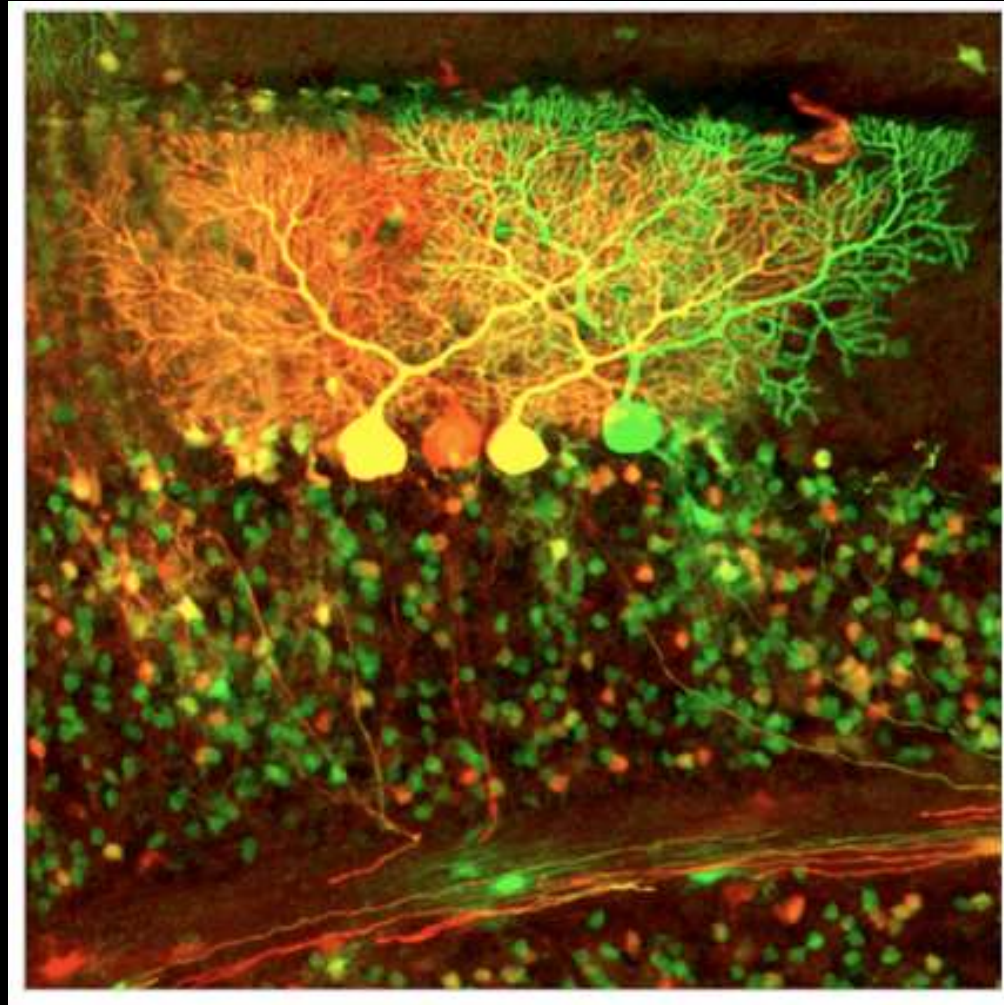


# Techniques in Probing Neural Circuits



Dayu Lin (Dayu.lin@nyumc.org)

# Basic strategies in studying Neural circuits

- **Visualize cells**
- **Monitor cell activity**
- **Manipulate cell and gene activity**

# Visualize cells

- Any cells
- Cells with certain genetic features
- Cells with certain developmental lineage
- Connected cells

# Visualize cells

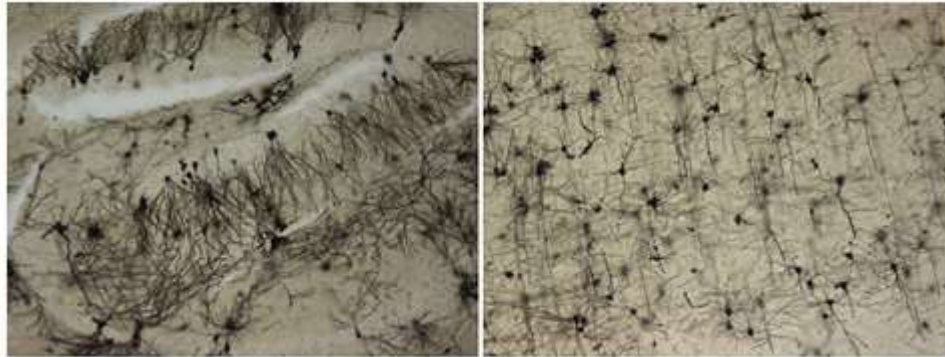
- Any cells
- Cells with certain genetic features
- Cells with certain developmental lineage
- Connected cells

# Methods to simply label cells

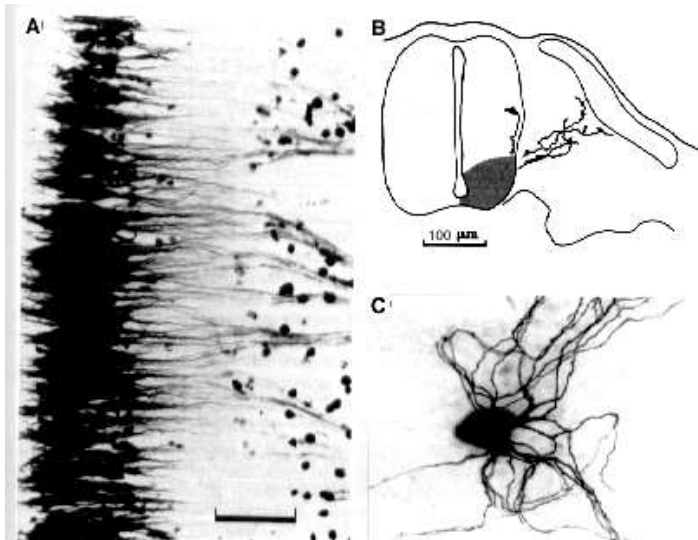
Golgi staining (Silver chromate,  $\text{Ag}_2\text{CrO}_4$ )

Hippocampus

Cortex



horseradish peroxidase (HRP)



Other tracers

subunit B of cholera toxin (CTB)

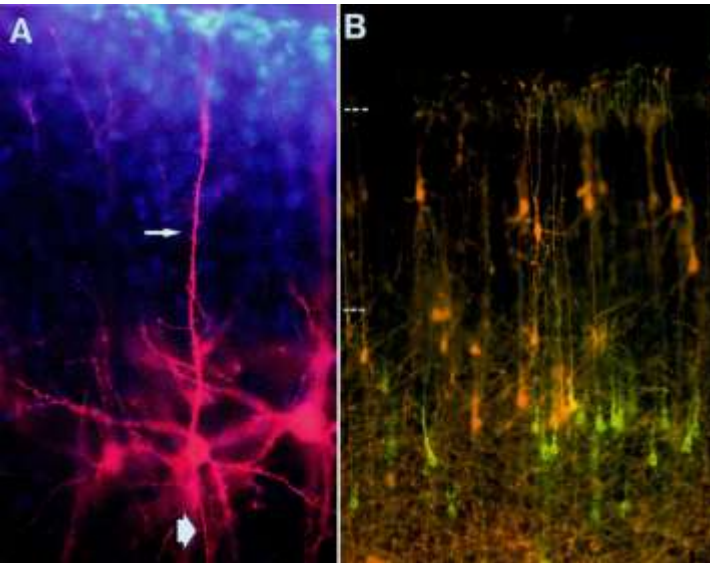
*Phaseolus vulgaris* leucoagglutinin (PHA-L)—a plant lectin

Biotinylated dextran amine (BDA)

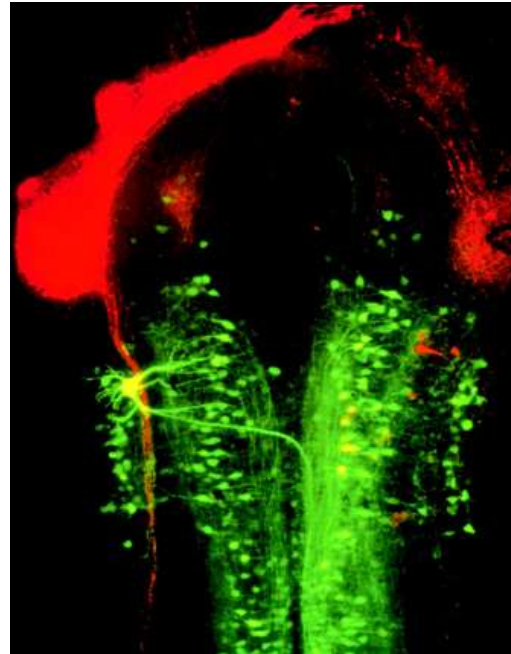
Biocytin and neurobiotin ( a small molecule)

# Conventional methods for neuroanatomy-Fluorescent dyes

Lipophilic dyes



Fluorescent dextrans  
(Lucifer Yellow, Texas Red, fluorescein)

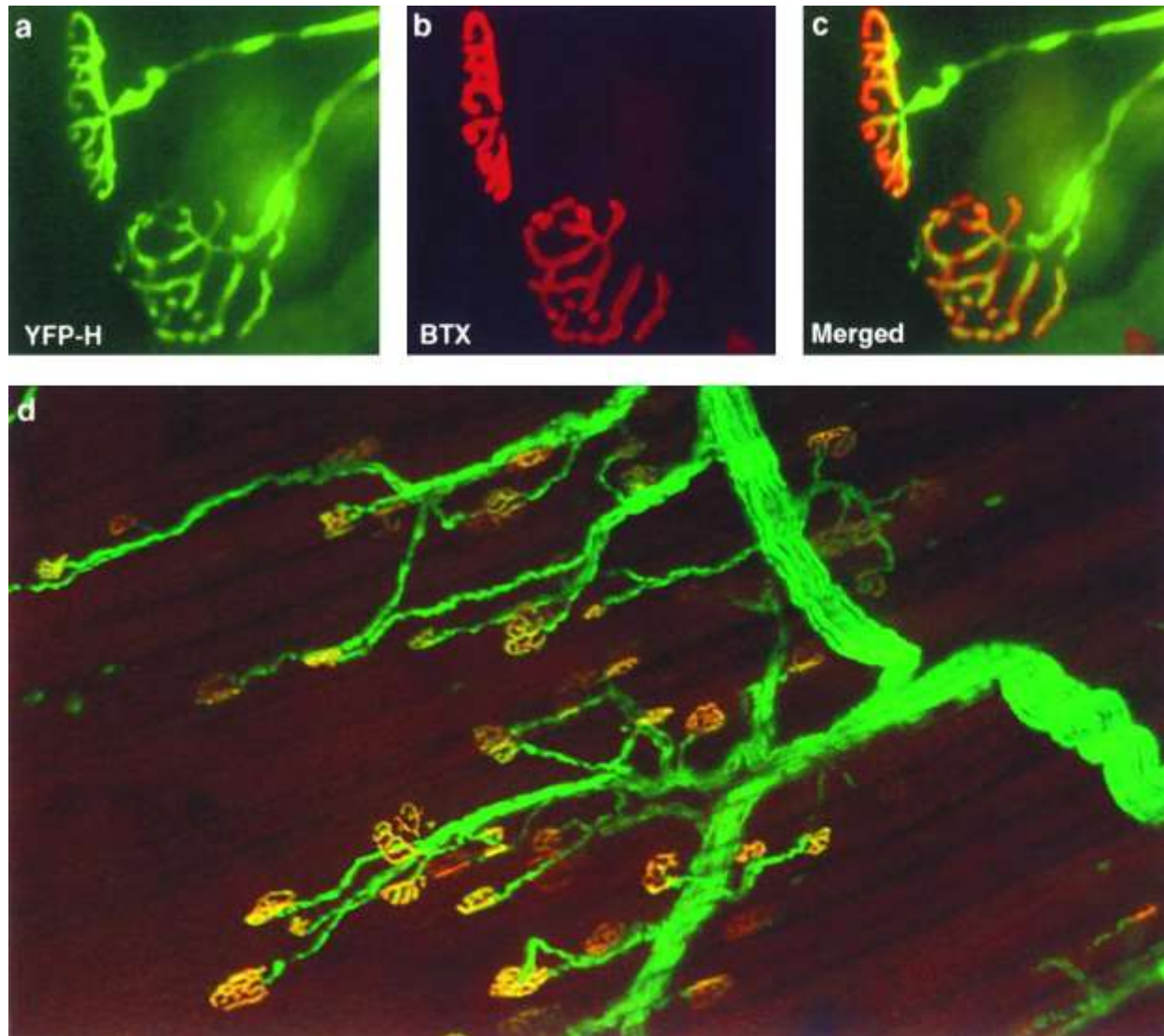


fluorescent microspheres





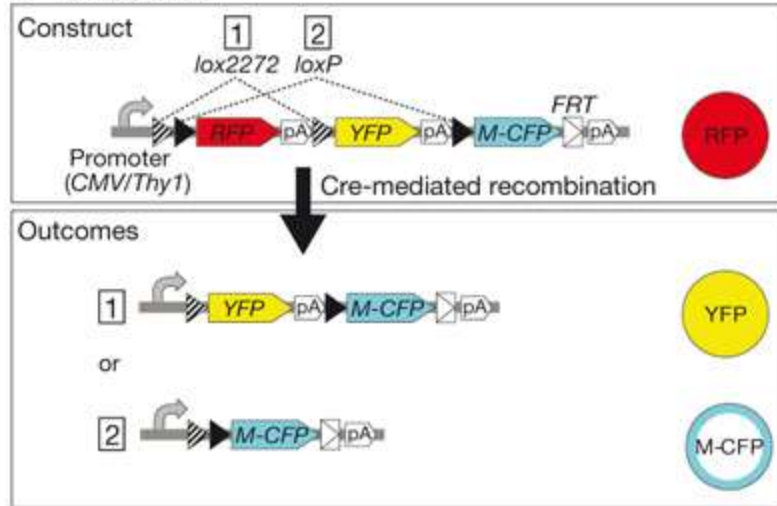
# Label cells reproducibly--Thy1-GFP mice



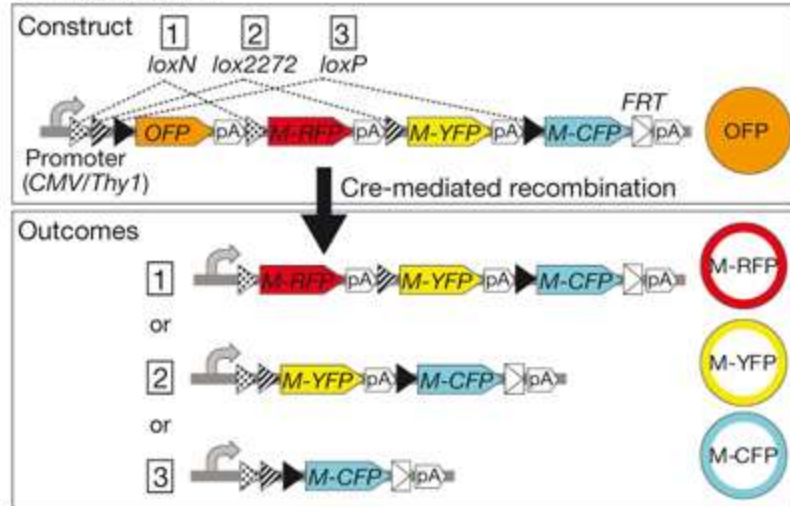
Feng et al, 2000, Neuron,  
Cite >1600

# Label many cells but differently—Brainbow mice

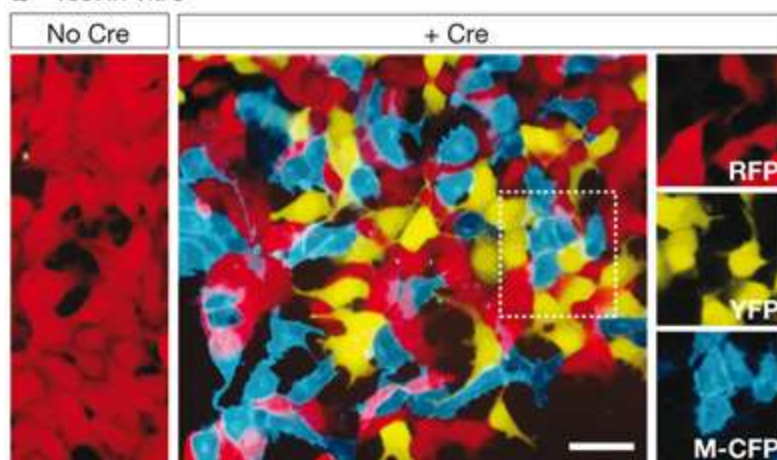
**a** Brainbow-1.0



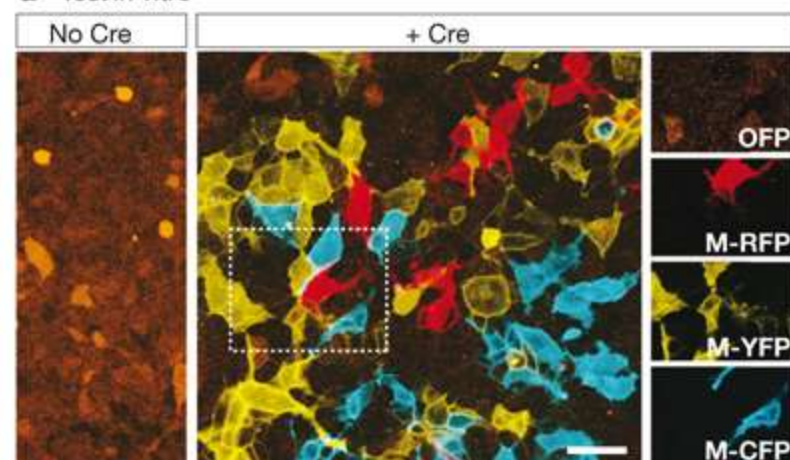
**c** Brainbow-1.1



**b** Test *in vitro*



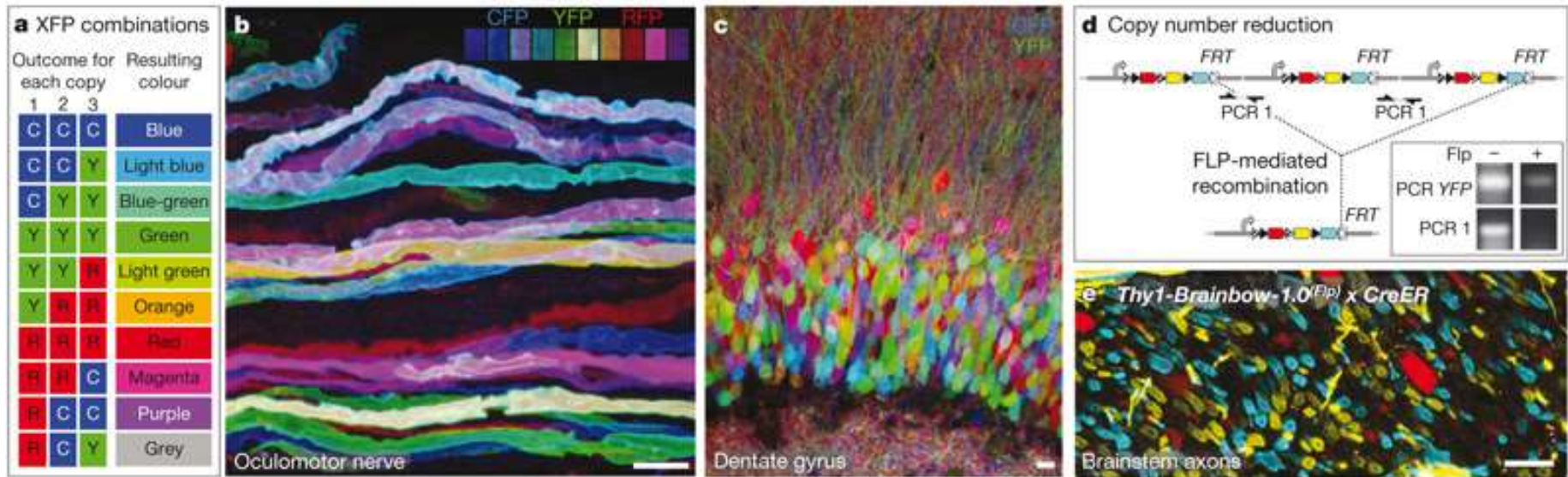
**d** Test *in vitro*



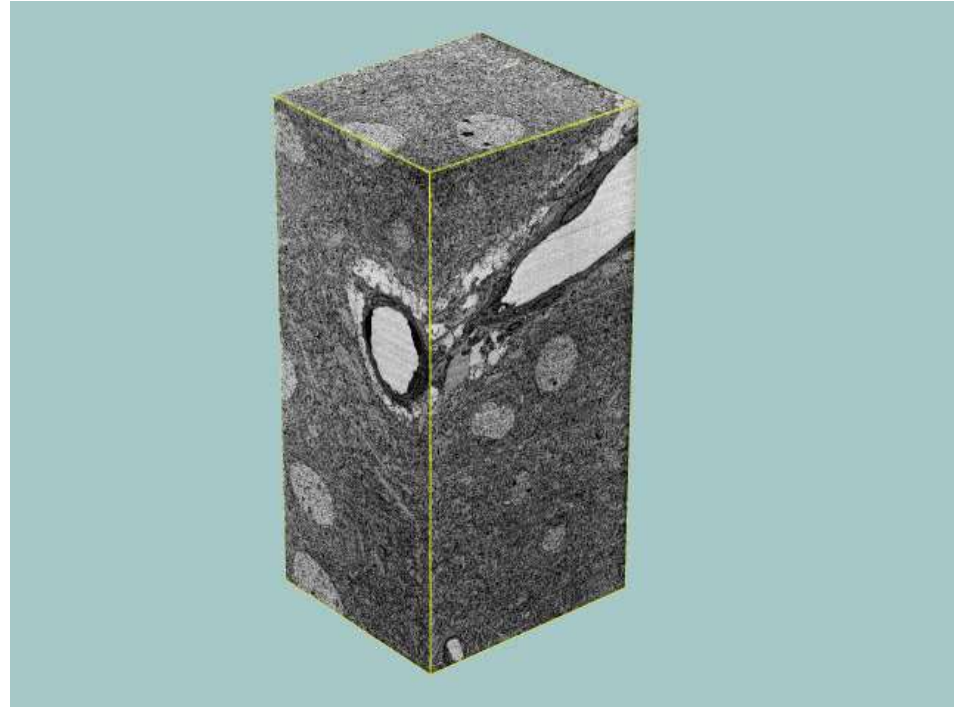
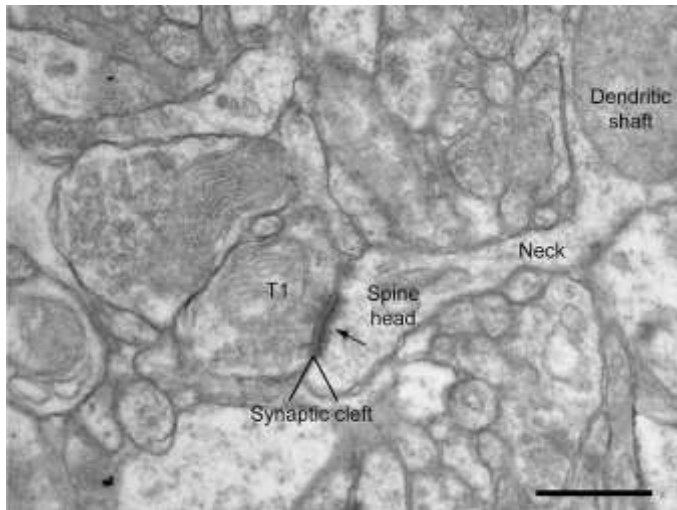
Livet et al, Nature, 2007, Cite ~800  
 Jeff W. Lichtman and Joshua R. Sanes



