orexin-A Orexin-B	Human	<i>HCRT</i>	17q21.1	1697 1699	602358	Hs0.158348	4847	N.A.
		Mouse	<i>Hcrt</i>	11qD		15171	Mm0.10096		
		Rat	<i>Hcrt</i>	10q32.1		25723	Rn0.7628		
OX ₁ receptor	Hypocretin (orexin) receptor 1	Human	<i>HCRT1</i>	1p35.2	N.A.	3061	Hs0.388226	4848	10009
		Mouse	<i>Hcrt1</i>	4qD2.2		23077	Mm0.246595		
		Rat	<i>Hcrt1</i>	5q36		25593	Rn0.88262		
OX ₂ receptor	Hypocretin (orexin) receptor 2	Human	<i>HCRT2</i>	6p12.1	N.A.	3062	Hs0.151624	4849	12968
		Mouse	<i>Hcrt2</i>	9qD		387285	Rn0.9893		
		Rat	<i>Hcrt2</i>	8q24		25605	Mm0.335300		

family of peptide hormones, the gene was termed hypocretin (*Hcrt*). Two peptide products predicted from proteolytic cleavage sites were confirmed by immunohistochemistry to be present in cell bodies and efferent fibers, and one of the synthetically generated peptides was able to elicit depolarizing currents in primary cultures of hypothalamic neurons (de Lecea et al., 1998). In an effort to “deorphanize” a panel of G-protein-coupled receptors, a second group (Sakurai et al., 1998) identified one such receptor capable of mediating Ca²⁺ transients in response to crude rat brain extracts. Purification and sequencing of the biological activity capable of activating this receptor revealed a sequence encoding a precursor peptide processed into two related peptides. Because the mRNA was found to be expressed in the lateral hypothalamus (LH), an area implicated in feeding regulation (Bernardis et al., 1993; Bernardis and Bellinger, 1996), and because intraventricular administration of both peptides dose-dependently induced food intake, the peptides were designated orexin-A and -B (OX-A¹ and

OX-B) after the Greek word *orexis*, for appetite. These deorphanized receptors were identified as orexin 1 and orexin 2 (OX₁ and OX₂) receptors (Sakurai et al., 1998). These findings ignited a large body of work aimed at deciphering the role of these signaling components in appetite control (Edwards et al., 1999; Sweet et al., 1999), energy homeostasis (Beck and Richy, 1999), and metabolism (Lubkin and Stricker-Krongrad, 1998; Takahashi et al., 1999) such that the term “orexin” seemed appropriate, yet the nomenclature debate had already begun (Nisoli et al., 1998). The subsequent discoveries of the genetic link to narcolepsy and the predominant role of orexin in arousal (Mieda et al., 2004), however, raised the possibility that orexin-induced feeding observed in preclinical models may be secondary to heightened wakefulness (Ida et al., 1999), further fueling the debate.

B. Nomenclature Recommendations

A parsimonious resolution to the nomenclature debate is to designate hypocretin the gene and mRNA name (human abbreviation: *HCRT*; rodent: *Hcrt*) and the precursor peptide and processed peptides after orexin (orexin A, OX-A, orexin B, OX-B) (Table 1). Likewise, the same is true for the two known G-protein-coupled receptors for these peptides; the *HCRT1* and *HCRT2* genes (*Hcrt1* and *Hcrt2* in rodents) encode the protein products OX₁ and OX₂ receptors, respectively. Although this nomenclature may be confusing to those new to the field, it does recognize the identification of the hypocretin mRNA and gene by molecular biology approaches (de Lecea et al., 1998) and the biochemical discovery of orexin peptides and their now deorphanized G-protein-

¹Abbreviations: 5-HT, 5-hydroxytryptamine (serotonin); CGS21680, 2-[p-(2-carboxyethyl)phenyl-ethylamino]-5'-N-ethylcarboxamidoadenosine; CPP, conditioned place preference; CRF, corticotropin-releasing factor; CSF, cerebrospinal fluid; DMH, dorsomedial nucleus of the hypothalamus; DMV, dorsal motor nucleus of the vagus nerve; DORA, dual orexin receptor antagonist; DR, dorsal raphe; DRG, dorsal root ganglion; EMPA, N-ethyl-2-[(6-methoxy-pyridin-3-yl)-(toluene-2-sulfonyl)-amino]-N-pyridin-3-ylmethyl-acetamide; GSK, GlaxoSmithKline; GSK1059865, 5-bromo-N-[(2S,5S)-1-(3-fluoro-2-methoxybenzoyl)-5-methylpiperidin-2-yl]methyl-pyridin-2-amine; *HCRT*, hypocretin gene; *HCRT1*, hypocretin (orexin) receptor 1 gene; *HCRT2*, hypocretin (orexin) receptor 2 gene; IP₃, inositol-1,4,5-trisphosphate; JNJ-10397049, [1-(2,4-dibromophenyl)-3-[(4S,5S)-2,2-dimethyl-4-phenyl-1,3-dioxan-5-yl]urea; LC, locus ceruleus; LDT, laterodorsal tegmental nuclei; LH, lateral hypothalamus; LPS, latency to persistent sleep; MK-4305, suvorexant; MK-6096, [2(R,5R)-5-[(5-fluoropyridin-2-yl)oxy]methyl]-2-methylpiperidin-1-yl][5-methyl-2-(pyrimidin-2-yl)phenyl]methanone; nNOS, neuronal nitric-oxide synthase; NPFF, neuropeptide FF; NPY, neuropeptide Y; NREM, non-rapid eye movement; OBpt-9, N-benzyl-N-(3,4-dimethoxybenzyl)glycyl-N²-(1-phenylethyl)glycinamide; OX, orexin; OX/Atx, transgenic orexinergic neuron ablation mutant; PFC, prefrontal cortex; PLC, phospholipase C; PPT, pedunculo pontine tegmental nuclei; PTSD, post-traumatic stress disorder; qEEG, quantitative electroencephalography; QRFP receptor, QRF peptide receptor (GPR103); REM, rapid eye movement; SB-334867, 1-(2-methylbenzoxazol-6-yl)-3-

[1,5]naphthyridin-4-yl urea; SB-408124, 1-(6,8-difluoro-2-methylquinolin-4-yl)-3-(4-dimethylamino-phenyl)-urea; SB-649868, N-[[[(2S)-1-[[5-(4-fluorophenyl)-2-methyl-4-thiazolyl]carbonyl]-2-piperidinyl]methyl]-4-benzofurancarboxamide; SCN, suprachiasmatic nuclei; SORA, selective OX₁ receptor and OX₂ receptor antagonist; TC-SOX₂29, (2S)-1-(3,4-dihydro-6,7-dimethoxy-2(1H)-isoquinolinyl)-3,3-dimethyl-2-[(4-pyridinylmethyl)amino]-1-butanone; TMN, tuberoammillary nuclei; VLPO, ventrolateral preoptic area; VTA, ventral tegmental area; WASO, wake after sleep onset.

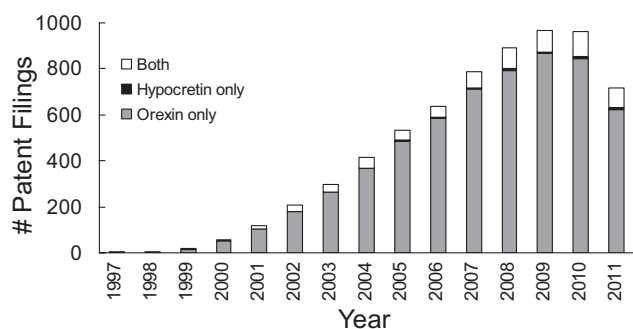


FIG. 1. "Orexin" is used preferentially in the patent literature over "hypocretin." The number of patent filings containing, in the full text, the words "orexin" (gray), "hypocretin" (black), or both (open) per year are shown. Results do not include patents in which the word "orexinergic" appears without either of the above.

coupled receptors (Sakurai et al., 1998). These distinct genetic and protein product designations are also now a matter of practical necessity. *HCRT*, *HCRT1*, and *HCRT2* are now the accepted gene symbols in all genetic databases, including GenBank and HUGO. As for orexin-related protein products, the formal nomenclature from the International Union of Basic and Clinical Pharmacology Nomenclature Committee designates "OX-A" and "OX-B" as pharmacological ligands for "OX₁" and "OX₂" receptors, respectively (<http://www.iuphar-db.org/DATABASE/FamilyMenuForward?familyId=51>). The biological literature primarily uses the "orexin" designation, with references as well to "hypocretin" and/or the accepted gene name. Where the term "orexin" has been used much more exclusively, however, is in both the chemistry and patent literature, which has expanded substantially in the past 15 years as the therapeutic potential of modulating the orexin system has become increasingly evident (Fig. 1). Although early patents were filed for orexin peptide ligand derivatives and increasingly thereafter for molecular probes (e.g., microarray and quantitative polymerase chain reaction probes and primers), compound patents became more prominent with a growing interest in the development of small-molecule therapeutics. As seen in Fig. 1, the number of "orexin"-only patent filings has far exceeded that of "hypocretin"-only patents since 1999, the year the genetic link with narcolepsy was made. At the peak of patent filings in 2009, only 6 of a total of 965 patents filed mentioned "hypocretin" only, whereas 866 were exclusively for "orexin" and 93 for both. These figures demonstrate that the orexin and orexin receptor protein designations are the most widely accepted pharmacological designations.

C. Orexin-A and -B Structures Are Highly Conserved

Both OX-A and OX-B neuropeptides are derived from the same prepro-orexin precursor encoded by the *HCRT* gene. The structure and organization of the hypocretin gene has been largely conserved through evolution. In all vertebrates examined, the gene is composed of two exons with the intron splice falling within the early

portion of the open reading frame encoding the secretory signal sequence (Fig. 2). *Hcrt* genes from multiple organisms, including teleost fish, avian, and mammalian species, are located within chromosomal loci having considerable synteny and, along with neighboring genes, may have experienced considerable pressure for functional conservation through evolution (Wong et al., 2011).

The organization of prepro-orexin precursor peptide and the sequence of mature OX-A and OX-B ligands that are derived from it are highly conserved. The translated 131-amino acid human prepro-orexin peptide consists of nearly contiguous sequences encoding the secretory signal sequence, 33-amino acid OX-A, and 28-amino acid OX-B (Sakurai et al., 1999), and this organization, along with cleavage site sequences, are exactly conserved among all vertebrate organisms examined, including frog, chicken, and fish. This includes consensus "Gly-basic-basic" cleavage and C-terminal Gly-Lys-Arg amidation motifs, separating the OX-A and OX-B sequences, and Gly-Arg-Arg sequences, marking the termination of OX-B (Wong et al., 2011). Among mammals, the sequence of the mature OX-A ligand is entirely conserved among all species examined and contains two disulfide bridges, one formed by cysteines 6 and 12 and another between cysteines 7 and 14. These four residues are also 100% conserved from humans to amphibians (Wong et al., 2011). Mature OX-A is further post-translationally modified with an N-terminal pyroglutamic acid (Sakurai et al., 1998). Mammalian OX-B sequences, on the other hand, are very well conserved but have two points of differentiation: a serine residue at the second amino acid position in rodents, canines, and bovines is replaced by a proline in the human sequence, and a serine in the 18th position is replaced by an asparagine in rodents (Wong

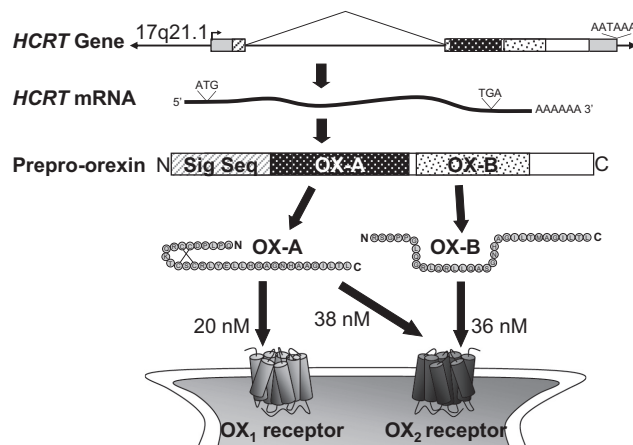


FIG. 2. OX-A and OX-B are encoded by the *HCRT* gene. Structures of the human gene [from UCSB genome browser (<http://genome.ucsc.edu>); intronic sequence is shown at 1/10th scale of exon sequence], mRNA, and protein gene products shown. IC₅₀ values for radioligand binding by OX-A and OX-B are depicted with the exception of the affinity of OX-B for OX₁ receptors (420 nM, not shown), which is ~10-fold less than for OX₂ receptor (36 nM) (Sakurai et al., 1998).

TABLE 2
Mammalian OX₁ receptor protein homology

The indicated annotated protein sequences were compared with the human OX₁ receptor sequence using a BLASTP algorithm with default comparative parameters (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Identity and similarity values are the percentage of identical or homologous amino acids shared with the core region of the human OX₁ receptor.

Protein	Identity	Similarity
%		
OX ₁ receptor		
Human	100	100
Rhesus	97	98
Chimp	94	96
Canine	93	95
Rabbit	93	95
Mouse	92	93
Rat	91	93
OX ₂ receptor		
Human	69	80

et al., 2011). Sequence diversity outside the secretory signal, OX-A, and OX-B sequences substantiates the functional importance of these defined regions.

OX-A and OX-B also share sequence similarity with one another, which is likely to underlie their ability to serve as ligands for both OX₁ and OX₂ receptors, albeit with differing affinities. In this regard, it is worth noting that mammalian OX-A and OX-B sequences are identical in the C-terminal portion of the mature peptides, including the nine-amino acid sequence Gly-Asn-His-Ala-Ala-Gly-Ile-Leu-Thr. They also share Arg-Leu and Leu-Leu sequences spaced two amino acids from one another and three amino acids N-terminal of the nine conserved positions mentioned above, suggesting that these residues may exist at one surface of an α -helical secondary structure (Sakurai et al., 1998; Wong et al., 2011). Because both peptides have measurable affinities for each of the OX₁ and OX₂ receptors, these observations indicate that these residues are essential for orexin receptor interaction. Human OX-A has nearly equal activity on both orexin receptors, with ligand binding affinities (IC₅₀) of 20 and 38 nM for OX₁ and OX₂ receptors, respectively, and EC₅₀ values of 30 and 34 nM in [Ca²⁺]_i mobilization assays of cells transfected to express human OX₁ and OX₂, respectively (Fig. 2) (Sakurai et al., 1998). OX-B, however, has markedly less activity toward OX₁ receptors with an IC₅₀ for radioligand binding of 420 nM and an EC₅₀ for [Ca²⁺]_i of 2500 nM. It is more selective for OX₂ receptors, exhibiting an IC₅₀ of 36 nM and EC₅₀ of 60 nM (Sakurai et al., 1998). This selectivity of OX-B for OX₂ has been used to interpret the relative roles of OX₂ and OX₁ receptors in biological functions, because differential responses to microinjection of these peptides into brain regions indicates OX₁ receptor function, whereas similar responses to both peptides may suggest OX₂ receptor function. Definitive receptor selective function, however, is demonstrated only with genetic and/or highly selective orexin receptor antagonist reagents.

D. Orexin Receptor Structure Is Evolutionarily Conserved

1. Orexin 1 Receptor. Orexin 1 and 2 receptors are found throughout mammalian species, and the core regions of these proteins are highly conserved. The greatest diversity occurs between rat and human OX₁ receptor sequences, but even these proteins are 91% identical and 93% homologous (Table 2). As seen in the predicted structure of the OX₁ receptor based on a β_2 -adrenergic receptor homology model (Fig. 3A), most of the divergent residues occur in the large cytoplasmic loop between transmembrane spanning helices five and six. Fewer residues within membrane spanning loops and the ligand-binding pocket diverge in rat, dog, and human sequences, and these changes are largely homologous. The exception is an Arg-to-His change at residue 205 within the second extracellular loop between transmem-

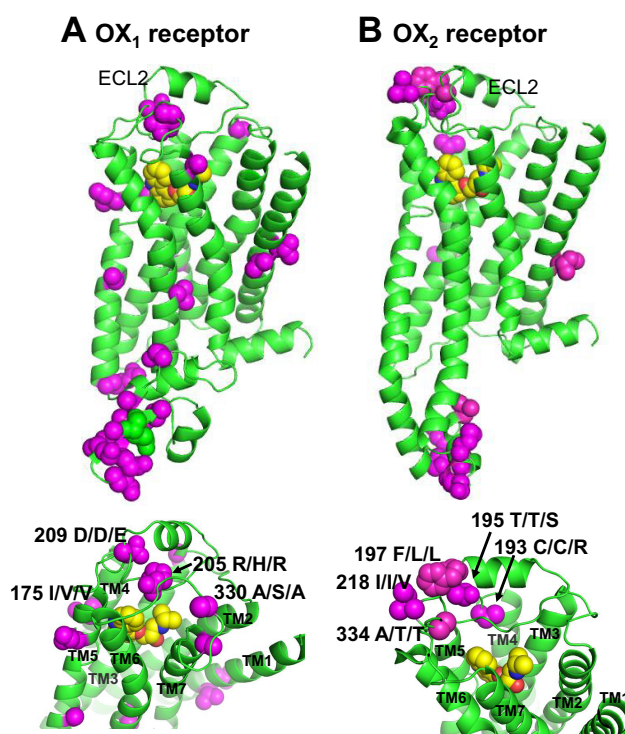


FIG. 3. Amino acid divergence in the structures of OX₁ and OX₂ receptors among human, rat, and canine sequences. Homology models were created using MOE software (Chemical Computing Group, Montreal, QC, Canada) based on the crystal structure of carazolol binding to the β_2 -adrenergic receptor (Protein Data Bank code 2rh1) (Cherezov et al., 2007; Rosenbaum et al., 2007) used as the template. Sequence alignment of the transmembrane helices and the extracellular loop 2 (ECL2) of OX₁ and OX₂ receptors with the 2rh1 structure was performed according to Malherbe et al. (2010) and the best model selected from 10 intermediates. Backbone coordinates remained identical to the crystal structure such that no minimization was performed (structural data comparison and figure generated by M. Katharine Holloway, Ph.D., Chemical Modeling and Informatics, Merck Research Laboratories). The predicted structure of OX₁ (A) and OX₂ (B) receptors along with their ligand binding sites (lower panels) showing the peptide backbone (green) for sequence conserved among human, rat, and dog. Space-filling residues exhibiting sequence divergence are shown in magenta and the amino acid position and sequence substitutions are indicated in the lower panel (human/dog/rat amino acids at these positions). TM, transmembrane helices. The binding site is occupied by carazolol (yellow) used in homology modeling to the β_2 -adrenergic receptor.

brane helices four and five of the predicted canine sequence, which may have the potential to affect ligand binding and antagonist activity. Differences in canine OX₁ receptor sequences relative to human and rodent sequences may be of interest given the possible difference in phenotypes displayed by disruptions in orexin signaling in these species. Truncation of the dog OX₂ receptor results in a narcoleptic phenotype accompanied by cataplexy (Lin et al., 1999), whereas deletion of the *Hcrt* gene or both orexin receptors in mice (Chemelli et al., 1999; Willie et al., 2003; Scammell et al., 2009) and orexin neuron loss in human narcoleptics is associated with this phenotype (Thannickal et al., 2003; Nishino et al., 2010). The Arg-to-His change in the dog OX₁ receptor, however, does not seem to affect ligand activation or the activity of two different dual orexin receptor antagonists, [2(*R*,5*R*)-5-[(5-fluoropyridin-2-yl)oxy]methyl]-2-methylpiperidin-1-yl][5-methyl-2-(pyrimidin-2-yl)phenyl]methanone (MK-6096) and DORA-22, toward the dog OX₁ receptor (Winrow et al., 2012), indicating that any differences in the biological function of this receptor in canines may not be due to differences in receptor activity but might be explained by differential or regional expression changes.

2. Orexin 2 Receptor. Mammalian OX₂ receptor sequences exhibit even greater conservation between species (Table 3), which is probably a consequence of its greater role in mediating orexin's effects on arousal and vigilance state (see section V.E). As with the OX₁ receptor, the greatest divergence among rat, dog, and human sequences occurs within the cytoplasmic loop between transmembrane helices five and six (Fig. 3B), suggesting that these positions are not critical for transduction of the ligand activation signal or subsequent G protein signaling. Within transmembrane regions and the predicted ligand-binding region, only conserved amino acid substitutions are observed. The most divergent position seems to be a Phe-for-Leu difference in rat and dog sequences located in the second extracellular loop between transmembrane helices four and five. Although this change represents an aromatic to aliphatic residue

substitution, both are hydrophobic and are predicted to be removed from the putative ligand binding pocket.

Among the regions of OX₂ receptors that are invariant in human, dog, and rat sequences are those critical for the interaction with OX-A and OX-B peptide ligands as well as small-molecule antagonists to the receptor. The ligand-binding pocket is formed by an interface between transmembrane helices, deep within the extracellular portion of both receptors. As indicated by functional studies, known small-molecule antagonists act in an orthosteric mode in that they compete for binding with residues critical for peptide ligand interaction. Transmembrane domain 3 seems to be the most critical, where exchange of OX₂ receptor sequence with that from OX₁ receptor switches the ligand binding properties of a chimeric receptor (Putula et al., 2011). The extracellular portion of transmembrane domain 3 also contains Gln134 and Thr135, residues essential for peptide ligand and small molecule interactions, respectively (Malherbe et al., 2010; Tran et al., 2011). It is noteworthy that substitution of Thr135 with an alanine in the OX₂ receptor, which matches the Ala135 naturally found in the OX₁ receptor, substantially attenuates small-molecule binding but induces a small increase in activity toward OX-B (Tran et al., 2011). Additional residues contained in the extracellular portions of transmembrane domains six and seven also contribute to the small-molecule pocket (Malherbe et al., 2010; Tran et al., 2011).

3. Evolutionary Origins. Evolutionarily, OX₂ receptors seem to be a more ancient addition to class B G-protein-coupled receptors relative to the OX₁ receptor, which seems to have arisen from a more recent gene duplication (Wong et al., 2011). *Hcrt* genes encoding OX₁ receptors have not been identified outside of the mammalian class (Fig. 4), suggesting that the function of these proteins represents a refinement of sleep, the control of vigilance or potentially other behavioral functions unique to mammals. Using the human orexin receptor sequences as queries in BLASTP searches, proteins exhibiting similarity to these receptors were identified, including neuropeptide FF receptors 1 and 2 (NPFF receptors 1 and 2), substance K receptors, GPR83, neuropeptide Y receptors, and QRF peptide receptors (QRFP receptor; also known as GPR103). Although NPFF receptors 1 and 2 share a greater number of identical amino acid positions with OX₂ receptors (31 and 33%) relative to QRFP receptor (27%), QRFP receptors retain a greater number of conserved or similar amino acid positions. When known QRFP receptors and NPFF receptors are included in phylogenetic analysis, QRFP receptors cluster nearer to orexin receptors (Fig. 4). This sequence similarity and lack of identity suggests that QRFP receptors diverged from orexin receptors earlier than NPFF receptors but experienced selective pressure to retain homologous sequence. Reported responses to central administration of QRF peptides, the ligands for QRFP receptor, include

TABLE 3
Mammalian OX₂ receptor protein homology

The indicated annotated protein sequences were compared with the human OX₂ receptor sequence using a BLASTP algorithm with default comparative parameters (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). Identity and similarity values are the percentage of identical or homologous amino acids shared with the core region of the human OX₂ receptor.

Protein	Identity	Similarity
%		
OX ₂ receptor		
Human	100	100
Chimp	99	99
Rhesus	98	99
Canine	97	98
Rabbit	97	98
Mouse	94	96
Rat	94	96
OX ₁ receptor		
Human	69	80

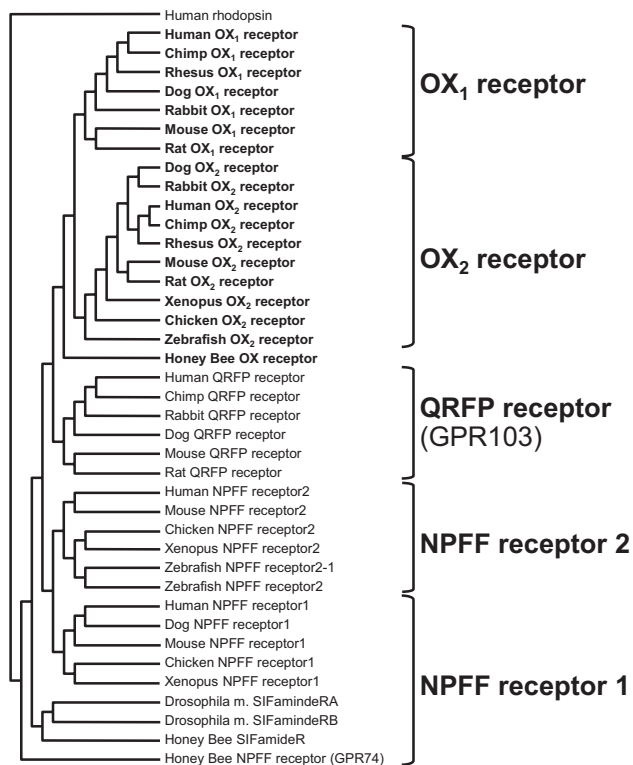


FIG. 4. Orexin receptor phylogeny and evolutionary conservation. Human OX₁ and OX₂ receptor sequences were used in BLASTP algorithm searches (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) to identify orthologous orexin receptor sequences from other species as well as nonorexin receptor sequences sharing sequence identity and homology. Human NPFF receptor 1, NPFF receptor 2, and GPR103 (QRFP) protein sequences were then used to identify orthologous versions of these sequences in other species. Sequences were then aligned, and phylogenetic trees generated by using the Cobalt Phylo Tree tools at NCBI (<http://www.ncbi.nlm.nih.gov/tools/cobalt>), using human rhodopsin as the out-group. Note: human NPFF receptor 1 and NPFF receptor 2 sequences share greater identity with orexin receptors, whereas GPR103 (QRFP) shares greater sequence similarity (residues of similar property) such that phylogenetic comparison, including GPR103 sequences from multiple species places them nearer orexin receptor sequences.

increases in arousal and locomotor activity; increased feeding, energy balance regulation, and the modulation of pain responses (Thuau et al., 2005; Moriya et al., 2006; Takayasu et al., 2006; Yamamoto et al., 2008; Lectez et al., 2009; Yamamoto et al., 2009), functions remarkably similar to that of orexin.

