

# Tools for Probing Local Circuits: High-Density Silicon Probes Combined with Optogenetics

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<http://dx.doi.org/10.1016/j.neuron.2015.01.028>

To understand how function arises from the interactions between neurons, it is necessary to use methods that allow the monitoring of brain activity at the single-neuron, single-spike level and the targeted manipulation of the diverse neuron types selectively in a closed-loop manner. Large-scale recordings of neuronal spiking combined with optogenetic perturbation of identified individual neurons has emerged as a suitable method for such tasks in behaving animals. To fully exploit the potential power of these methods, multiple steps of technical innovation are needed. We highlight the current state of the art in electrophysiological recording methods, combined with optogenetics, and discuss directions for progress. In addition, we point to areas where rapid development is in progress and discuss topics where near-term improvements are possible and needed.

## Introduction

How does the brain orchestrate perceptions, thoughts, and actions from the activity of its neurons? Addressing these challenging questions requires methods capable of isolating, identifying and manipulating statistically representative fractions of the neurons of the investigated circuits at single-neuron and single-spike resolution during behavior (Alivisatos et al., 2013; Buzsáki, 2004; Carandini, 2012; Marblestone et al., 2013; Nicolelis et al., 1997). Electrical recording of extracellular action potentials ("spikes") is among the oldest neuronal recording techniques (Adrian and Moruzzi, 1939) and its physical principles are well understood (Buzsáki et al., 2013; Einevoll et al., 2013; Logothetis, 2003; Nádasdy et al., 1998). Electrical recordings have the additional advantage of simultaneously detecting the superimposed synaptic activity of neurons in the form of local field potentials (LFPs).

While the number of simultaneously recorded neurons has doubled every seven years over the past several decades (Stevenson and Kording, 2011), the widespread adoption of large-scale recording methods by the neuroscience community has generally lagged behind. However, significant recent technological innovations are now bringing large-scale recording methods into the mainstream, thereby enabling progressively more advanced experiments and analyses – and associated challenges. To meet the expectations of the BRAIN Initiative (<http://www.braininitiative.nih.gov/index.htm>), this trend of innovation coupled with translation over multiple technologies becomes increasingly important.

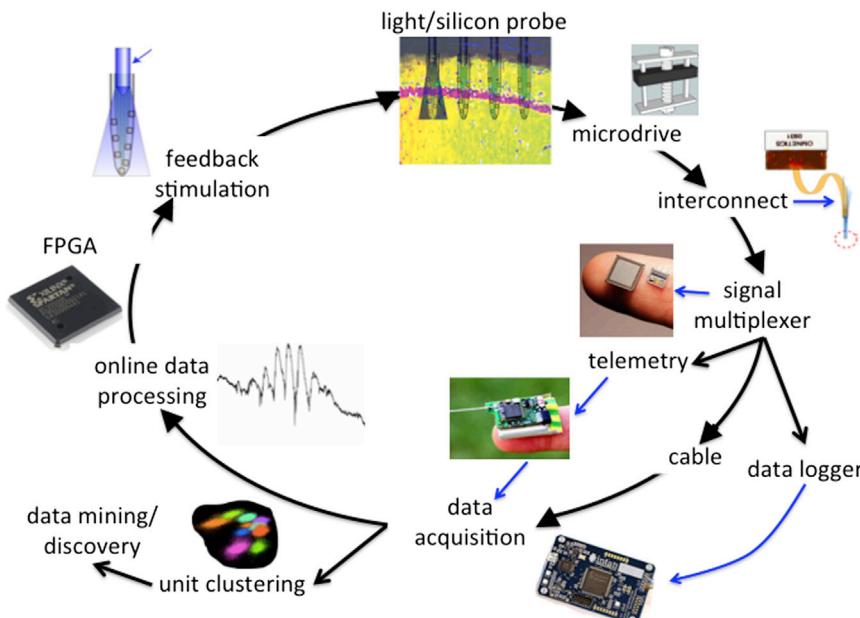
Relating the activity patterns of circuit components to behavior is a powerful method for inferring their role in organizing behavior. Testing their causal role, however, requires additional steps. Optogenetics has recently emerged as the appropriate method for fast manipulation of genetically identified neuron types. While optogenetic methods have the sufficient temporal speed for interacting