# Membrane bound GFP facilitated neuroanatomy— Brainbow mice



# Electron microscope (EM)--Connectome



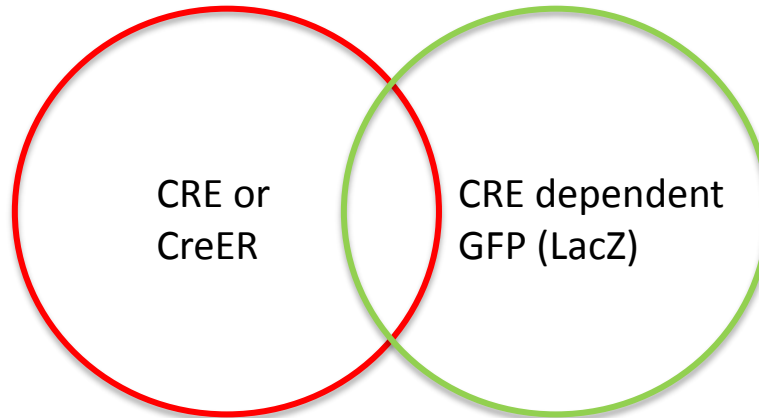
<http://www.cgl.ucsf.edu/chimera/animations/ratbrain/ratbrain.html>

# Visualize cells

- Any cells
- Cells with certain genetic features
- Cells with certain developmental lineage
- Connected cells

# Label genetically defined cell type

mice

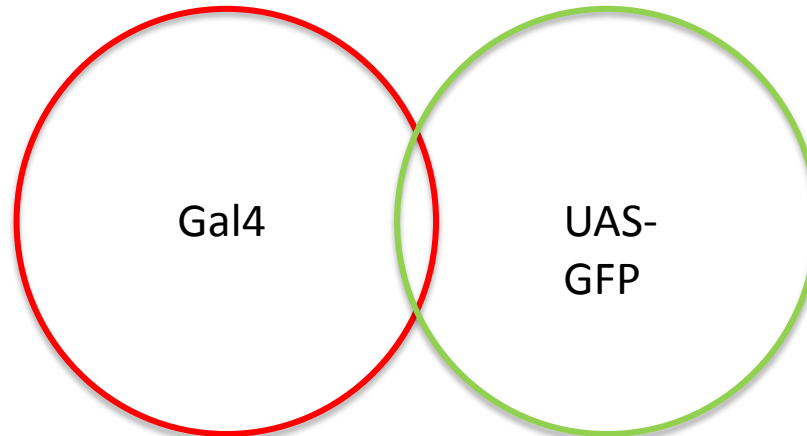


<http://cre.jax.org/index.html>

<http://www.gensat.org/cre.jsp>

<http://connectivity.brain-map.org/transgenic>

fly



<http://flystocks.bio.indiana.edu/>

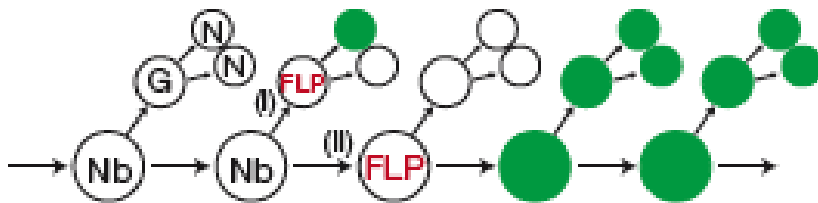
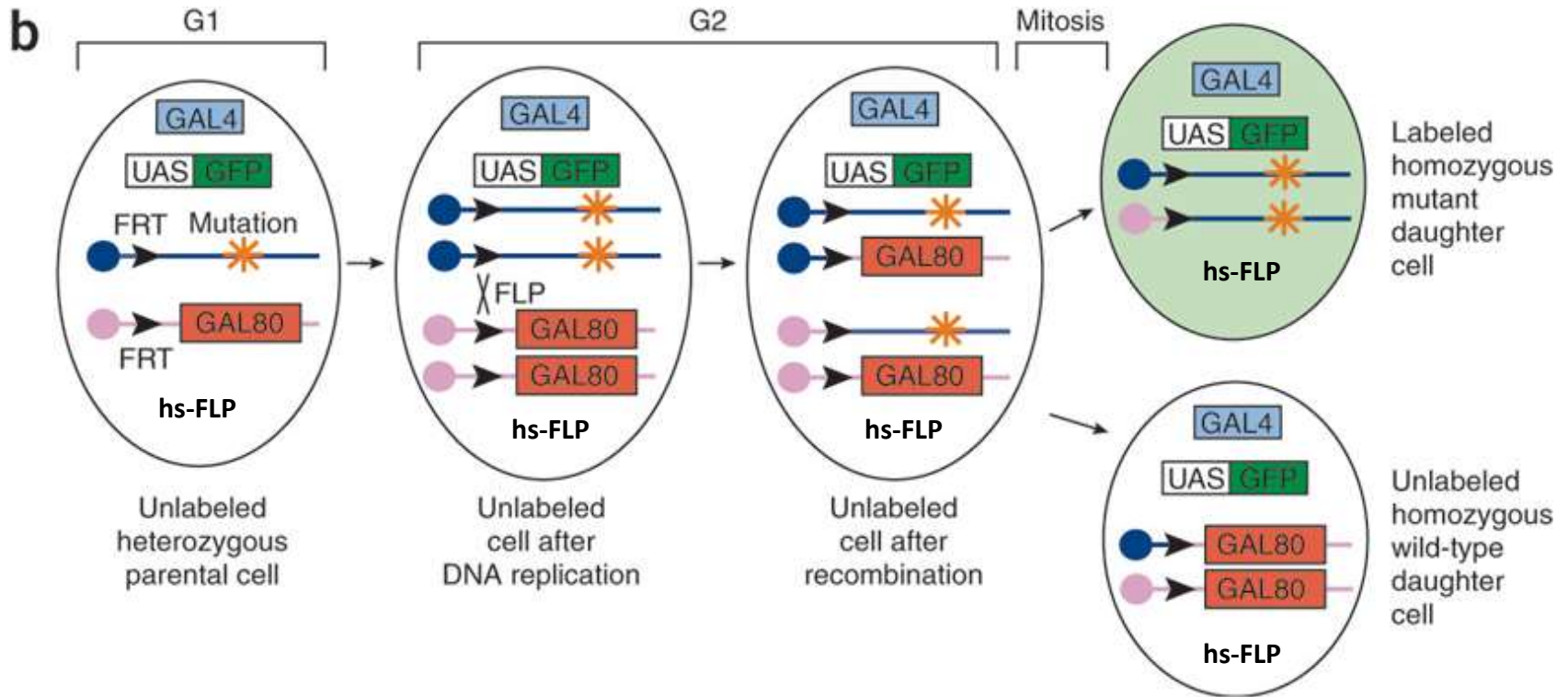
# Visualize cells

- Any cells
- Cells with certain genetic features
- Cells with certain developmental lineage
- Connected cells



# MARCM

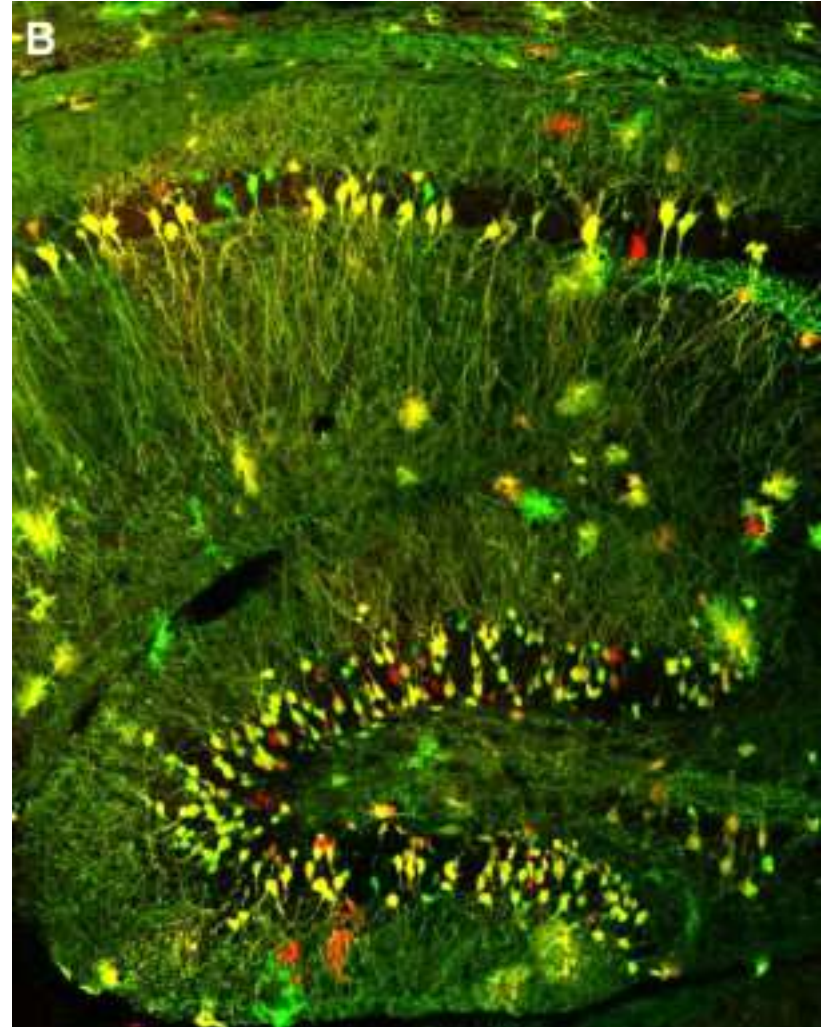
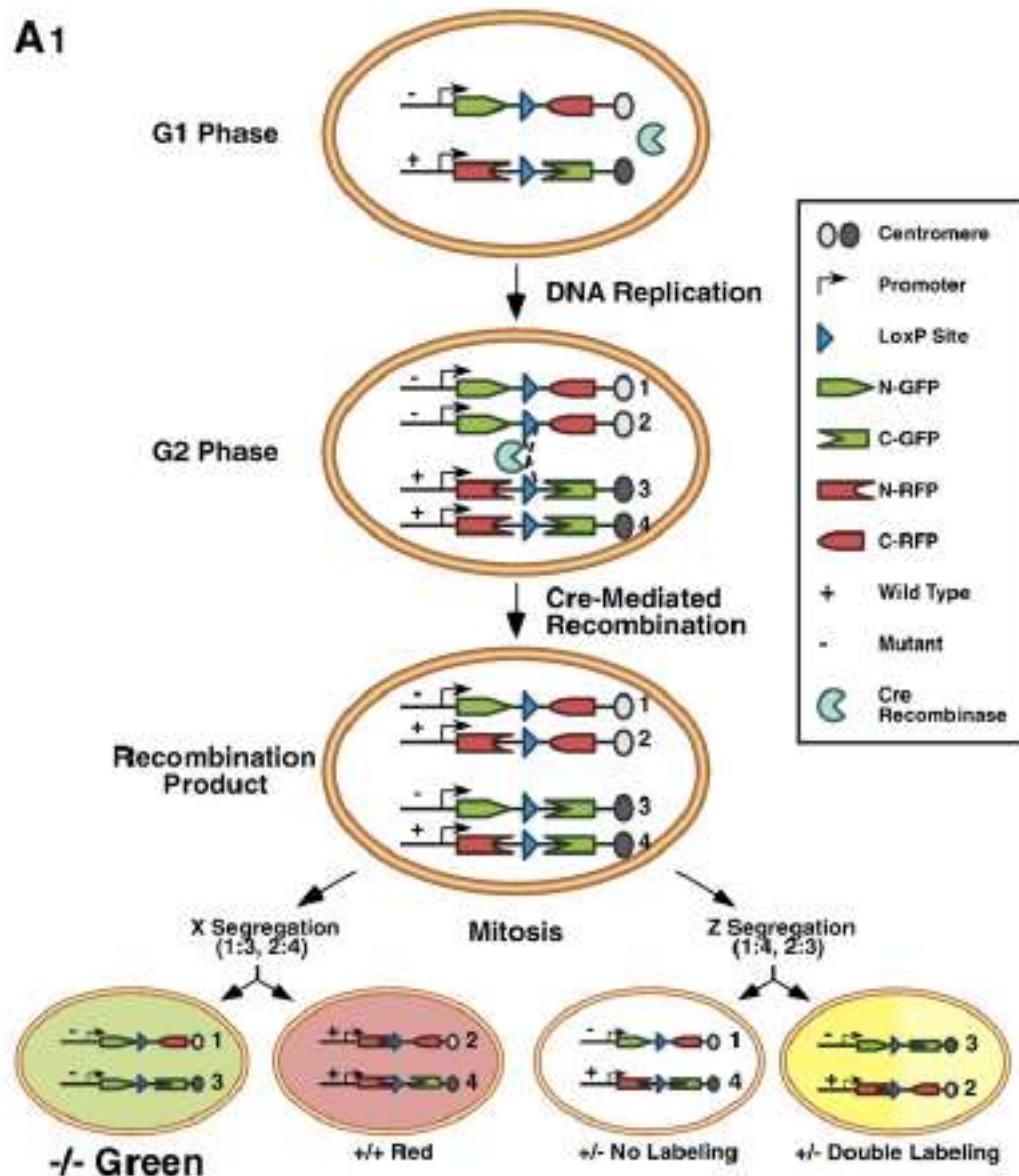
## Mosaic Analysis with a Repressible Cell Marker– Label one or set of mutant cell with GFP



T Lee, Trends Neuroscience, 2001  
Cite>500

# MADM: Mosaic Analysis with Double Markers

A1



# Visualize cells

- Any cells
- Cells with certain genetic features
- Cells with certain developmental lineage
- **Connected cells**

# Label cells transneuronally

WGA: wheat germ agglutinin (an antegrade and retrograde transneuronal tracer)

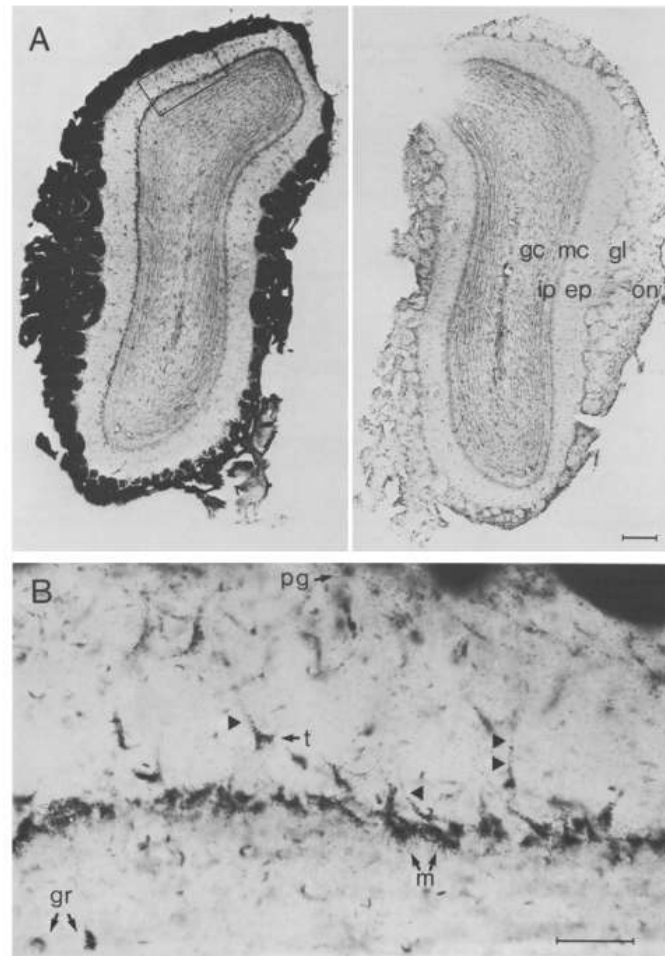


Fig. 1

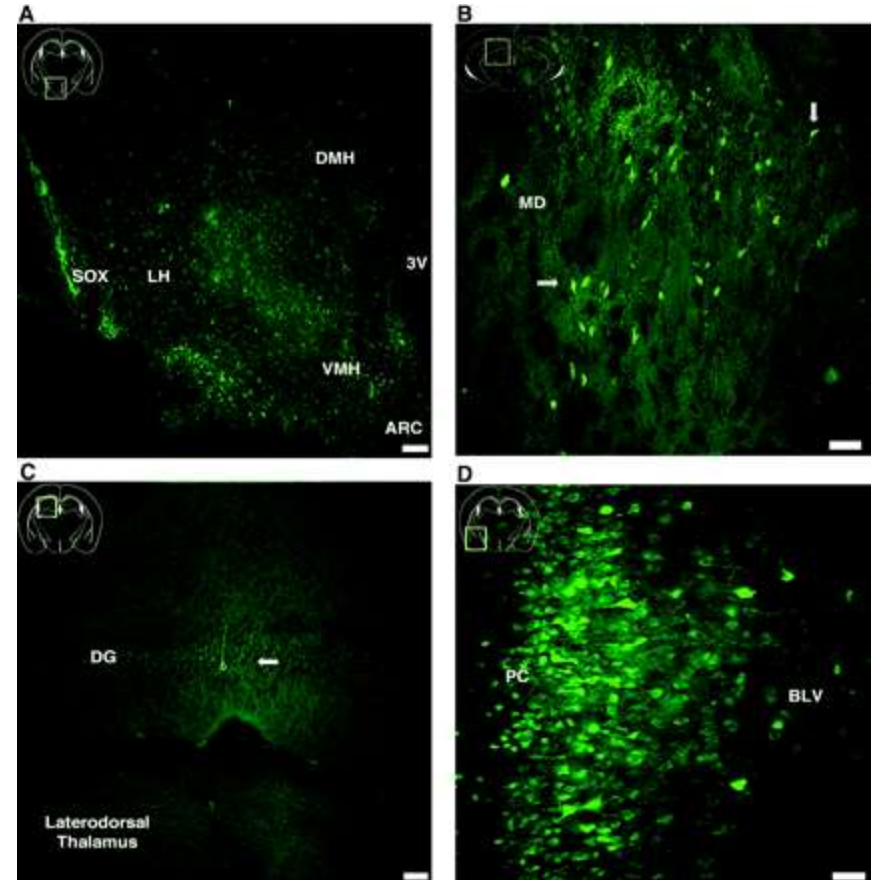
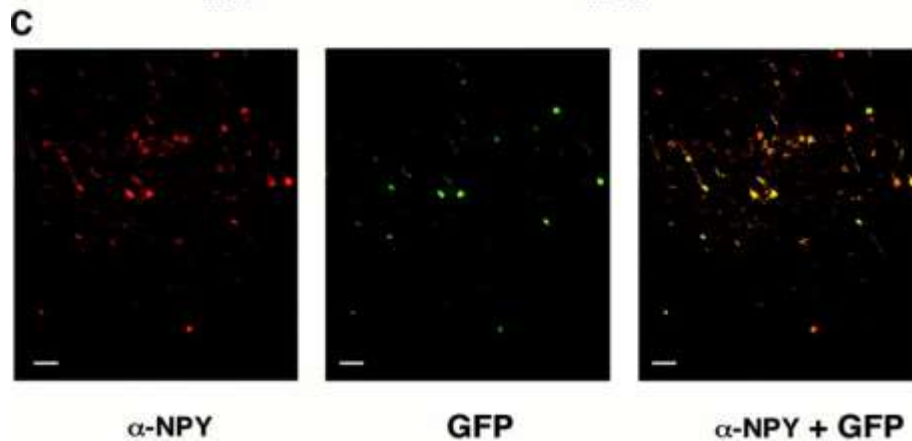
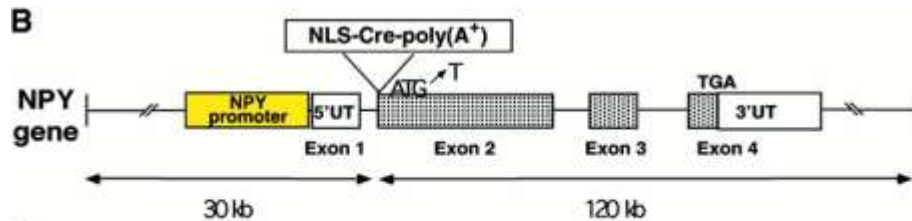
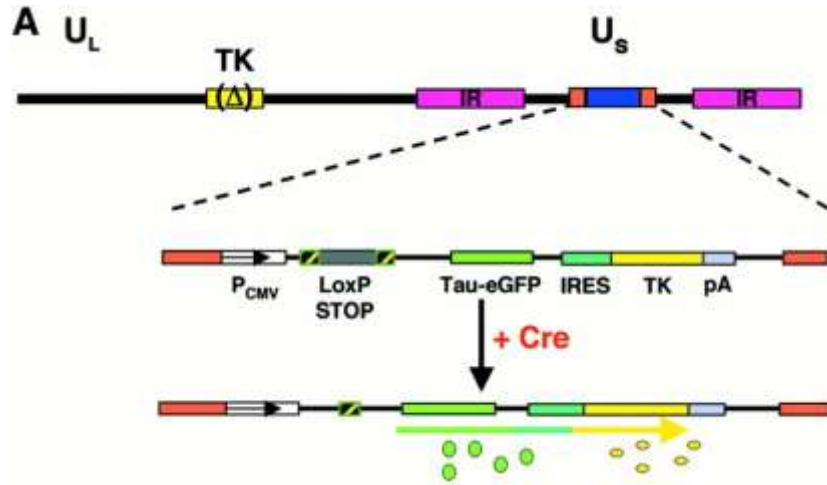