E. Orexin Receptor G Protein Signaling Modulating Neuronal Excitability

Orexin activation of OX₁ and OX₂ receptors results in target cell activation, generally reflected in increased intracellular Ca²⁺ levels and postsynaptic excitation that can last several minutes (Sakurai et al., 1998; van den Pol et al., 1998; Hagan et al., 1999; Bourgin et al., 2000; Van Den Pol et al., 2001; Liu et al., 2002; Arrigoni et al., 2010). Presynaptic modulation of neurotransmitter release and postsynaptic modulation of responses to other neurotransmitters have also been observed, particularly for OX₂ receptors, the cellular signaling mechanisms of which seem more diverse. The specific intra-

cellular signaling pathways in which each of these receptors participate probably depend upon the cellular expression and subcellular localization (e.g., presynaptic versus postsynaptic) of other second-messenger signaling components.

Studies performed both in native primary neurons and in immortalized cell systems expressing recombinant orexin receptors indicate that these G-protein-coupled receptors increase intracellular Ca²⁺ through Gα_{q/11} activation (Lund et al., 2000; Smart et al., 2000; Kukkonen and Akerman, 2001; Holmqvist et al., 2002; Zhu et al., 2003). Both orexin receptors seem to signal primarily through Gα_{q/11}, but some evidence suggests that both receptors are also capable of modulating cyclic nucleotide levels through Gα_s and Gα_{i/o} with varying ligand potencies, even though their predominant intracellular effect is to increase intracellular Ca²⁺ levels (Karteris et al., 2001; Holmqvist et al., 2002; Zhu et al., 2003). The most direct mechanism through which Gα_{q/11} induces Ca²⁺ levels in orexin responsive cells is the activation of phospholipase C (PLC), triggering the liberation of inositol-1,4,5-trisphosphate (IP₃) and the release of Ca²⁺ from intracellular stores through IP₃ receptors (Smart et al., 1999). However, the full Ca²⁺ response mediated by OX₂ receptors requires extracellular Ca²⁺ conducted through either non-voltage-gated cation channels potentiating the PLC response (Lund et al., 2000; Kukkonen and Akerman, 2001; Holmqvist et al., 2002) or through L- and N-type Ca²⁺ channels, as indicated by a block in OX-A responsiveness of dopaminergic neurons of the ventral tegmental area (VTA) by Ω-conotoxin and nitrendipine (Uramura et al., 2001). Still other work has suggested a mechanism through which diacylglycerol, liberated by PLC, may directly activate transient receptor potential channels responsible for Ca²⁺ influx (Larsson et al., 2005; Näsman et al., 2006; Louhivuori et al., 2010).

The capacity of OX₂ receptors to both increase intracellular Ca²⁺ and regulate cAMP levels enables orexin signaling to modulate both the presynaptic release of neurotransmitters and the postsynaptic response to other transmitters. Orexin can regulate the presynaptic release of serotonin, GABA or glutamate (van den Pol et al., 1998; Liu et al., 2002). Postsynaptically in the VTA, the substantial Ca²⁺ responses induced by orexin can induce long-duration changes in N-methyl-D-aspartate receptor expression, thereby potentiating the response of these neurons for several hours (Borgland et al., 2006; Borgland et al., 2008). Orexin neurons also release dynorphin, which attenuates inhibitory postsynaptic potentials, an effect that ultimately potentiates orexin-induced postsynaptic activation of neurons in tuberomammillary nuclei (TMN) (Eriksson et al., 2004; Kantor et al., 2009; Williams and Behn, 2011). This synergistic effect of neuropeptides may provide an explanation for the more pronounced obesity and REM dysregulation phenotype of animal models in which

orexin-secreting neurons are genetically ablated relative to *Hcrt* knockout animals harboring a mutation in the gene encoding the prepro-orexin peptide (Hara et al., 2005; Kantor et al., 2009).

III. Orexin Receptor Reagents and Potential Therapeutics

A. Orexin Receptor Agonists/Potentiators

Advances in the study of orexin receptor agonism have been limited; fewer small-molecule tools are available compared with many antagonist compounds developed to date. Lack of an established positive control complicates any screening strategy to uncover a small-molecule agonist. Studies have thus been limited primarily to synthetic and modified versions of the endogenous neuropeptide agonists OX-A and OX-B.

In contrast to the narcoleptic phenotype in dogs and humans with dysfunctional orexin receptors or deficiencies in endogenous peptide ligands, (Lin et al., 1999; Nishino et al., 2000; Peyron et al., 2000; Wu et al., 2002), studies in rodents have demonstrated that intracerebroventricular administration of the orexin ligands serves to induce arousal and increase wakefulness (Hagan et al., 1999; Akanmu and Honda, 2005). Deadwyler et al. (2007) further reported that systemic and intranasal delivery of OX-A can decrease the effects of sleep deprivation on cognitive performance in nonhuman primates. The observation that systemic administration elicits an effect in these studies supports the finding that orexin-A crosses the blood-brain barrier by diffusion (Kastin and Akerstrom, 1999). No reports of behavioral responses to exogenously applied orexin peptides in humans, however, have appeared in the literature.

In addition to shedding light on the function of the endogenous orexin ligands, work with synthetic, modified peptides has allowed a greater understanding of ligand-receptor interactions. For example, the orexin-B analog [Ala¹¹,D-Leu¹⁵]orexin-B was generated by replacing L-leucine residues at positions 11 and 15. These particular substitutions resulted in an enhancement of selectivity toward the OX₂ receptor (compared with the OX₁ receptor) by approximately 400-fold (Asahi et al., 2003). In the same study, a systematic approach to residue replacement revealed that three leucine residues are important for OX-B's selectivity for the OX₂ receptor and determined the minimal peptide sequence required for orexin receptor activation.

In a recent patent disclosure, Yanagisawa (2010) reports a small-molecule agonist for OX₂ receptors. This represents the only published account of nonpeptidic orexin ligands to date. The chemical series reportedly induces a robust Ca²⁺ response in OX₂ receptor-expressing Chinese hamster ovary cells. There is one report of a small-molecule allosteric OX₂ receptor potentiator, a compound that binds to a site on the receptor other than the ligand-binding site and potentiates the response of

the receptor to its cognate ligand (Lee et al., 2010). This compound, *N*-benzyl-*N*-(3,4-dimethoxybenzyl)glycyl-*N*²-(1-phenylethyl)glycinamide (OBPt-9), was identified via a microarray-based, two-color, cell-binding screen and was shown to potentiate the response to orexin-A in both OX₁ receptor- and OX₂ receptor-expressing cells (Lee et al., 2010). Further characterization of these molecules should prove informative.

B. Dual Orexin Receptor Antagonists

Intensive efforts to identify orexin receptor antagonists began soon after the discovery of these excitatory neuropeptides and their receptors. At least four structurally distinct dual orexin receptor antagonists (DORAs) have entered human trials including almorexant [Actelion Pharmaceuticals, now GlaxoSmithKline (GSK), Brentford, Middlesex, UK], *N*-[[[(2*S*)-1-[[5-(4-fluorophenyl)-2-methyl-4-thiazolyl]carbonyl]-2-piperidinyl]methyl]-4-benzofurancarboxamide (SB-649868; GSK), suvorexant (Merck, Whitehouse Station, NJ), and MK-6096 (Merck). Others have reported the discovery and characterization of additional DORAs as well as selective OX₁ receptor and OX₂ receptor antagonists (SORAs) (Coleman and Renger, 2010). Both Actelion and Merck have reported clinical proof of concept for treating primary insomnia with their respective receptor antagonists.

In 2007, Actelion disclosed data for almorexant, a potent DORA, showing this compound to be effective in promoting sleep in preclinical species. When administered to rats, almorexant dose-dependently increased REM and NREM sleep, effects beginning within an hour of dosing and persisting for up to 12 h after treatment (Brisbare-Roch et al., 2007). With repeat dosing at 100 mg · kg⁻¹ · day⁻¹ in rats, no tolerance to the sleep effects were seen, and no rebound was observed upon discontinuation of drug treatment (Brisbare-Roch et al., 2008). It is noteworthy that when dosed during the inactive period, when endogenous levels of orexin are at their lowest, the compound had little effect beyond normal sleep (Brisbare-Roch et al., 2007). In dogs, 100 mg/kg almorexant significantly reduced mobility scores relative to vehicle-treated dogs; the subjects could be easily aroused by the presence of a familiar individual but quickly returned to sleep once the stimulus was withdrawn (Jenck et al., 2007). Cataplexy was not observed in either species at any dose (Brisbare-Roch et al., 2007).

In double-blind, placebo-controlled clinical studies, almorexant was well tolerated, with reports of somnolence, dizziness, disturbed attention, and fatigue at doses above 200 mg. The incidence of somnolence increased with dose over the range of 100 to 1000 mg. As in preclinical studies, no cataplexy-related side effects were reported (Hoever et al., 2010). In double-blind studies with zolpidem as an active control, almorexant increased sleep efficiency, reduced sleep latency, and increased total sleep time at doses greater than 200 mg in healthy volunteers. A phase II study in patients with

primary insomnia demonstrated the efficacy of both the 200- and 400-mg doses in improving sleep efficiency. Significant effects on secondary endpoints including latency to persistent sleep (LPS) and wake after sleep onset (WASO) were observed at the 400-mg dose (Dingemans et al., 2007). Actelion initiated a phase III study in adults with primary insomnia (RESTORA1) in 2007. In this study, almorexant met the primary endpoint of superiority compared with placebo on both objective and subjective measures of WASO. However, an undisclosed human tolerability issue resulted in termination of Phase III clinical development in January 2009 (Almorexant in adult subjects, NCT00608985, <http://www.clinicaltrials.gov>).

Researchers at GSK also discovered a series of piperidine-derived antagonists, including SB-649868 as potent DORAs. SB-649868 was reported to inhibit both OX₁ and OX₂ receptor activity and entered clinical trials in 2005. Preclinically, SB-649868 was shown to be sleep-promoting in rodent and primate studies (Di Fabio et al., 2011). In 2007, GSK announced that SB-649868 had advanced to phase II clinical trials. In initial single rising dose studies, SB-649868 was well tolerated and exhibited proportional increases in exposure across the dose range. After administration of SB-649868 to healthy volunteers, there were statistically significant improvements in total sleep time, reduced LPS, and WASO at both doses relative to placebo. Neither dose produced cognitive impairment the morning after evening drug administration (Bettica et al., 2009; Renzulli et al., 2011). Clinical studies, however, revealed that SB-649868 increased exposure of coadministered simvastatin in a drug-drug interaction study, consistent with the potent inhibition of CYP3A4 in vitro (Bettica et al., 2011). Phase II studies of SB-649868 were placed on clinical hold in late 2007 because of the emergence of a reported preclinical toxicity, and GSK entered into a collaborative agreement with Actelion in 2008 to codevelop almorexant and other potential back-up compounds.

Merck has developed a diverse portfolio of orexin receptor antagonists in several distinct structural classes (Coleman et al., 2011a). After completing a screening campaign to identify new leads, researchers at Merck disclosed proline bis-amides including DORA-1 as potent dual orexin receptor antagonists. Intracerebroventricular administration of orexin B to rats placed in a beam-break box produced significant increases in locomotor activity over several hours. When rats were pretreated with DORA-1 by intraperitoneal injection 30 min before neuropeptide administration, DORA-1 produced dose-dependent reductions in this locomotor activity relative to baseline (Bergman et al., 2008).

Merck has also reported *N,N*-disubstituted-1,4-diazepanes, including suvorexant (MK-4305) and DORA-12, to be potent dual orexin receptor antagonists. Both compounds are selective antagonists with excellent activity in cell-based assays (Cox et al., 2009, 2010). Suvorexant

inhibits orexin induced Ca²⁺ levels in cells expressing human OX₁ or OX₂ receptors with IC₅₀ values of 50 and 56 nM, respectively, but has >6000-fold selectivity against a panel of 170 receptors and enzymes. Preclinically, suvorexant is orally bioavailable, has good brain penetrance, and demonstrates orexin receptor occupancy in rat brain (Cox et al., 2010; Winrow et al., 2011). An example of the sleep-promoting effects of DORA-12 in mice is seen in Fig. 5A. Active phase treatment is associated with dose-dependent reductions in active wake and augmentation of NREM and REM sleep, effects that diminish abruptly with the onset of the animals' normal inactive phase. These DORA-12-induced changes are mediated through OX₁ and OX₂ receptors, because these effects are absent in genetically modified mice lacking these receptors (Fig. 5B). In other rodent sleep studies, suvorexant dose dependently reduced active wake and increased REM and NREM sleep when administered orally at 10, 30, and 100 mg/kg. Suvorexant was also highly efficacious in promoting sleep in canines and rhesus monkeys (Winrow et al., 2011). Based on these and other efficacy studies as well as a favorable pharmacokinetic and safety profile, suvorexant was selected to advance into clinical development.

Suvorexant was well tolerated in phase I studies, with peak plasma levels achieved at 1.5 to 4 h after dosing and a terminal plasma half-life of 8 to 14 h. In healthy volunteers, dose-dependent observations of somnolence were evident. Next-day residual sedation was not observed when suvorexant was administered at doses of 10 and 50 mg in the evening, whereas these same doses provided significant increases in overall sleep efficiency and reductions in WASO and LPS in a dose-dependent manner. Results from the phase IIb study demonstrated that suvorexant was superior to placebo in improving sleep efficiency on the first night of treatment as well as at the end of 4 weeks in patients with primary insomnia. These improvements in sleep efficiency were noted at all doses (10, 20, 40, and 80 mgs). Suvorexant also showed improvements in the secondary endpoints of reduced WASO at all doses and reduced LPS at 80 mg (Herring et al., 2010). In 2010, Merck announced that suvorexant had entered into phase III trials; it is currently the most advanced orexin antagonist in active clinical development.

Merck has disclosed an additional series of 2,5-disubstituted piperidine carboxamides including DORA-22 and MK-6096 as potent dual orexin receptor antagonists. The structure and preclinical pharmacology of MK-6096, a second dual orexin receptor antagonist in clinical development from Merck, has been disclosed (Coleman et al., 2012; Winrow et al., 2012). MK-6096 is structurally distinct from suvorexant and is highly efficacious in promoting sleep in rats (3–30 mg/kg) and dogs (0.25–0.5 mg/kg) (Winrow et al., 2012). MK-6096 was reported to have entered phase II clinical studies in 2009.

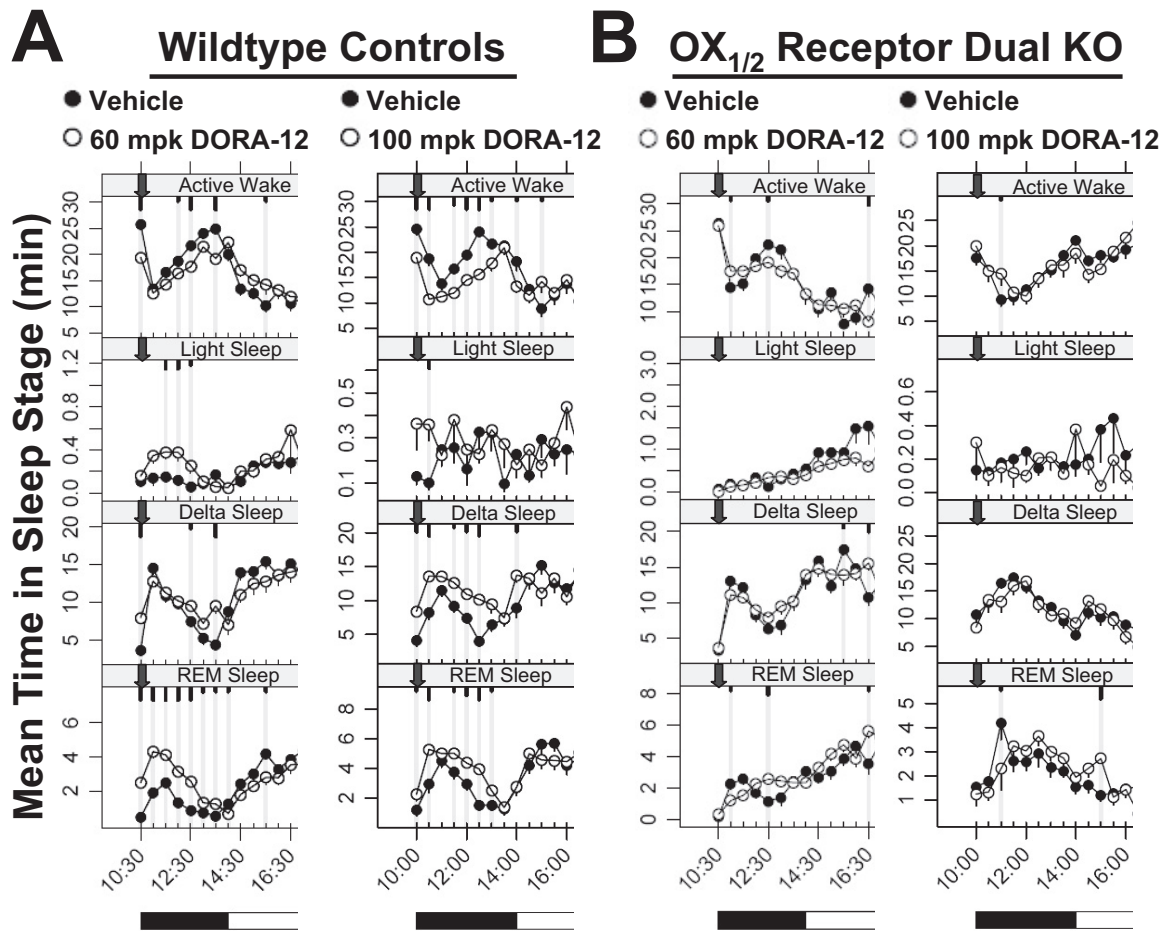


FIG. 5. The sleep-promoting effects of DORA-12 are absent in OX_{1/2} receptor double-knockouts. Electroencephalogram and electromyogram were monitored in wild-type (A) and age-matched mice with targeted ablation of both OX₁ and OX₂ receptors (B) by radiotelemetry to determine mean time spent in the indicated sleep stages as described previously (Winrow et al., 2012). At times indicated by arrows, vehicle [20% vitamin E TPGS (D- α -tocopheryl polyethylene glycol 1000 succinate), by mouth, closed symbols] or DORA-12 at 60 or 100 mg/kg (open symbols) were administered in a balanced 5-day crossover paradigm. Treatment occurred during the late active phase, approximately 4 h before the onset of the inactive phase (10:00/10:30 AM; zeitgeber time, 08:00/08:30; black arrows). Closed and open bars below each plot represent dark (active) and light (inactive) phases, respectively. Plotted points are the mean time spent in each sleep state during 30-min intervals after treatment over 5 days of consecutive treatment as determined by automated scoring and analysis as described previously (Winrow et al., 2012). Error bars (where visible) depicting the S.E.M. for each point are included, and time points exhibiting significant differences between vehicle and DORA-12 responses are indicated by gray vertical lines and tick marks (short, $p < 0.05$; medium, $p < 0.01$; long, $p < 0.001$; linear mixed effects model for repeated measures applied t test).

C. Orexin 2 Receptor-Selective Antagonists

Although dual orexin receptor antagonists have been shown to promote sleep in multiple species, SORAs have been used extensively to evaluate the relative role of each receptor subtype in the control of arousal and sleep, as well as other behaviors and physiology. An OX₂ SORA [1-(2,4-dibromophenyl)-3-[(4*S*,5*S*)-2,2-dimethyl-4-phenyl-1,3-dioxan-5-yl] urea (JNJ-10397049)] has been evaluated for sleep/wake effects, both alone and in conjunction with 1-(6,8-difluoro-2-methyl-quinolin-4-yl)-3-(4-dimethyl-amino-phenyl)-urea (SB-408124), an OX₁ SORA (McAtee et al., 2004; Dugovic et al., 2009). Both compounds are brain-penetrant, with JNJ-10397049 producing ~80% cortical OX₂ receptor occupancy for more than 6 h, an OX₂ receptor occupancy matched by almorexant at this dose. Sleep-promoting effects of the OX₂ SORA JNJ-10397049 at 30 mg/kg were due to lengthening sleep bouts, whereas treatment with almorexant was associated with sleep-

promoting effects marked by an increased number of REM and NREM sleep bouts. Both almorexant and JNJ-10397049 reduced histamine levels in the LH, whereas neither the histamine levels nor sleep parameters were affected by the relatively OX₁ receptor-selective SB-408124 (Dugovic et al., 2009).

Researchers at Roche have reported EMPA (*N*-ethyl-2-[(6-methoxy-pyridin-3-yl)-(toluene-2-sulfonyl)-amino]-*N*-pyridin-3-ylmethyl-acetamide) as a selective OX₂ SORA structurally distinct from JNJ-10397049 that exhibits greater affinity for human OX₂ relative to OX₁ receptors (approximately 900-fold selective) (Malherbe et al., 2009). Autoradiography using [³H]EMPA in rat brain slices shows high specific binding in hypothalamus, tuberomammillary nuclei, hippocampus, and nucleus accumbens. In vivo, EMPA dose-dependently reversed OX-B-induced hyperlocomotion in mice, achieving full reversal at a dose of 300 mg/kg i.p. It is noteworthy

that EMPA induced no psychomotor deficits when evaluated in a rat Rotorod assay at doses as high as 30 mg/kg i.p. (Malherbe et al., 2009). These results are consistent with earlier results reported for structurally distinct DORAs, including almorexant and SB-649868.

D. Orexin 1 Receptor-Selective Antagonists

The need for potent, selective preclinical research tools is highlighted by the wealth of studies using 1-(2-methylbenzoxazol-6-yl)-3-[1,5]naphthyridin-4-yl urea (SB-334867), described as a highly selective OX₁ SORA developed by SmithKline Beecham (now part of Glaxo-SmithKline). More than 160 papers have reported the use of SB-334867 as an OX₁ SORA in behavioral models evaluating addiction, feeding, sleep, and other behaviors. This compound antagonizes OX-A-induced [Ca²⁺]_i signal mediated by OX₁ receptors expressed on Chinese hamster ovary cells (K_B , 40 nM), and shows ~50-fold selectivity relative to OX₂ receptors evaluated in the same assay (K_B , 1995 nM) (Haynes et al., 2000; Smart et al., 2001) (Table 4). These results are consistent with binding affinities observed in our laboratories, which have found it to be ~45-fold selective for OX₁ over OX₂ receptors (K_i = 18 and 835 nM, respectively) (A. Gotter, P. Coleman, J. Renger, and C. Winrow, unpublished observations). However, examination of SB-334867 potency against a panel of 170 other enzymes and receptors showed significant interactions with at least seven other targets at concentrations less than 10 μ M. These included activities toward the adenosine A_{2A} receptor (K_i , 0.67 μ M), 5-HT_{2C} receptor (K_i , 1.2 μ M), monoamine transporter (K_i , 1.44 μ M), norepinephrine transporter (K_i , 1.58 μ M), adenosine transporter (K_i , 2.45 μ M), adenosine A₃ receptor (K_i , 3 μ M), and 5-HT_{2B} receptor (K_i , 3.47 μ M) (A. Gotter, P. Coleman, J. Renger, and C. Winrow, unpublished observations). The favorable pharmacokinetic properties and commercial availability of SB-334867 have made this a popular tool compound for studying orexin signaling in vivo, and there is the potential for substantial central nervous system exposure with this compound. For example, in rats administered 20 mg/kg i.p. (25% 2-hydroxypropyl β -cyclodextrin), we observed a sustained concentration of 3.4 μ M in plasma at both 30 minutes and 2 h after dosing. Given the selectivity profile of SB-334867, there exists a possibility for antagonism for not only OX₁ receptors but also OX₂ and several other targets at these doses. It is for these reasons that some caution should be exercised in interpretations of behavioral observations attributed to selective OX₁ receptor antagonism in studies using SB-334867 exclusively.