with neuronal circuits, their spatial resolution in currently used instantiations is typically very poor. Another limitation is the lack of multisite light delivery methods for flexible circuit control in freely behaving small rodents. Thus, new techniques are needed to validate hypotheses derived from the correlation measures between the spatiotemporal coordination of neurons and behavior.

To harness the maximum potential of combining large-scale recording methods, progressive innovation is needed at various levels of device integration. Our Primer focuses on technologies developed for small-size animals. However, the discussed methods can be adapted to nonhuman primates as well, and several recent innovations with specific solutions for primates have been reviewed recently (Cavanaugh et al., 2012; Diester et al., 2011; Gerits et al., 2012; Gray et al., 2007; Schwarz et al., 2014). Three issues are addressed: (1) state-of-art current methods and barriers, (2) approaches to make the current upper limit more accessible to mainstream neuroscience, and (3) a look ahead to technology innovations required to increase the upper limit of recording sites by one or two orders of magnitude. A flow chart of large-scale recording and the closed-loop targeted feedback perturbation of neurons in their native networks is illustrated in Figure 1.

## Sensing Neuronal Activity from the Extracellular Space

Neuronal activity gives rise to transmembrane current and the superposition of multiple potentials spreading through the extracellular medium can be measured as voltage ( $V_e$ ), referred to as local field potential (LFP). The somato-dendritic tree of the neurons forms an electrically conducting interior surrounded by a highly insulating membrane with hundreds to thousands of synapses located along it. They can conduct either inward (excitatory) or outward (hyperpolarizing, most often inhibitory) currents. The amplitude and time-varying patterns of the LFP



**Figure 1. Flowchart of Large-Scale Silicon Probe Recordings of Unit and LFP Activity Combined with Optogenetic Manipulation of Circuits**

The components of the chart are discussed in this primer. While large-scale recordings and optogenetic perturbations can be implemented separately, their combination provides a powerful tool for circuit analysis. FPGA, field-programmable gate array; digital signal processing (DSP).

depend on the proportional contributions of the multiple current sources and are modified by the various properties of the brain tissue (Buzsáki et al., 2012; Einevoll et al., 2013). Because the amplitude of  $V_e$  inversely scales with the distance between the source and the recording site, the larger the distance, the less informative is the measured LFP regarding its origin.

Action potentials generate the largest-amplitude currents across the somatic membrane and can be detected as extracellular “unit” or spike activity. In addition to the distance between the neuron and the recording electrode, the magnitude of the extracellular spike depends on the size and shape of the neuron (Buzsáki et al., 2012; Einevoll et al., 2013). **Extracellular spikes are negative near the soma (sink), corresponding to the fastest rate of  $\text{Na}^+$  influx into the cell body (Figure 2).**

In cortical structures, the bodies and apical dendrites of densely packed pyramidal neurons lie parallel to each other, making the separation of neurons on the basis of their extracellular spikes difficult. On the other hand, such architecturally regular geometry is favorable for estimating afferent-induced synaptic inputs to the circuit since afferents in cortical layers run perpendicular to the dendritic axis, and this arrangement facilitates the combination of synchronously active synaptic/transmembrane currents by superposition (Figure 2B). In summary, both identification of spikes of different neurons and the inference of population synaptic activity from the extracellular voltage require high-resolution spatiotemporal sampling of the extracellular space, ideally equal to or smaller than the diameter of neuronal cell bodies.