# Transneuronal viral tracing

## Anterograde: Herpes simplex virus (HSV)

## Retrograde: Pseudorabies Virus (PRV)

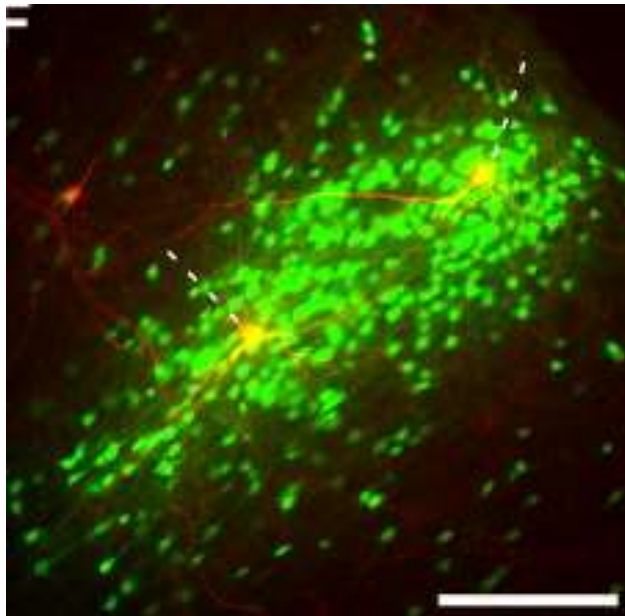
TK: thymidine kinase



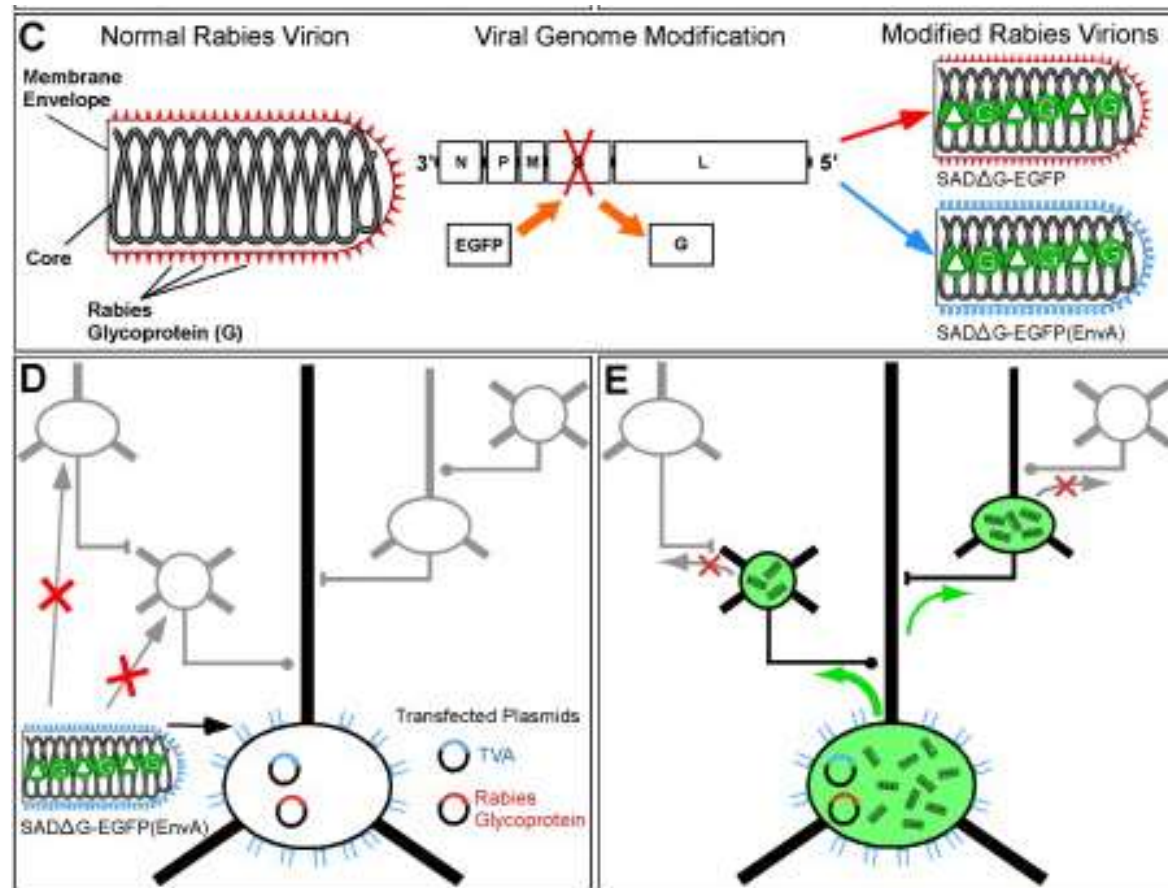
DeFalco ... Jeffery Friedman. 2001, Science  
~ 250 citation



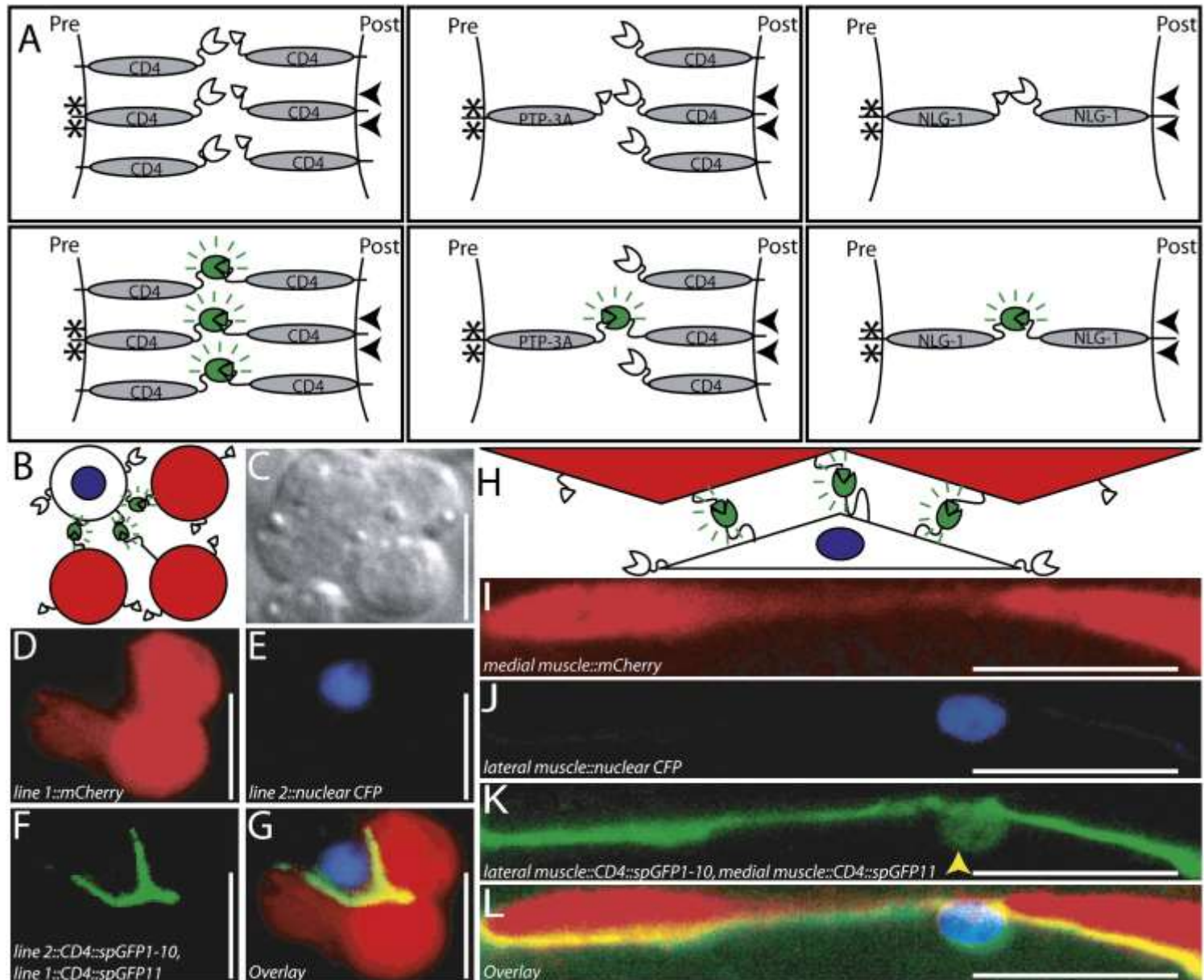
# Monosynaptic trans-neuronal tracing



G protein: essential for viral particle amplification  
EnvA: make virus only infect TVA expressing cells



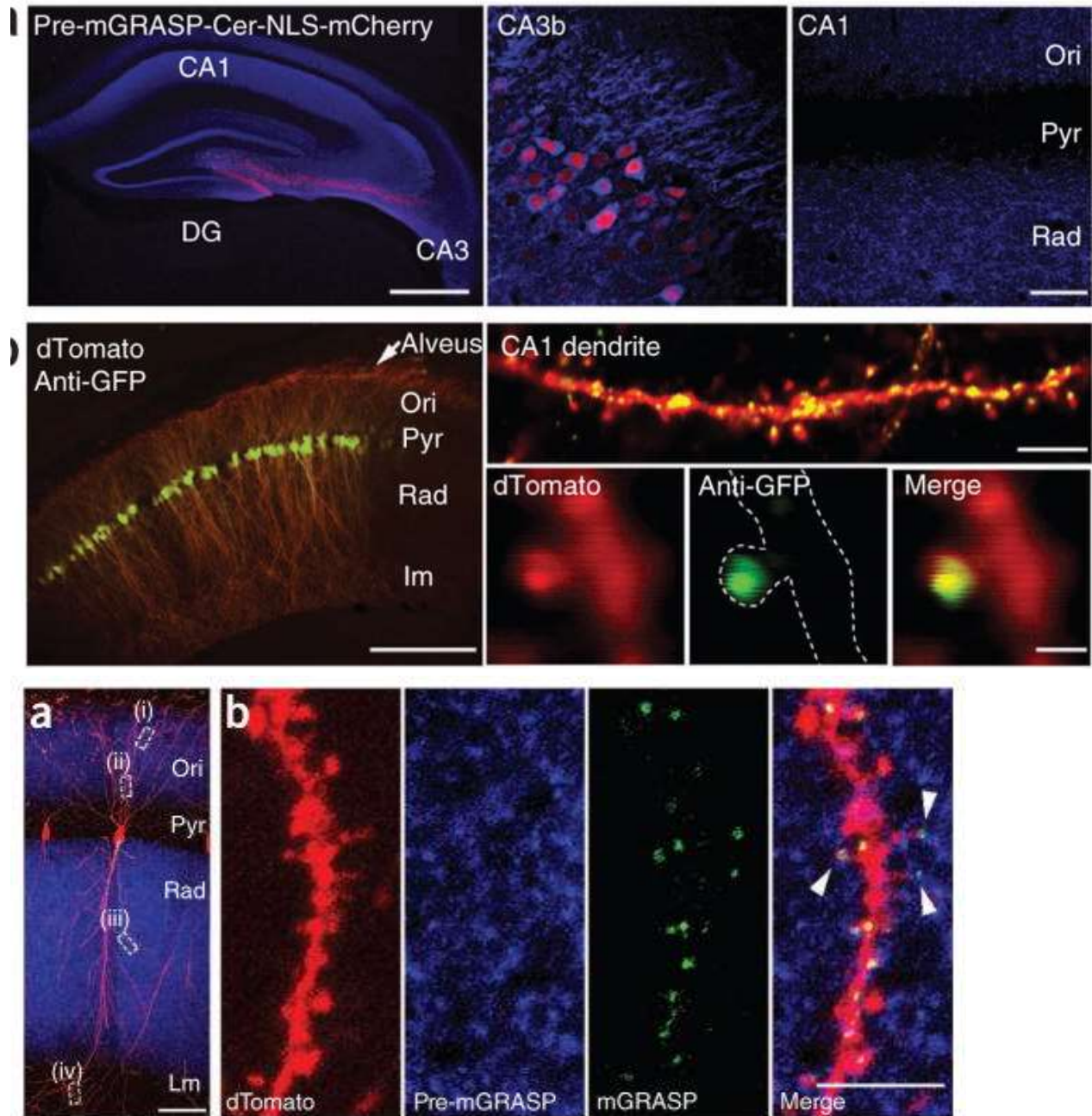
# GFP Reconstitution Across Synaptic Partners (GRASP) Defines Cell Contacts and Synapses