Two other OX₁ receptor-selective antagonists appearing in the literature are SB-408124 and 5-bromo-*N*-[(2*S*,5*S*)-1-(3-fluoro-2-methoxybenzoyl)-5-methylpiperidin-2-yl]methyl-pyridin-2-amine (GSK1059865). Disclosed in 2004 by SmithKline Beecham, SB-408124 has a published OX₁/OX₂ receptor selectivity of 63-fold (K_b , 22 versus 1405 nM

in Ca²⁺ mobilization assays), slightly better than SB-334867 (Langmead et al., 2004). Like SB-334867, however, SB-408124 exhibits significant activity toward other receptors, notably 5-HT_{2B} (0.32 μ M), dopamine D₁ (1.78 μ M), 5-HT_{2C} (1.88 μ M), adenosine A_{2A} (2.77 μ M), and α_{2b} -adrenergic receptors (3.29 μ M). SB-408124 has good pharmacokinetic properties, including bioavailability (~80%) and brain penetration (1.7%) in rats (A. Gotter, P. Coleman, J. Renger, and C. Winrow, unpublished observations). However, these favorable properties allow SB-408124 to reach levels that may also affect other receptors. GSK1059865, a recently identified OX₁-selective antagonist (Gozzi et al., 2011), may offer an improvement over SB compounds, as it has a reported OX₁/OX₂ receptor selectivity of ~79-fold (K_b , 1.6 versus 126 nM in IP₃ accumulation assays), with no significant activity at concentrations under 1 μ M toward a panel of 113 different receptors with the exception of the κ -opioid receptor (K_i , 320 nM) (A. Gotter, P. Coleman, J. Renger, and C. Winrow, unpublished observations). More selective OX₁ SORAs will undoubtedly be developed, but currently available reagents used in combination with DORAs, 2-SORAs, and genetic models have nonetheless provided valuable insight into the function of this receptor in the control of vigilance state and other orexin-mediated physiology and behavior.

IV. Orexin Neurons Are a Key Component of Pathways Regulating Sleep

The identification of mutant *hcrtr2* genes encoding truncated versions of OX₂ receptors responsible for genetically transmitted canine narcolepsy (Lin et al., 1999) and the description of the narcoleptic phenotype of *Hcrt* knockout mice 1 month later (Chemelli et al., 1999) set off extensive investigations into the mechanisms through which orexins promote arousal and control of vigilance state. These findings also led to focused efforts to develop small-molecule antagonists to probe the function of orexin receptors in sleep and to validate these receptors as targets for the development of pharmacological therapeutics.

A. Neurological Pathways Regulating Sleep

The identification of orexin neuropeptides and their cognate receptors has provided an understanding of the network of neuronal pathways involved in the interplay between sleep and arousal-promoting centers and how these areas interact to control vigilance state. The predominant sleep-promoting influence in the brain is provided by the ventrolateral preoptic (VLPO) and adjacent median preoptic areas. Neurons of the VLPO are most active during NREM sleep, partially active during REM sleep, and silent during wakefulness. They send inhibitory projections to arousal promoting areas, including the tuberomammillary nuclei (TMN), laterodorsal and pedunculopontine tegmental (LDT, PPT) nuclei, the locus ceruleus (LC), dorsal raphe (DR) nuclei, the ventral tegmental area (VTA) (Saper et al., 2005b, 2010; España

TABLE 4
Selectivity and potency of major orexin receptor antagonists

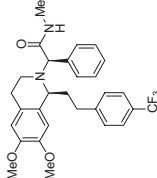
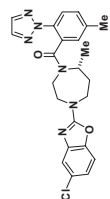
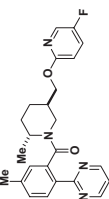
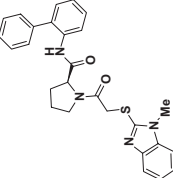
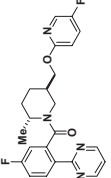
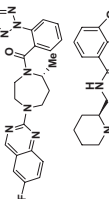
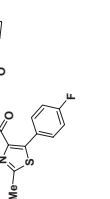
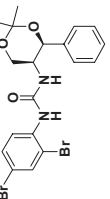
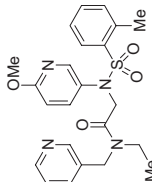
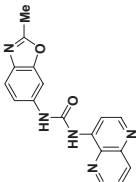
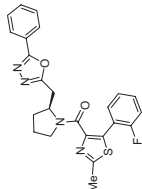
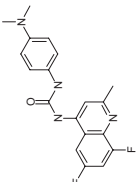
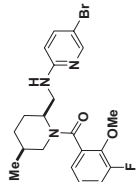
Antagonist	Structure	Potency			Selectivity			Origin	Reference	
		OX ₁ receptor		K _b	OX ₂ Receptor		OX ₁ Receptor			
		K _i	IC ₅₀		K _i	IC ₅₀				K _b
<i>nM</i>										
Dual Ox receptor antagonists										
Almorexant (Act-078573)		13			8			1.6×	Actelion	Brisbare-Roch et al., 2007
Suvorexant		0.55		50	0.35	56		0.9–1.6×	Merck	Coleman et al., 2010
MK-6096		2.5		11	0.31		11	1.0–8.1×	Merck	Coleman et al., 2012 Winrow et al., 2012
DORA-1		0.2	4		3	17		0.1–0.2×	Merck	Bergman et al., 2008
DORA-22		9.7	32		0.61		10	3.2–15×	Merck	Winrow et al., 2012
DORA-12		1.8	27		0.17	27		1.0–10.5×	Merck	Cox et al., 2010
SB-649868		0.76	0.32		1.3	0.4		0.6–0.8×	GlaxoSmithKline	Di Fabio et al., 2011
OX ₂ receptor antagonists										
JNJ-1037049		3162			5			~630×	Johnson & Johnson	McAtee et al., 2004

TABLE 4—Continued

Antagonist	Structure	Potency				Selectivity		Origin	Reference
		OX ₁ receptor		OX ₂ Receptor		OX ₂ Receptor	OX ₁ Receptor		
		K _i	IC ₅₀	K _b	K _i				
		<i>nM</i>							
EMPA		>900			1	>900×		Hoffmann-La Roche	Malherbe et al., 2009
OX ₁ receptor antagonists ^a									
SB-334867				40			~50×	GlaxoSmithKline	Smart et al., 2001
SB-674042				1			~130×	GlaxoSmithKline	Langmead et al., 2004
SB-408124				22			~64×	GlaxoSmithKline	Langmead et al., 2004
GSK1059865		1.6				126	~79×	GlaxoSmithKline	Gozzi et al., 2011

All values are determined in assays using recombinant human OX₁ or OX₂ receptors expressed in mammalian cell lines. K_i was determined in radioligand binding. IC₅₀ was determined from cell-based Ca²⁺ mobilization assay. K_b was determined by antagonism of orexin activation of either cellular Ca²⁺ mobilization or IP₃ signaling cellular assay.

^a Compounds are included here as OX₁ receptor antagonists based upon historical designations and at least partial selectivity for the OX₁ receptor.

and Scammell, 2011) and notably, orexinergic neurons of the LH (Sakurai et al., 2005b; Yoshida et al., 2006). This inhibitory influence is primarily mediated by GABAergic influences. In fact, potentiation of GABA signaling from the VLPO is thought to underlie the mechanism of action of currently marketed sleep agents (e.g., zolpidem and eszopiclone) (España and Scammell, 2011).

Orexin-secreting neurons of the LH project to brainstem nuclei involved in promoting arousal (Fig. 6). The activity of these nuclei, which include the TMN, LDT/PPT, DR, and LC, is dependent upon the balance of influence imposed by both inhibitory signals from the VLPO and excitatory ones provided by orexin. Ultimately, the integration of these signals determines arousal and behavioral state. A primary orexinergic projection is sent to the TMN, which preferentially express OX_2 over OX_1 receptors (Trivedi et al., 1998; Marcus et al., 2001). TMN neurons project broadly to the prefrontal cortex (PFC), thalamus, and other subcortical structures and are normally active during wake and progressively less active during NREM and REM sleep. They represent the primary source of histaminergic (HA) neurons in the brain, and HA receptor agonists and antagonists promote and attenuate arousal, respectively (Monti et al., 1986; Lin et al., 1988; Mochizuki and Scammell, 2003). Up-regulation of this histaminergic activity underlies the mechanism of a new class of wake-promoting drugs that inhibit histamine H_3 receptors to counter the reduced HA levels observed in narcoleptics (Lin et al., 2008; España and Scammell, 2011).

Orexin neurons also send projections to cholinergic tegmental nuclei (the LDT and PPT) as well as noradrenergic LC and serotonergic DR neurons. OX_1 receptors are preferentially expressed in the LC, whereas both orexin receptors are detectable in LDT, PPT, and DR

(Fig. 6) (Trivedi et al., 1998; Marcus et al., 2001). Along with arousal-promoting influences, these brainstem regions are also responsible for gating between vigilance states, particularly in and out of REM sleep. Orexin influences on neurons of the LDT and PPT also regulate muscular atonia that accompanies REM sleep, through both direct and indirect effects on ventromedial neurons of the medulla, which in turn inhibit spinal motor neurons through GABA projections. More extensive discussions of these mechanisms regulating vigilance state and sleep-dependent motor activity have been reviewed elsewhere (Saper et al., 2010; España and Scammell, 2011; Scammell and Winrow, 2011).

B. Orexin-A and -B Promote Arousal and Modulate Vigilance State

Orexins provide an arousal signal that is both necessary and sufficient for normal sleep/wake regulation. Over the course of the 24-h circadian cycle, changes in arousal match oscillating levels of orexin. In nocturnal animal models, orexin levels rise over the night-time hours, peaking late in the active phase, whereas in primates, OX-A levels in CSF accumulate over daytime hours, peaking just before the dark phase (Taheri et al., 2000; Zeitzer et al., 2003). OX-A applied exogenously (via intracerebroventricular injection) to rats results in increased locomotor activity, grooming and wakefulness, whereas the mean time spent in NREM and REM sleep is diminished. These effects are greatest when OX-A is administered during the inactive phase when endogenous orexin levels are at their lowest and barely detectable when applied during periods of wakefulness, as might be expected of an arousal signal (Hagan et al., 1999; Piper et al., 2000). Exogenously applied OX-A also promotes arousal in mice with selected genetic ablation of orexinergic neurons. In this case, the levels of wakefulness and NREM and REM suppression in mutant animals exceed those seen in wild-type animals treated identically (Mieda et al., 2004), indicating that downstream orexin signaling components, including orexin receptors, are intact and are up-regulated in these mutants and that the neuropeptide alone is sufficient as a wakefulness signal. Similar results are seen in narcoleptic dogs where OX-A rescues the cataplexy and hypersomnolence phenotype of an animal harboring a mutation in the *Hcrt* gene encoding prepro-orexin but has no effect in dogs with mutations in the gene for the OX_2 receptor (Fujiki et al., 2003). Artificial activation of orexin-secreting neurons is sufficient to drive arousal. In an elegant set of experiments, Adamantidis et al. (2007) and later Carter et al. (2009) used mice in which the expression of channelrhodopsin-2 was driven by the *Hcrt* promoter such that orexin-containing neurons could be photically stimulated by a fiber optic means. Activation of orexigenic neurons in the LH of these mice induced transitions from NREM or REM sleep into wakefulness (Adamantidis et al., 2007), but in the face of increased

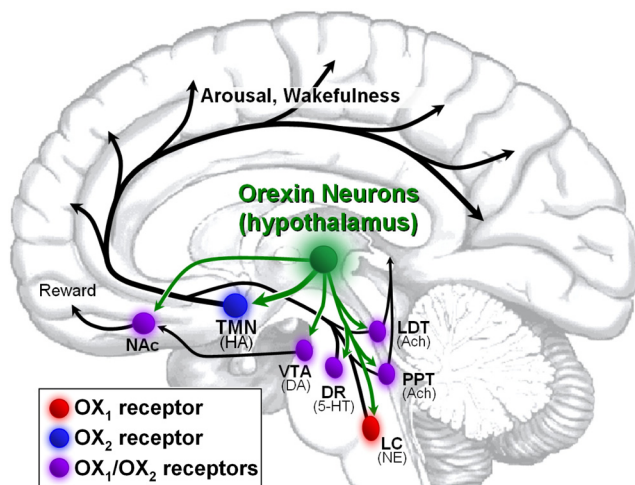


FIG. 6. Orexin and OX_R efferent pathways associated with arousal, vigilance state, and reward pathways. NAc, nucleus accumbens; HA, histaminergic; DA, dopaminergic; ACh, cholinergic; NE, noradrenergic; 5-HT, serotonergic. Green, orexinergic neuron projections; red, preferential OX_1 receptor expression; blue, preferential OX_2 receptor expression; violet, both OX_1 and OX_2 receptor expression.

sleep pressure induced by sleep deprivation, these wake-promoting effects as well as orexin-dependent activation of TMN and LC neurons were diminished, indicating that homeostatic influences converge downstream of orexin neuron activation (Carter et al., 2009).

C. Modulation of Orexin Signaling

In general, the timing and extent of sleep and wakefulness is driven by two different influences; the circadian clock that aligns the timing of an organism's physiology and behavior with daily environmental timing cues, and the homeostatic drive for sleep aligning the restorative properties of rest with physiological and energy needs. The endogenous circadian pacemaker resides in the suprachiasmatic nuclei (SCN) of the hypothalamus, and controls daily cycles of arousal, locomotor activity, gut motility, and the timing of sleep even in the absence of external cues such as ambient lighting changes (Huang et al., 2011). When grown in culture, neurons of the SCN in themselves have periods of electrical activity of ~24 h (Welsh et al., 1995). The activity of the SCN is primarily entrained to the environment by ambient lighting cues (Klein et al., 1991). The output from the SCN to the sleep network is conveyed through the ventral subparaventricular zone to the dorsomedial nucleus of the hypothalamus (DMH). From here the DMH projects to VLPO neurons via inhibitory GABA projections and sends excitatory connections to orexin-containing neurons of the LH (Saper et al., 2005b). The DMH, which is largely active during wakefulness (Saper et al., 2005a), also receives appetite, feeding, and body temperature inputs, such that some level of homeostatic regulation may occur at this level (Saper et al., 2005b). Together, the DMH and the VLPO coordinate the activity of orexin neurons to regulate arousal aligned with circadian environmental cues.

The specific mechanism through which the homeostatic drive for sleep is mediated is less clear, however, but is thought to be associated with energy homeostasis and the need to conserve metabolic energy. The need for sleep accumulates with wakefulness such that the pressure for sleep accumulates with sleep deprivation. Evidence for orexin neurons being an integration site for homeostatic influences are the observations that orexin cell firing is increased by direct acetylcholine, glutamate, and ghrelin application and decreased by leptin, glucose, norepinephrine, 5-HT, and GABA (Burdakov, 2004). One attractive candidate for this regulation is adenosine, because during prolonged wakefulness, ATP is degraded to ADP, AMP, and eventually adenosine, which accumulates in parts of the brain (Porkka-Heiskanen et al., 2000). Orexin neurons express adenosine A₁ receptors, and application of the adenosine A₁R antagonist 1,3-dipropyl-8-phenylxanthine into the LH increases wake and suppresses both REM and NREM sleep (Thakkar et al., 2008). Microinfusion of the adenosine A_{2A} agonist 2-[p-(2-carboxyethyl)phenyl-ethyl-

amino]-5'-N-ethylcarboxamidoadenosine (CGS21680) into the ventral striatum both promotes sleep measures and attenuates c-Fos production in orexin-producing cells, suggesting not only that adenosine's sleep promoting effect may be facilitated by reduced orexinergic cell firing but also that orexin signaling is not necessarily the exclusive mechanism for adenosine-mediated sleep promotion (Sato et al., 2006).

Corticotropin-releasing factor (CRF) and dopamine also seem to modulate the influence of orexin on arousal. CRF signaling in response to stress represents a possible interaction with orexin signaling and sleep, because elevated levels of the hormone are associated with arousal, and intracerebroventricular administration of CRF induces wakefulness and locomotor activity (Sakurai and Mieda, 2011). Any interaction with CRF signaling, however, appears to be upstream of orexin, because orexin-induced arousal persists in CRF receptor knockouts as well as in the presence of CRF receptor antagonists (Fenzl et al., 2011). Evidence for the modulation of orexin signaling in arousal and vigilance state by dopamine has also recently emerged (Sakurai et al., 2010; Sakurai and Mieda, 2011). In orexin peptide knockout mice, pharmacological D₁ receptor activation decreases the prevalence of sleep attacks relative to wild-type animals. Conversely, the hypersomnolence of these animals is exacerbated with D₁ antagonism, whereas D₂ receptor modulation has little to no effect on arousal (Burgess et al., 2010). Cataplectic attacks, however, are affected by D₂ receptor activity; pharmacological D₂ activation and inhibition are associated with substantial increases and decreases, respectively, in behavioral arrest (Burgess et al., 2010). These studies further illustrate the existence of distinct pathways for arousal control and the regulation of vigilance state as well as the dopamine receptor subtype involvement in each.

V. Genetic and Pharmacological Dissection of Orexin-Mediated Arousal

Our current understanding of the role of specific orexin signaling components in arousal and vigilance state is based on both pharmacological manipulations using orexin receptor modulators and the evaluation of animals mutant for orexin receptors, the *Hcrt* gene encoding the prepro-orexin precursor and targeted ablation of orexinergic neurons. Combining the interpretations of both approaches has proven invaluable toward uncovering the role of orexin and its cognate receptors in the control of sleep and arousal.

A. Canine Narcolepsy

In dogs, narcolepsy is manifested as active-phase sleep attacks and, during the inactive phase, as short sleep latency and fragmented sleep with rapid EEG/polysomnographic transitions between sleep stages. In severe cases, cataplectic attacks are characterized by atonia with hind-limb buckling during which the animal

retains consciousness, often with open eyes that are able to follow visual stimuli. Consciousness may give way to sleep, typically REM. These episodes are typically triggered by palatable food presentation or play, much as positive emotions do in human patients (Nishino, 2005).

Canine narcolepsy is classified into two forms: familial (genetic), associated with mutations in the OX_2 receptor gene, and sporadic, which is typically associated with loss of orexin-secreting neurons. The genetic form is transmitted in an autosomal-recessive fashion with complete penetrance. To date, OX_2 receptor mutations include truncations within transmembrane region five or just after transmembrane region six in Doberman pinschers and Labrador retrievers (Lin et al., 1999) and a point mutation in a dachshund family resulting in a glutamate to lysine change at amino acid position 54, rendering a receptor incapable of being bound and activated by OX-A or OX-B (Hungs et al., 2001). CSF and brain levels of OX-A and OX-B are normal in these mutants, and orexin-containing neurons are normal in appearance (Ripley et al., 2001). Although exogenously applied OX-A can reverse narcoleptic symptoms in sporadic narcoleptic dogs deficient in orexin neurons, high doses of intracerebroventricular and intravenous administration of OX-A have no effect on arousal or cataplectic symptoms of OX_2 receptor mutant narcoleptic Doberman pinschers (Fujiki et al., 2003). Sporadic canine narcolepsy (poodles, beagles, dachshunds, collies, fox terriers) does not seem to be associated with a single, highly penetrant gene mutation but is typically associated with symptoms more pervasive than the genetic form (Nishino, 2005). Unlike humans in which a leukocyte antigen gene is associated with sporadic narcolepsy, a canine allele specifically associated with the disorder remains elusive (Peyron et al., 2000; Nishino, 2005; De la Herrán-Arita et al., 2011).

B. Genetically Engineered Mouse and Rat Models

Genetic manipulation of rats and mice has been used to mimic the clinical etiology of narcolepsy associated with orexinergic neuron loss and to dissect the role of individual orexin signaling components. Efforts have included transgenic models in which the expression of a cellular toxic transgene is used to specifically ablate orexin-producing neurons to imitate the autoimmune loss of these neurons in patients with narcolepsy. Genetic models also include targeted gene knockouts of the *Hcrt* gene encoding the prepro-orexin peptide processed into OX-A and OX-B ligands (orexin peptide knockouts), the *Hcrtr1* and *Hcrtr2* genes encoding OX_1 and OX_2 receptors (OX_1 receptor and OX_2 receptor knockouts), and double receptor mutant mice lacking both of these receptors ($OX_{1/2}$ receptor double knockouts). Together, with pharmacological manipulation using selective receptor agonists and antagonists described in sections III and IV, these mutant animals have provided invaluable information regarding not only the role of orexin signal-

ing in narcolepsy/cataplexy but also the general neuronal pathways regulating sleep and vigilance state (De la Herrán-Arita et al., 2011). Both approaches uncovered the importance of each of these signaling components in arousal, vigilance state, and the regulation of sleep. Overall, the importance of these signaling components to arousal may be summarized as follows: Ox neurons > orexin peptide KOs \geq $OX_{1/2}$ receptor double KO > OX_2 receptor KOs \gg OX_1 receptor KOs. It should be noted, however, that in many cases, these animal models have been evaluated by different laboratories using different methods (e.g., EEG versus visual observation) often using different criteria to make interpretations regarding the impact of a given mutation on arousal and/or cataplectic behavior. Efforts have been made, however, to standardize the criteria for the cataplectic behavior in mice. Based on observations from orexin peptide knockouts, the following criteria have been established on behalf of the International Working Group on Rodent Models of Narcolepsy (Chemelli et al., 1999; Willie et al., 2003; Scammell et al., 2009): 1) abrupt atonia lasting over 10 s, 2) animal immobility during the episode, 3) quantitative EEG (qEEG) dominated by θ activity during the episode, and 4) behavioral arrest preceded by a period of active wake lasting > 40 s. In addition, cataplectic behavior should be reversible, with administration of clomipramine or other anticataplectic (e.g., monoamine reuptake inhibitors such as desipramine) (Willie et al., 2003; Scammell et al., 2009). Nevertheless, this expression of the importance of these components on arousal can be concluded on the basis of the studies discussing these genetic models below, along with what has been determined from both dogs and humans.

C. Orexin Neurons Are Critical for Both Arousal and Vigilance State Gating

In theory, the best preclinical models for the majority of human cases exhibiting narcolepsy with cataplexy are transgenic mice and rats lacking orexin-producing neurons. These animals (Ox/Atx mice and rats) express an apoptotic Ataxin-3 transgene whose expression is driven by the human *Hcrt* gene promoter, resulting in programmed cell death of orexin-containing neurons while leaving neighboring neurons, including melanin-concentrating hormone-containing neurons of the LH, intact (Hara et al., 2001; Beuckmann et al., 2004). Despite incomplete loss of orexin neurons in hemizygous animals, OX-A levels within CSF, cortex, and brainstem are as much as 100-fold lower than that of wild-type animals (Zhang et al., 2007). The phenotype of Ox/Atx mice and rats resembles that seen in human patients with narcolepsy/cataplexy active-phase hypersomnolence, behavioral arrest/tonia episodes triggered by excited ambulation or grooming, and frequent wake-to-REM transitions never observed in wild-type animals that often enter REM through an intervening NREM transition (wake to NREM to REM). Inactive phase sleep is fragmented with reduced

REM latency and frequent, but short bouts of wake and NREM with transitions occurring rapidly (Hara et al., 2001; Beuckmann et al., 2004). Remarkably, these animals remain sensitive to ectopically expressed or exogenously administered OX-A and OX-B, which reverse behavioral arrests and REM abnormalities, including wake-to-REM transitions of Ox/Atx mice, in some cases to greater effect than in wild-type mice (Mieda et al., 2004). One divergence from the accepted criteria for cataplexy, however, is the presence of quantitative EEG spectral pattern resembling wake during episodes of behavioral arrest in Ox/Atx rats (Beuckmann et al., 2004), as opposed to θ or REM activity characteristic of cataplexy. It remains to be seen whether this is a species-dependent divergence from mice or if more complete loss of orexin neurons in homozygous animals may affect these animals even more profoundly.

D. The Role of the Hcrt Gene and Hcrt Gene Product

Mice with a targeted deletion of the *Hcrt* gene encoding the prepro-orexin precursor peptide display a narcolepsy phenotype similar to transgenic Ox/Atx animals, with cataplectic episodes satisfying the criteria described above (Chemelli et al., 1999; Fujiki et al., 2009; Scammell et al., 2009). Furthermore, the abrupt behavioral arrests in orexin peptide knockouts are reversed by clomipramine, whereas caffeine only increased wakefulness and actually exacerbated cataplectic symptoms, demonstrating the specificity of this treatment (Willie et al., 2003). Although orexin peptide knockouts clearly exhibit cataplexy, their phenotype is not as pervasive as that observed in Ox/Atx animals. Direct comparisons revealed an even greater number of vigilance state transitions and time spent in REM for Ox/Atx animals (Kantor et al., 2009). These results indicate that orexin-containing neurons provide additional signals beyond orexin itself, perhaps glutamate or dynorphin, which can contribute to narcoleptic symptoms (Sakurai et al., 2005a; Kantor et al., 2009). Modeling of orexin neuron activity suggests that dynorphin is capable of modulating orexin responses, delaying its arousal effects at the sleep/wake transition by affecting the sensitization and firing rate of orexin-sensitive neurons (Williams and Behn, 2011). Nevertheless, orexin peptide knockout animals have provided a useful model in which to study the cataplectic episodes in detail, where both scheduled palatable food and running wheel presentation effectively increase the frequency of cataplectic episodes, presumably mimicking the effect of positive emotions in human patients (España et al., 2007; Clark et al., 2009). Consistent with this observation is the finding that both positive and negative olfactory stimuli (female and coyote urine) are also sufficient to induce narcoleptic episodes in male orexin peptide knockout mice (Morawska et al., 2011). These behavioral patterns indicate that orexin peptide knockouts are a particularly good model for early onset narcolepsy with cataplexy. Mutation of the human

HCRT gene affecting peptide trafficking and processing is known to be associated with severe cataplexy (5–20 episodes/day) (Peyron et al., 2000).