# Mammalian GRASP

CA3:  
pre-mGRASP-mCerulean  
NLS-mCherry

CA1:  
post-mGRASP  
(recognized with GFP antibody)  
dTomato



# Basic strategies in studying Neural circuits

- Visualize cells
- **Monitor cell activity**
- Manipulate cell activity

# **Monitor Cell activity**

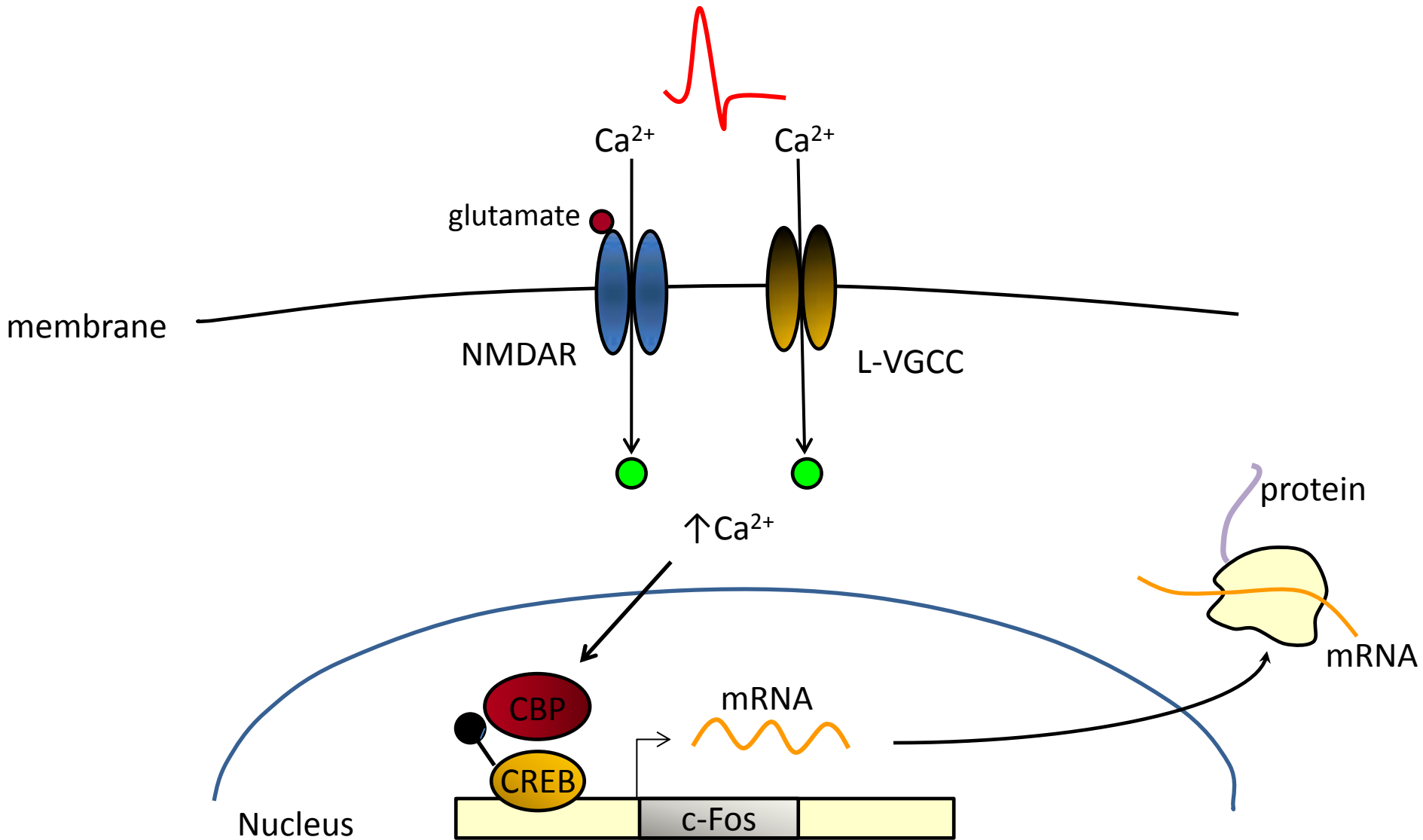
Immediate early gene mapping

Electrophysiology

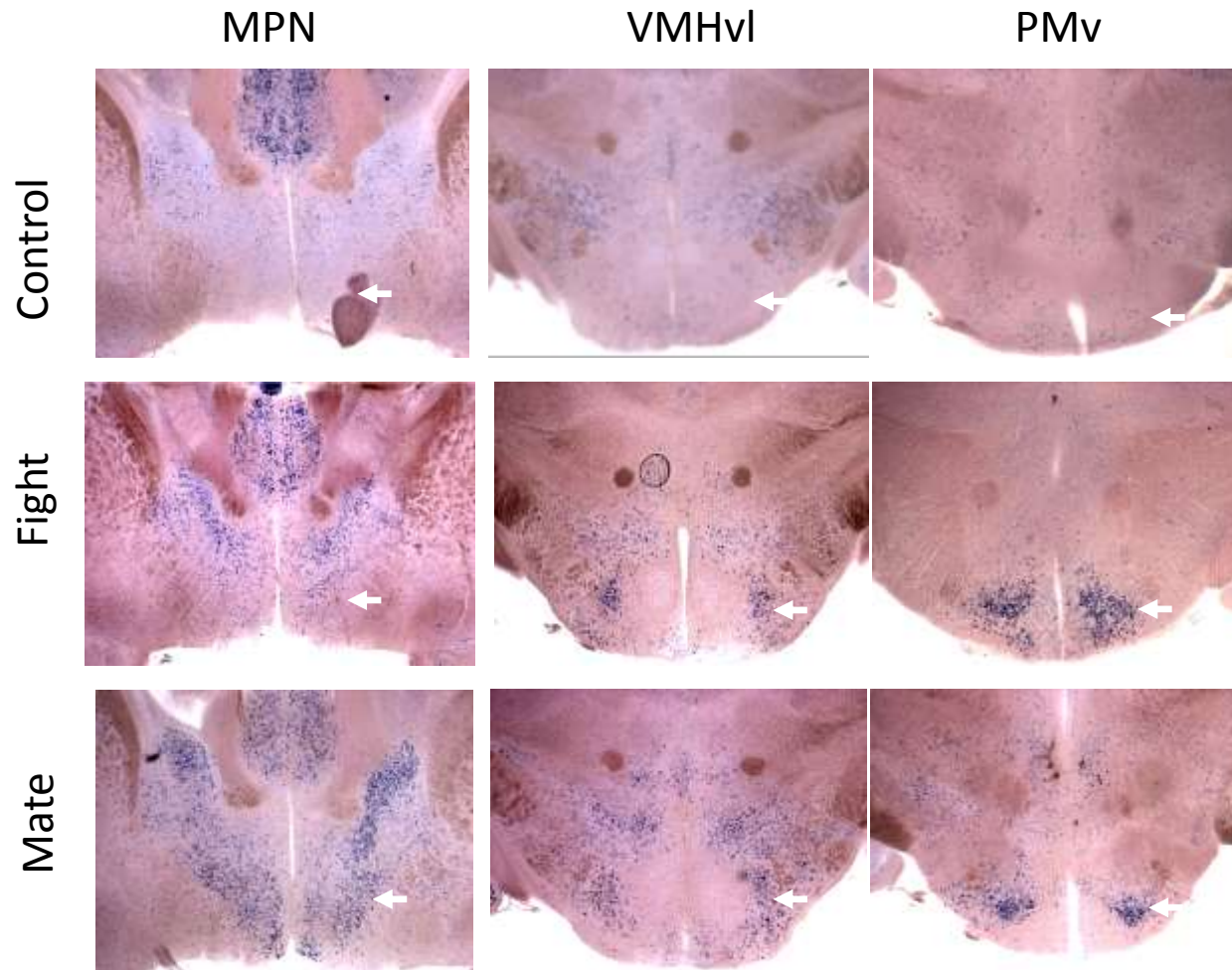
Functional Imaging



# Immediate early gene response



# Fos expression patterns after fighting and mating



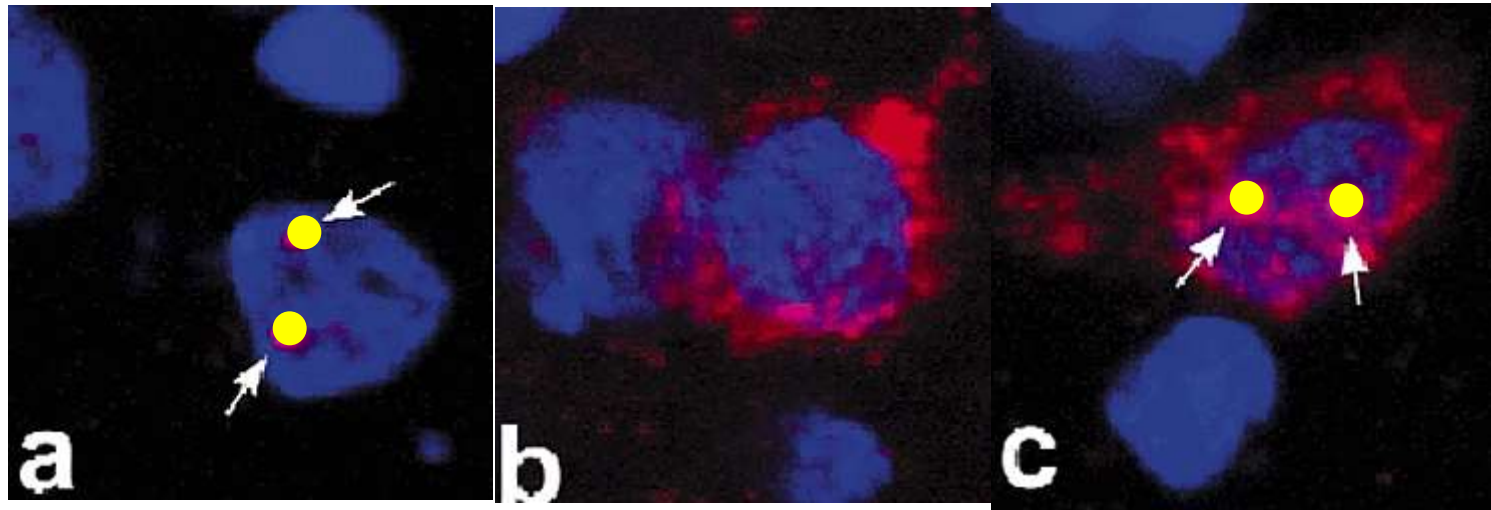
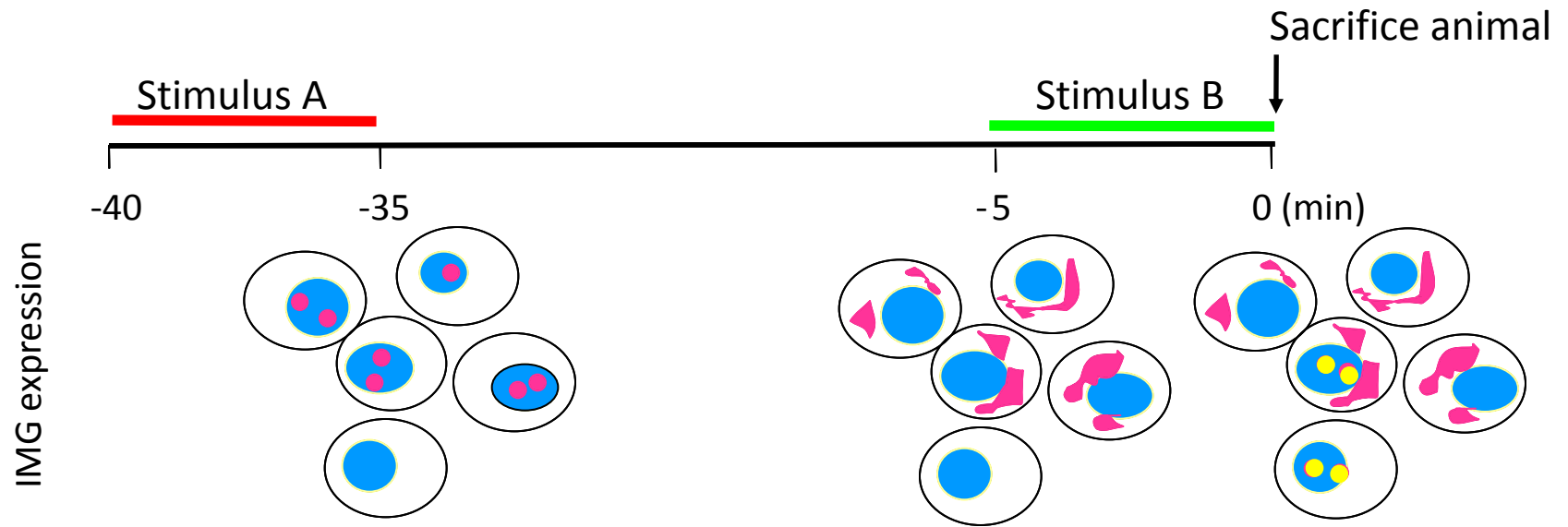
PMv: premammillary nucleus

MPN: Medial preoptic nucleus

VMH: Ventromedial hypothalamic nucleus

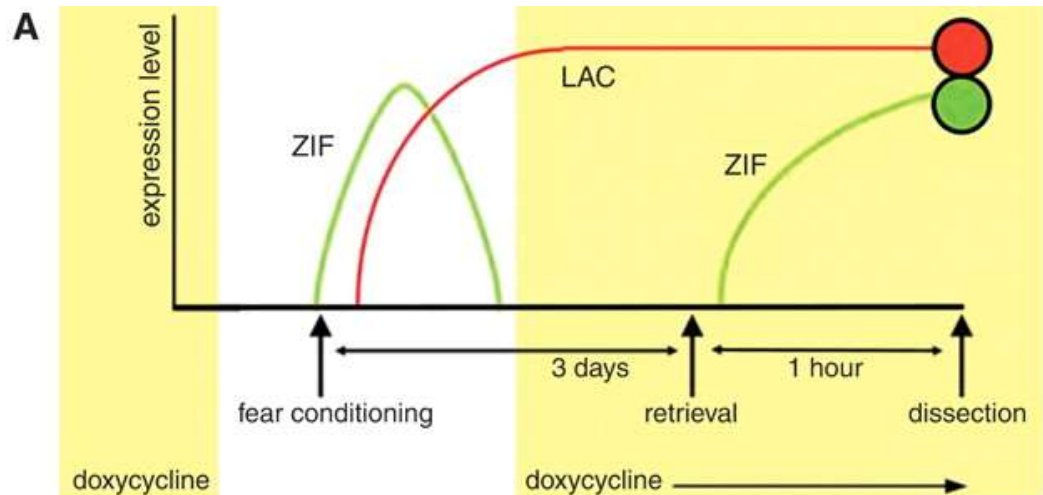
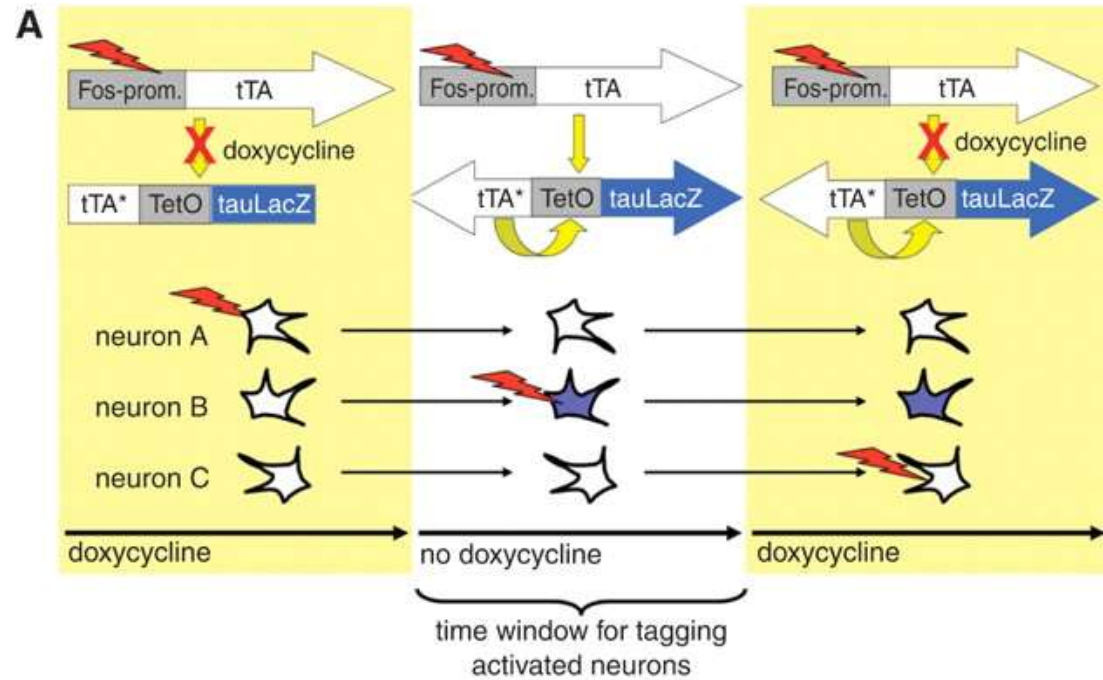
# Immediate early gene (IMG) CatFISH —

cellular compartment analysis of temporal activity by fluorescence in situ hybridization

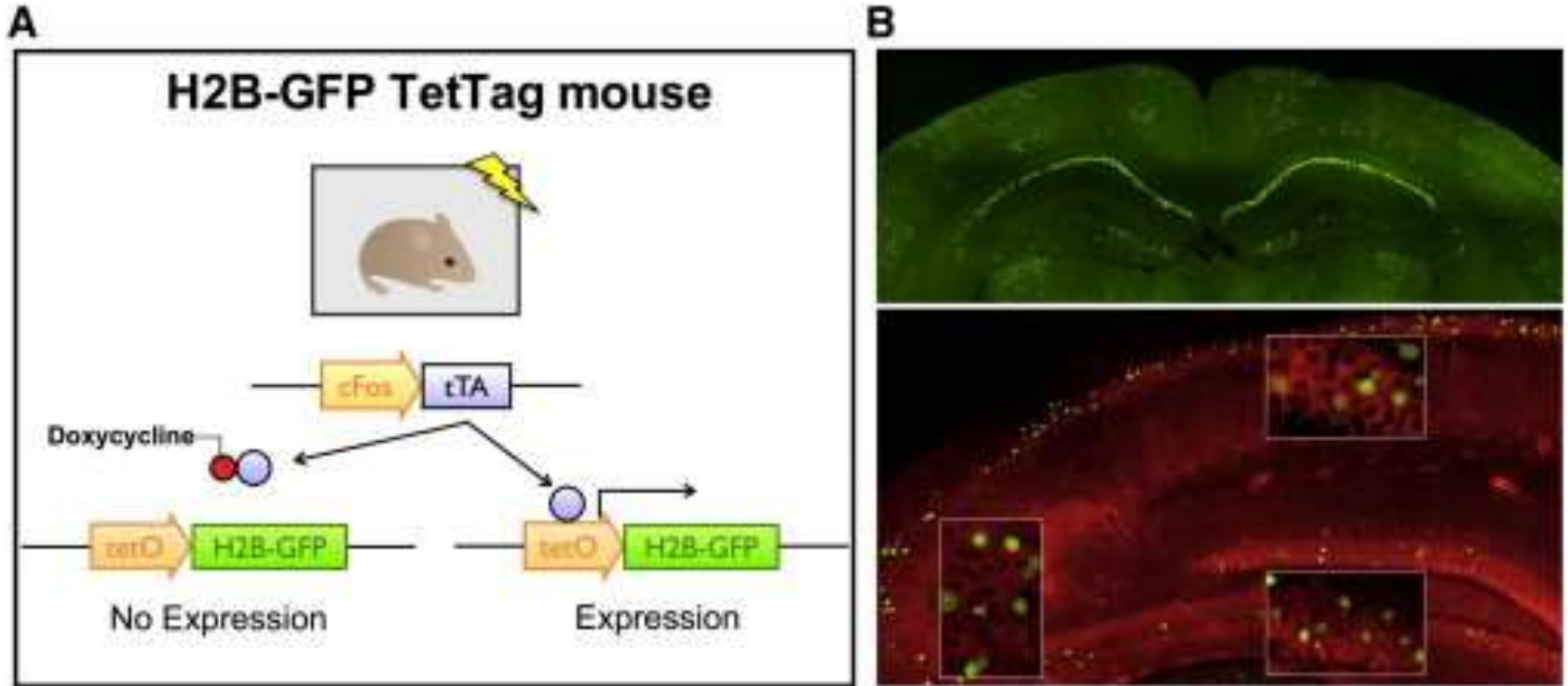


# Long term tagging activated cells

*tTA*: tetracycline transactivator  
*tetO*: *tTA* binding sites



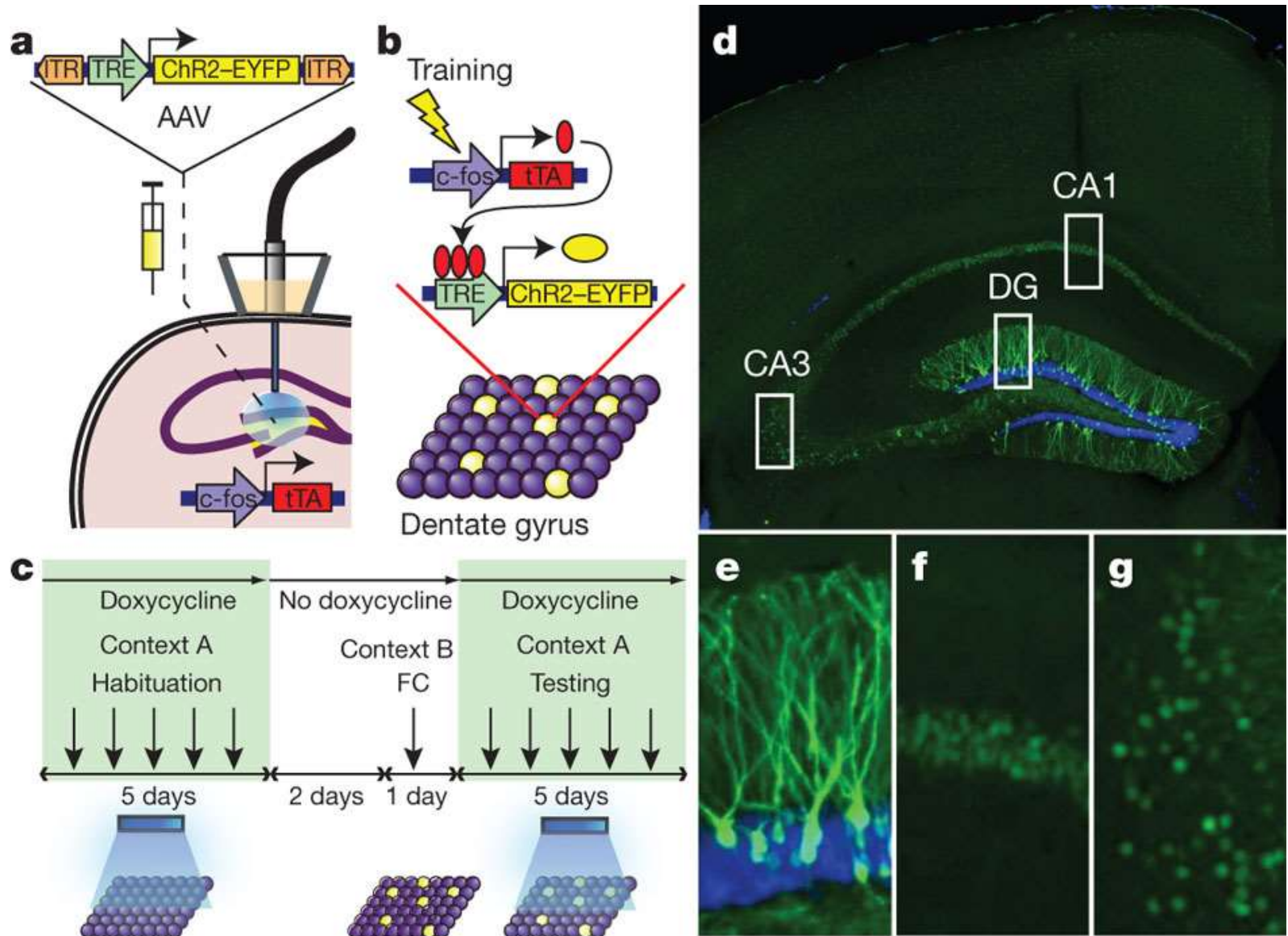
# TetTag mouse with long lasting GFP



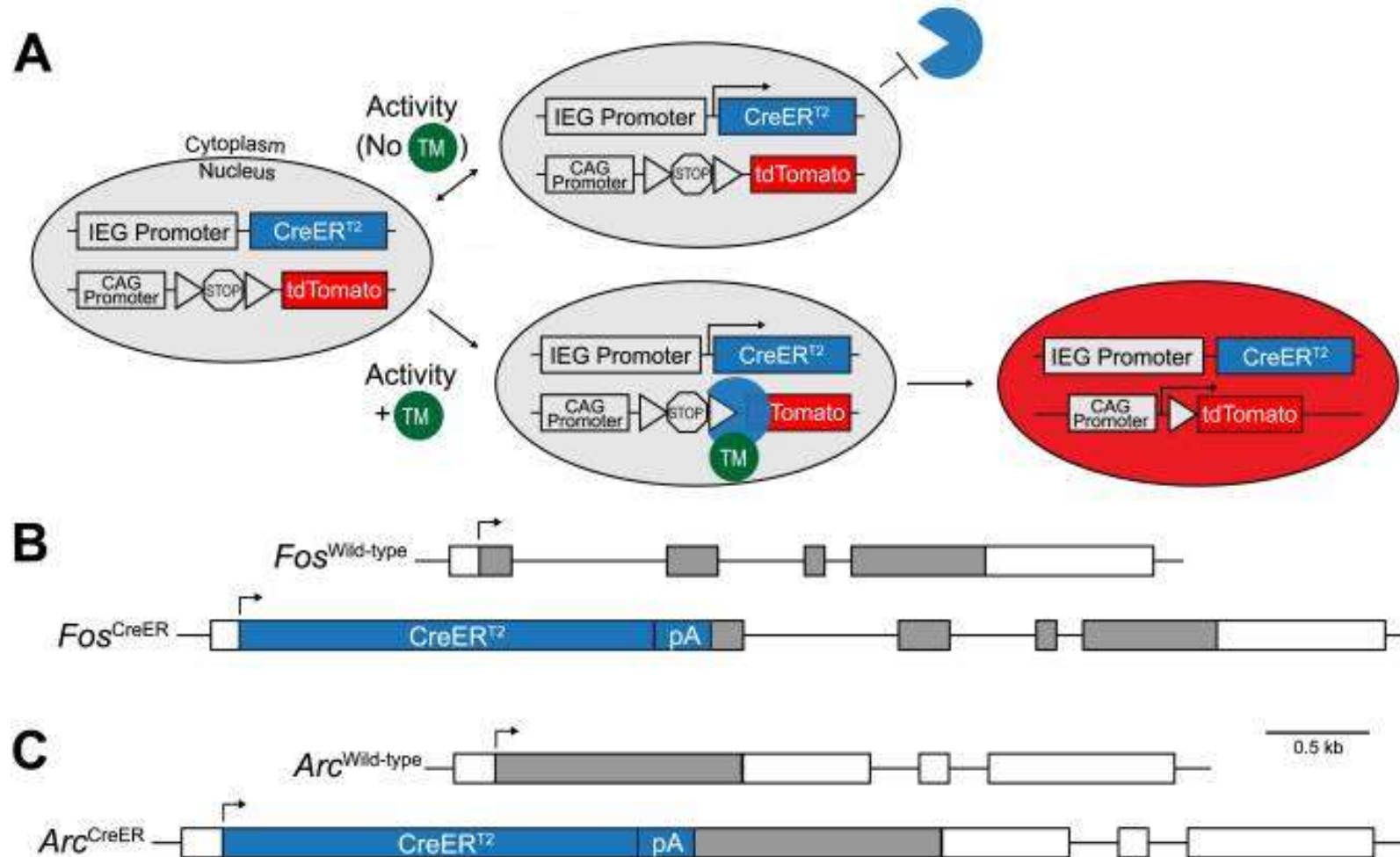
Taylor K et al. Current Biology, 2013



# TetTag system to label active cells with ChR2



# Labeling of transiently activated neurons using TRAP



# Electrophysiology

Methods:

Intracellular recording (Patch clamp or sharp electrode)

Cell attach recording

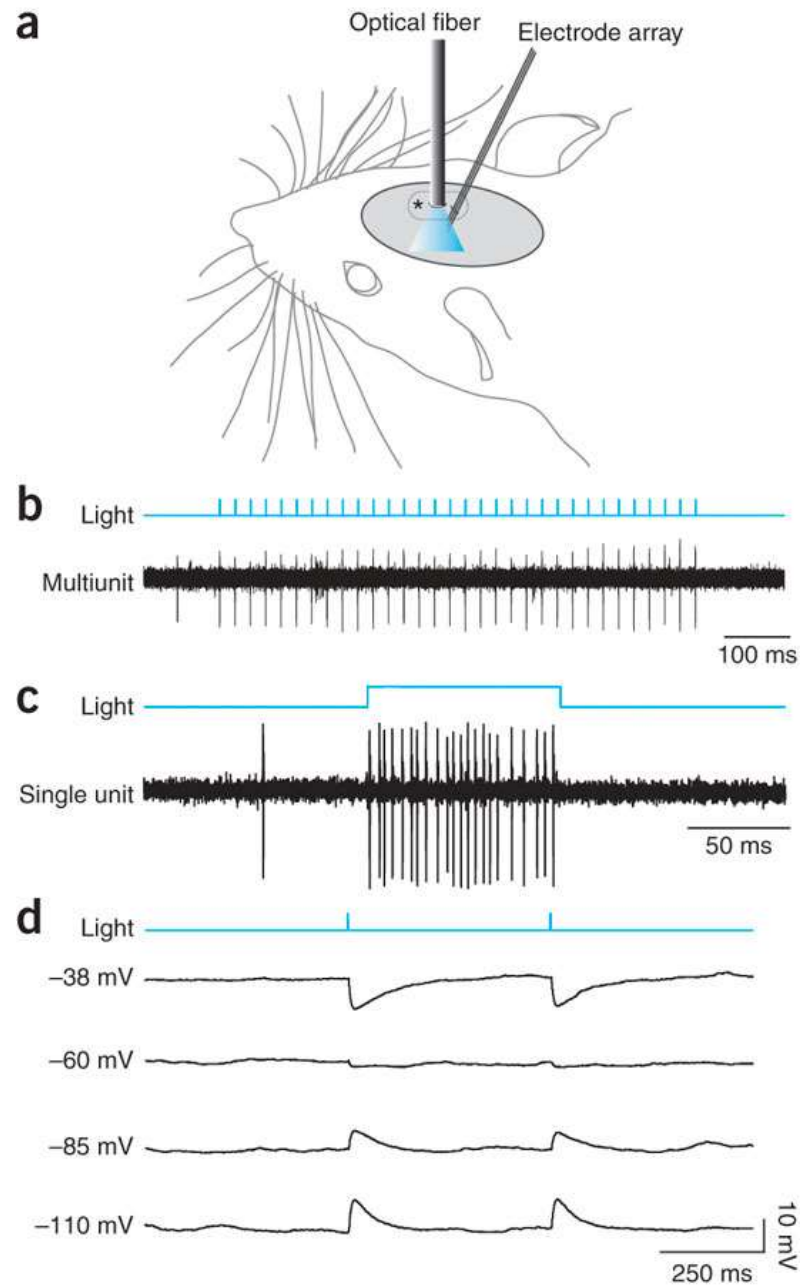
Extracellular recording (single or multiple electrodes)

Preparation:

In vitro (slice or cells)

In vivo (anaesthetized animals, head fixed awake animal, free moving animals)

# Combining ChR2 and electrophysiology



# Functional imaging --detecting the activity of neurons by optical imaging

**What kind of changes will occur when a brain area (or cell) becomes active?**

Membrane potential

Calcium concentration

Chloride concentration

Sodium concentration

Potassium concentration

PH level (synapse)

Oxygen level

**How to convert non-optical changes to optical changes?**

Voltage sensitive dye

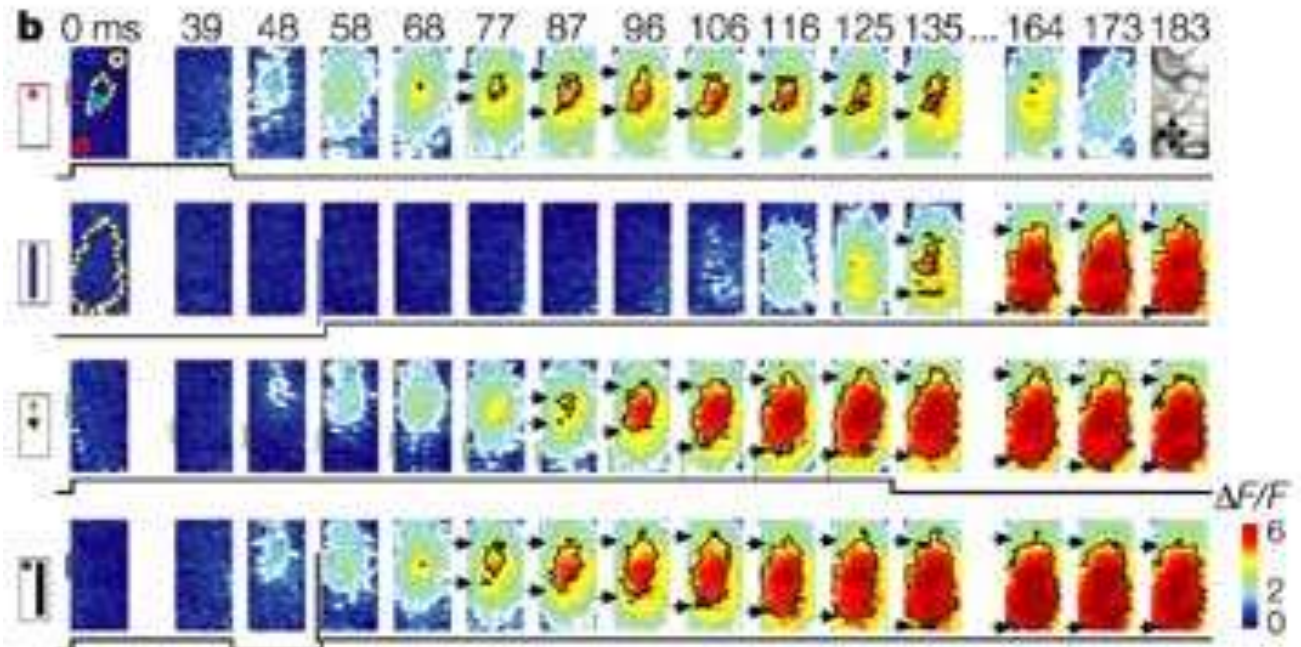
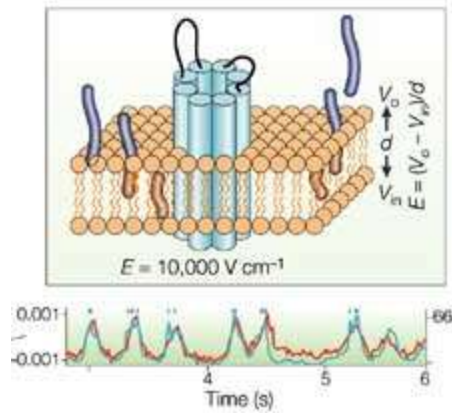
Calcium sensitive dye

Synapto-pHluorin

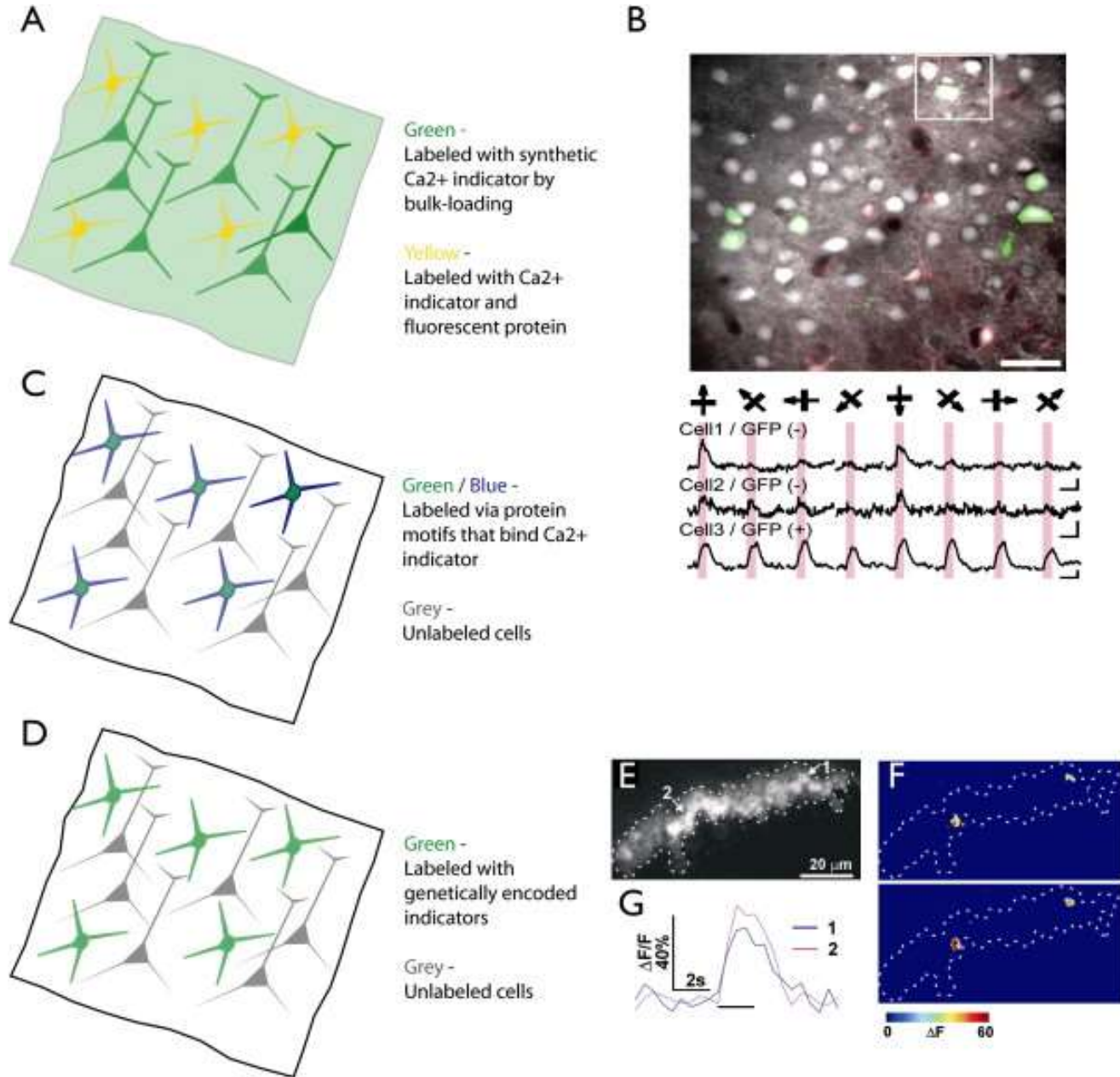
Camgaroo- ratiometric GECI



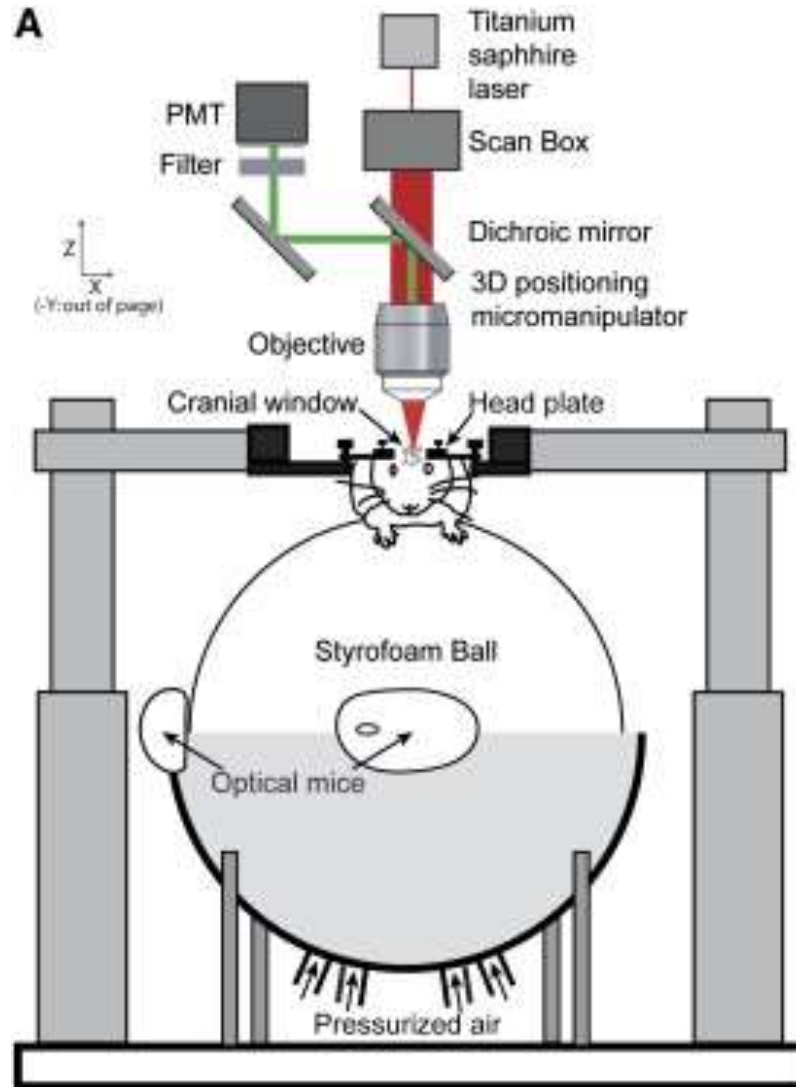
# Functional imaging--Voltage sensitive dye



# Functional Imaging –Calcium imaging



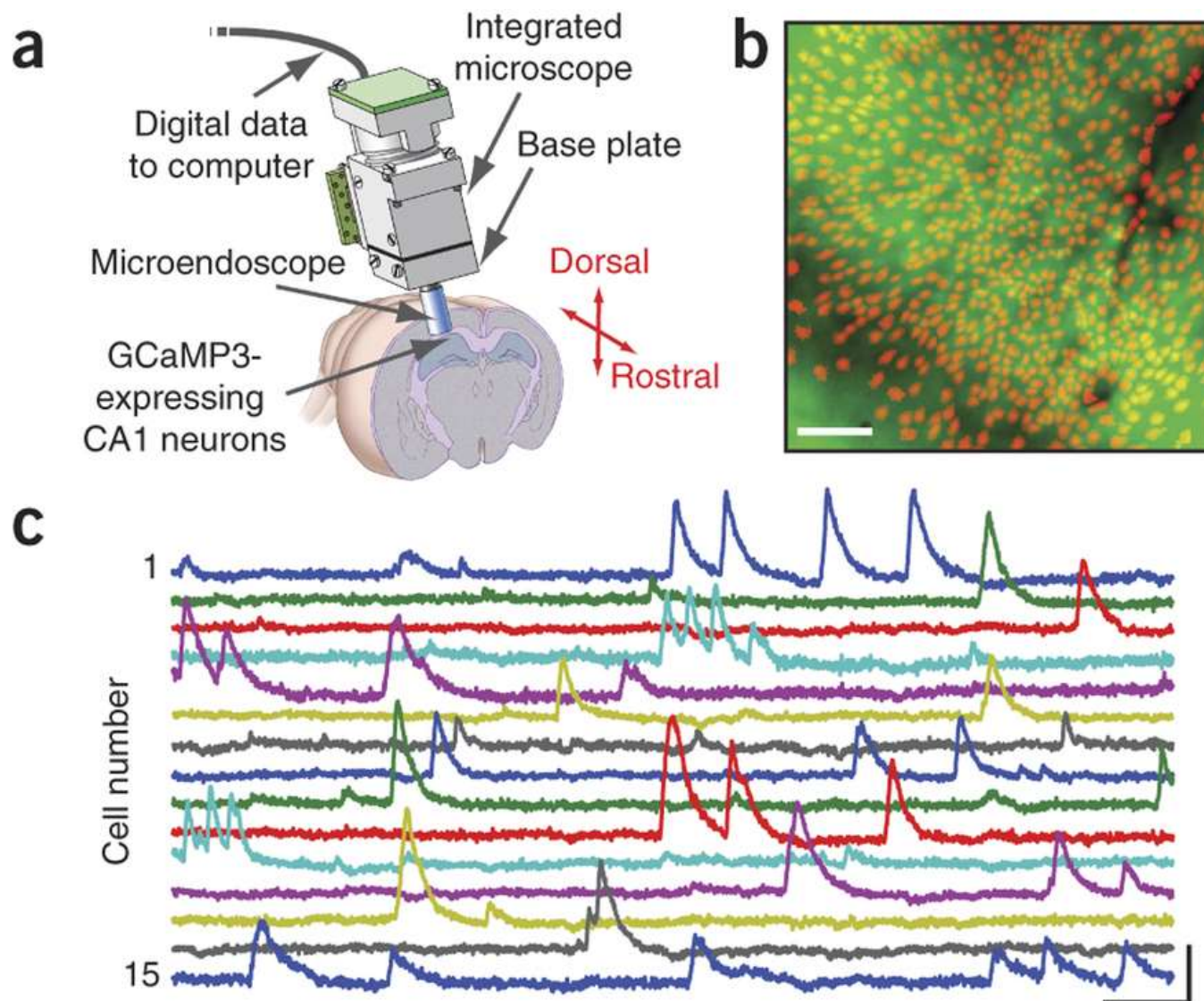
# Head Fixed 2-photon calcium imaging



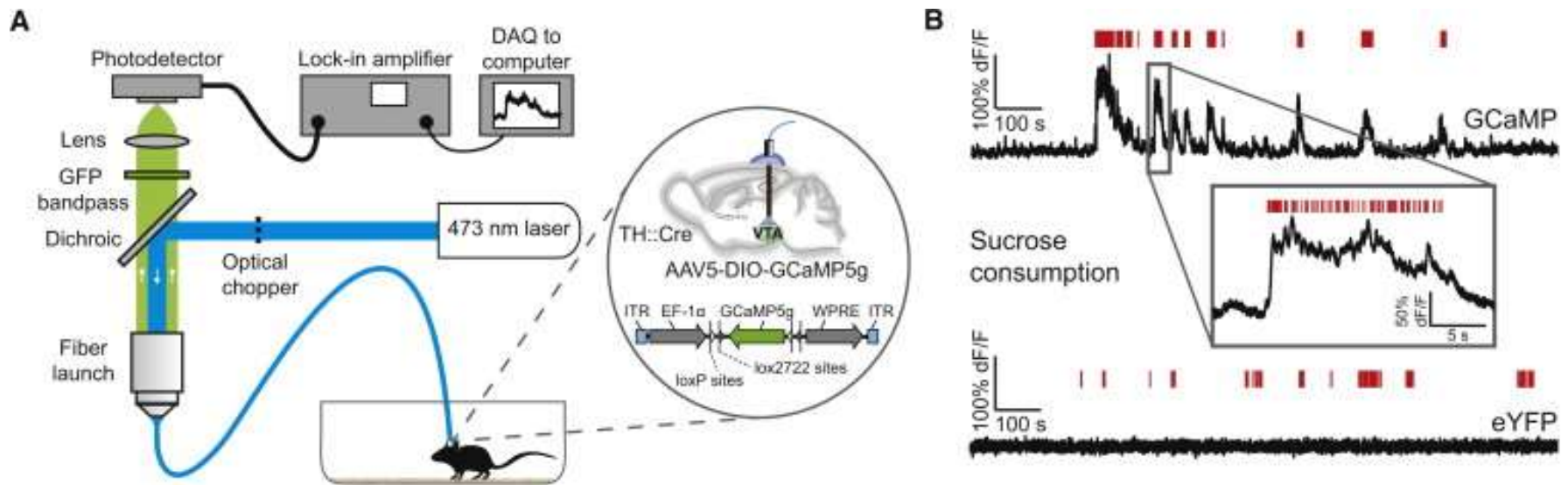
[Daniel A. Dombeck](#)... David Tank  
Nature Neuroscience, 2010, Cite 230