E. Role of OX₂ Receptors in the Control of Arousal, Vigilance State

Arousal responses to orexin are primarily mediated by OX₂ receptors. Intracerebroventricular administration of OX-A, OX-B, or [Ala¹¹]OX-B, a modified signaling peptide having 120-fold selectivity for OX₂ receptors, similarly promote wakefulness and decrease the amount of time spent in REM and slow-wave sleep in a dose-dependent manner in rats (Hagan et al., 1999; Piper et al., 2000; Akanmu and Honda, 2005). Conversely, OX₂ receptor-selective antagonists have sleep-promoting effects similar to antagonists having equal potencies for both OX₁ and OX₂ receptors, including attenuated active wake, increased REM and NREM sleep, and decreased latencies to NREM and REM (Dugovic et al., 2008). As might be expected, OX₂ receptor knockouts display a narcoleptic phenotype similar to that of orexin peptide knockouts, including hypersomnolence, fragmented wakefulness and NREM sleep, increased active phase NREM sleep, and limited wake-to-REM transitions (Willie et al., 2003). The behavioral arrests exhibited by these mice, however, fall short of the cataplexy criteria established by Scammell et al. (2009). Although clomipramine reverses behavioral arrests exhibited by OX₂ receptor knockouts, these episodes are far less frequent than those observed in *Hcrt* knockouts, are typically more gradual in their onset, and are preceded by quiet wakefulness rather than “emotive” behaviors such as grooming or climbing that precede abrupt behavioral arrests. These gradual arrests are characterized by EEG power spectra similar to that of NREM sleep, as opposed to θ -rich REM sleep typical of abrupt cataplectic episodes (Willie et al., 2003). As such, the behavioral arrests displayed by OX₂ receptor knockouts do not seem cataplectic in nature but suggest that other signaling mechanisms may be required for these episodes.

F. Role of Orexin 1 Receptor in the Control of Vigilance State

As the only other known orexin receptor, the incomplete phenotype of OX₂ receptor knockouts relative to orexin peptide knockout animals indicates that the OX₁ receptor does participate in the mechanism of cataplexy. However, constitutive OX₁ receptor mutants reportedly display only a mild sleep phenotype with some increase in fragmentation (Kisanuki et al., 2000). Likewise, OX₁ receptor-selective antagonism with compounds of imperfect selectivity elicits little to no effect on sleep architecture (Dugovic et al., 2008) but has been reported to both increase extracellular dopamine in the PFC and to attenuate the sleep promoting effects of OX₂ receptor antagonism (Dugovic et al., 2009). Given its expression on locus ceruleus neurons involved in the control of REM

and its activating effect on those neurons, a role for OX_1 receptor in gating transitions into REM remains possible (Bourgin et al., 2000; Ohno and Sakurai, 2008). In support of this assertion is the observation that small interfering RNA-mediated knockdown of OX_1 receptor expression in locus ceruleus is associated with inappropriate increases in REM sleep during the active period of rats for up to 4 days after treatment, a time-course coincident with reduced OX_1 receptor mRNA levels. Remarkably, neither wakefulness, NREM sleep, nor the qEEG power spectra of treated animals was affected, suggesting that this effect was specific for vigilance state gating and not a general effect on arousal (Chen et al., 2010).

G. Mechanisms Underlying Narcolepsy/Cataplexy

1. Hypersomnolence. Disruption in the arousal effects of orexin are clearly mediated through deficiencies in OX_2 receptor activity, most likely through histaminergic neurons of the tuberomammillary TMN. In fact, adenovirus-mediated focal expression of OX_2 receptors selectively within the TMN of OX_2 receptor knockout mice is sufficient to rescue the arousal deficits of these narcoleptic mutants, whereas the sleep fragmentation phenotype was unaffected, indicating that the control of vigilance state gating by orexin resides in brain nuclei exclusive to the TMN (Mochizuki et al., 2011). The hypersomnolence, sleep attacks, and decreased latency to NREM and REM sleep displayed by both OX_2 receptor knockout mice (Willie et al., 2001) and OX_2 receptor mutant dogs with genetically transmitted narcolepsy (Lin et al., 1999) are clear illustrations of this receptor's role in disrupted arousal pathways. The sleep promoting effects of selective OX_2 receptor antagonism is associated with attenuated extracellular histamine levels in the LH (Dugovic et al., 2009), whereas histamine H_1 receptor blockade with pyrilamine blocks arousal induced by OX-A (Yamanaka et al., 2002).

Dopamine also seems to modulate orexin-mediated arousal and the prevalence of hypersomnia in narcoleptic models. In orexin peptide knockouts, sleep attack prevalence was decreased with pharmacological D_1 receptor activation and increased with inhibition of these receptors, whereas D_2 receptor modulation had little to no effect (Burgess et al., 2010). These findings together with the observation that OX_1 receptor antagonism has the potential to elevate PFC dopamine levels and attenuate OX_2 receptor antagonist-induced sleep (Dugovic et al., 2009) suggests that dopaminergic signaling has the potential to modulate hypersomnolence associated with diminished OX_2 receptor activity.

2. Sleep Stage Instability. A symptom common to all of the model organisms in which components of orexin signaling are disrupted is sleep fragmentation associated with vigilance state instability (Saper et al., 2010). OX_1 receptor knockouts exhibit mild sleep fragmentation whereas OX_2 receptor knockouts have intermediate

instability approaching that of orexin peptide knockouts or Ox/Atx transgenic animals. In the latter cases, the disruptions are more pronounced with rapid transitions between states and inappropriate wake-to-REM transitions. Orexin peptide knockout animals have also been evaluated in constant dark conditions to examine both the circadian control of sleep and sleep architecture in the absence of the masking effects of light. Although the circadian timing of sleep/wake cycles was normal, the absence of light/dark arousal cues revealed unusually rapid and random transitions between sleep states, including wake to REM and short duration bout time, suggesting behavioral state instability with a low threshold for transition (Mochizuki et al., 2004). Using a state space analysis technique, Diniz Behn et al. (2010) analyzed high-resolution quantitative EEG spectra in relation to EEG/EMG polysomnographic state of orexin peptide knockout animals to track the rate of movement between sleep states on a second-by-second basis. Although most state transitions were normal relative to wild-type animals, orexin peptide knockouts seemed to experience less stability such that drifting out of wake occurred more readily, ultimately explaining the sleep fragmentation described previously. Overall, Ox mutants spent more time near the wake-to-NREM transition boundary and less time in deep NREM or θ -rich wake than wild-type counterparts. During cataplectic episodes, orexin peptide knockouts also exhibited greater θ activity than that typically observed in REM (Diniz Behn et al., 2010).

3. Cataplexy. Many of the characteristics of cataplexy indicate that these episodes are related to a REM-like intrusion into wakefulness. Bilateral muscle atonia and a prevalence of θ qEEG power along with the circumstantial propensity for REM sleep and the dysregulation of vigilance state boundary control associated with narcolepsy substantiate this assertion (Beuckmann and Yanagisawa, 2002). However, cataplexy may not be as simple as an intrusion of REM sleep into wakefulness. At the cellular level, histaminergic neurons normally quiescent during both REM and NREM sleep are active during cataplectic episodes (John et al., 2004), which is consistent with the observation that canines and human patients maintain consciousness and are aware of their surroundings (Siegel and Boehmer, 2006). Cholinergic neurons of pedunculopontine nuclei that are normally active during wake and REM have attenuated activity during cataplexy, further indicating that this state may be distinct from REM (Thankachan et al., 2009). Muscarinic acetylcholine-mediated signaling from pedunculopontine nuclei neurons also seems to be involved, because pharmacologically induced acetylcholine activity in this region is associated with increases in behavioral arrest number without affecting mean arrest time in narcoleptic OX_1 and OX_2 receptor double-knockout animals (Kalogiannis et al., 2010), although the EEG characteristics of behavioral arrest episodes in these animals remain to be determined. Dopamine also seems to influ-

ence the prevalence of cataplectic attacks. Unlike hypersomnolence, which is attenuated by D₁ receptor subtype activation, cataplectic attacks in orexin peptide knockouts were substantially increased with D₂ activation and attenuated with antagonism of this receptor (Burgess et al., 2010).

In Ox/Atx mice, the prevalence of behavioral arrest episodes was attenuated by adenoassociated virus-mediated expression of prepro-orexin selectively in the zona incerta region of the hypothalamus (Liu et al., 2011). Remarkably, this ectopic orexin expression was not associated with a reduction in hypersomnolence, sleep attacks, or any detectable changes in sleep architecture, indicating that activity of the zona incerta has a specific role in stabilizing motor tone that is typically disrupted in cataplectic attacks. Retrograde tracer mapping found these neurons to receive inputs from the amygdala and to innervate the locus ceruleus, suggesting a possible pathway connecting emotional state with muscle paralysis associated with REM sleep (Liu et al., 2011). Together, these studies begin to round out our understanding of the role of orexin signaling in narcolepsy/cataplexy and, in so doing, its function in arousal and vigilance state.

H. Genetic versus Pharmacological Manipulation: Complementary Interpretations

Independently, experiments using mutant animals and pharmacological manipulation of orexin receptor activity are largely complementary but have distinct caveats associated with these approaches. These differences stem from the constitutive nature of gene knockouts relative to acute transient pharmacological manipulation. Constitutive gene knockouts and Ox/ATX transgenics more closely mimic narcolepsy and varying aspects of the disorder than what might be expected from pharmacological manipulation. For example, behavioral arrests have been observed in *Hcrt* and OX₂ receptor knockouts (Chemelli et al., 1999; Willie et al., 2003) and in mice in which both orexin receptors have been mutated (Kalogiannis et al., 2011). Although it is currently unclear whether the behavioral episodes observed in the later model constitute cataplexy (Scammell et al., 2009), cataplectic episodes have not been observed in response to even high doses of dual orexin receptor antagonists across multiple species (Brisbare-Roch et al., 2007).

The fundamental difference between constitutive genetic manipulation and pharmacological perturbation is the likely development of molecular, cellular, neuronal, and/or behavioral mechanisms that compensate for the loss of a targeted gene product or orexin neuron loss. Indeed, the hypersomnolence exhibited by animals with a targeted disruption of the *Hcrt* gene is still punctuated by periods of wakefulness (Chemelli et al., 1999), and pharmacological orexin receptor antagonism is capable of acutely promoting sleep in wild-type animals to levels exceeding that seen in untreated mutant mice lacking

both receptors (Winrow et al., 2012). Because of the likelihood of compensatory mechanisms, interpretations regarding the function of a gene product from genetically manipulated animal models should be made with this caveat in mind. In this regard, the availability of specific reagents targeting orexin receptors provides useful tools to specifically explore the function of orexin signaling within intact animals. Still, the interpretation of pharmacological perturbations depends not only on reagent specificity but also on the time of day at which they are applied, given oscillating endogenous orexin levels. Because OX-A and OX-B levels are highest during waking periods and reach a nadir during the inactive phase (Taheri et al., 2000; Zeitzer et al., 2003), the effectiveness of orexin receptor antagonists is expected to be greatest during periods of behavioral activity, a prediction confirmed experimentally (Brisbare-Roch et al., 2007; Li and Nattie, 2010; Winrow et al., 2011).

Further work combining genetic and pharmacological approaches will be particularly valuable. The most obvious experiments will examine the effectiveness of potential narcolepsy therapeutics in these knockout and Ox/ATX transgenic models of the disorder. These are anticipated to include both orexin agonists and other wake-promoting compounds such as those modulating histamine. Other studies have found orexin receptor knockouts to lack the response to orexin receptor antagonists, demonstrating their selectivity (Winrow et al., 2011, 2012). The use of small-molecule reagents specific for other neuronal targets in these animal models also has the potential to more clearly define the role of orexin in additional neurotransmitter pathways involved in modulating neurophysiology and behavior.

VI. Orexin Function beyond Sleep and Arousal

A. Central Modulation of Behavior and Physiology by Orexin Signaling

1. Feeding. The name orexin was ascribed by Sakurai et al. (1998) in their initial report on the behavioral effects of administration of the neuropeptide in rodents to reflect the peptide's apparent effects on feeding. In these animals treated with synthetic orexin peptide, increased wakefulness accompanied by feeding behavior was interpreted to reflect an increase in appetite. It is noteworthy that many of the patents covering orexin receptor antagonists indicate that these molecules may have potential therapeutic applications for obesity and metabolic disorders. Early efforts included the characterization of orexin receptor expression and screening for orexin receptor antagonists with the aim of identifying new drugs for treating metabolic disorders (Alvaro et al., 2009, 2011; Liu, 2009). Subsequently, the focus of many investigators shifted to the impact of orexin receptor modulation for regulating sleep and wake using small-molecule antagonists. Although the wake-promoting effects of orexin and the sleep-promoting effects of

orexin receptor antagonists have been widely documented, the role of this neurotransmitter in modulating feeding behavior is less established.

In multiple rodent studies, administration of exogenous OX-A has been demonstrated to increase food intake, which is frequently associated with elevated blood glucose levels. On the other hand, pretreatment with the partial OX₁ SORA, SB-334867, blocks the effects of OX-A on food intake (Haynes et al., 2000; Rodgers et al., 2000, 2001; Yamada et al., 2000; Thorpe and Kotz, 2005; Thorpe et al., 2005; Yi et al., 2009). The effect of OX-A on food intake, however, is diminished or absent in aged rats, and Western blots suggest that decreased OX₁ receptor levels may be responsible for the diminished feeding response (Takano et al., 2004; Kotz et al., 2005). It is noteworthy that acute treatment with SB-334867 or an anti-orexin antibody reduced levels of natural feeding, often leading to a reduction in body weight (Haynes et al., 2000, 2002; Yamada et al., 2000; Rodgers et al., 2001; Ishii et al., 2004, 2005). SB-334867 has further been observed to have anorectic action and to accelerate satiety (Haynes et al., 2000; Rodgers et al., 2001; Ishii et al., 2005). Together, these studies have suggested that orexin receptor antagonists may be useful for the treatment of obesity and eating disorders.

Gene expression studies suggest that orexin levels may predetermine eating preferences as well as respond to eating habits. For example, baseline orexin expression in the perifornical LH is higher in rats prone to overeating a high-fat diet (Morganstern et al., 2010), and expression stimulated by a high-fat diet was closely associated with elevated triglycerides (Wortley et al., 2003). Another study showed that food deprivation induced mRNA expression of transcripts encoding prepro-orexin, OX₁, and OX₂ receptors in the hypothalamus. In addition, food deprivation led to changes in the G-protein coupling with orexin receptors, altering the relative levels of coupling to G_q, G_s, G_o, and G_i (Karteris et al., 2005). Interactions between the orexin pathway and NPY, nitric oxide, serotonin, acetylcholine, and GABA signaling mechanisms have been implicated (Dube et al., 2000; Niimi et al., 2001; Orlando et al., 2001; Farr et al., 2005; Thorpe et al., 2006; Frederick-Duus et al., 2007). Most orexin neurons are glucose-sensitive, and their activity can be modulated by the peptide hormones leptin and ghrelin (Muroya et al., 2001; Yamanaka et al., 2003; González et al., 2008; Louis et al., 2010).

Multiple studies point to the involvement of orexin signaling in reward-based feeding. Administration of orexin has been shown to affect multiple reward-seeking behaviors, including operant responding for high-fat pellets and sucrose, conditioned place preference (CPP) for food, cue-induced reinstatement of extinguished sucrose-seeking, and food-reinforced responding under both variable and progressive ratio schedules of reinforcement (Harris et al., 2005; Cason et al., 2010; Sharf et al., 2010). These studies use the partial OX₁ SORA SB-334867 to

suggest the involvement of OX₁ receptors in reward-based feeding. Studies also suggest orexin's involvement in both dieting and overconsumption of palatable foods, once again suggesting a link to the treatment of obesity. During dieting however, OX₁ receptors seem to contribute to operant self-administration but are not involved in relapse to food-seeking behaviors (Nair et al., 2008; Choi et al., 2010).

Rodent and human genetics have helped tease out the role of orexin in energy homeostasis. Orexin/ataxin-3 mice exhibit late-onset obesity despite the fact that food intake is decreased, gaining more weight than wild-type mice on a high-fat diet (Hara et al., 2001, 2005). Transgenic orexin overexpression, on the other hand, renders mice resistant to diet-induced obesity (Funato et al., 2009). Genetic studies and the administration of the OX₂ receptor-selective agonist, [Ala¹¹,D-Leu¹⁵]OX-B suggest that this receptor is involved primarily in inhibiting the obesity phenotype. In human narcolepsy, weight gain and reduced energy expenditure often accompany the increased risk of type 2 diabetes. Patients exhibiting narcolepsy with cataplexy have a higher incidence of obesity and a higher body mass index than narcoleptics without cataplexy or the general population (Honda et al., 1986; Schuld et al., 2000, 2002; Sonka et al., 2010). The direct role of orexin on feeding in these patients remains in question given the secondary effect of hypersomnolence and the reduced physical activity and energy expenditure associated with the profound phenotype of those with narcolepsy/cataplexy. In support of this idea is the observation that orexin/ataxin-3 mice exhibit reduced wakefulness in response to food deprivation relative to wild-type animals, whose change in vigilance presumably reflects a foraging drive to identify additional food sources (Yamanaka et al., 2003).

In sum, the orexin system is well situated to govern the integration of multiple pathways (metabolic, motivational, sleep/wake) required to maintain energy homeostasis and drive the feeding-sleep cycle (Rodgers et al., 2002; Sakurai, 2002; Burdakov and Alexopoulos, 2005; Rolls et al., 2010). Despite carefully controlled studies, however, many of orexin's short-term effects on feeding seem secondary to its role in promoting arousal. This is supported by the observation that OX-A does not promote feeding when administered during the normal active phase (España et al., 2002). More work will confirm this short-term effect on feeding as well as the possibility that orexin mediates the rewarding properties of food.

2. Reward Pathways and Addiction. Orexin neurons project from the LH to the VTA and to components of the mesocorticolimbic reward pathway, including the nucleus accumbens, amygdala, and the medial PFC (Fadel and Deutch, 2002). Furthermore, orexin receptors are expressed in brain regions associated with reward pathways, including the VTA and nucleus accumbens (see Fig. 6) (Trivedi et al., 1998; Marcus et al., 2001). Lesion and intracranial self-stimulation experiments have long

indicated the LH as important in brain reward function (Olds and Milner, 1954; Velley et al., 1983). Several landmark studies performed between 2003 and 2006 strongly suggest a role for orexin signaling in drug abuse and addictive reward processing. Georgescu et al. (2003) reported activation of orexin neurons in the LH and induction of orexin gene expression in response to long-term morphine administration or morphine withdrawal. Furthermore, the authors found that withdrawal symptoms were attenuated in orexin knockout mice. In 2005, Harris et al. (2005) showed that orexin neurons in the LH were strongly activated by a CPP for morphine, cocaine, or food. Activation of LH orexin cells or intra-VTA administration of OX-A was sufficient to reinstate extinguished drug seeking behavior; this reinstatement was blocked by systemic pretreatment with an OX₁ SORA. That same year, Boutrel et al. (2005) reported that intracerebroventricular administration of OX-A reinstated cocaine-seeking and that OX-A elevated intracranial self-stimulation thresholds. They proposed the involvement of orexin in stress pathways linked to addiction, because OX-A reinstatement was blocked by inhibition of either the noradrenergic or the CRF system. Borgland et al. (2006) expanded upon these studies and linked orexin signaling with synaptic plasticity in VTA dopamine neurons and behavioral sensitization to cocaine. On the basis of these studies, a vast body of preclinical research has evolved around orexin signaling in addictive behaviors (Aston-Jones et al., 2010; Lawrence, 2010; Sharf et al., 2010; Zhou et al., 2011). Together, these studies have indicated the potential for blocking drug-seeking behaviors through antagonism of orexin signaling, suggesting orexin receptor antagonists as possible therapeutics for treating addiction to a variety of abusive drugs, including cocaine, amphetamine, alcohol, and morphine.

Harris and Aston-Jones (2006) were the first to formally propose the dichotomy theory of reward versus arousal. That is, they proposed that a functional heterogeneity exists among orexin neurons and their associated circuitries such that the effects of orexins on drug-seeking are mediated via two behavioral systems: 1) reward processing and 2) arousal/stress. Furthermore, these two systems show regional specificity within the hypothalamus; i.e., the LH orexin system affects drug-seeking behavior through activation of reward pathways (mostly through the VTA) and the perifornical-dorsomedial hypothalamus controls drug behavior via activation of arousal and stress pathways (Harris and Aston-Jones, 2006). Finally, the dichotomy in orexin function is likely also attributable to differences in OX₁ and OX₂ receptor signaling (OX₁ in reward seeking and OX₂ in arousal). This dichotomy of function was the focus of a more recent publication in which the authors used functional magnetic resonance imaging to show that the two receptors had distinct neuroanatomical patterns of functional inhibition. Gozzi et al. (2011) spatially

monitored the modulatory effects of OX₁ or OX₂ receptor blockade on the regional cerebral blood flow increases produced by D-amphetamine. The study used the highly selective OX₁ and OX₂ SORAs GSK1059865 and JNJ10397049, respectively. The two compounds showed distinct patterns of inhibition; GSK1059865 modulated the functional response to D-amphetamine in a focal striatal pattern, whereas JNJ10397049 induced widespread attenuation of the response predominantly in the cortex. In an attempt to correlate behavior with these divergent imaging profiles, the authors investigated the effect of the compounds on sleep and cocaine-induced CPP in rats. Robust sleep effects were observed for JNJ10397049 but not GSK1059865, whereas GSK1059865 dose-dependently reduced expression of cocaine-induced CPP (Gozzi et al., 2011). In sum, the authors proposed that OX₁ receptor activity in the striatum is specifically involved in drug-related reward behaviors, whereas cortical activity of OX₂ receptors is essential for regulating sleep/wake.

The majority of in vivo experiments investigating the role of orexin in addiction have used the partially selective OX₁ SORA SB-334867 to suggest that OX₁ receptors are involved in drug-seeking behaviors. That said, the high doses of SB-334867 used in some of these studies and the compound's limited specificity suggest that blockade of OX₂ receptors may in fact have contributed to the results of these studies (Scammell and Winrow, 2011; Shoblock et al., 2011). Indeed, more recent experiments with OX₂ SORAs JNJ10397049 and (2S)-1-(3,4-dihydro-6,7-dimethoxy-2(1*H*)-isoquinoliny)-3,3-dimethyl-2-[(4-pyridinylmethyl)amino]-1-butanone (TCSOX₂29) argue for a direct role of OX₂ receptor signaling in drug behaviors involving ethanol and morphine (Li et al., 2011; Shoblock et al., 2011). Shoblock et al. (2011) expound upon the potential for an OX₂ SORA that might provide treatment for addiction and also serve to treat the comorbid insomnia that often accompanies drug use and contributes to relapse.

Patients with narcolepsy are frequently clinically treated with addictive drugs yet rarely demonstrate addictive behaviors (Nishino and Mignot, 1997). Dimitrova et al. (2011) tested the hypothesis that persons with orexin deficiency exhibit a diminished tendency toward addictive behaviors. They found similar risk-taking behavior in narcoleptics relative to control groups and no significant differences in substance or alcohol abuse. Although these results counter the hypothesis that orexin deficiency affects reward processing, these investigators suggested that orexin-deficient subjects use a different neurological mechanism for these behaviors (Dimitrova et al., 2011). In fact, a recent functional magnetic resonance imaging study suggests altered activity in brain reward circuits in patients exhibiting narcolepsy with cataplexy (Ponz et al., 2010).

The aforementioned studies indicate that orexins and orexin receptors play a role in many models of addiction

but that results are variable depending upon the particular addictive substance, animal model, and orexin receptor antagonist used; moreover, results are probably mediated by distinct neuronal populations and circuits. Further studies will more clearly define the role of orexin in these mechanisms, as well as its possible interconnections with arousal pathways.

3. Anxiety. Much work has been done preclinically in rodent behavioral models to elucidate the role of orexin signaling in anxiety. OX-A was shown to have an anxiogenic effect in the mouse light-dark exploration test and the mouse and rat elevated plus-maze test (Suzuki et al., 2005). The authors of the preclinical study suggest that CRF, 5-HT, and NPY may be involved in mediating the anxiogenic effects of OX-A. Indeed, others have substantiated the involvement of these systems in mediating stress-related orexinergic effects by showing a reduction in orexin-mediated stress behaviors by pretreatment with the CRF antagonist α -helical CRF (Ida et al., 2000), an altered sleep response to restraint stress in serotonin transporter knockout mice (Rachalski et al., 2009), and an inhibition of orexin-induced activation of the hypothalamic-pituitary axis by an NPY antagonist (Jászberényi et al., 2001). Li et al. (2010) investigated the effects of administration of OX-A and OX-B into the paraventricular nucleus of the thalamus, a site characterized by orexin-containing fibers and known to innervate forebrain areas associated with fear and anxiety. Administration of OX-A and OX-B caused an anxiogenic response in the rat elevated plus-maze, whereas κ -opioid or CRF receptor antagonists attenuated the effects of OX-A (Li et al., 2010). Furthermore, endogenous orexins may be involved in producing anxiety in these models, because the OX₂ SORA TCSOX₂29, but not the partial OX₁ SORA SB-334867, reduced the anxiogenic response in a foot shock-induced anxiety model. The use of elevated plus-maze and light-dark exploration in the Syrian golden hamster showed an anxiogenic response for orexins in the amygdala and an interaction with GABAergic signals (Avolio et al., 2011). In fact, decreased GABA levels have been reported in those suffering from panic disorder (Goddard et al., 2001). In a rat model of sodium lactate-induced panic attacks, activation of orexin-synthesizing neurons is required for the panic response in panic-prone rats. Accordingly, treatment with *Hcrt* small interfering RNA or an OX₁ antagonist (SB-334867 or SB-408124) blocks the behavioral and cardiovascular effects elicited by sodium lactate challenge associated with the panicked state (Johnson et al., 2010).