# 1-photon Calcium imaging in awake behaving animals

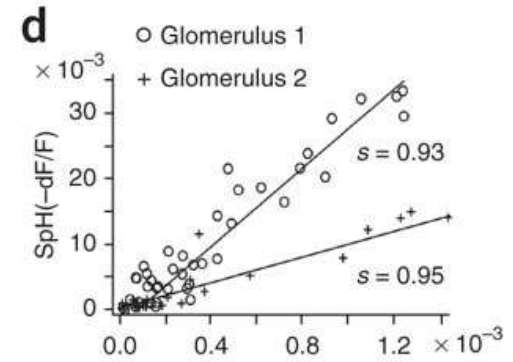
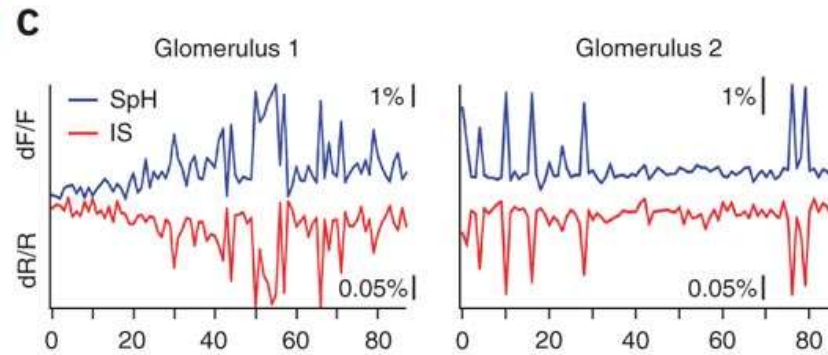
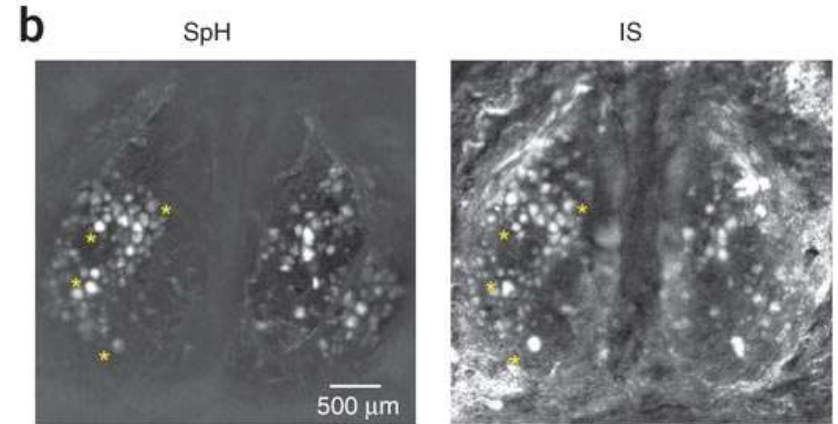
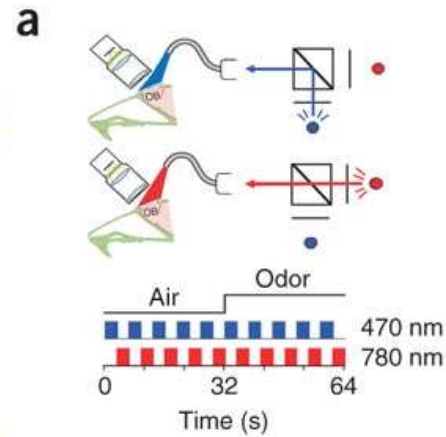
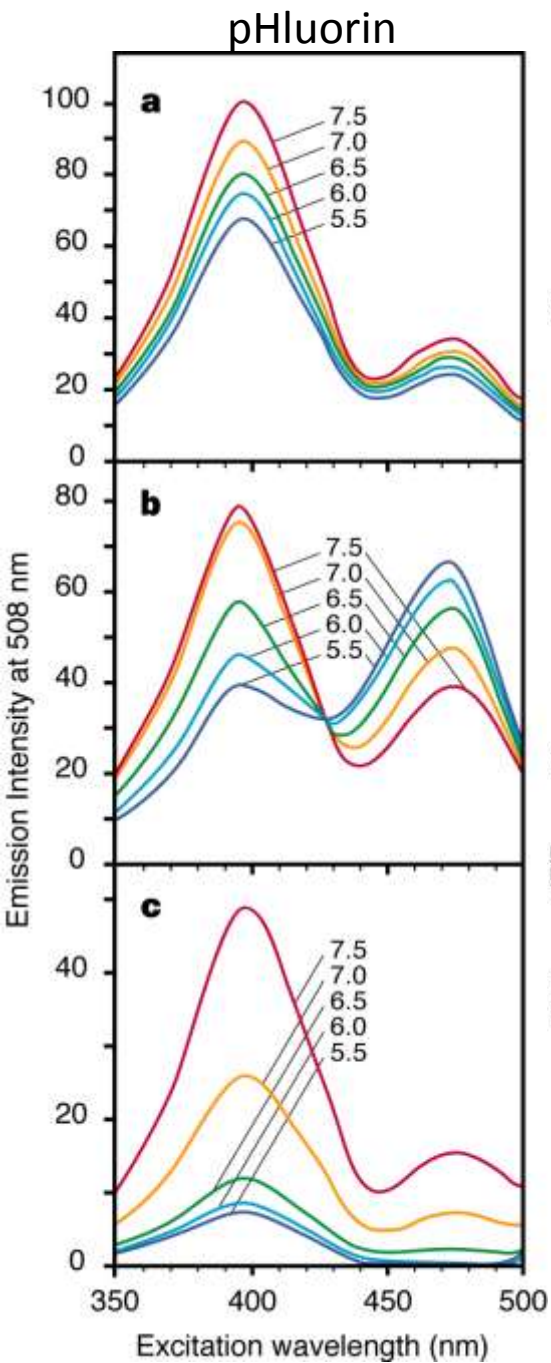


# Population calcium imaging in awake behaving animals –fiber photometry





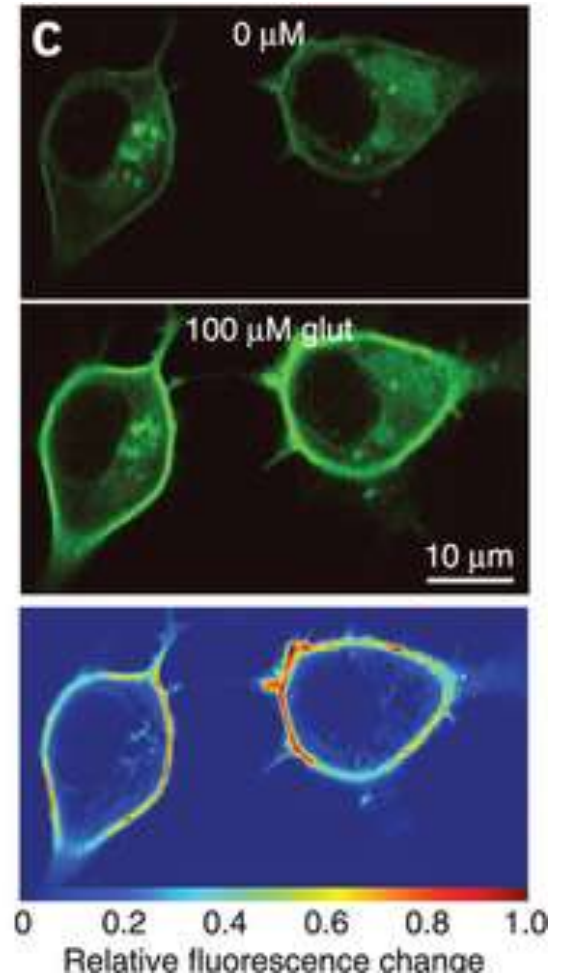
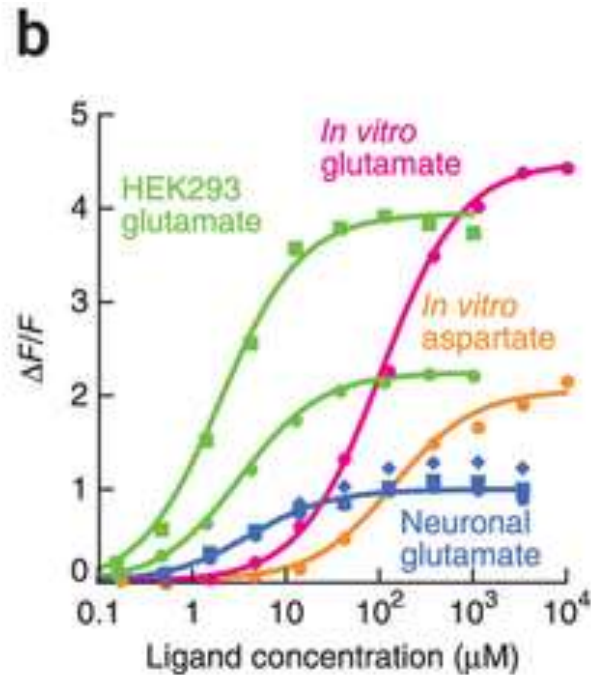
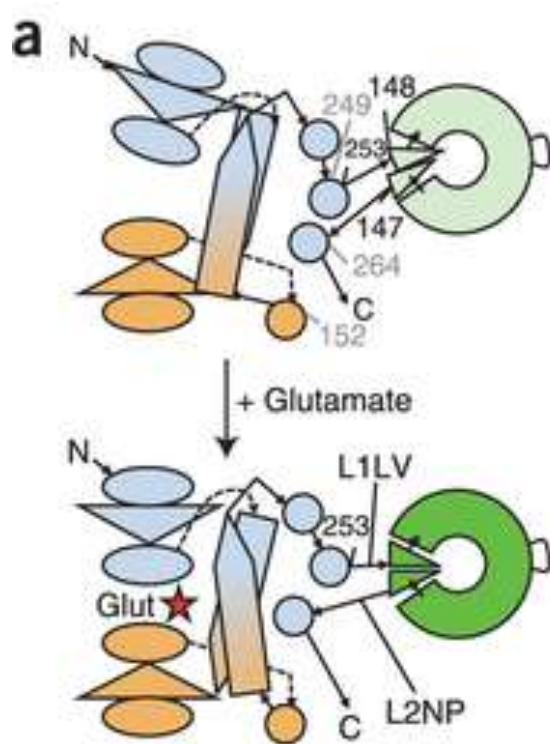
# Synapto-pHluorin and Intrinsic signal imaging





# iGluSnFR as an glutamate sensor

Bacterial periplasmic binding proteins (PBPs)



# Basic strategies in studying Neural circuits

- Visualize cells
- Monitor cell activity
- **Manipulate cell/gene activity**

# Manipulate cell/gene activity

## Region

1. Electrical Lesion
2. Pharmacological inactivation
3. Electrical stimulation
4. Pharmacological activation

## Cells

1. Inactivate
2. Activate
3. Killing

## Genes

1. Traditional gene targeting
2. Transgenic
3. CRISPR/Cas9

## Receptors

1. Antagonist
2. Agonist
3. Ligand uncaging

# Manipulate cell/gene activity

## Region

1. Electrical Lesion
2. Pharmacological inactivation
3. Electrical stimulation
4. Pharmacological activation

## Cells

1. Inactivate
2. Activate
3. Killing

## Genes

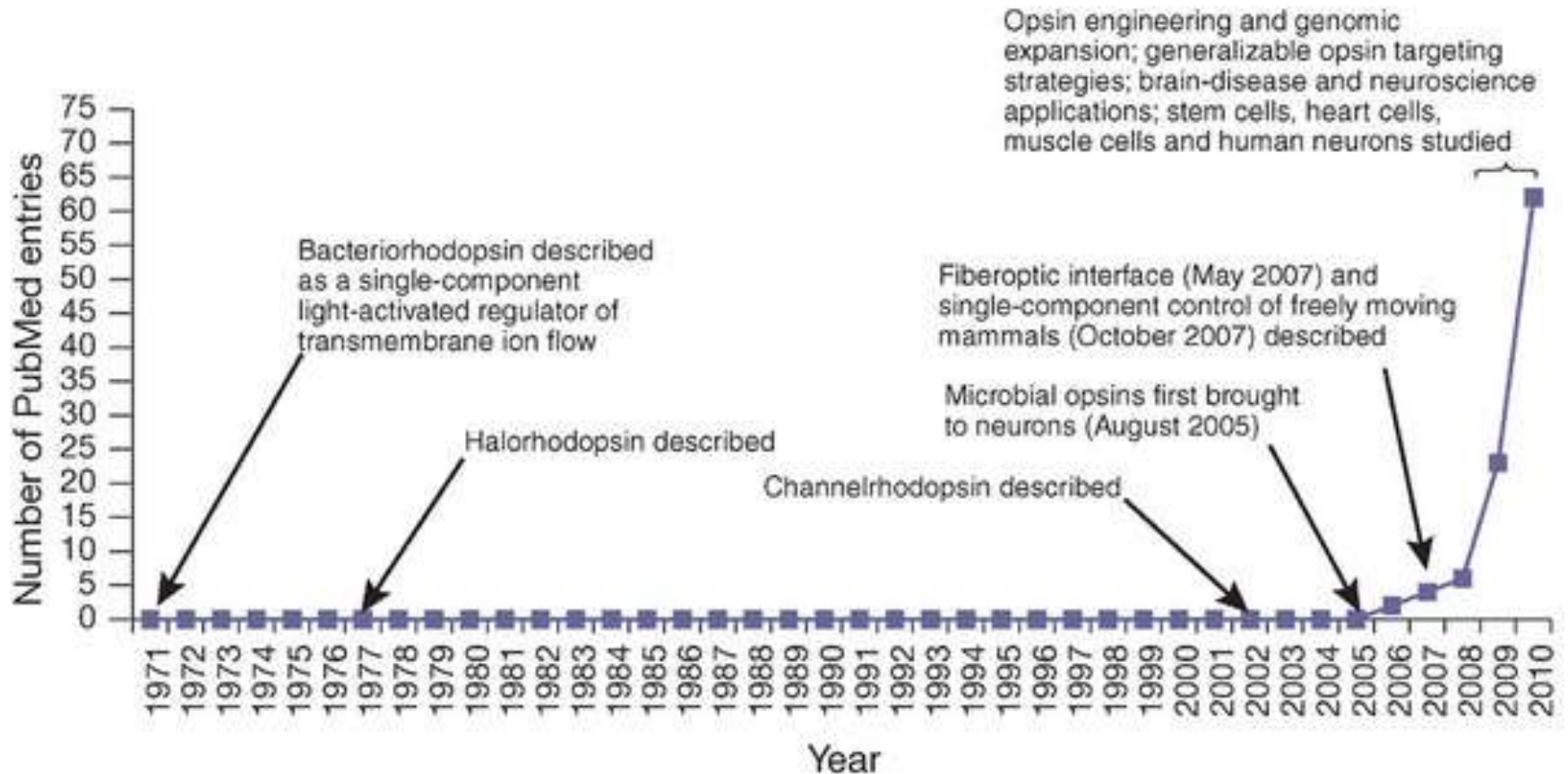
1. Traditional gene targeting
2. Transgenic
3. CRISPR/Cas9

## Receptors

1. Antagonist
2. Agonist
3. Ligand uncaging

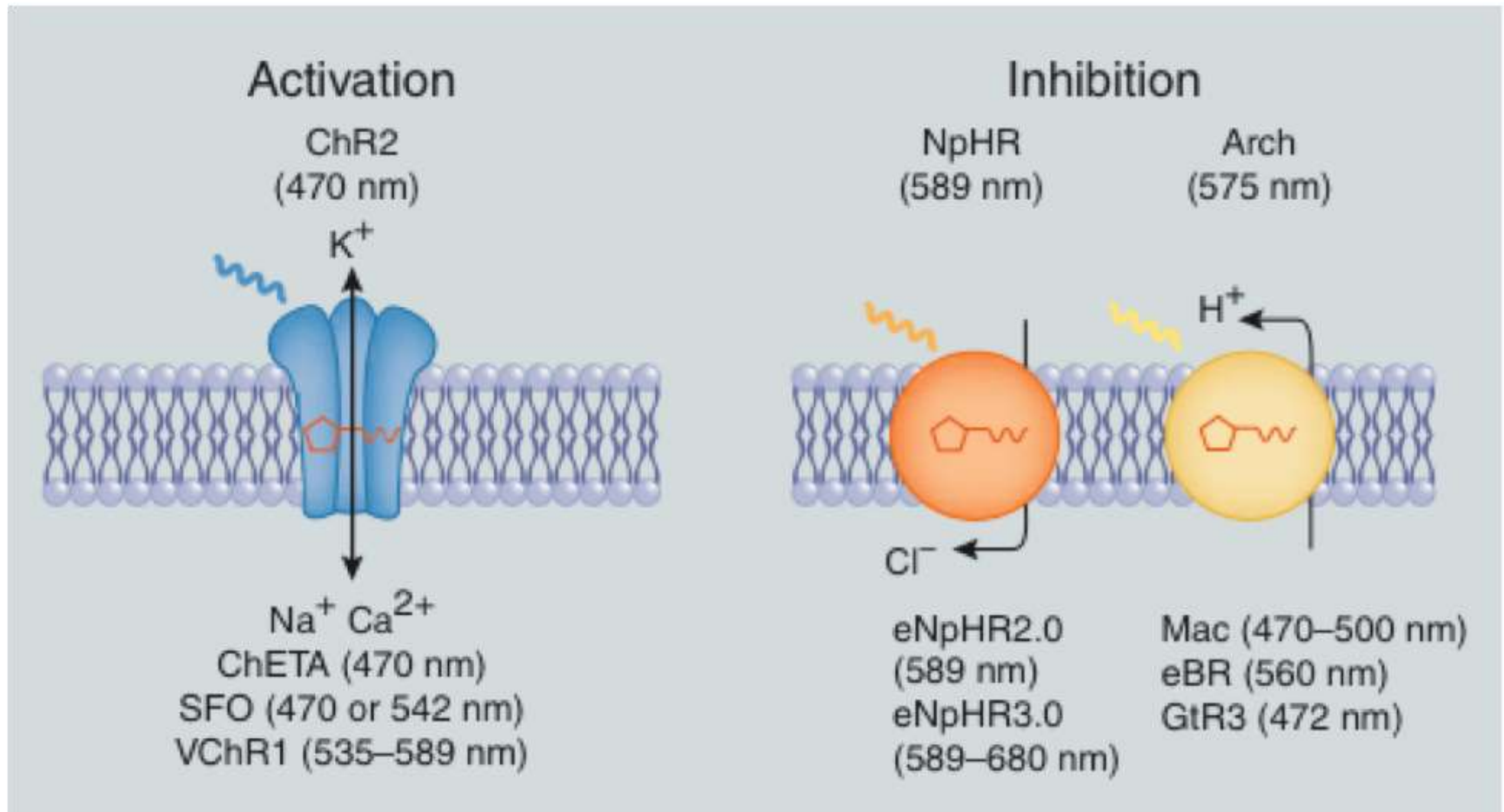


# Light mediated cell control -- optogenetics



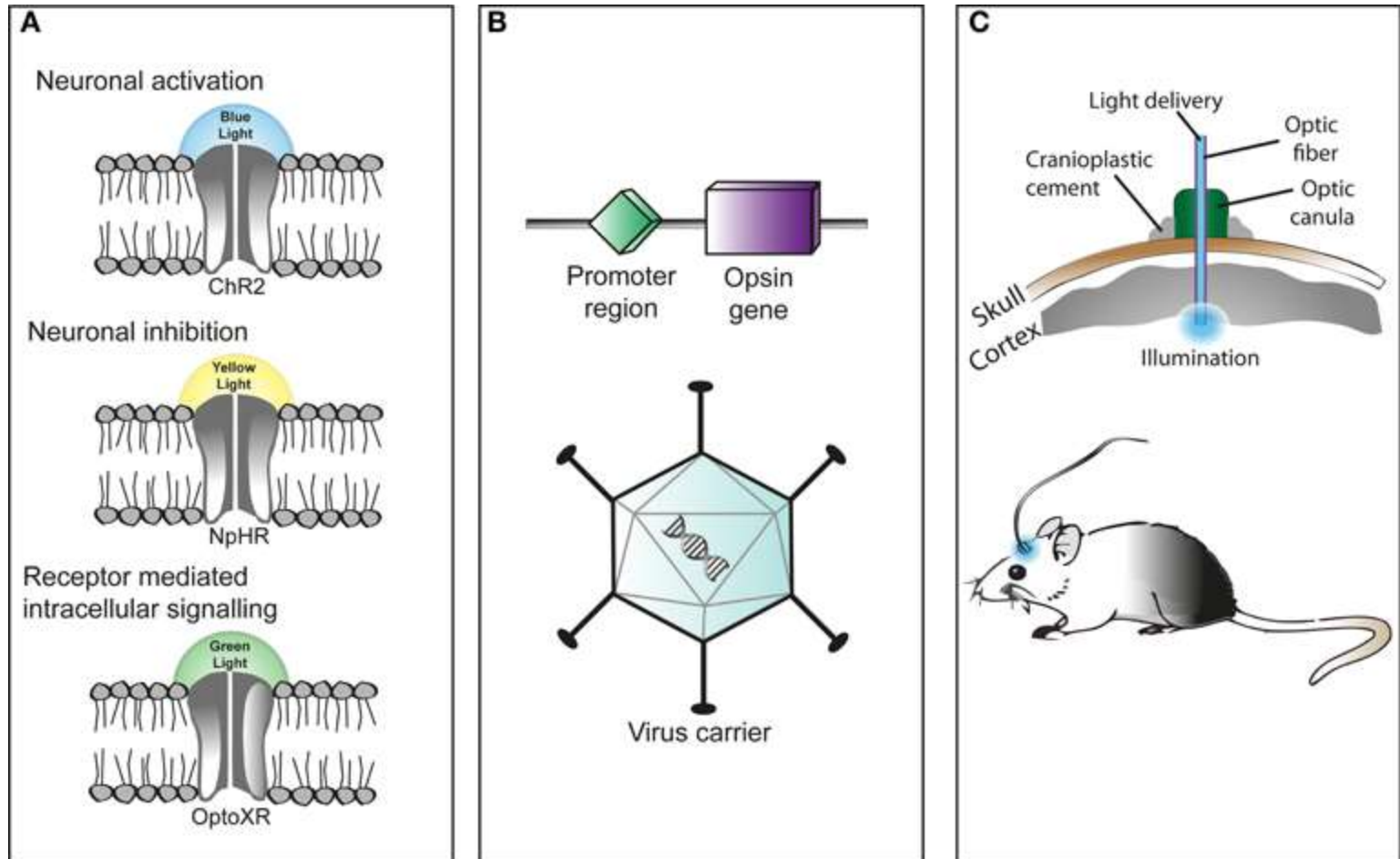
Karl Deisseroth and Ed Boyden  
2005 Nature ~2000 citations

# Optogenetics



Karl Deisseroth and Ed Boyden  
2005 Nature ~2000 citations

# In vivo optogenetic control of cell activity

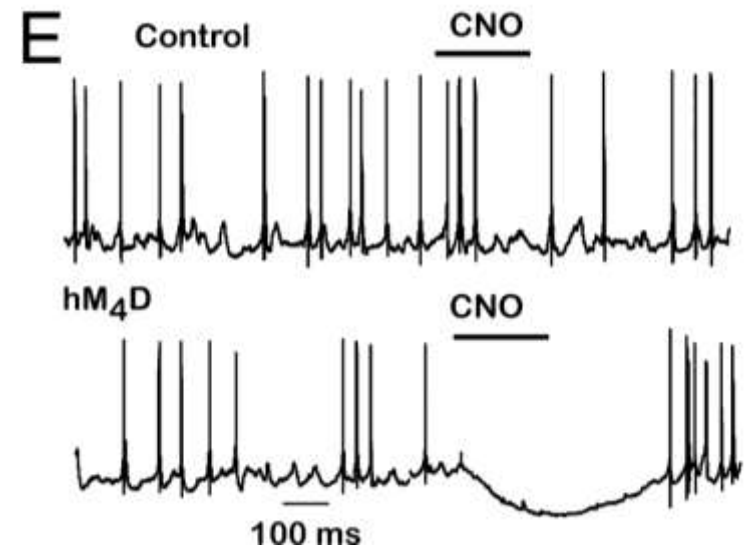
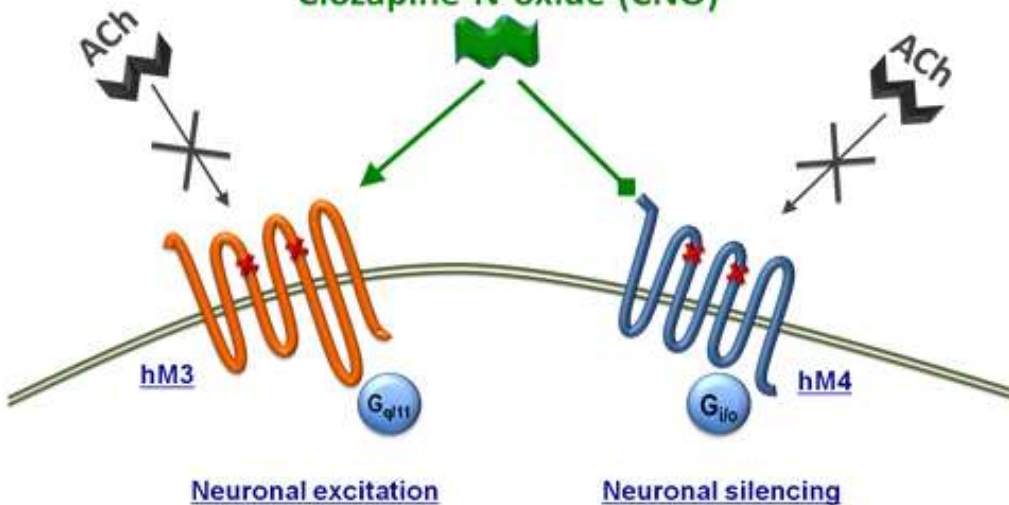


# Ligand mediated cell control --pharmacogenetics

## DEARDD

PHARMACOGENETICS  
Designer Receptors Exclusively Activated by Designer Drugs  
DREADDs

Clozapine-N-oxide (CNO)

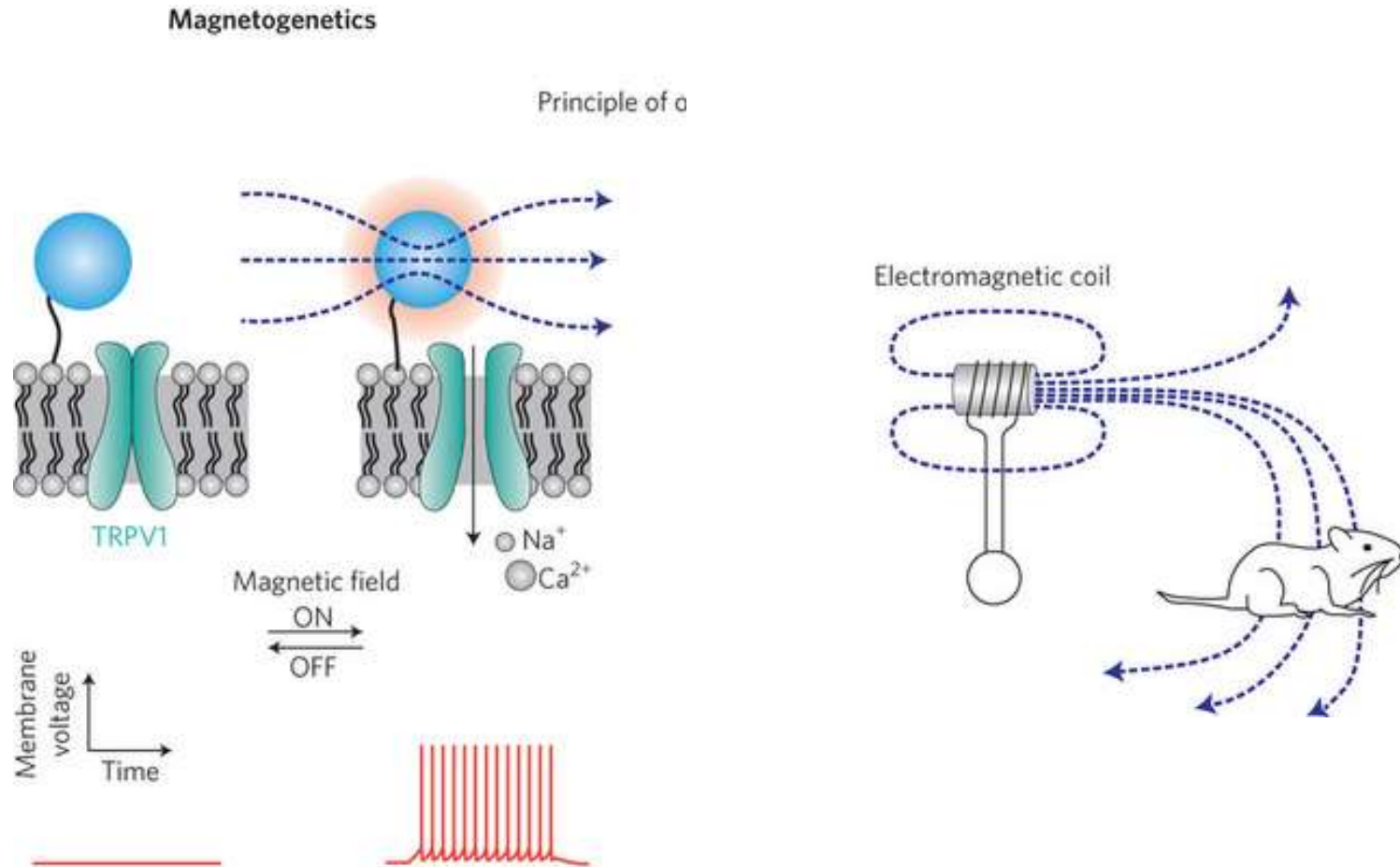


Armbruster et al., 2007, PNAS

Adapted from Wess et al., 2013, Trends in Pharmacological Sciences

2007 PNAS Armbruster BN et al.  
Cite ~390

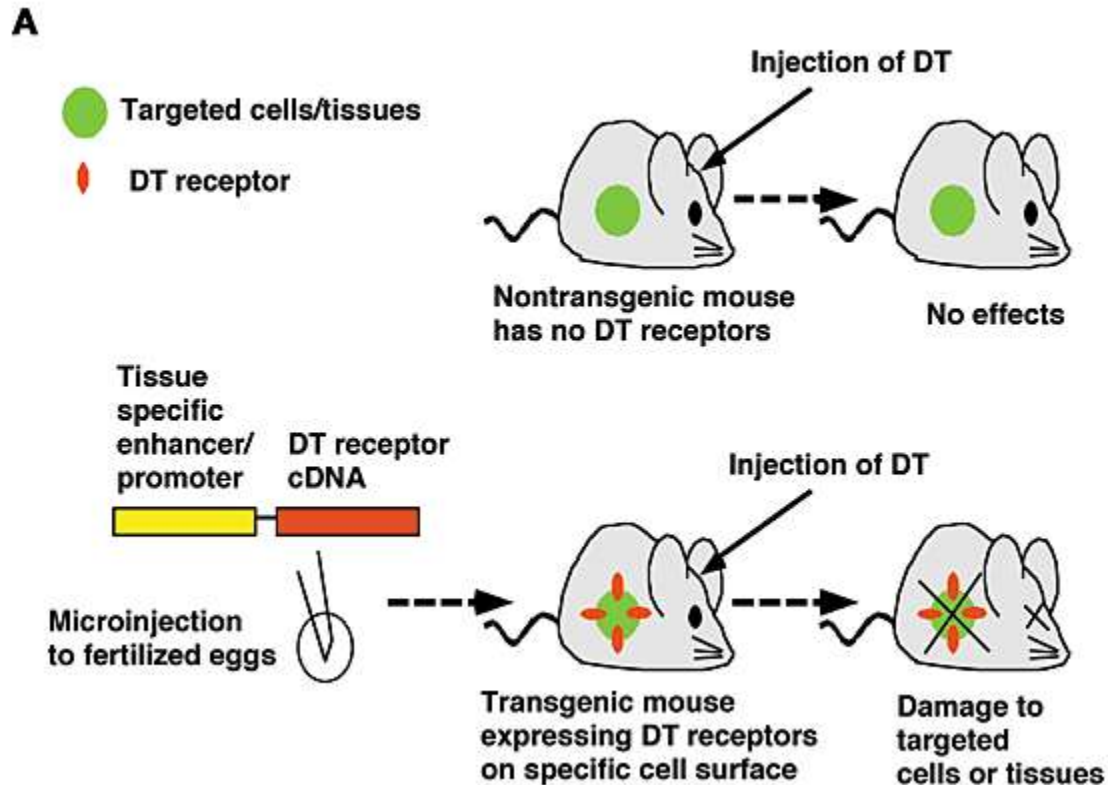
# Magnetic field mediated cell control -- Meganogenetic





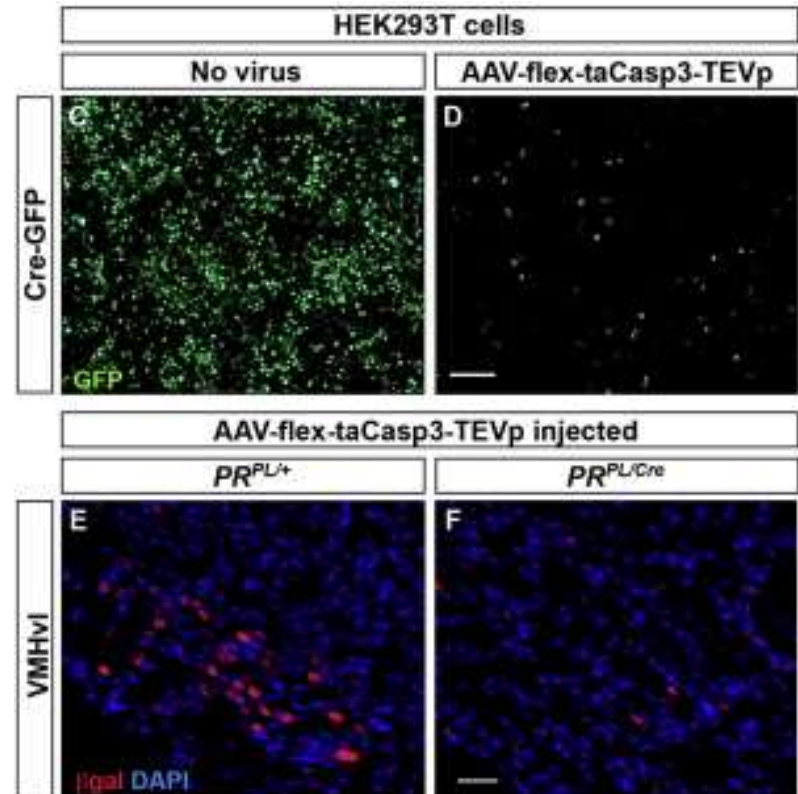
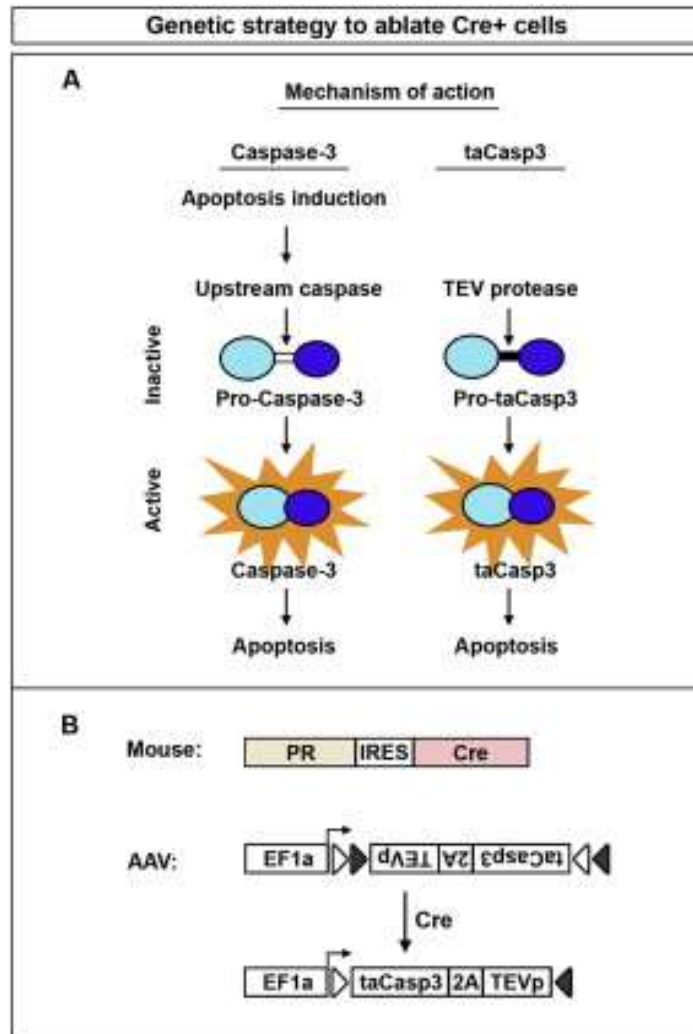
# Diphtheria toxin receptor (DTR) mediated cell killing

One molecular of DT can kill a cell – Cell, 1978, Yamaizumi, M.