In contrast to the reports of anxiogenic responses mediated by orexin, rat studies using a startle paradigm ascribe an anxiolytic effect to the neuropeptides (Singareddy et al., 2006). Exogenous administration of OX-A or OX-B produced an anxiolytic effect in situations reflective of unconditioned, but not conditioned, anxiety. Suzuki et al. suggest that differences in the range of response to orexin (anxiogenic versus anxiolytic) may be

attributable to the dose of orexin, interacting signaling pathways, and perhaps differential roles of the orexin receptor subtypes in mediating anxiety-related behaviors (Suzuki et al., 2005).

Clinically, several recent studies have reported on the link between orexinergic signaling and anxiety and stress disorders. In a study of 10 male veterans with post-traumatic stress disorder (PTSD) and 10 healthy men, it was reported that CSF and plasma OX-A levels are significantly lower in the patients suffering from PTSD; furthermore, levels of OX-A in the CSF negatively correlate with the level of severity of the disorder (Strawn et al., 2010). In contrast, Johnson et al. (2010) report higher CSF levels of orexin in subjects exhibiting panic anxiety symptoms compared with subjects without anxiety. These conflicting reports may reflect the differential responses to orexin as described in preclinical anxiety studies, the different forms of anxiety highlighted in the two clinical studies, or may be secondary to disrupted sleep/wake cycles observed in patients with these illnesses. Fortuyn et al. (2010) studied the prevalence of anxiety and mood disorders in narcolepsy. They found that although mood disorders are not increased among narcoleptics, anxiety behaviors, including panic attacks and social phobias, are present in more than half of narcoleptic subjects, 35% of whom are also diagnosed with anxiety disorder. Another more recent study, however, found that rates of both depression and anxiety are higher in narcoleptic subjects (Dimitrova et al., 2011).

4. Depression / Mood. The activation of orexin-secreting and orexin receptor-expressing neurons in response to stress and orexin activation of stress-related systems, including norepinephrine, dopamine, and CRF, may point to a possible role in PTSD and depression (Berridge et al., 2010). OX-A administration was reported to reduce immobility in the rat forced-swim test and increased hippocampal neurogenesis, with pretreatment of SB-334867 blocking these effects (Ito et al., 2008). Lutter et al. (2008) showed that intact orexin signaling was necessary for the efficacy of caloric restriction in a mouse model of depression. After caloric restriction, wild-type mice show less immobility in the forced swim test compared with orexin peptide knockout mice. Likewise, in a social defeat model, caloric restriction is efficacious in wild-type mice but not in orexin peptide knockout mice. Behavioral profiling studies conducted on mice lacking OX₁ and OX₂ receptors as well as in wild-type animals treated with the partially selective OX₁ receptor antagonist SB-334867 suggest that a balance of orexin receptor activity may differentially affect behavioral despair activities (Scott et al., 2011). In tail suspension and forced swim test models of despair behavior, OX₁ receptor-null mice displayed a significant reduction in immobility, suggesting an antidepressant phenotype, and similar results were observed in wild-type animals treated with SB-334867. On the other hand, OX₂ receptor-null mice displayed an increase in

behavioral despair (Scott et al., 2011). Although this study detected no genotype-dependent changes in anxiety measures, these apparent counter-acting orexin receptor activities are reminiscent of that suggested for orexin signaling in anxiety described above.

Clinical reports on orexin levels in depression are varied across specific affective disorders. In mild-to-moderately depressed subjects, CSF OX-A diurnal variation seems dampened, overall levels seeming higher in patients and modest but significant reductions after treatment with the antidepressant sertraline, a serotonin-reuptake inhibitor (Salomon et al., 2003). In contrast, suicidal patients with major depressive disorder had lower OX-A levels in CSF compared with other suicidal patients (Brundin et al., 2007). A more recent study of patients with mania failed to show an association between severity of disease and levels of orexin. Evaluation of OX-A in CSF from five patients exhibiting manic episodes failed to reveal significant differences compared with age-matched patients with major depressive disorder or healthy control subjects without any psychiatric or neurological disorder (Schmidt et al., 2010). Overall, these studies indicate a potential role for orexin signaling in preclinical depression models; however, additional studies will be needed to better understand the contributions of orexin signaling to mood and anxiety.

B. Orexin Influences on Peripheral Physiology

A growing body of evidence indicates that orexin signaling has the capacity to influence peripheral physiology. Many of these effects seem to be mediated by hypothalamic orexinergic neuron activity, modulating either autonomic nervous system tone or perhaps secondary to the peptides' effects on arousal and vigilance state. Clear demonstrations of the physiological role of orexin receptor activity in peripheral tissues are scarce, but the emergence of increasingly selective orexin receptor antagonists, however, should provide useful reagents to resolve this issue, because their concentrations are typically 10- to 100-fold higher in the periphery relative to the brain after administration. The potential roles for orexin signaling in the periphery, and whether these effects might be autonomic or secondary to its behavioral effects, remain to be determined.

1. Metabolism and Gastrointestinal Motility. The vast body of research related to orexins and feeding behaviors has prompted a closer look at the role of orexin signaling in feeding-related responses in the periphery. Orexin ligand and receptor expression in the gastrointestinal tract have been investigated in rat, human, dog, and a limited number of other species. Orexin-like immunoreactivity has been observed throughout the gastrointestinal tract and the pancreas in various cell types (e.g., neurons, nerve fibers, smooth muscle, endocrine cells) and is often colocalized with signaling molecules such as vasoactive intestinal peptide, neuronal nitric-

oxide synthase, substance P, insulin, or gastrin, although a full understanding of the selectivity of some antibodies has been a challenge (Kirchgessner, 2002; Nakabayashi et al., 2003; Voisin et al., 2003; Heinonen et al., 2008; Dall'Aglia et al., 2008, 2009, 2010). Orexin-containing projections extend from the lateral hypothalamus to the dorsal motor nucleus of the vagus (DMV) and project to target peripheral tissues including stomach, intestine, and pancreas. In a study using whole-cell patch-clamp recordings, it was shown that responsiveness to the orexins is organized in a viscerotopical manner, DMV neurons projecting to gastric fundus and corpus being much more responsive than those projecting to the more distal duodenum and caecum (Grabauskas and Moises, 2003).

Orexin signaling is hypothesized to be a trigger of the gastrointestinal secretion and motility associated with the cephalic phase of feeding in response to the sight, smell, taste, or anticipation of food (Takahashi et al., 1999; Okumura and Takakusaki, 2008). In rats, intracisternal injection of OX-A, but neither intracisternal OX-B nor intraperitoneal OX-A, stimulated gastric acid secretion (Takahashi et al., 1999). This central action was blocked by atropine or vagotomy, suggesting that orexin is affecting the gastrointestinal tract from the brain via the vagal system. Furthermore, the effects of exogenously added or endogenous OX-A on gastric acid secretion were inhibited by the orexin receptor antagonist SB-334867 (Ehrström et al., 2005b; Yamada et al., 2005).

Intestinal bicarbonate secretion is considered another key feature of the cephalic phase of feeding. In contrast to gastric secretion, however, bicarbonate secretion in the duodenum is peripherally rather than centrally mediated and has been shown in rats to be affected by nutritional status (Bengtsson et al., 2007). At the cellular level, fasting was shown to reduce the expression of OX₁ receptor mRNA in duodenal enterocytes and to inhibit OX-A induced calcium mobilization (Bengtsson et al., 2009). It has been widely demonstrated in rodent and guinea pig that OX-A and OX-B modulate gastrointestinal motility (Kirchgessner and Liu, 1999; Kobashi et al., 2002; Krowicki et al., 2002; Näslund et al., 2002; Ehrström et al., 2003; Katayama et al., 2003; Bülbül et al., 2010a,b). The effects of orexins vary depending upon location within the gastrointestinal tract and represent a combination of central and peripheral actions (Baccari, 2010). Experiments in rat and one clinical report suggest effects on gastric emptying rate as well (Ehrström et al., 2005a,b; Bülbül et al., 2010a).

Intracerebroventricular injection of OX-A stimulated pancreatic exocrine secretion, demonstrating that in addition to acting on gastric acid secretion, OX-A in the brain modulates rat pancreatic fluid output (Miyasaka et al., 2002). Data suggest that glucose-sensing neurons in the lateral hypothalamus release orexin, which acts on neurons of the DMV to modulate pancreatic vagal

pathways (Wu et al., 2004). Studies linking orexin to glucose homeostasis suggest that the orexin system is poised to connect critical physiological systems including metabolism, goal-oriented (reward) behaviors, and arousal (Tsujino and Sakurai, 2009; Tsuneki et al., 2010).

Given its critical function in controlling arousal and sleep, and its presumed role in maintaining energy homeostasis, orexin signaling may be expected to respond to changes in plasma pH associated with fluctuations in energy expenditure. Indeed, changes in evoked and spontaneous firing activity of orexin neurons in organotypic culture are observed in response to changes in extracellular pH (Williams et al., 2007; Gestreau et al., 2008). The role of orexin in modulating pulmonary changes in response to plasma hypoxia and hypercapnia, however, is less clear. Observations of ventilatory responses to these conditions in orexin peptide knockouts and after central administration of orexin peptides has been mixed; some studies have found an involvement of orexin in mediating increased ventilatory responses to hypercapnia (Deng et al., 2007; Nakamura et al., 2007; Dias et al., 2009, 2010; Nattie and Li, 2010), decreased pulmonary activity to hypoxia (Deng et al., 2007; Nakamura et al., 2007), or no detectable response to either condition (Zhang et al., 2005). Conversely, a study of patients with narcolepsy-cataplexy detected attenuated responses to hypoxia but not hypercapnia (Han et al., 2010), which is exactly opposite that found in rodent models. Clearly, more work needs to be done to confirm a role for orexin in mediating ventilatory responses to changing blood gas levels as opposed to controlling sleep/wake cycles to maintain energy homeostasis.

2. Potential Roles in Nociception/Pain. Orexinergic neurons from the hypothalamus project to numerous supraspinal sites involved in the modulation of pain, including the thalamus, DR, LC, reticular formation, periaqueductal gray, and the trigeminal nucleus caudalis (Peyron et al., 1998). OX-A and OX-B fibers have been identified in the rat spinal cord, and orexinergic projections from the hypothalamus have been shown in rat, mouse, and human to innervate multiple layers of the spinal dorsal horn (van den Pol, 1999; Date et al., 2000). There, they terminate in laminae I and II, the location of peripheral sensory afferent terminals that synapse onto central nociceptive neurons. Despite differential OX₁ and OX₂ receptor expression, both receptors are present in the aforementioned supraspinal sites involved in descending nociceptive modulation (Marcus et al., 2001). OX-A and OX₁ receptor immunoreactivity are localized to the superficial laminae of the spinal dorsal horn and in the dorsal root ganglion (DRG) neurons of peripheral sensory afferents (Bingham et al., 2001; Hervieu et al., 2001).

Work in multiple pain models suggests that supraspinal orexins may be involved in the modulation of pain. Intracerebroventricular administration of OX-A pro-

duces an antinociceptive effect in multiple rodent pain models, including acute, chemical, inflammatory, and neuropathic pain models. The antinociceptive effect is blocked by administration of SB-334867, suggesting the effects may be mediated by OX₁ receptors. In fact, OX-B is less potent than OX-A in attenuating the pain response in these models (Bingham et al., 2001; Yamamoto et al., 2003a; Mobarakeh et al., 2005b). OX-A may modulate supraspinal histamine release, because the effects of OX-A in acute and chemical pain models are enhanced in H₁ or H₂ receptor knockout mice or with the administration of H₁ or H₂ antagonists (Mobarakeh et al., 2005a).

In addition, OX-A and OX-B have a functional role in spinal sensory transmission, according to in vivo and in vitro electrophysiological studies, respectively (Grudt et al., 2002; Peng et al., 2008). OX-B may elicit its effect by facilitating glycine release presynaptically. Behavioral studies support these electrophysiological findings, because intrathecal administration of OX-A and, to a lesser extent, OX-B inhibits withdrawal responses and spontaneous nociceptive behaviors in acute, chemical, inflammatory, neuropathic, and postsurgical pain models (Yamamoto et al., 2002, 2003a,b; Cheng et al., 2003; Kajiyama et al., 2005; Mobarakeh et al., 2005a; Jeong and Holden, 2009). The antinociceptive effects of the orexins are blocked by intrathecally administered SB-334867, once again suggesting that pain modulation may be mediated through OX₁ receptors.

In a few instances, administration of SB-334867 alone enhanced sensitivity in the in vivo models, supporting the idea of enhanced or tonic OX₁ receptor-mediated inhibitory tone during pain (Bingham et al., 2001; Cheng et al., 2003; Jeong and Holden, 2009). Consistent with this hypothesis of orexin-mediated inhibitory tone, orexin peptide knockout mice show enhanced thermal hypersensitivity in an inflammatory pain model (Watanabe et al., 2005). Furthermore, knockout mice as well as orexin/ataxin transgenic mice, with degeneration and dysfunction of orexinergic neurons, show attenuation of stress-induced analgesia (Xie et al., 2008).

Although the role of the orexins and their receptors in the periphery has not been extensively studied, two studies point to a role for OX₁ receptors on peripheral DRG neuron function. Application of OX-A to cultured DRG neurons increases action potentials and levels of intracellular calcium. These effects are blocked by SB-334867 and by the protein kinase C inhibitor chelerythrine, suggesting that peripheral sensory transmission may be modulated by activation of OX₁ receptors and subsequent protein kinase C-dependent calcium signaling (Yan et al., 2008; Ozcan et al., 2010).

Orexin signaling in the trigeminal nucleus caudalis seems to modulate nociceptive transmission in the context of facial pain accompanying headache and migraine. OX-A inhibits spontaneous and stimulus-evoked responses in trigeminal nucleus caudalis neurons; an-

tagonist treatment suggests the effect might be mediated via OX₁ receptors. On the other hand, OX-B facilitates these same responses suggesting an involvement of OX₂ as well (Bartsch et al., 2004; Holland et al., 2005, 2006). In a rat model of dural vasodilation thought to be representative of migraine pathophysiology, intravenous administration of OX-A but not OX-B resulted in the inhibition of vasodilation, and pretreatment with SB-334867 blocked this effect (Holland et al., 2005). Increased levels of OX-A have been reported in the CSF of chronic migraine sufferers, although the significance of this finding is currently unknown (Sarchielli et al., 2008). Clinical studies have identified polymorphisms within the orexin receptor genes that are associated with cluster headache and migraine. A 1246G→A polymorphism in *HCRTR2* is significantly associated with cluster headaches but not with treatment response, whereas the same polymorphism is not associated with migraine (Rainero et al., 2004, 2007; Schürks et al., 2006, 2007a,b; Pinessi et al., 2007). More recently, a 1222G→A polymorphism within the *HCRTR1* gene has been found to be associated with an increased risk of migraine without aura (Rainero et al., 2011). Further investigations are needed, however, to determine whether these genetic correlations reflect variation around these chromosomal loci or real expression or biochemical changes affecting orexin receptor activity and or signaling function.

3. Influence on Cardiovascular Physiology. Preclinical studies, mostly in rat, have focused on the cardiovascular effects of exogenously added orexins. In the majority of studies, central administration (intracerebroventricular or intracisternal) of extraphysiological doses of orexin peptides, particularly OX-A, acts to increase blood pressure and heart rate, although differences in experimental paradigms have also found unchanged or decreased cardiovascular parameters with orexin administration (Samson et al., 1999, 2007; Shirasaka et al., 1999; Chen et al., 2000; White et al., 2006). The partial OX₁ receptor antagonists SB-408124 and SB-334867 and the OX₂ receptor antagonists [Ala¹¹,D-Leu¹⁵]OX-B and TCISOX₂29 either partially or fully blocked the effects of exogenous orexins, suggesting that both receptor subtypes are involved (White et al., 2006; Samson et al., 2007; Huang et al., 2010). Direct administration to specific brain regions evoked either increases or decreases in blood pressure and heart rate, suggesting that stimulation of different structures may indeed differentially modulate cardiovascular function; however, differences in protocol, dose, and state of anesthesia probably also contribute to the disparate results (Chen et al., 2000; Antunes et al., 2001; Machado et al., 2002; Sato-Suzuki et al., 2002; Smith et al., 2002, 2007; Ciriello and de Oliveira, 2003; Ciriello et al., 2003; de Oliveira et al., 2003; Shahid et al., 2011). In fact, the doses used in several of the studies is relatively high compared with the endogenous OX-A levels (0.2–0.4 pmol/ml) in rat CSF (Fujiki et al., 2001), and so the relevance to the role of intrinsic orexins in cardiovascular function is unknown. More relevant to the safety profile of

orexin receptor antagonists in the clinic is the finding that central administration of the aforementioned OX₁ or OX₂ SORAs elicited no changes in heart rate or blood pressure when administered alone in preclinical models (Hirota et al., 2003; White et al., 2006; Samson et al., 2007; Huang et al., 2010; Shahid et al., 2011). Furthermore, preclinical studies in rats and dogs showed no effects on heart rate and blood pressure with oral administration of the dual orexin receptor antagonist, almorexant (ACT-078573) (Brisbare-Roch et al., 2007; Furlong et al., 2009). Oral ascending single-dose and multiple-day dosing of almorexant in the clinic has also caused no changes in heart rate or blood pressure (Brisbare-Roch et al., 2007; Hoever et al., 2010).

The effects of genetic manipulation on cardiovascular function have been studied in both orexin peptide knock-out mice and orexin neuron-ablated orexin/ataxin-3 transgenic mice and rats, leading to a conflicting data set regarding the effects on heart rate and blood pressure (Kayaba et al., 2003; Schwimmer et al., 2006; Zhang et al., 2006; Schwimmer et al., 2010; Bastianini et al., 2011). The relevance of either of these genetic models to the pharmacological antagonism of orexin receptors in humans is unknown.

VII. Conclusion

Relative to the current therapies, orexin receptor antagonism represents a novel mechanism for the treatment of insomnia, and provides potential advantages over the current standard of care, which includes “z-drugs” (zolpidem, zaleplon, zopiclone, eszopiclone) that interact with the allosteric site of GABA_A receptors. Although they are not benzodiazepines themselves, z-drugs interact with the benzodiazepine binding site on these receptor subunits with improved GABA_A subtype specificity. GABA_A receptors are inhibitory ligand-gated chloride channels for which conductance is potentiated by z-drugs to provide an inhibitory influence on postsynaptic activity and ultimately CNS depression in areas in which these receptors are expressed (Costa and Guidotti, 1979; Sullivan and Guilleminault, 2009). GABA_A receptors exhibit wide expression and function in a number of pathways, including those associated with arousal, anxiety, psychomotor tone, and cognition. As such, z-drugs have the potential to affect behavior and physiology beyond sleep (Ashton, 1994; Hoque and Chesson, 2009). Whereas z-drugs attenuate sleep latency and promote NREM sleep, they also suppress REM and slow-wave components of normal sleep (Lancel, 1999; Bettica et al., 2012). In contrast, sleep architecture induced by DORAs is characterized by increased mean NREM and REM time (Brisbare-Roch et al., 2007; Winrow et al., 2011, 2012; Bettica et al., 2012). The most salient effect of zolpidem in rats is the suppression of both the mean time spent in REM sleep and the number of REM sleep bouts, with comparatively slight reductions in active wake time (Renger et al., 2004). On the other hand, dual orexin receptor antagonists such as suvorexant, MK-6096, DORA-22,

SB-649868, and almorexant promote sleep that includes increased NREM and REM sleep in multiple species including humans (Brisbare-Roch et al., 2007; Di Fabio et al., 2011; Winrow et al., 2011, 2012; Bettica et al., 2012). GABA_A receptor-induced sleep is also associated with significant impairment of locomotor coordination not seen with DORAs, even when administered at 10-fold above its effective dose (Steiner et al., 2011).

The pharmacology associated with orexin receptors is relatively simple compared with GABA_A receptors, which are composed of five subunits arranged in a heteromultimeric complex. As such, the myriad effects potentially elicited by GABA_A receptor modulation depend upon subtype specificity, where targeting the $\alpha 1$ subtype has sedative and anticonvulsive effects but is also associated with amnesia and dependence. The $\alpha 2$, $\alpha 3$, and $\alpha 5$ subtypes seem differentially involved in anxiolysis, muscle relaxation, and amnesia (Nutt and Stahl, 2010; Tan et al., 2011). GABA_A-mediated amnesia may be an underlying mechanism for anxiolytic and antidepressant effects, because loss of memory for unpleasant events may actually be favorable for these psychiatric outcomes. Amnesia may also underlie the rare side effects of walking, eating, and driving while asleep observed in human patients taking zolpidem (Hoque and Chesson, 2009; Tan et al., 2011). In contrast, the orexin receptor antagonist almorexant has no impact on spatial learning and memory tasks or avoidance retention in rodent models at 300 mg/kg, which is 10-fold above the effective dose required to induce sleep (Dietrich and Jenck, 2010).

Continued clinical investigation will be required to understand the advantages and potential shortcomings of orexin receptor antagonism relative to z-drugs. Parameters to be monitored include sleep-stage dysregulation, next-day sleepiness, and cognitive performance and other physiological responses secondary to the modulation of sleep/wake cycles. Preclinical genetic models mimicking human narcolepsy with chronic constitutive loss of orexin signaling suggest that orexin receptor antagonism may be associated with deficiencies in sleep-stage regulation and active phase cataplexy. As indicated in section V.H, however, effects observed in constitutive knockout models are not necessarily equal to those induced by transient pharmacological manipulation. Nevertheless, cataplexy in response to orexin antagonism is a concern to be closely monitored in both the clinic and in preclinical models. Next-day sleepiness and impaired cognitive impairment are not expected for orexin antagonists but are nonetheless monitored closely in clinical trials. Short-acting orexin receptor antagonists administered before the onset of rest, however, are likely to avoid most all of these potential issues and perhaps even improve some of these measures by virtue of their ability to improve sleep maintenance.

Compared with the current standard of care, the role of orexin signaling in the control of arousal and vigilance state suggests that orexin receptor antagonism represents a selective mechanism capable of effectively promoting sleep associated with increases in both NREM and REM sleep. Orexin signaling is both sufficient to induce arousal and necessary for the normal maintenance of vigilance state (España and Scammell, 2011). Orexinergic neurons are restricted to the lateral hypothalamus and selectively project to arousal-promoting histaminergic neurons of the TMN as well as brainstem nuclei involved in sleep/wake control (Trivedi et al., 1998; Marcus et al., 2001). As an arousal signal, orexin neurons have daily oscillations in activity that give rise to accumulating peptide levels during waking hours and fall silent during the normal inactive period (Taheri et al., 2000; Zeitzer et al., 2003). As such, orexin receptor antagonists have maximal effects late in the active period, precisely at the most therapeutically relevant time, but have little to no effect during the inactive phase (Brisbare-Roch et al., 2007; Li and Nattie, 2010; Winrow et al., 2011). As described in the latter sections of this review, orexin signaling has the potential to affect physiology and behavior beyond the regulation of sleep. How much of this function (e.g., feeding, metabolism, cardiovascular physiology) is a secondary consequence of orexin-mediated arousal relative to direct mechanisms is a matter of ongoing debate and remains to be determined. Taken together, orexin antagonism represents a novel and selective mechanism for the therapeutic treatment of insomnia as well as offering potential opportunities for alternative indications.

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References

- Adamantidis AR, Zhang F, Aravanis AM, Deisseroth K, and de Lecea L (2007) Neural substrates of awakening probed with optogenetic control of hypocretin neurons. *Nature* **450**:420–424.
- Akanmu MA and Honda K (2005) Selective stimulation of orexin receptor type 2 promotes wakefulness in freely behaving rats. *Brain Res* **1048**:138–145.
- Alvaro G, Amantini D, and Belvedere S (2009), inventors; GlaxoSmithKline, Alvaro G, Amantini D, and Belvedere S, assignees. Novel compounds. U.S. patent application no. 2009022670. 2009 Jan 22.
- Alvaro G, Amantini D, and Stasi LP (2011), inventors; GlaxoSmithKline, Alvaro G, Amantini D, and Stasi LP, assignees. Pyridine derivatives used to treat orexin related disorders. U.S. patent application no. 20110053979. 2011 Mar 3.
- Antunes VR, Brailoiu GC, Kwok EH, Scruggs P, and Dun NJ (2001) Orexins/hypocretins excite rat sympathetic preganglionic neurons in vivo and in vitro. *Am J Physiol Regul Integr Comp Physiol* **281**:R1801–R1807.
- Arrigoni E, Mochizuki T, and Scammell TE (2010) Activation of the basal forebrain by the orexin/hypocretin neurones. *Acta Physiol (Oxf)* **198**:223–235.

- Asahi S, Egashira S, Matsuda M, Iwaasa H, Kanatani A, Ohkubo M, Ihara M, and Morishima H (2003) Development of an orexin-2 receptor selective agonist, [Ala(11), D-Leu(15)]orexin-B. *Bioorg Med Chem Lett* **13**:111–113.
- Ashton H (1994) Guidelines for the rational use of benzodiazepines. When and what to use. *Drugs* **48**:25–40.
- Aston-Jones G, Smith RJ, Sartor GC, Moorman DE, Massi L, Tahsili-Fahadan P, and Richardson KA (2010) Lateral hypothalamic orexin/hypocretin neurons: A role in reward-seeking and addiction. *Brain Res* **1314**:74–90.
- Avolio E, Alò R, Carelli A, and Canonaco M (2011) Amygdalar orexinergic-GABAergic interactions regulate anxiety behaviors of the Syrian golden hamster. *Behav Brain Res* **218**:288–295.
- Baccari BC (2010) Orexins and gastrointestinal functions. *Curr Protein Pept Sci* **11**:148–155.
- Bartsch T, Levy MJ, Knight YE, and Goadsby PJ (2004) Differential modulation of nociceptive dural input to [hypocretin] orexin A and B receptor activation in the posterior hypothalamic area. *Pain* **109**:367–378.
- Bastianini S, Silvani A, Berteotti C, Elghozi JL, Franzini C, Lenzi P, Lo Martire V, and Zoccoli G (2011) Sleep related changes in blood pressure in hypocretin-deficient narcoleptic mice. *Sleep* **34**:213–218.
- Beck B and Richey S (1999) Hypothalamic hypocretin/orexin and neuropeptide Y: divergent interaction with energy depletion and leptin. *Biochem Biophys Res Commun* **258**:119–122.
- Bengtsson MW, Mäkelä K, Herzig KH, and Flemström G (2009) Short food deprivation inhibits orexin receptor 1 expression and orexin-A induced intracellular calcium signaling in acutely isolated duodenal enterocytes. *Am J Physiol Gastrointest Liver Physiol* **296**:G651–G658.
- Bengtsson MW, Mäkelä K, Sjöblom M, Uotila S, Akerman KE, Herzig KH, and Flemström G (2007) Food-induced expression of orexin receptors in rat duodenal mucosa regulates the bicarbonate secretory response to orexin-A. *Am J Physiol Gastrointest Liver Physiol* **293**:G501–G509.
- Bergman JM, Roecker AJ, Mercer SP, Bednar RA, Reiss DR, Ransom RW, Meacham Harrell C, Pettibone DJ, Lemaire W, Murphy KL, et al. (2008) Proline bis-amides as potent dual orexin receptor antagonists. *Bioorg Med Chem Lett* **18**:1425–1430.
- Bernardis LL and Bellinger LL (1996) The lateral hypothalamic area revisited: ingestive behavior. *Neurosci Biobehav Rev* **20**:189–287.
- Bernardis LL, Ciesla A, and Bellinger LL (1993) Hypophagic rats with dorsomedial hypothalamic lesions produce lighter and smaller pups with a lower survival rate at weaning than offspring of sham-operated controls. *Physiol Behav* **53**:59–64.
- Berridge CW, España RA, and Vittoz NM (2010) Hypocretin/orexin in arousal and stress. *Brain Res* **1314**:91–102.
- Bettica P, Nucci G, Pyke C, Squassante L, Zamuner S, Ratti E, Gomeni R, and Alexander R (2011) Phase I studies on the safety, tolerability, pharmacokinetics and pharmacodynamics of SB-649868, a novel dual orexin receptor antagonist. *J Psychopharmacol* <http://dx.doi.org/10.1177/0269881111408954>.
- Bettica PU, Lichtenfeld U, Squassante L, Shabbir S, Zuechner D, Dreykluft P, Lehmann R, Danker-Hopfe H, and Ratti E (2009) The orexin antagonist SB-649868 promotes and maintains sleep in healthy volunteers and in patients with primary insomnia (Abstract 0774). *Sleep* **32** (Suppl):A252–A253.
- Bettica P, Squassante L, Groeger JA, Gennery B, Winsky-Sommerer R, and Dijk DJ (2012) Differential effects of a dual orexin receptor antagonist (SB-649868) and zolpidem on sleep initiation and consolidation, SWS, REM sleep, and EEG power spectra in a model of situational insomnia. *Neuropsychopharmacology* **37**:1224–1233.
- Beuckmann CT, Sinton CM, Williams SC, Richardson JA, Hammer RE, Sakurai T, and Yanagisawa M (2004) Expression of a poly-glutamine-ataxin-3 transgene in orexin neurons induces narcolepsy-cataplexy in the rat. *J Neurosci* **24**:4469–4477.
- Beuckmann CT and Yanagisawa M (2002) Orexins: from neuropeptides to energy homeostasis and sleep/wake regulation. *J Mol Med (Berl)* **80**:329–342.
- Bingham S, Davey PT, Babbs AJ, Irving EA, Sammons MJ, Wyles M, Jeffrey P, Cutler L, Riba I, Johns A, et al. (2001) Orexin-A, an hypothalamic peptide with analgesic properties. *Pain* **92**:81–90.
- Borgland SL, Storm E, and Bonci A (2008) Orexin B/hypocretin 2 increases glutamatergic transmission to ventral tegmental area neurons. *Eur J Neurosci* **28**:1545–1556.
- Borgland SL, Taha SA, Sarti F, Fields HL, and Bonci A (2006) Orexin A in the VTA is critical for the induction of synaptic plasticity and behavioral sensitization to cocaine. *Neuron* **49**:589–601.
- Bourgin P, Huitrón-Réndiz S, Spier AD, Fabre V, Morte B, Criado JR, Sutcliffe JG, Henriksen SJ, and de Lecea L (2000) Hypocretin-1 modulates rapid eye movement sleep through activation of locus coeruleus neurons. *J Neurosci* **20**:7760–7765.
- Boutrel B, Kenny PJ, Specio SE, Martin-Fardon R, Markou A, Koob GF, and de Lecea L (2005) Role for hypocretin in mediating stress-induced reinstatement of cocaine-seeking behavior. *Proc Natl Acad Sci USA* **102**:19168–19173.
- Brisbare-Roch C, Clozel M, and Jenck F (2008) Effects of repeated oral administration of the orexin receptor antagonist almorexant in male rats and dogs. *Sleep* **31**:A38.
- Brisbare-Roch C, Dingemans J, Koberstein R, Hoever P, Aissaoui H, Flores S, Mueller C, Nayler O, van Gerven J, de Haas SL, et al. (2007) Promotion of sleep by targeting the orexin system in rats, dogs and humans. *Nat Med* **13**:150–155.
- Brundin L, Björkqvist M, Petersén A, and Träskman-Bendz L (2007) Reduced orexin levels in the cerebrospinal fluid of suicidal patients with major depressive disorder. *Eur Neuropsychopharmacol* **17**:573–579.
- Bülbül M, Babygirija R, Ludwig K, and Takahashi T (2010a) Central orexin-A increases gastric motility in rats. *Peptides* **31**:2118–2122.
- Bülbül M, Tan R, Gemici B, Özdem S, Üstünel I, Acar N, and Işgüt-Uysal VN (2010b) Endogenous orexin-A modulates gastric motility by peripheral mechanisms in rats. *Peptides* **31**:1099–1108.
- Burdakov D and Alexopoulos H (2005) Metabolic state signalling through central hypocretin/orexin neurons. *J Cell Mol Med* **9**:795–803.
- Burdakov D (2004) Electrical signaling in central orexin/hypocretin circuits: tuning arousal and appetite to fit the environment. *Neuroscientist* **10**:286–291.
- Burgess CR, Tse G, Gillis L, and Peever JH (2010) Dopaminergic regulation of sleep and cataplexy in a murine model of narcolepsy. *Sleep* **33**:1295–1304.
- Carter ME, Adamantidis A, Ohtsu H, Deisseroth K, and de Lecea L (2009) Sleep homeostasis modulates hypocretin-mediated sleep-to-wake transitions. *J Neurosci* **29**:10939–10949.
- Cason AM, Smith RJ, Tahsili-Fahadan P, Moorman DE, Sartor GC, and Aston-Jones G (2010) Role of orexin/hypocretin in reward-seeking and addiction: implications for obesity. *Physiol Behav* **100**:419–428.
- Chemelli RM, Willie JT, Sinton CM, Elmquist JK, Scammell T, Lee C, Richardson JA, Williams SC, Xiong Y, Kisanuki Y, et al. (1999) Narcolepsy in orexin knockout mice: molecular genetics of sleep regulation. *Cell* **98**:437–451.
- Chen CT, Hwang LL, Chang JK, and Dun NJ (2000) Pressor effects of orexins injected intracranially and to rostral ventrolateral medulla of anesthetized rats. *Am J Physiol Regul Integr Comp Physiol* **278**:R692–R697.
- Chen L, McKenna JT, Bolortuya Y, Winston S, Thakkar MM, Basheer R, Brown RE, and McCarley RW (2010) Knockdown of orexin type 1 receptor in rat locus coeruleus increases REM sleep during the dark period. *Eur J Neurosci* **32**:1528–1536.
- Cheng JK, Chou RC, Hwang LL, and Chiou LC (2003) Antiallodynic effects of intrathecal orexins in a rat model of postoperative pain. *J Pharmacol Exp Ther* **307**:1065–1071.
- Cherezov V, Rosenbaum DM, Hanson MA, Rasmussen SG, Thian FS, Kobilka TS, Choi HJ, Kuhn P, Weis WI, Kobilka BK, et al. (2007) High-resolution crystal structure of an engineered human beta2-adrenergic G protein-coupled receptor. *Science* **318**:1258–1265.
- Choi DL, Davis JF, Fitzgerald ME, and Benoit SC (2010) The role of orexin-A in food motivation, reward-based feeding behavior and food-induced neuronal activation in rats. *Neuroscience* **167**:11–20.
- Ciriello J and de Oliveira CV (2003) Cardiac effects of hypocretin-1 in nucleus ambiguus. *Am J Physiol Regul Integr Comp Physiol* **284**:R1611–R1620.
- Ciriello J, Li Z, and de Oliveira CV (2003) Cardioacceleratory responses to hypocretin-1 injections into rostral ventromedial medulla. *Brain Res* **991**:84–95.
- Clark EL, Baumann CR, Cano G, Scammell TE, and Mochizuki T (2009) Feeding-elicited cataplexy in orexin knockout mice. *Neuroscience* **161**:970–977.
- Coleman PJ, Cox CD, and Roecker AJ (2011a) Discovery of dual orexin receptor antagonists (DORAs) for the treatment of insomnia. *Curr Top Med Chem* **11**:696–725.
- Coleman PJ and Renger JJ (2010) Orexin receptor antagonists: a review of promising compounds patented since 2006. *Expert Opin Ther Pat* **20**:307–324.
- Coleman PJ, Schreier JD, Cox CD, Breslin MJ, Whitman DB, Bogusky MJ, McGaughey GB, Bednar RA, Lemaire W, Doran SM, et al. (2012) Discovery of [(2R,5R)-5-([(5-fluoropyridin-2-yl)oxy]methyl)-2-methylpiperidin-1-yl][5-methyl-2-(pyrimidin-2-yl)phenyl]methanone (MK-6096): a dual orexin receptor antagonist with potent sleep-promoting properties. *ChemMedChem* **7**:415–424.
- Costa E and Guidotti A (1979) Molecular mechanisms in the receptor action of benzodiazepines. *Annu Rev Pharmacol Toxicol* **19**:531–545.
- Cox CD, Breslin MJ, Whitman DB, Schreier JD, McGaughey GB, Bogusky MJ, Roecker AJ, Mercer SP, Bednar RA, Lemaire W, et al. (2010) Discovery of the dual orexin receptor antagonist [(7R)-4-(5-chloro-1,3-benzoxazol-2-yl)-7-methyl-1,4-diazepan-1-yl][5-methyl-2-(2H-1,2,3-triazol-2-yl)phenyl]methanone (MK-4305) for the treatment of insomnia. *J Med Chem* **53**:5320–5332.
- Cox CD, McGaughey GB, Bogusky MJ, Whitman DB, Ball RG, Winrow CJ, Renger JJ, and Coleman PJ (2009) Conformational analysis of N,N-disubstituted-1,4-diazepane orexin receptor antagonists and implications for receptor binding. *Bioorg Med Chem Lett* **19**:2997–3001.
- Dall'Aglio C, Pascucci L, Mercati F, Giontella A, Pedini V, and Ceccarelli P (2009) Immunohistochemical identification and localization of orexin A and orexin type 2 receptor in the horse gastrointestinal tract. *Res Vet Sci* **86**:189–193.
- Dall'Aglio C, Pascucci L, Mercati F, Giontella A, Pedini V, Scocco P, and Ceccarelli P (2008) Identification of orexin A- and orexin type 2 receptor-positive cells in the gastrointestinal tract of neonatal dogs. *Eur J Histochem* **52**:229–235.
- Dall'Aglio C, Pedini V, Scocco P, Boiti C, and Ceccarelli P (2010) Immunohistochemical evidence of Orexin-A in the pancreatic beta cells of domestic animals. *Res Vet Sci* **89**:147–149.
- Date Y, Mondal MS, Matsukura S, and Nakazato M (2000) Distribution of orexin-A and orexin-B (hypocretins) in the rat spinal cord. *Neurosci Lett* **288**:87–90.
- De la Herrán-Arita AK, Guerra-Crespo M, and Drucker-Colin R (2011) Narcolepsy and orexins: an example of progress in sleep research. *Front Neurol* **2**:26.
- de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett FS 2nd, et al. (1998) The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* **95**:322–327.
- de Oliveira CV, Rosas-Arellano MP, Solano-Flores LP, and Ciriello J (2003) Cardiovascular effects of hypocretin-1 in nucleus of the solitary tract. *Am J Physiol Heart Circ Physiol* **284**:H1369–H1377.
- Deadwyler SA, Porrino L, Siegel JM, and Hampson RE (2007) Systemic and nasal delivery of orexin-A (Hypocretin-1) reduces the effects of sleep deprivation on cognitive performance in nonhuman primates. *J Neurosci* **27**:14239–14247.
- Deng BS, Nakamura A, Zhang W, Yanagisawa M, Fukuda Y, and Kuwaki T (2007) Contribution of orexin in hypercapnic chemoreflex: evidence from genetic and pharmacological disruption and supplementation studies in mice. *J Appl Physiol* **103**:1772–1779.
- Di Fabio R, Pellacani A, Faedo S, Roth A, Piccoli L, Gerrard P, Porter RA, Johnson CN, Thewlis K, Donati D, et al. (2011) Discovery process and pharmacological characterization of a novel dual orexin 1 and orexin 2 receptor antagonist useful for sleep disorders. *Bioorg Med Chem Lett* **21**:5562–5567.
- Dias MB, Li A, and Nattie E (2010) The orexin receptor 1 (OX1R) in the rostral medullary raphe contributes to the hypercapnic chemoreflex in wakefulness, during the active period of the diurnal cycle. *Respir Physiol Neurobiol* **170**:96–102.
- Dias MB, Li A, and Nattie EE (2009) Antagonism of orexin receptor-1 in the retrotrapezoid nucleus inhibits the ventilatory response to hypercapnia predominantly in wakefulness. *J Physiol* **587**:2059–2067.

- Dietrich H and Jenck F (2010) Intact learning and memory in rats following treatment with the dual orexin receptor antagonist almoxerant. *Psychopharmacology (Berl)* **212**:145–154.
- Dimitrova A, Fronczek R, Van der Ploeg J, Scammell T, Gautam S, Pascual-Leone A, and Lammers GJ (2011) Reward-seeking behavior in human narcolepsy. *J Clin Sleep Med* **7**:293–300.
- Dingemans J, Dorffner G, Hajak G, Benes H, Danker-Hopfe H, Polo O, Saletu B, Barbanjo MJ, Pillar G, Penzel T, et al. (2007) Proof-of-concept study in primary insomnia patients with ACT-078573, a dual orexin receptor antagonist (Abstract). *Sleep Biol Rhythms* **5** (Suppl):P0653 (A194).
- Diniz Behn CG, Klerman EB, Mochizuki T, Lin SC, and Scammell TE (2010) Abnormal sleep/wake dynamics in orexin knockout mice. *Sleep* **33**:297–306.
- Dube MG, Horvath TL, Kalra PS, and Kalra SP (2000) Evidence of NPY Y5 receptor involvement in food intake elicited by orexin A in satiated rats. *Peptides* **21**:1557–1560.
- Dugovic C, Shelton J, Sutton S, Yun S, Li X, Dvorak C, Carruthers N, Atack J, and Lovenberg T (2008) Sleep-inducing effects mediated by selective blockade of orexin OX2 receptors during the light phase in the rat (Abstract). *Sleep* **31**:A33.
- Dugovic C, Shelton JE, Aluisio LE, Fraser IC, Jiang X, Sutton SW, Bonaventure P, Yun S, Li X, Lord B, et al. (2009) Blockade of orexin-1 receptors attenuates orexin-2 receptor antagonism-induced sleep promotion in the rat. *J Pharmacol Exp Ther* **330**:142–151.
- Edwards CM, Abusnana S, Sunter D, Murphy KG, Ghatei MA, and Bloom SR (1999) The effect of the orexins on food intake: comparison with neuropeptide Y, melanin-concentrating hormone and galanin. *J Endocrinol* **160**:R7–R12.
- Ehrström M, Gustafsson T, Finn A, Kirchgessner A, Grybäck P, Jacobsson H, Hellström PM, and Näslund E (2005a) Inhibitory effect of exogenous orexin A on gastric emptying, plasma leptin, and the distribution of orexin and orexin receptors in the gut and pancreas in man. *J Clin Endocrinol Metab* **90**:2370–2377.
- Ehrström M, Levin F, Kirchgessner AL, Schmidt PT, Hilsted LM, Grybäck P, Jacobsson H, Hellström PM, and Näslund E (2005b) Stimulatory effect of endogenous orexin A on gastric emptying and acid secretion independent of gastrin. *Regul Pept* **132**:9–16.
- Ehrström M, Näslund E, Ma J, Kirchgessner AL, and Hellström PM (2003) Physiological regulation and NO-dependent inhibition of migrating myoelectric complex in the rat small bowel by OXA. *Am J Physiol Gastrointest Liver Physiol* **285**:G688–G695.
- Eriksson KS, Sergeeva OA, Selbach O, and Haas HL (2004) Orexin (hypocretin)/dopamine neurons control GABAergic inputs to tuberomammillary neurons. *Eur J Neurosci* **19**:1278–1284.
- España RA, McCormack SL, Mochizuki T, and Scammell TE (2007) Running promotes wakefulness and increases cataplexy in orexin knockout mice. *Sleep* **30**:1417–1425.
- España RA, Plahn S, and Berridge CW (2002) Circadian-dependent and circadian-independent behavioral actions of hypocretin/orexin. *Brain Res* **943**:224–236.
- España RA and Scammell TE (2011) Sleep neurobiology from a clinical perspective. *Sleep* **34**:845–858.
- Fadel J and Deutch AY (2002) Anatomical substrates of orexin-dopamine interactions: lateral hypothalamic projections to the ventral tegmental area. *Neuroscience* **111**:379–387.
- Farr SA, Banks WA, Kumar VB, and Morley JE (2005) Orexin-A-induced feeding is dependent on nitric oxide. *Peptides* **26**:759–765.
- Fenzl T, Romanowski CP, Flachskamm C, Deussing JM, and Kimura M (2011) Wake-promoting effects of orexin: Its independent actions against the background of an impaired corticotropin-releasing hormone receptor system. *Behav Brain Res* **222**:43–50.
- Fortuyn HA, Lappenschaar MA, Furer JW, Hodiament PP, Rijnders CA, Renier WO, Buitelaar JK, and Overeem S (2010) Anxiety and mood disorders in narcolepsy: a case-control study. *Gen Hosp Psychiatry* **32**:49–56.
- Frederick-Duus D, Guyton MF, and Fadel J (2007) Food-elicited increases in cortical acetylcholine release require orexin transmission. *Neuroscience* **149**:499–507.
- Fujiki N, Cheng T, Yoshino F, and Nishino S (2009) Specificity of direct transition from wake to REM sleep in orexin/ataxin-3 transgenic narcoleptic mice. *Exp Neurol* **217**:46–54.
- Fujiki N, Yoshida Y, Ripley B, Honda K, Mignot E, and Nishino S (2001) Changes in CSF hypocretin-1 (orexin A) levels in rats across 24 hours and in response to food deprivation. *Neuroreport* **12**:993–997.
- Fujiki N, Yoshida Y, Ripley B, Mignot E, and Nishino S (2003) Effects of IV and ICV hypocretin-1 (orexin A) in hypocretin receptor-2 gene mutated narcoleptic dogs and IV hypocretin-1 replacement therapy in a hypocretin-ligand-deficient narcoleptic dog. *Sleep* **26**:953–959.
- Funato H, Tsai AL, Willie JT, Kisanuki Y, Williams SC, Sakurai T, and Yanagisawa M (2009) Enhanced orexin receptor-2 signaling prevents diet-induced obesity and improves leptin sensitivity. *Cell Metab* **9**:64–76.
- Furlong TM, Vianna DM, Liu L, and Carrive P (2009) Hypocretin/orexin contributes to the expression of some but not all forms of stress and arousal. *Eur J Neurosci* **30**:1603–1614.
- Georgescu D, Zachariou V, Barrot M, Mieda M, Willie JT, Eisch AJ, Yanagisawa M, Nestler EJ, and DiLeone RJ (2003) Involvement of the lateral hypothalamic peptide orexin in morphine dependence and withdrawal. *J Neurosci* **23**:3106–3111.
- Gestreau C, Bévengut M, and Dutschmann M (2008) The dual role of the orexin/hypocretin system in modulating wakefulness and respiratory drive. *Curr Opin Pulm Med* **14**:512–518.
- Goddard AW, Mason GF, Almai A, Rothman DL, Behar KL, Petroff OA, Charney DS, and Krystal JH (2001) Reductions in occipital cortex GABA levels in panic disorder detected with 1h-magnetic resonance spectroscopy. *Arch Gen Psychiatry* **58**:556–561.
- González JA, Jensen LT, Fugger L, and Burdakov D (2008) Metabolism-independent sugar sensing in central orexin neurons. *Diabetes* **57**:2569–2576.
- Gozzi A, Turrini G, Piccoli L, Massagrande M, Amantini D, Antolini M, Martinelli P, Cesari N, Montanari D, Tessari M, et al. (2011) Functional magnetic resonance imaging reveals different neural substrates for the effects of orexin-1 and orexin-2 receptor antagonists. *Plos One* **6**:e16406.
- Grabauskas G and Moises HC (2003) Gastrointestinal-projecting neurones in the dorsal motor nucleus of the vagus exhibit direct and viscerotopically organized sensitivity to orexin. *J Physiol* **549**:37–56.
- Grudt TJ, van den Pol AN, and Perl ER (2002) Hypocretin-2 (orexin-B) modulation of superficial dorsal horn activity in rat. *J Physiol* **538**:517–525.
- Hagan JJ, Leslie RA, Patel S, Evans ML, Wattam TA, Holmes S, Benham CD, Taylor SG, Routledge C, Hemmati P, et al. (1999) Orexin A activates locus coeruleus cell firing and increases arousal in the rat. *Proc Natl Acad Sci USA* **96**:10911–10916.
- Han F, Mignot E, Wei YC, Dong SX, Li J, Lin L, An P, Wang LH, Wang JS, He MZ, et al. (2010) Ventilatory chemoresponsiveness, narcolepsy-cataplexy and human leukocyte antigen DQB1*0602 status. *Eur Respir J* **36**:577–583.
- Hara J, Beuckmann CT, Nambu T, Willie JT, Chemelli RM, Sinton CM, Sugiyama F, Yagami K, Goto K, Yanagisawa M, et al. (2001) Genetic ablation of orexin neurons in mice results in narcolepsy, hypophagia, and obesity. *Neuron* **30**:345–354.
- Hara J, Yanagisawa M, and Sakurai T (2005) Difference in obesity phenotype between orexin-knockout mice and orexin neuron-deficient mice with same genetic background and environmental conditions. *Neurosci Lett* **380**:239–242.
- Harris GC and Aston-Jones G (2006) Arousal and reward: a dichotomy in orexin function. *Trends Neurosci* **29**:571–577.
- Harris GC, Wimmer M, and Aston-Jones G (2005) A role for lateral hypothalamic orexin neurons in reward seeking. *Nature* **437**:556–559.
- Haynes AC, Chapman H, Taylor C, Moore GB, Cawthorne MA, Tadayyon M, Clapham JC, and Arch JR (2002) Anorectic, thermogenic and anti-obesity activity of a selective orexin-1 receptor antagonist in ob/ob mice. *Regul Pept* **104**:153–159.
- Haynes AC, Jackson B, Chapman H, Tadayyon M, Johns A, Porter RA, and Arch JR (2000) A selective orexin-1 receptor antagonist reduces food consumption in male and female rats. *Regul Pept* **96**:45–51.
- Heinonen MV, Purhonen AK, Makela KA, and Herzog KH (2008) Functions of orexins in peripheral tissues. *Acta Physiol (Oxf)* **192**:471–485.
- Herrington WJ, Budd KS, Hutzelmann J, Snyder E, Snively D, Liu K, Lines C, Michelson D, and Roth T (2010) Efficacy and tolerability of the dual orexin receptor antagonist mk-4305 in patients with primary insomnia: randomized, controlled, adaptive crossover polysomnography study (Abstract A0591). *Sleep* **33** (Suppl): A199.
- Hervieu GJ, Cluderay JE, Harrison DC, Roberts JC, and Leslie RA (2001) Gene expression and protein distribution of the orexin-1 receptor in the rat brain and spinal cord. *Neuroscience* **103**:777–797.
- Hirota K, Kushikata T, Kudo M, Kudo T, Smart D, and Matsuki A (2003) Effects of central hypocretin-1 administration on hemodynamic responses in young-adult and middle-aged rats. *Brain Res* **981**:143–150.
- Hoever P, de Haas S, Winkler J, Schoemaker RC, Chiossi E, van Gerven J, and Dingemans J (2010) Orexin receptor antagonism, a new sleep-promoting paradigm: an ascending single-dose study with almoxerant. *Clin Pharmacol Ther* **87**:593–600.
- Holland PR, Akerman S, and Goadsby PJ (2005) Orexin 1 receptor activation attenuates neurogenic dural vasodilation in an animal model of trigeminovascular nociception. *J Pharmacol Exp Ther* **315**:1380–1385.
- Holland PR, Akerman S, and Goadsby PJ (2006) Modulation of nociceptive dural input to the trigeminal nucleus caudalis by activation of the orexin 1 receptor in the rat. *Eur J Neurosci* **24**:2825–2833.
- Holmgvist T, Akerman KE, and Kukkonen JP (2002) Orexin signaling in recombinant neuron-like cells. *FEBS Lett* **526**:11–14.
- Honda Y, Doi Y, Ninomiya R, and Ninomiya C (1986) Increased frequency of non-insulin-dependent diabetes mellitus among narcoleptic patients. *Sleep* **9**:254–259.
- Hoque R and Chesson AL, Jr. (2009) Zolpidem-induced sleepwalking, sleep related eating disorder, and sleep-driving: fluorine-18-fluorodeoxyglucose positron emission tomography analysis, and a literature review of other unexpected clinical effects of zolpidem. *J Clin Sleep Med* **5**:471–476.
- Huang SC, Dai YW, Lee YH, Chiou LC, and Hwang LL (2010) Orexins depolarize rostral ventrolateral medulla neurons and increase arterial pressure and heart rate in rats mainly via orexin 2 receptors. *J Pharmacol Exp Ther* **334**:522–529.
- Huang W, Ramsey KM, Marcheva B, and Bass J (2011) Circadian rhythms, sleep, and metabolism. *J Clin Invest* **121**:2133–2141.
- Hungs M, Fan J, Lin L, Lin X, Maki RA, and Mignot E (2001) Identification and functional analysis of mutations in the hypocretin (orexin) genes of narcoleptic canines. *Genome Res* **11**:531–539.
- Ida T, Nakahara K, Katayama T, Murakami N, and Nakazato M (1999) Effect of lateral cerebroventricular injection of the appetite-stimulating neuropeptide, orexin and neuropeptide Y, on the various behavioral activities of rats. *Brain Res* **821**:526–529.
- Ida T, Nakahara K, Murakami T, Hanada R, Nakazato M, and Murakami N (2000) Possible involvement of orexin in the stress reaction in rats. *Biochem Biophys Res Commun* **270**:318–323.
- Ishii Y, Blundell JE, Halford JC, Upton N, Porter R, Johns A, and Rodgers RJ (2004) Differential effects of the selective orexin-1 receptor antagonist SB-334867 and lithium chloride on the behavioural satiety sequence in rats. *Physiol Behav* **81**:129–140.
- Ishii Y, Blundell JE, Halford JC, Upton N, Porter R, Johns A, and Rodgers RJ (2005) Satiety enhancement by selective orexin-1 receptor antagonist SB-334867: influence of test context and profile comparison with CCK-8S. *Behav Brain Res* **160**:11–24.
- Ito N, Yabe T, Gamo Y, Nagai T, Oikawa T, Yamada H, and Hanawa T (2008) I.e.v. administration of orexin-A induces an antidepressant-like effect through hippocampal cell proliferation. *Neuroscience* **157**:720–732.
- Jászberényi M, Bujdosó E, and Telegdy G (2001) The role of neuropeptide Y in orexin-induced hypothalamic-pituitary-adrenal activation. *J Neuroendocrinol* **13**:438–441.
- Jenck F, Fischer C, Qiu C, Hess P, Kolberstein R, and Brisbare-Roch C (2007)