Michiko Saito et al. 2001, Nature Biotechnology  
Cite ~250

# Caspase 3 mediated cell killing



# Manipulate cell/gene activity

## Region

1. Electrical Lesion
2. Pharmacological inactivation
3. Electrical stimulation
4. Pharmacological activation

## Cells

1. Inactivate
2. Activate
3. Killing

## Genes

1. Traditional gene targeting
2. Transgenic
3. CRISPR/Cas9

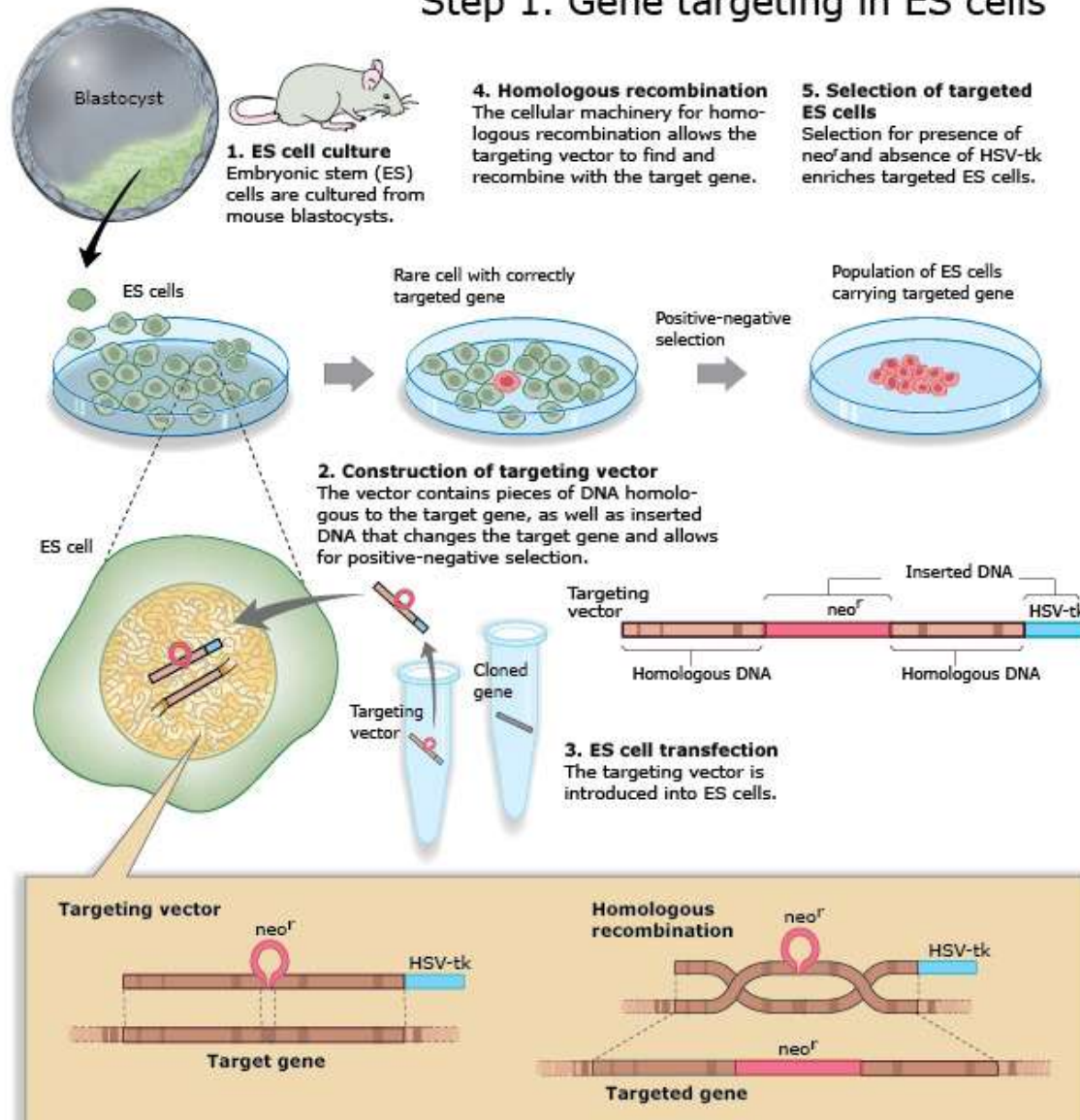
## Receptors

1. Antagonist
2. Agonist
3. Ligand uncaging

# Traditional gene targeting

Rely on rare homologous recombination in ES cells

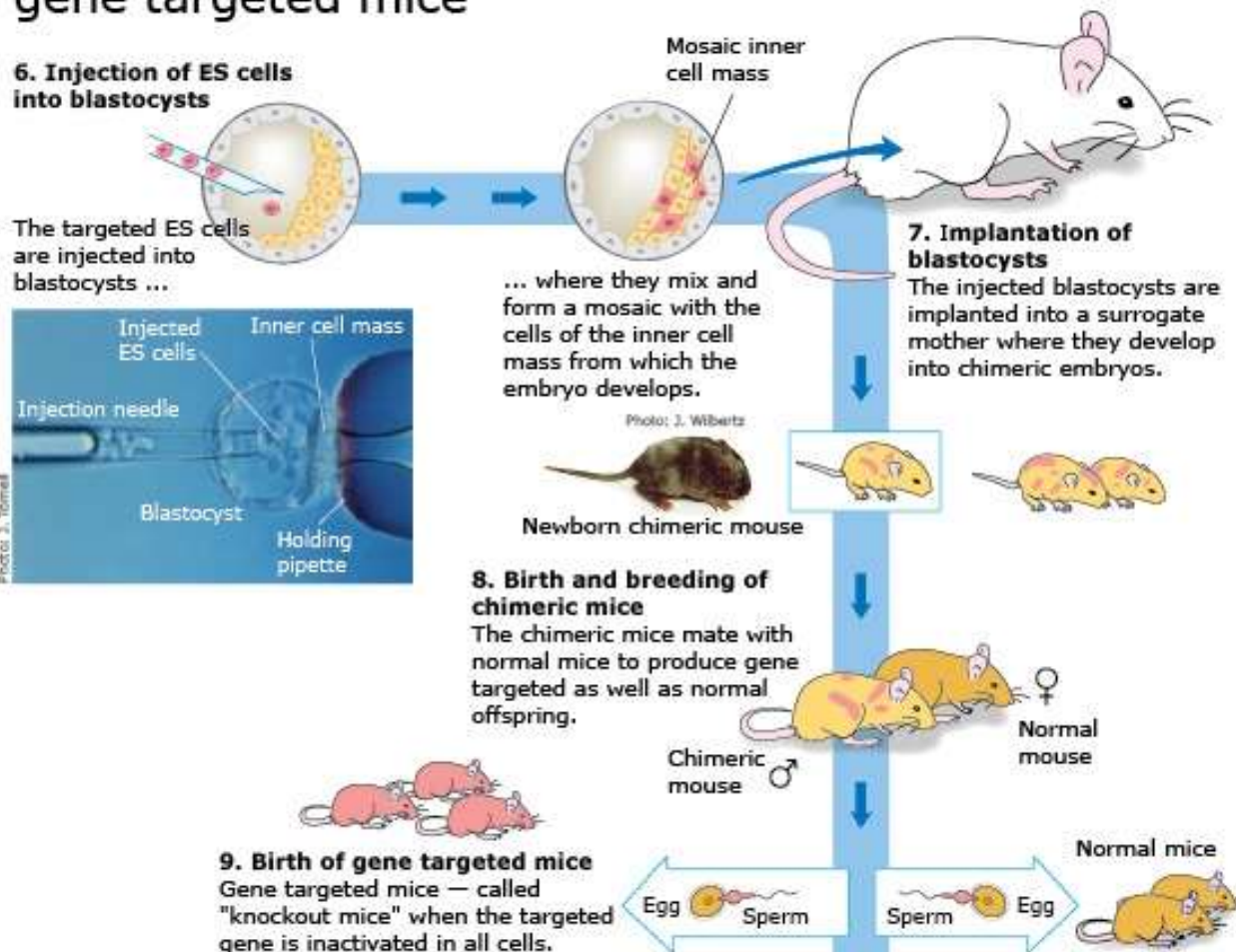
## Step 1. Gene targeting in ES cells



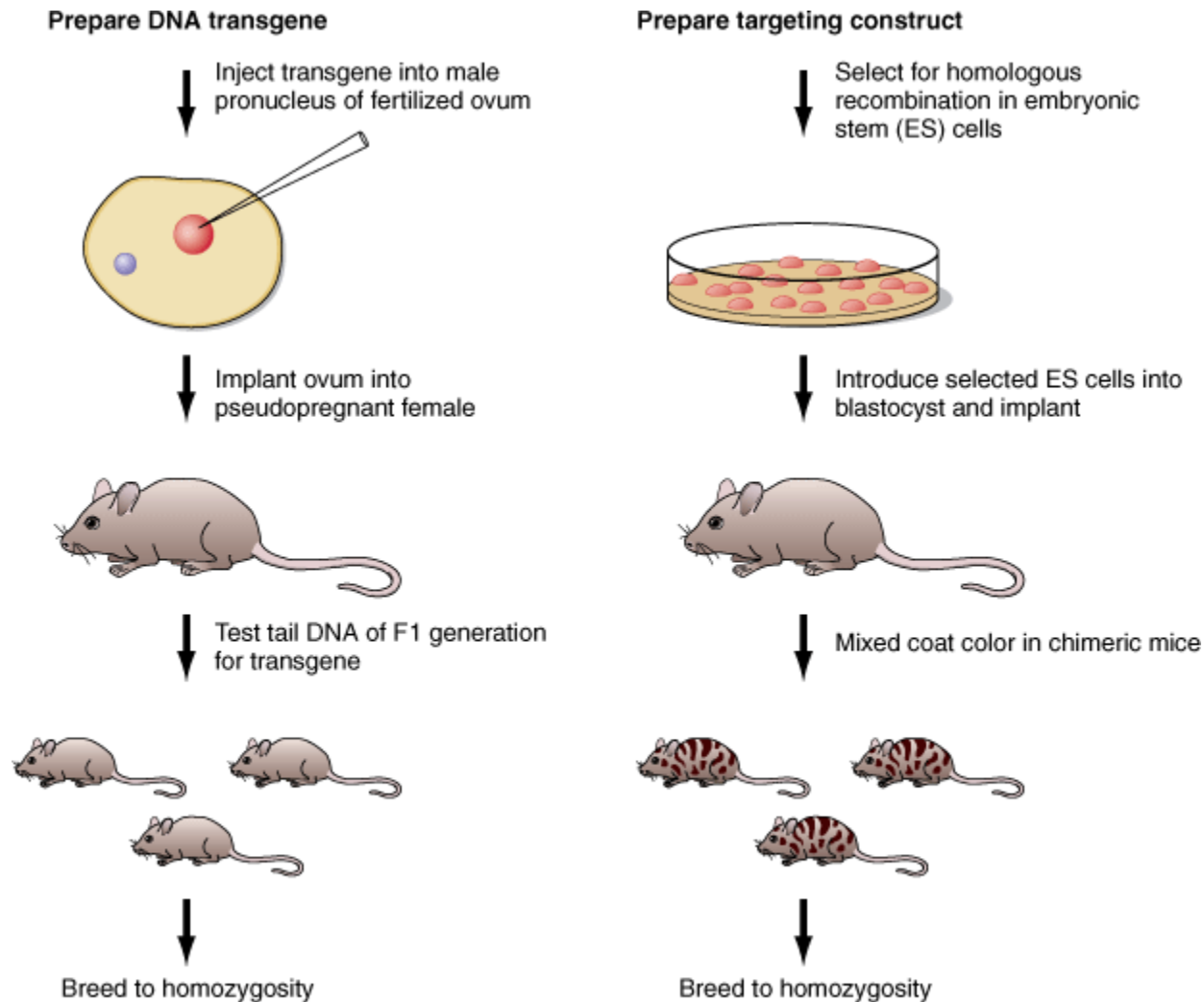
Mario R. Capecchi,  
Martin J. Evans and  
Oliver Smithies  
Nobel Prize 2007

# Traditional gene targeting

## Step 2. From gene targeted ES cells to gene targeted mice



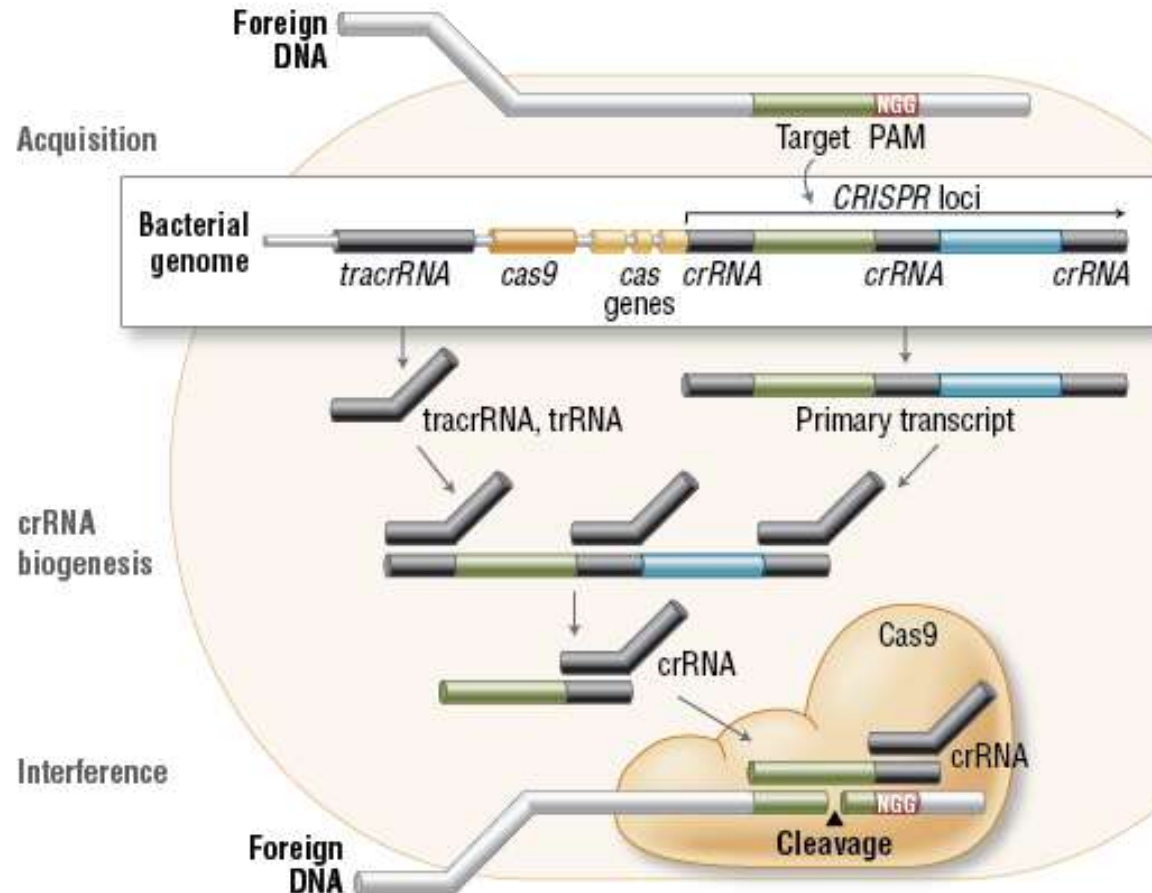
# Transgenic vs. gene targeting



Source: Fauci AS, Kasper DL, Braunwald E, Hauser SL, Longo DL, Jameson JL, Loscalzo J: *Harrison's Principles of Internal Medicine*, 17th Edition: <http://www.accessmedicine.com>  
Copyright © The McGraw-Hill Companies, Inc. All rights reserved.



# CRISPR/Cas9: Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) and CRISPR Associated (Cas) system-- bacterial self-defense mechanism



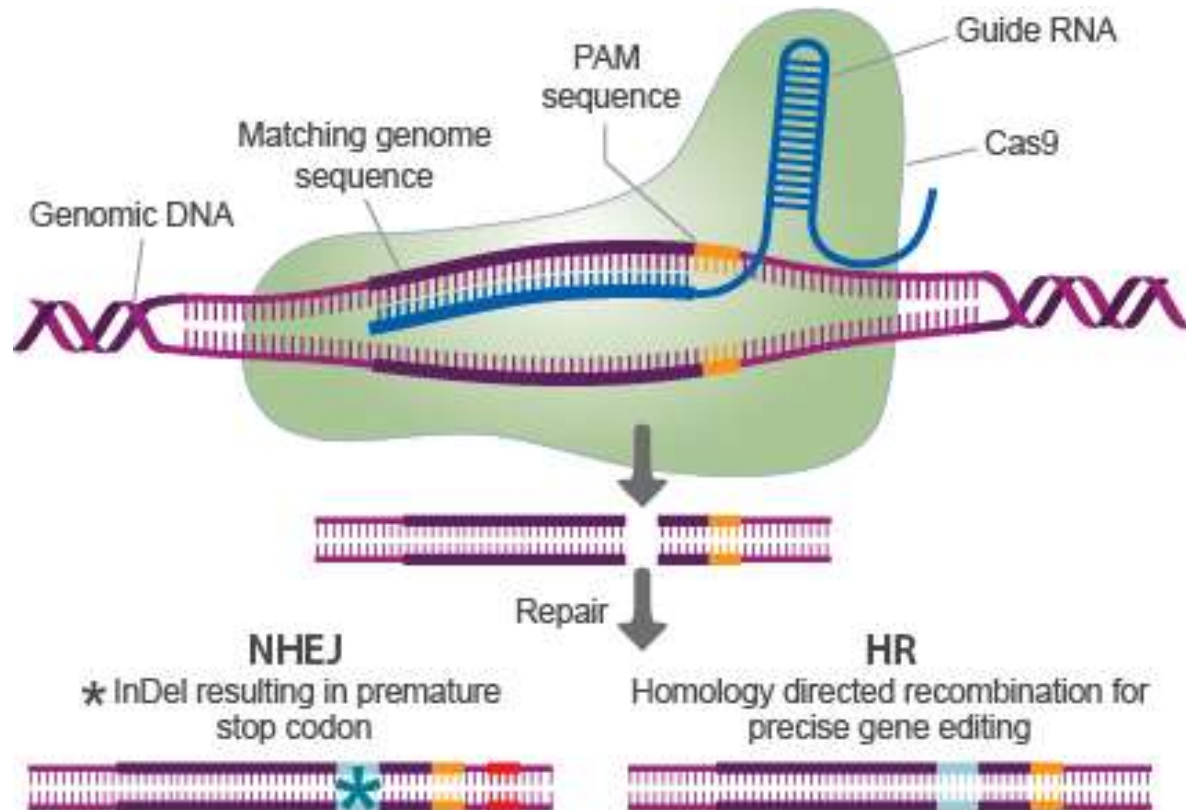
Ishino, Y., et al. (1987) *J. Bacteriol.* 169, 5429–5433.

PAM: protospacer-associated motif

crRNA: CRISPR targeting RNA

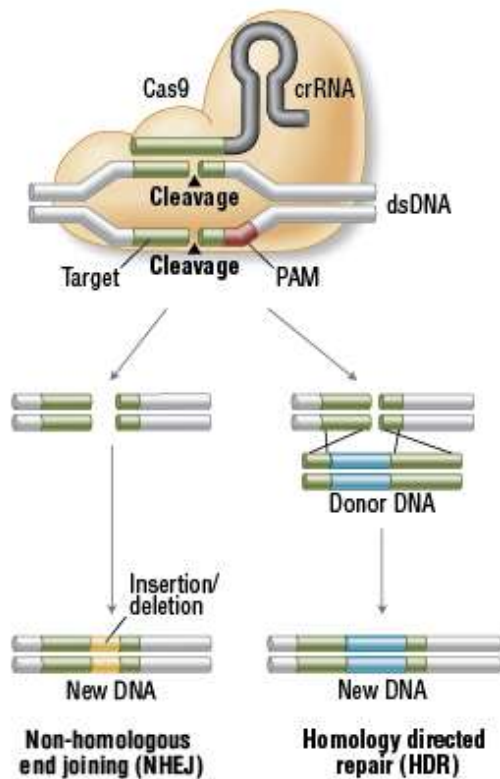
Required for the Cas( to recognized the sequence 2-5 conserved bp

# CRISPR/Cas9 mediated gene targeting -- a much more efficient method

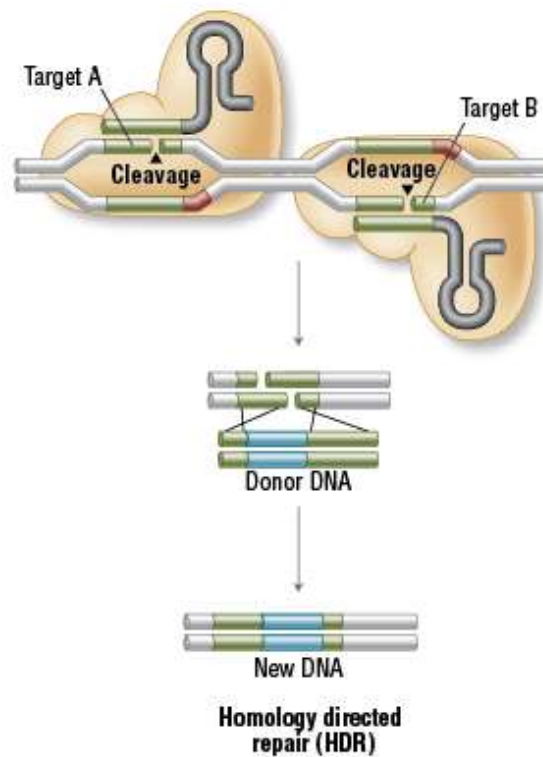


# Other variations of CRISPR/Cas9

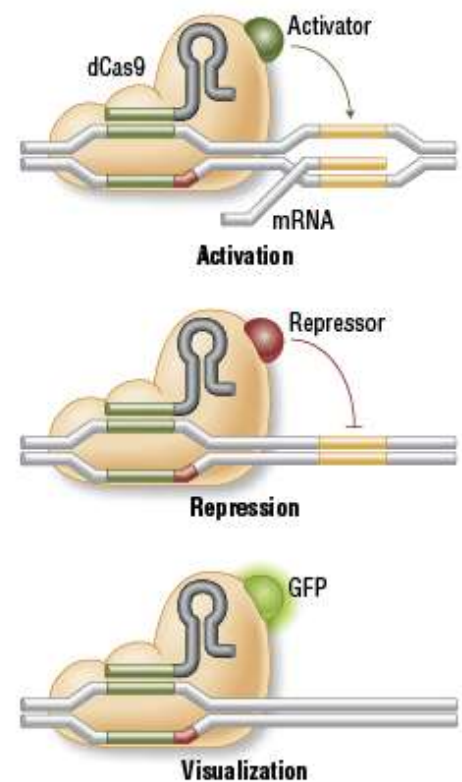
**A. Genome Engineering With Cas9 Nuclease**



**B. Genome Engineering By Double Nicking With Paired Cas9 Nickases**

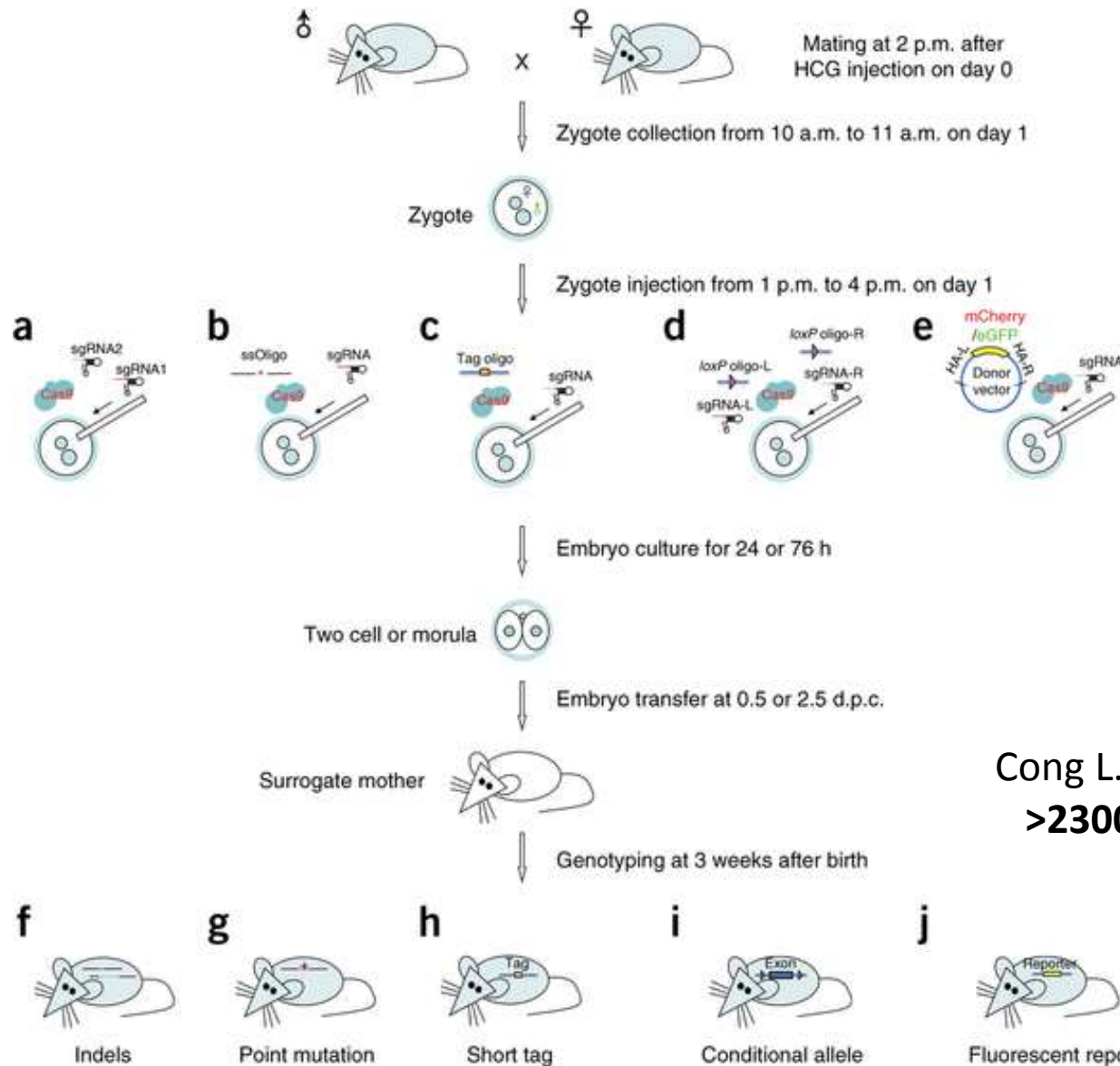


**C. Localization With Defective Cas9 Nuclease**



# CRISPR/Cas9 mediated gene targeting

## Time: 1-3 months



Injection of zygotes



Cong L... Zheng F. (2013) *Science*  
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