- Somnolence induced by pharmacological blockade of both orexin OX1 and OX2 receptors in dogs (Abstract PO271). *Sleep Biol Rhythms* **5** (Suppl 1):A79.
- Jeong Y and Holden JE (2009) The role of spinal orexin-1 receptors in posterior hypothalamic modulation of neuropathic pain. *Neuroscience* **159**:1414–1421.
- John J, Wu MF, Boehmer LN, and Siegel JM (2004) Cataplexy-active neurons in the hypothalamus: implications for the role of histamine in sleep and waking behavior. *Neuron* **42**:619–634.
- Johnson PL, Truitt W, Fitz SD, Minick PE, Dietrich A, Sanghani S, Traskman-Bendz L, Goddard AW, Brundin L, and Shekhar A (2010) A key role for orexin in panic anxiety. *Nature Med* **16**:111–115.
- Kajiyama S, Kawamoto M, Shiraishi S, Gaus S, Matsunaga A, Suyama H, and Yuge O (2005) Spinal orexin-1 receptors mediate anti-hyperalgesic effects of intrathecally-administered orexins in diabetic neuropathic pain model rats. *Brain Res* **1044**:76–86.
- Kalogiannis M, Grupke SL, Potter PE, Edwards JG, Chemelli RM, Kisanuki YY, Yanagisawa M, and Leonard CS (2010) Narcoleptic orexin receptor knockout mice express enhanced cholinergic properties in laterodorsal tegmental neurons. *Eur J Neurosci* **32**:130–142.
- Kalogiannis M, Hsu E, Willie JT, Chemelli RM, Kisanuki YY, Yanagisawa M, and Leonard CS (2011) Cholinergic modulation of narcoleptic attacks in double orexin receptor knockout mice. *PLoS ONE* **6**:e18697.
- Kantor S, Mochizuki T, Janisiewicz AM, Clark E, Nishino S, and Scammell TE (2009) Orexin neurons are necessary for the circadian control of REM sleep. *Sleep* **32**:1127–1134.
- Karteris E, Machado RJ, Chen J, Zervou S, Hillhouse EW, and Randeve HS (2005) Food deprivation differentially modulates orexin receptor expression and signaling in rat hypothalamus and adrenal cortex. *Am J Physiol Endocrinol Metab* **288**:E1089–E1100.
- Karteris E, Randeve HS, Grammatopoulos DK, Jaffe RB, and Hillhouse EW (2001) Expression and coupling characteristics of the CRH and orexin type 2 receptors in human fetal adrenals. *J Clin Endocrinol Metab* **86**:4512–4519.
- Kastin AJ and Akerstrom V (1999) Orexin A but not orexin B rapidly enters brain from blood by simple diffusion. *J Pharmacol Exp Ther* **289**:219–223.
- Katayama Y, Homma T, Honda K, and Hirai K (2003) Actions of orexin-A in the myenteric plexus of the guinea-pig small intestine. *Neuroreport* **14**:1515–1518.
- Kayaba Y, Nakamura A, Kasuya Y, Ohuchi T, Yanagisawa M, Komuro I, Fukuda Y, and Kuwaki T (2003) Attenuated defense response and low basal blood pressure in orexin knockout mice. *Am J Physiol Regul Integr Comp Physiol* **285**:R581–R593.
- Kirchgessner AL (2002) Orexins in the brain-gut axis. *Endocr Rev* **23**:1–15.
- Kirchgessner AL and Liu M (1999) Orexin synthesis and response in the gut. *Neuron* **24**:941–951.
- Kisanuki YY, Chemelli RM, Sinton CM, Williams SC, Richardson JA, Hammer RE, and Yanagisawa M (2000) The role of orexin receptor type-1 (OX1R) in the regulation of sleep (Abstract). *Sleep* **23**:A91.
- Klein DC, Moore RY, and Reppert SM (1991) *Suprachiasmatic Nucleus: The Mind's Clock*. Oxford University Press, New York, NY.
- Kobashi M, Furudono Y, Matsuo R, and Yamamoto T (2002) Central orexin facilitates gastric relaxation and contractility in rats. *Neurosci Lett* **332**:171–174.
- Kotz CM, Mullett MA, and Wang C (2005) Diminished feeding responsiveness to orexin A (hypocretin 1) in aged rats is accompanied by decreased neuronal activation. *Am J Physiol Regul Integr Comp Physiol* **289**:R359–R366.
- Krowicki ZK, Burmeister MA, Berthoud HR, Scullion RT, Fuchs K, and Hornby PJ (2002) Orexins in rat dorsal motor nucleus of the vagus potentially stimulate gastric motor function. *Am J Physiol Gastrointest Liver Physiol* **283**:G465–G472.
- Kukkonen JP and Akerman KE (2001) Orexin receptors couple to Ca²⁺ channels different from store-operated Ca²⁺ channels. *Neuroreport* **12**:2017–2020.
- Lancel M (1999) Role of GABA_A receptors in the regulation of sleep: initial sleep responses to peripherally administered modulators and agonists. *Sleep* **22**:33–42.
- Langmead CJ, Jerman JC, Brough SJ, Scott C, Porter RA, and Herdon HJ (2004) Characterisation of the binding of [³H]-SB-674042, a novel nonpeptide antagonist, to the human orexin-1 receptor. *Br J Pharmacol* **141**:340–346.
- Larsson KP, Peltonen HM, Bart G, Louhivuori LM, Penttonen A, Antikainen M, Kukkonen JP, and Akerman KE (2005) Orexin-A-induced Ca²⁺ entry: evidence for involvement of trpc channels and protein kinase C regulation. *J Biol Chem* **280**:1771–1781.
- Lawrence AJ (2010) Regulation of alcohol-seeking by orexin (hypocretin) neurons. *Brain Res* **1314**:124–129.
- Lectez B, Jeandel L, El-Yamani FZ, Arthaud S, Alexandre D, Mardargent A, Jégou S, Mounien L, Bizet P, Magoul R, et al. (2009) The orexigenic activity of the hypothalamic neuropeptide 26RFa is mediated by the neuropeptide Y and proopiomelanocortin neurons of the arcuate nucleus. *Endocrinology* **150**:2342–2350.
- Lee J, Reddy MM, and Kodadek T (2010) Discovery of an orexin receptor positive potentiator. *Chem Sci* **1**:48–54.
- Li A and Nattie E (2010) Antagonism of rat orexin receptors by almorexant attenuates central chemoreception in wakefulness in the active period of the diurnal cycle. *J Physiol* **588**:2935–2944.
- Li Y, Li S, Wei C, Wang H, Sui N, and Kirouac GJ (2010) Orexins in the paraventricular nucleus of the thalamus mediate anxiety-like responses in rats. *Psychopharmacology* **212**:251–265.
- Li Y, Wang H, Qi K, Chen X, Li S, Sui N, and Kirouac GJ (2011) Orexins in the midline thalamus are involved in the expression of conditioned place aversion to morphine withdrawal. *Physiol Behav* **102**:42–50.
- Lin J, Dauviliers Y, Arnulf I, Analet C, Parmentier R, Ligneau X, Lecomte J, and Schwartz J (2008) An inverse agonist of the histamine H3-receptor improves wakefulness in narcolepsy: study in orexin-/- mice and patients (Abstract 0009). *Sleep* **31** (Suppl):A3.
- Lin JS, Sakai K, and Jouvet M (1988) Evidence for histaminergic arousal mechanisms in the hypothalamus of cat. *Neuropharmacology* **27**:111–122.
- Lin L, Faraco J, Li R, Kadotani H, Rogers W, Lin X, Qiu X, de Jong PJ, Nishino S, and Mignot E (1999) The sleep disorder canine narcolepsy is caused by a mutation in the hypocretin (orexin) receptor 2 gene. *Cell* **98**:365–376.
- Liu J (2009), inventor. Tetrahydroisoquinoline derivatives. U.S. patent application no. 20090192188. 2009 Jul 30.
- Liu M, Blanco-Centurion C, Konadhode R, Begum S, Pelluru D, Gerashchenko D, Sakurai T, Yanagisawa M, van den Pol AN, and Shiromani PJ (2011) Orexin gene transfer into zona incerta neurons suppresses muscle paralysis in narcoleptic mice. *J Neurosci* **31**:6028–6040.
- Liu RJ, van den Pol AN, and Aghajanian GK (2002) Hypocretins (orexins) regulate serotonin neurons in the dorsal raphe nucleus by excitatory direct and inhibitory indirect actions. *J Neurosci* **22**:9453–9464.
- Louhivuori LM, Jansson L, Nordström T, Bart G, Näsman J, and Akerman KE (2010) Selective interference with TRPC3/6 channels disrupts OX1 receptor signalling via NCX and reveals a distinct calcium influx pathway. *Cell Calcium* **48**:114–123.
- Louis GW, Leininger GM, Rhodes CJ, and Myers MG Jr (2010) Direct innervation and modulation of orexin neurons by lateral hypothalamic LepRb neurons. *J Neurosci* **30**:11278–11287.
- Lubkin M and Stricker-Krongrad A (1998) Independent feeding and metabolic actions of orexins in mice. *Biochem Biophys Res Commun* **253**:241–245.
- Lund PE, Shariatmadari R, Uustare A, Detheux M, Parmentier M, Kukkonen JP, and Akerman KE (2000) The orexin OX1 receptor activates a novel Ca²⁺ influx pathway necessary for coupling to phospholipase C. *J Biol Chem* **275**:30806–30812.
- Lutter M, Krishnan V, Russo SJ, Jung S, McClung CA, and Nestler EJ (2008) Orexin signaling mediates the antidepressant-like effect of calorie restriction. *J Neurosci* **28**:3071–3075.
- Machado BH, Bonagamba LG, Dun SL, Kwok EH, and Dun NJ (2002) Pressor response to microinjection of orexin/hypocretin into rostral ventrolateral medulla of awake rats. *Regul Pept* **104**:75–81.
- Malherbe P, Borroni E, Gobbi L, Knust H, Nettekoven M, Pinard E, Roche O, Rogers-Evans M, Wettstein JG, and Moreau JL (2009) Biochemical and behavioural characterization of EMPA, a novel high-affinity, selective antagonist for the OX(2) receptor. *Br J Pharmacol* **156**:1326–1341.
- Malherbe P, Roche O, Marcuz A, Kratzstein C, Wettstein JG, and Bissantz C (2010) Mapping the binding pocket of dual antagonist almorexant to human orexin 1 and orexin 2 receptors: comparison with the selective OX1 antagonist SB-674042 and the selective OX2 antagonist N-ethyl-2-[(6-methoxy-pyridin-3-yl)-(toluene-2-sulfonyl)-amino]-N-pyridin-3-ylmethyl-acetamide (EMPA). *Mol Pharmacol* **78**:81–93.
- Marcus JN, Aschkenasi CJ, Lee CE, Chemelli RM, Saper CB, Yanagisawa M, and Elmquist JK (2001) Differential expression of orexin receptors 1 and 2 in the rat brain. *J Comp Neurol* **435**:6–25.
- McAtee LC, Sutton SW, Rudolph DA, Li X, Aluisio LE, Phuong VK, Dvorak CA, Lovenberg TW, Carruthers NI, and Jones TK (2004) Novel substituted 4-phenyl-[1,3]dioxanes: potent and selective orexin receptor 2 (OX2R) antagonists. *Bioorg Med Chem Lett* **14**:4225–4229.
- Mieda M, Willie JT, Hara J, Sinton CM, Sakurai T, and Yanagisawa M (2004) Orexin peptides prevent cataplexy and improve wakefulness in an orexin neuron-ablated model of narcolepsy in mice. *Proc Natl Acad Sci USA* **101**:4649–4654.
- Miyasaka K, Masuda M, Kanai S, Sato N, Kurosawa M, and Funakoshi A (2002) Central Orexin-A stimulates pancreatic exocrine secretion via the vagus. *Pancreas* **25**:400–404.
- Mobarakeh JI, Takahashi K, Sakurada S, Nishino S, Watanabe H, Kato M, Naghdi N, and Yanai K (2005a) Enhanced antinociception by intracerebroventricularly administered orexin A in histamine H1 or H2 receptor gene knockout mice. *Pain* **118**:254–262.
- Mobarakeh JI, Takahashi K, Sakurada S, Nishino S, Watanabe H, Kato M, and Yanai K (2005b) Enhanced antinociception by intracerebroventricularly and intrathecally-administered orexin A and B (hypocretin-1 and -2) in mice. *Peptides* **26**:767–777.
- Mochizuki T, Arrigoni E, Marcus JN, Clark EL, Yamamoto M, Honer M, Borroni E, Lowell BB, Elmquist JK, and Scammell TE (2011) Orexin receptor 2 expression in the posterior hypothalamus rescues sleepiness in narcoleptic mice. *Proc Natl Acad Sci USA* **108**:4471–4476.
- Mochizuki T, Crocker A, McCormack S, Yanagisawa M, Sakurai T, and Scammell TE (2004) Behavioral state instability in orexin knock-out mice. *J Neurosci* **24**:6291–6300.
- Mochizuki T and Scammell TE (2003) Orexin/hypocretin: wired for wakefulness. *Curr Biol* **13**:R563–R564.
- Monti JM, Pellejero T, and Jantos H (1986) Effects of H1- and H2-histamine receptor agonists and antagonists on sleep and wakefulness in the rat. *J Neural Transm* **66**:1–11.
- Morawska M, Buchi M, and Fendt M (2011) Narcoleptic episodes in orexin-deficient mice are increased by both attractive and aversive odors. *Behav Brain Res* **222**:397–400.
- Morganstern I, Chang GQ, Karatayev O, and Leibowitz SF (2010) Increased orexin and melanin-concentrating hormone expression in the perifornical lateral hypothalamus of rats prone to overconsuming a fat-rich diet. *Pharmacol Biochem Behav* **96**:413–422.
- Moriya R, Sano H, Umeda T, Ito M, Takahashi Y, Matsuda M, Ishihara A, Kanatani A, and Iwaasa H (2006) RFamide peptide QRFP43 causes obesity with hyperphagia and reduced thermogenesis in mice. *Endocrinology* **147**:2916–2922.
- Muroya S, Uramura K, Sakurai T, Takigawa M, and Yada T (2001) Lowering glucose concentrations increases cytosolic Ca²⁺ in orexin neurons of the rat lateral hypothalamus. *Neurosci Lett* **309**:165–168.
- Nair SG, Golden SA, and Shaham Y (2008) Differential effects of the hypocretin 1 receptor antagonist SB 334867 on high-fat food self-administration and reinstatement of food seeking in rats. *Br J Pharmacol* **154**:406–416.
- Nakabayashi M, Suzuki T, Takahashi K, Totsune K, Muramatsu Y, Kaneko C, Date F, Takeyama J, Darnel AD, Moriya T, et al. (2003) Orexin-A expression in human peripheral tissues. *Mol Cell Endocrinol* **205**:43–50.
- Nakamura A, Zhang W, Yanagisawa M, Fukuda Y, and Kuwaki T (2007) Vigilance

- state-dependent attenuation of hypercapnic chemoreflex and exaggerated sleep apnea in orexin knockout mice. *J Appl Physiol* **102**:241–248.
- Näslund E, Ehrström M, Ma J, Hellström PM, and Kirchgessner AL (2002) Localization and effects of orexin on fasting motility in the rat duodenum. *Am J Physiol Gastrointest Liver Physiol* **282**:G470–G479.
- Näsman J, Bart G, Larsson K, Louhivuori L, Peltonen H, and Akerman KE (2006) The orexin OX1 receptor regulates Ca^{2+} entry via diacylglycerol-activated channels in differentiated neuroblastoma cells. *J Neurosci* **26**:10658–10666.
- Nattie E and Li A (2010) Central chemoreception in wakefulness and sleep: evidence for a distributed network and a role for orexin. *J Appl Physiol* **108**:1417–1424.
- Niimi M, Sato M, and Taminato T (2001) Neuropeptide Y in central control of feeding and interactions with orexin and leptin. *Endocrine* **14**:269–273.
- Nishino S (2005) The canine model of narcolepsy, in *Hypocretins: Integrators of Physiological Functions* (de Lecea L and Sutcliffe G eds) p. 39, Springer, New York.
- Nishino S and Mignot E (1997) Pharmacological aspects of human and canine narcolepsy. *Prog Neurobiol* **52**:27–78.
- Nishino S, Okuro M, Kotorii N, Aneqawa E, Ishimaru Y, Matsumura M, and Kanbayashi T (2010) Hypocretin/orexin and narcolepsy: new basic and clinical insights. *Acta Physiol (Oxf)* **198**:209–222.
- Nishino S, Ripley B, Overeem S, Lammers GJ, and Mignot E (2000) Hypocretin (orexin) deficiency in human narcolepsy. *Lancet* **355**:39–40.
- Nisoli E, Carruba MO, Valerio A, and Borsani G (1998) Hypocretins or hypocretins? *Nat Med* **4**:645.
- Nutt DJ and Stahl SM (2010) Searching for perfect sleep: the continuing evolution of GABA_A receptor modulators as hypnotics. *J Psychopharmacol* **24**:1601–1612.
- Ohno K and Sakurai T (2008) Orexin neuronal circuitry: role in the regulation of sleep and wakefulness. *Front Neuroendocrinol* **29**:70–87.
- Okumura T and Takakusaki K (2008) Role of orexin in central regulation of gastrointestinal functions. *J Gastroenterol* **43**:652–660.
- Olds J and Milner P (1954) Positive reinforcement produced by electrical stimulation of septal area and other regions of rat brain. *J Comp Physiol* **47**:419–427.
- Orlando G, Brunetti L, Di Nisio C, Michelotto B, Recinella L, Ciabattoni G, and Vacca M (2001) Effects of cocaine- and amphetamine-regulated transcript peptide, leptin and orexins on hypothalamic serotonin release. *Eur J Pharmacol* **430**:269–272.
- Ozcan M, Ayar A, Serhatlioglu I, Alcin E, Sahin Z, and Kelestimur H (2010) Orexins activates protein kinase C-mediated Ca^{2+} signaling in isolated rat primary sensory neurons. *Physiol Res* **59**:255–262.
- Peng HY, Chang HM, Chang SY, Tung KC, Lee SD, Chou D, Lai CY, Chiu CH, Chen GD, and Lin TB (2008) Orexin-A modulates glutamatergic NMDA-dependent spinal reflex potentiation via inhibition of NR2B subunit. *Am J Physiol Endocrinol Metab* **295**:E117–E129.
- Peyron C, Faraco J, Rogers W, Ripley B, Overeem S, Charnay Y, Nevsimanova S, Aldrich M, Reynolds D, Albin R, et al. (2000) A mutation in a case of early onset narcolepsy and a generalized absence of hypocretin peptides in human narcoleptic brains. *Nature Med* **6**:991–997.
- Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, and Kilduff TS (1998) Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* **18**:9996–10015.
- Pinessi L, Binello E, De Martino P, Gallone S, Gentile S, Rainero I, Rivoiro C, Rubino E, Savi L, Valfrè W, et al. (2007) The 1246G→A polymorphism of the HCRTR2 gene is not associated with migraine. *Cephalalgia* **27**:945–949.
- Piper DC, Upton N, Smith MI, and Hunter AJ (2000) The novel brain neuropeptide, orexin-A, modulates the sleep-wake cycle of rats. *Eur J Neurosci* **12**:726–730.
- Ponz A, Khatami R, Poryazova R, Werth E, Boesiger P, Bassetti CL, and Schwartz S (2010) Abnormal activity in reward brain circuits in human narcolepsy with cataplexy. *Ann Neurol* **67**:190–200.
- Porkka-Heiskanen T, Strecker RE, and McCarley RW (2000) Brain site-specificity of extracellular adenosine concentration changes during sleep deprivation and spontaneous sleep: an in vivo microdialysis study. *Neuroscience* **99**:507–517.
- Putula J, Turunen PM, Johansson L, Näsman J, Ra R, Korhonen L, and Kukkonen JP (2011) Orexin/hypocretin receptor chimera reveals structural features important for orexin peptide distinction. *FEBS Lett* **585**:1368–1374.
- Rachalski A, Alexandre C, Bernard JF, Saurini F, Lesch KP, Hamon M, Adrien J, and Fabre V (2009) Altered sleep homeostasis after restraint stress in 5-HTT knock-out male mice: a role for hypocretins. *J Neurosci* **29**:15575–15585.
- Rainero I, Gallone S, Valfrè W, Ferrero M, Angilella G, Rivoiro C, Rubino E, De Martino P, Savi L, Ferrone M, et al. (2004) A polymorphism of the hypocretin receptor 2 gene is associated with cluster headache. *Neurology* **63**:1286–1288.
- Rainero I, Rubino E, Gallone S, Fenoglio P, Picci LR, Giobbe L, Ostacoli L, and Pinessi L (2011) Evidence for an association between migraine and the hypocretin receptor 1 gene. *J Headache Pain* **12**:193–199.
- Rainero I, Rubino E, Valfrè W, Gallone S, De Martino P, Zampella E, and Pinessi L (2007) Association between the G1246A polymorphism of the hypocretin receptor 2 gene and cluster headache: a meta-analysis. *J Headache Pain* **8**:152–156.
- Renger JJ, Dunn SL, Motzel SL, Johnson C, and Koblan KS (2004) Sub-chronic administration of zolpidem affects modifications to rat sleep architecture. *Brain Res* **1010**:45–54.
- Renzulli C, Nash M, Wright M, Thomas S, Zamuner S, Pellegatti M, Bettica P, and Boyle G (2011) Disposition and metabolism of [14 C]SB-649868, an orexin 1 and 2 receptor antagonist, in humans. *Drug Metab Dispos* **39**:215–227.
- Ripley B, Fujiki N, Okura M, Mignot E, and Nishino S (2001) Hypocretin levels in sporadic and familial cases of canine narcolepsy. *Neurobiol Dis* **8**:525–534.
- Rodgers RJ, Halford JC, Nunes de Souza RL, Canto de Souza AL, Piper DC, Arch JR, and Blundell JE (2000) Dose-response effects of orexin-A on food intake and the behavioural satiety sequence in rats. *Regul Pept* **96**:71–84.
- Rodgers RJ, Halford JC, Nunes de Souza RL, Canto de Souza AL, Piper DC, Arch JR, Upton N, Porter RA, Johns A, and Blundell JE (2001) SB-334867, a selective orexin-1 receptor antagonist, enhances behavioural satiety and blocks the hyperphagic effect of orexin-A in rats. *Eur J Neurosci* **13**:1444–1452.
- Rodgers RJ, Ishii Y, Halford JC, and Blundell JE (2002) Orexins and appetite regulation. *Neuropeptides* **36**:303–325.
- Rolls A, Schaich Borg J, and de Lecea L (2010) Sleep and metabolism: role of hypothalamic neuronal circuitry. *Best Pract Res Clin Endocrinol Metab* **24**:817–828.
- Rosenbaum DM, Cherezov V, Hanson MA, Rasmussen SG, Thian FS, Kobilka TS, Choi HJ, Yao XJ, Weis WI, Stevens RC, et al. (2007) GPCR engineering yields high-resolution structural insights into beta2-adrenergic receptor function. *Science* **318**:1266–1273.
- Sakurai T (2002) Roles of orexins in the regulation of feeding and arousal. *Sleep Med* **3** (Suppl 2):S3–S9.
- Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, et al. (1998) Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* **92**:573–585.
- Sakurai T and Mieda M (2011) Connectomics of orexin-producing neurons: interface of systems of emotion, energy homeostasis and arousal. *Trends Pharmacol Sci* **32**:451–462.
- Sakurai T, Mieda M and Masashi Y (2005a) Rodent models of human narcolepsy-cataplexy, in *Hypocretins: Integrators of Physiological Functions* (de Lecea L and Sutcliffe G eds) p. 27, Springer, New York.
- Sakurai T, Mieda M, and Tsujino N (2010) The orexin system: roles in sleep/wake regulation. *Ann NY Acad Sci* **1200**:149–161.
- Sakurai T, Moriguchi T, Furuya K, Kajiwara N, Nakamura T, Yanagisawa M, and Goto K (1999) Structure and function of human prepro-orexin gene. *J Biol Chem* **274**:17771–17776.
- Sakurai T, Nagata R, Yamanaka A, Kawamura H, Tsujino N, Muraki Y, Kageyama H, Kunita S, Takahashi S, Goto K, et al. (2005b) Input of orexin/hypocretin neurons revealed by a genetically encoded tracer in mice. *Neuron* **46**:297–308.
- Salomon RM, Ripley B, Kennedy JS, Johnson B, Schmidt D, Zeitzer JM, Nishino S, and Mignot E (2003) Diurnal variation of cerebrospinal fluid hypocretin-1 (Orexin-A) levels in control and depressed subjects. *Biol Psychiatry* **54**:96–104.
- Samson WK, Bagley SL, Ferguson AV, and White MM (2007) Hypocretin/orexin type 1 receptor in brain: role in cardiovascular control and the neuroendocrine response to immobilization stress. *Am J Physiol Regul Integr Comp Physiol* **292**:R382–R387.
- Samson WK, Gosnell B, Chang JK, Resch ZT, and Murphy TC (1999) Cardiovascular regulatory actions of the hypocretins in brain. *Brain Res* **831**:248–253.
- Saper CB, Fuller PM, Pedersen NP, Lu J, and Scammell TE (2010) Sleep state switching. *Neuron* **68**:1023–1042.
- Saper CB, Lu J, Chou TC, and Gooley J (2005a) The hypothalamic integrator for circadian rhythms. *Trends Neurosci* **28**:152–157.
- Saper CB, Scammell TE, and Lu J (2005b) Hypothalamic regulation of sleep and circadian rhythms. *Nature* **437**:1257–1263.
- Sarchielli P, Rainero I, Coppola F, Rossi C, Mancini M, Pinessi L, and Calabresi P (2008) Involvement of corticotrophin-releasing factor and orexin-A in chronic migraine and medication-overuse headache: findings from cerebrospinal fluid. *Cephalalgia* **28**:714–722.
- Sato-Suzuki I, Kita I, Seki Y, Oguri M, and Arita H (2002) Cortical arousal induced by microinjection of orexins into the paraventricular nucleus of the rat. *Behav Brain Res* **128**:169–177.
- Satoh S, Matsumura H, Kanbayashi T, Yoshida Y, Urakami T, Nakajima T, Kimura N, Nishino S, and Yoneda H (2006) Expression pattern of FOS in orexin neurons during sleep induced by an adenosine A(2A) receptor agonist. *Behav Brain Res* **170**:277–286.
- Scammell TE, Willie JT, Guilleminault C, Siegel JM, and International Working Group on Rodent Models of Narcolepsy (2009) A consensus definition of cataplexy in mouse models of narcolepsy. *Sleep* **32**:111–116.
- Scammell TE and Winrow CJ (2011) Orexin receptors: pharmacology and therapeutic opportunities. *Annu Rev Pharmacol Toxicol* **51**:243–266.
- Schmidt FM, Brügel M, Kratzsch J, Strauss M, Sander C, Baum P, Thierry J, Hegerl U, and Schönknecht P (2010) Cerebrospinal fluid hypocretin-1 (orexin A) levels in mania compared to unipolar depression and healthy controls. *Neurosci Lett* **483**: 20–22.
- Schuld A, Blum WF, and Pollmächer T (2002) Low cerebrospinal fluid hypocretin (orexin) and altered energy homeostasis in human narcolepsy. *Ann Neurol* **51**: 660–661.
- Schuld A, Hebebrand J, Geller F, and Pollmacher T (2000) Increased body-mass index in patients with narcolepsy. *Lancet* **355**:1274–1275.
- Schürks M, Kurth T, Geissler I, Tessmann G, Diener HC, and Roskopf D (2006) Cluster headache is associated with the G1246A polymorphism in the hypocretin receptor 2 gene. *Neurology* **66**:1917–1919.
- Schürks M, Kurth T, Geissler I, Tessmann G, Diener HC, and Roskopf D (2007a) The G1246A polymorphism in the hypocretin receptor 2 gene is not associated with treatment response in cluster headache. *Cephalalgia* **27**:363–367.
- Schürks M, Limmroth V, Geissler I, Tessmann G, Savidou I, Engelbergs J, Kurth T, Diener HC, and Roskopf D (2007b) Association between migraine and the G1246A polymorphism in the hypocretin receptor 2 gene. *Headache* **47**:1195–1199.
- Schwimmer H, Stauss HM, Abboud F, Nishino S, Mignot E, and Zeitzer JM (2010) Effects of sleep on the cardiovascular and thermoregulatory systems: a possible role for hypocretins. *J Appl Physiol* **109**:1053–1063.
- Schwimmer H, Zeitzer JM, Masashi Y, Nishino S, Shengwen Z, and Mignot E (2006) Hypocretin/orexin effects on cardiovascular regulation during sleep (Abstract 0094). *Sleep* **29**:A31.
- Scott MM, Marcus JN, Pettersen A, Birnbaum SG, Mochizuki T, Scammell TE, Nestler EJ, Elmqvist JK, and Lutter M (2011) Hcrtr1 and 2 signaling differentially regulates depression-like behaviors. *Behav Brain Res* **222**:289–294.
- Shahid IZ, Rahman AA, and Pilowsky PM (2011) Intrathecal orexin A increases sympathetic outflow and respiratory drive, enhances baroreflex sensitivity and blocks the somato-sympathetic reflex. *Br J Pharmacol* **162**:961–973.
- Sharf R, Sarhan M, and Dileone RJ (2010) Role of orexin/hypocretin in dependence and addiction. *Brain Res* **1314**:130–138.

- Shirasaka T, Nakazato M, Matsukura S, Takasaki M, and Kannan H (1999) Sympathetic and cardiovascular actions of orexins in conscious rats. *Am J Physiol* **277**:R1780–R1785.
- Shoblock JR, Welty N, Aluisio L, Fraser I, Motley ST, Morton K, Palmer J, Bonaventure P, Carruthers NI, Lovenberg TW, et al. (2011) Selective blockade of the orexin-2 receptor attenuates ethanol self-administration, place preference, and reinstatement. *Psychopharmacology* **215**:191–203.
- Siegel JM and Boehmer LN (2006) Narcolepsy and the hypocretin system—where motion meets emotion. *Nat Clin Pract Neurol* **2**:548–556.
- Singareddy R, Uhde T, and Commissaris R (2006) Differential effects of hypocretins on noise-alone versus potentiated startle responses. *Physiol Behav* **89**:650–655.
- Smart D, Jerman JC, Brough SJ, Neville WA, Jewitt F, and Porter RA (2000) The hypocretins are weak agonists at recombinant human orexin-1 and orexin-2 receptors. *Br J Pharmacol* **129**:1289–1291.
- Smart D, Jerman JC, Brough SJ, Rushton SL, Murdock PR, Jewitt F, Elshourbagy NA, Ellis CE, Middlemiss DN, and Brown F (1999) Characterization of recombinant human orexin receptor pharmacology in a Chinese hamster ovary cell-line using FLIPR. *Br J Pharmacol* **128**:1–3.
- Smart D, Sabido-David C, Brough SJ, Jewitt F, Johns A, Porter RA, and Jerman JC (2001) SB-334867-A: the first selective orexin-1 receptor antagonist. *Br J Pharmacol* **132**:1179–1182.
- Smith PM, Connolly BC, and Ferguson AV (2002) Microinjection of orexin into the rat nucleus tractus solitarius causes increases in blood pressure. *Brain Res* **950**:261–267.
- Smith PM, Samson WK, and Ferguson AV (2007) Cardiovascular actions of orexin-A in the rat subfornical organ. *J Neuroendocrinol* **19**:7–13.
- Sonka K, Kemlink D, Busková J, Pretl M, Srůtková Z, Maurovich Horvat E, Vodicka P, Poláková V, and Nevšimalová S (2010) Obesity accompanies narcolepsy with cataplexy but not narcolepsy without cataplexy. *Neuro Endocrinol Lett* **31**:631–634.
- Steiner MA, Lecourt H, Strasser DS, Brisbane-Roch C, and Jenck F (2011) Differential effects of the dual orexin receptor antagonist almorexant and the GABA(A)-alpha1 receptor modulator zolpidem, alone or combined with ethanol, on motor performance in the rat. *Neuropsychopharmacology* **36**:848–856.
- Strawn JR, Pyne-Geithman GJ, Ekhardt NN, Horn PS, Uhde TW, Shutter LA, Baker DG, and Geraciotti TD Jr (2010) Low cerebrospinal fluid and plasma orexin-A (hypocretin-1) concentrations in combat-related posttraumatic stress disorder. *Psychoneuroendocrinology* **35**:1001–1007.
- Sullivan SS and Guilleminault C (2009) Emerging drugs for insomnia: new frontiers for old and novel targets. *Expert Opin Emerg Drugs* **14**:411–422.
- Suzuki M, Beuckmann CT, Shikata K, Ogura H, and Sawai T (2005) Orexin-A (hypocretin-1) is possibly involved in generation of anxiety-like behavior. *Brain Res* **1044**:116–121.
- Sweet DC, Levine AS, Billington CJ, and Kotz CM (1999) Feeding response to central orexins. *Brain Res* **821**:535–538.
- Taheri S, Sunter D, Dakin C, Moyes S, Seal L, Gardiner J, Rossi M, Gbatei M, and Bloom S (2000) Diurnal variation in orexin A immunoreactivity and prepro-orexin mRNA in the rat central nervous system. *Neurosci Lett* **279**:109–112.
- Takahashi N, Okumura T, Yamada H, and Kohgo Y (1999) Stimulation of gastric acid secretion by centrally administered orexin-A in conscious rats. *Biochem Biophys Res Commun* **254**:623–627.
- Takano S, Kanai S, Hosoya H, Ohta M, Uematsu H, and Miyasaka K (2004) Orexin-A does not stimulate food intake in old rats. *Am J Physiol Gastrointest Liver Physiol* **287**:G1182–G1187.
- Takayasu S, Sakurai T, Iwasaki S, Teranishi H, Yamanaka A, Williams SC, Iguchi H, Kawasawa YI, Ikeda Y, Sakakibara I, et al. (2006) A neuropeptide ligand of the G protein-coupled receptor GPR103 regulates feeding, behavioral arousal, and blood pressure in mice. *Proc Natl Acad Sci USA* **103**:7438–7443.
- Tan KR, Rudolph U, and Lüscher C (2011) Hooked on benzodiazepines: GABA receptor subtypes and addiction. *Trends Neurosci* **34**:188–197.
- Thakkar MM, Engemann SC, Walsh KM, and Sahota PK (2008) Adenosine and the homeostatic control of sleep: effects of A1 receptor blockade in the perifornical lateral hypothalamus on sleep-wakefulness. *Neuroscience* **153**:875–880.
- Thankachan S, Kaur S, and Shiromani PJ (2009) Activity of pontine neurons during sleep and cataplexy in hypocretin knock-out mice. *J Neurosci* **29**:1580–1585.
- Thannickal TC, Siegel JM, Nienhuis R, and Moore RY (2003) Pattern of hypocretin (orexin) soma and axon loss, and gliosis, in human narcolepsy. *Brain Pathol* **13**:340–351.
- Thorpe AJ, Doane DF, Sweet DC, Beverly JL, and Kotz CM (2006) Orexin A in the rostralateral hypothalamic area induces feeding by modulating GABAergic transmission. *Brain Res* **1125**:60–66.
- Thorpe AJ and Kotz CM (2005) Orexin A in the nucleus accumbens stimulates feeding and locomotor activity. *Brain Res* **1050**:156–162.
- Thorpe AJ, Teske JA, and Kotz CM (2005) Orexin A-induced feeding is augmented by caloric challenge. *Am J Physiol Regul Integr Comp Physiol* **289**:R367–R372.
- Thuau R, Guilhaudis L, Ségalas-Milazzo I, Chartrel N, Oulyadi H, Boivin S, Fournier A, Leprieux J, Dévoust D, and Vaudry H (2005) Structural studies on 26RFa, a novel human RFamide-related peptide with orexigenic activity. *Peptides* **26**:779–789.
- Tran DT, Bonaventure P, Hack M, Mirzadegan T, Dvorak C, Letavic M, Carruthers N, Lovenberg T, and Sutton SW (2011) Chimeric, mutant orexin receptors show key interactions between orexin receptors, peptides and antagonists. *Eur J Pharmacol* **667**:120–128.
- Trivedi P, Yu H, MacNeil DJ, Van der Ploeg LH, and Guan XM (1998) Distribution of orexin receptor mRNA in the rat brain. *FEBS Lett* **438**:71–75.
- Tsujino N and Sakurai T (2009) Orexin/hypocretin: a neuropeptide at the interface of sleep, energy homeostasis, and reward system. *Pharmacol Rev* **61**:162–176.
- Tsuneki H, Wada T, and Sasaoka T (2010) Role of orexin in the regulation of glucose homeostasis. *Acta Physiol (Oxf)* **198**:335–348.
- Uramura K, Funahashi H, Muroya S, Shioda S, Takigawa M, and Yada T (2001) Orexin-a activates phospholipase C- and protein kinase C-mediated Ca^{2+} signaling in dopamine neurons of the ventral tegmental area. *Neuroreport* **12**:1885–1889.
- van den Pol AN (1999) Hypothalamic hypocretin (orexin): robust innervation of the spinal cord. *J Neurosci* **19**:3171–3182.
- van den Pol AN, Gao XB, Obrietan K, Kilduff TS, and Belousov AB (1998) Presynaptic and postsynaptic actions and modulation of neuroendocrine neurons by a new hypothalamic peptide, hypocretin/orexin. *J Neurosci* **18**:7962–7971.
- Van Den Pol AN, Patrylo PR, Ghosh PK, and Gao XB (2001) Lateral hypothalamus: early developmental expression and response to hypocretin (orexin). *J Comp Neurol* **433**:349–363.
- Velley L, Chaminade C, Roy MT, Kempf E, and Cardo B (1983) Intrinsic neurons are involved in lateral hypothalamic self-stimulation. *Brain Res* **268**:79–86.
- Voisin T, Rouet-Benzineb P, Reuter N, and Laburthe M (2003) Orexins and their receptors: structural aspects and role in peripheral tissues. *Cell Mol Life Sci* **60**:72–87.
- Watanabe S, Kuwaki T, Yanagisawa M, Fukuda Y, and Shimoyama M (2005) Persistent pain and stress activate pain-inhibitory orexin pathways. *Neuroreport* **16**:5–8.
- Welsh DK, Logothetis DE, Meister M, and Reppert SM (1995) Individual neurons dissociated from rat suprachiasmatic nucleus express independently phased circadian firing rhythms. *Neuron* **14**:697–706.
- White MM, Bagley SL, and Samson WK (2006) Orexin/hypocretin receptor subtypes: cardiovascular and behavioral effects. *FASEB J* **20**:A366.
- Williams KS and Behn CG (2011) Dynamic interactions between orexin and dynorphin may delay onset of functional orexin effects: a modeling study. *J Biol Rhythms* **26**:171–181.
- Williams RH, Jensen LT, Verkhatsky A, Fugger L, and Burdakov D (2007) Control of hypothalamic orexin neurons by acid and CO₂. *Proc Natl Acad Sci USA* **104**:10685–10690.
- Willie JT, Chemelli RM, Sinton CM, Tokita S, Williams SC, Kisanuki YY, Marcus JN, Lee C, Elmquist JK, Kohlmeier KA, et al. (2003) Distinct narcolepsy syndromes in OREXIN receptor-2 and Orexin null mice: molecular genetic dissection of non-REM and REM sleep regulatory processes. *Neuron* **38**:715–730.
- Willie JT, Chemelli RM, Sinton CM, and Yanagisawa M (2001) To eat or to sleep? Orexin in the regulation of feeding and wakefulness. *Annu Rev Neurosci* **24**:429–458.
- Winrow CJ, Gotter AL, Cox CD, Doran SM, Tannenbaum PL, Breslin MJ, Garson SL, Fox SV, Harrell CM, Stevens J, et al. (2011) Promotion of sleep by suvorexant—a novel dual orexin receptor antagonist. *J Neurogenet* **25**:52–61.
- Winrow CJ, Gotter AL, Cox CD, Tannenbaum PL, Garson SL, Doran SM, Breslin MJ, Schreier JD, Fox SV, Harrell CM, et al. (2012) Pharmacological characterization of MK-6096—a dual orexin receptor antagonist for insomnia. *Neuropharmacology* **62**:978–987.
- Wong KK, Ng SY, Lee LT, Ng HK, and Chow BK (2011) Orexins and their receptors from fish to mammals: a comparative approach. *Gen Comp Endocrinol* **171**:124–130.
- Wortley KE, Chang GQ, Davydova Z, and Leibowitz SF (2003) Peptides that regulate food intake: orexin gene expression is increased during states of hypertriglyceridemia. *Am J Physiol Regul Integr Comp Physiol* **284**:R1454–R1465.
- Wu MF, John J, Maidment N, Lam HA, and Siegel JM (2002) Hypocretin release in normal and narcoleptic dogs after food and sleep deprivation, eating, and movement. *Am J Physiol Regul Integr Comp Physiol* **283**:R1079–R1086.
- Wu X, Gao J, Yan J, Ouyang C, and Li Y (2004) Hypothalamus-brain stem circuitry responsible for vagal efferent signaling to the pancreas evoked by hypoglycemia in rat. *J Neurophysiol* **91**:1734–1747.
- Xie X, Wisor JP, Hara J, Crowder TL, LeWinter R, Khroyan TV, Yamanaka A, Diano S, Horvath TL, Sakurai T, et al. (2008) Hypocretin/orexin and nociceptin/orphanin FQ coordinately regulate analgesia in a mouse model of stress-induced analgesia. *J Clin Invest* **118**:2471–2481.
- Yamada H, Okumura T, Motomura W, Kobayashi Y, and Kohgo Y (2000) Inhibition of food intake by central injection of anti-orexin antibody in fasted rats. *Biochem Biophys Res Commun* **267**:527–531.
- Yamada H, Takahashi N, Tanno S, Nagamine M, Takakusaki K, and Okumura T (2005) A selective orexin-1 receptor antagonist, SB334867, blocks 2-DG-induced gastric acid secretion in rats. *Neurosci Lett* **376**:137–142.
- Yamamoto T, Miyazaki R, and Yamada T (2009) Intracerebroventricular administration of 26RFa produces an analgesic effect in the rat formalin test. *Peptides* **30**:1683–1688.
- Yamamoto T, Nozaki-Taguchi N, and Chiba T (2002) Analgesic effect of intrathecally administered orexin-A in the rat formalin test in the rat hot plate test. *Br J Pharmacol* **137**:170–176.
- Yamamoto T, Saito O, Shono K, Aoe T, and Chiba T (2003a) Anti-mechanical allodynic effect of intrathecal and intracerebroventricular injection of orexin-A in the rat neuropathic pain model. *Neurosci Lett* **347**:183–186.
- Yamamoto T, Saito O, Shono K, and Hirasawa S (2003b) Activation of spinal orexin-1 receptor produces anti-allodynic effect in the rat carrageenan test. *Eur J Pharmacol* **481**:175–180.
- Yamamoto T, Wada T, and Miyazaki R (2008) Analgesic effects of intrathecally administered 26RFa, an intrinsic agonist for GPR103, on formalin test and carrageenan test in rats. *Neuroscience* **157**:214–222.
- Yamanaka A, Beuckmann CT, Willie JT, Hara J, Tsujino N, Mieda M, Tominaga M, Yagami K, Sugiyama F, Goto K, et al. (2003) Hypothalamic orexin neurons regulate arousal according to energy balance in mice. *Neuron* **38**:701–713.
- Yamanaka A, Tsujino N, Funahashi H, Honda K, Guan JL, Wang QP, Tominaga M, Goto K, Shioda S, and Sakurai T (2002) Orexins activate histaminergic neurons via the orexin 2 receptor. *Biochem Biophys Res Commun* **290**:1237–1245.
- Yan JA, Ge L, Huang W, Song B, Chen XW, and Yu ZP (2008) Orexin affects dorsal root ganglion neurons: a mechanism for regulating the spinal nociceptive processing. *Physiol Res* **57**:797–800.
- Yanagisawa M (2010) Small-molecule agonists for type-2 orexin receptor. U.S. Patent application no. 20100150840. 2010 Jun 17.

- Yi CX, Serlie MJ, Ackermans MT, Foppen E, Buijs RM, Sauerwein HP, Fliers E, and Kalsbeek A (2009) A major role for perifornical orexin neurons in the control of glucose metabolism in rats. *Diabetes* **58**:1998–2005.
- Yoshida K, McCormack S, España RA, Crocker A, and Scammell TE (2006) Afferents to the orexin neurons of the rat brain. *J Comp Neurol* **494**:845–861.
- Zeitler JM, Buckmaster CL, Parker KJ, Hauck CM, Lyons DM, and Mignot E (2003) Circadian and homeostatic regulation of hypocretin in a primate model: implications for the consolidation of wakefulness. *J Neurosci* **23**:3555–3560.
- Zhang S, Lin L, Kaur S, Thankachan S, Blanco-Centurion C, Yanagisawa M, Mignot E, and Shiromani PJ (2007) The development of hypocretin (orexin) deficiency in hypocretin/ataxin-3 transgenic rats. *Neuroscience* **148**:34–43.
- Zhang W, Fukuda Y, and Kuwaki T (2005) Respiratory and cardiovascular actions of orexin-A in mice. *Neurosci Lett* **385**:131–136.
- Zhang W, Sakurai T, Fukuda Y, and Kuwaki T (2006) Orexin neuron-mediated skeletal muscle vasodilation and shift of baroreflex during defense response in mice. *Am J Physiol Regul Integr Comp Physiol* **290**:R1654–R1663.
- Zhou L, Wei-Lun S, and See RE (2011) Orexin receptor targets for anti-relapse medication development in drug addiction. *Pharmaceuticals* **4**:804–821.
- Zhu Y, Miwa Y, Yamanaka A, Yada T, Shibahara M, Abe Y, Sakurai T, and Goto K (2003) Orexin receptor type-1 couples exclusively to pertussis toxin-insensitive G-proteins, while orexin receptor type-2 couples to both pertussis toxin-sensitive and -insensitive G-proteins. *J Pharmacol Sci* **92**:259–266.