### Hardware Components for Large-Scale Monitoring of Neuronal Activity Recording Electrodes

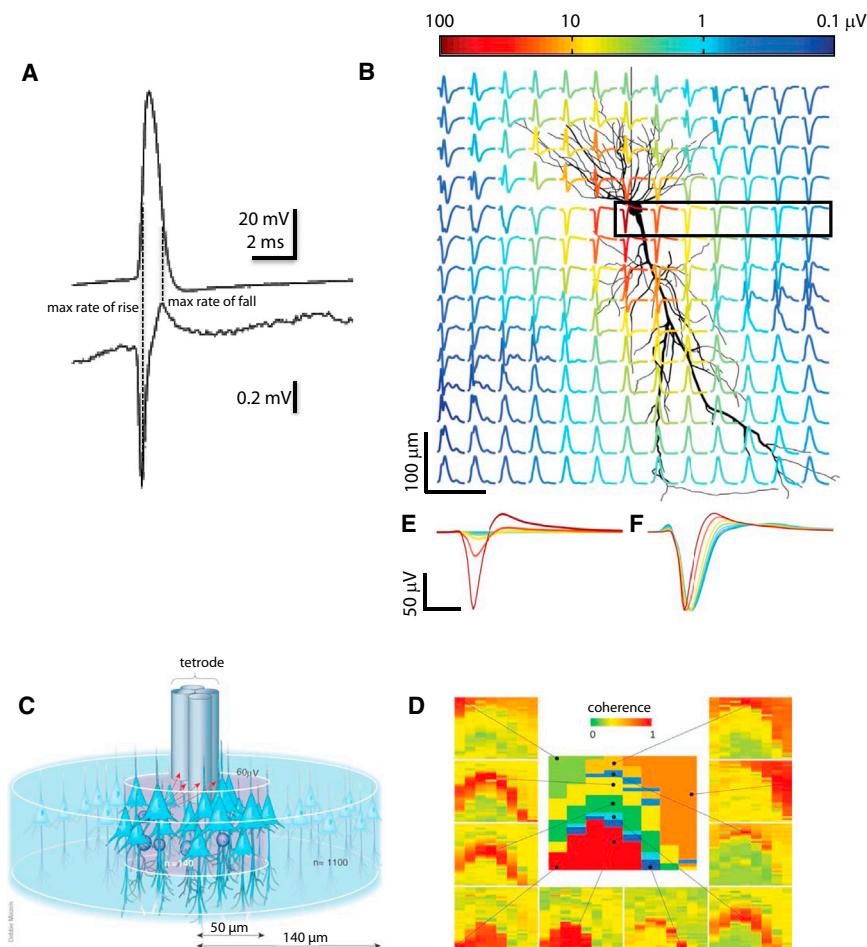
Action potentials can be detected as voltage by placing a conductor, such as a wire, in close proximity to the neuron (Adrian and Moruzzi, 1939). Because neurons of the same class

generate largely identical action potentials, identifying an individual neuron from its extracellular spikes is possible by moving the electrode tip substantially closer to the cell body of one neuron than to the neighboring neurons. To record another neuron or dissociate the spike signals of two different nearby neurons, more than one electrode is needed.

The use of two or more recording sites in close proximity allows for the triangulation of distances between the spike-emitting cell bodies and the electrodes because the spike amplitude waveform morphology changes as a function of distance and direction from the neuron (Drake et al., 1988). Often, this task is accomplished with two or four twisted wires (dubbed “stereotrodes” or “tetrodes” (McNaughton et al., 1983; Wilson and McNaughton, 1993)). Ideally, the tips are separated in three-dimensional space so that the point-source localization of several neurons is possible (Drake et al., 1988; Perlin and Wise, 2004; Wilson and McNaughton, 1993).

Typical types of wire tetrodes can “hear” hippocampal CA1 pyramidal cells in rats as far as 140  $\mu\text{m}$  laterally. A cylinder with this radius contains  $\sim$ 1000 neurons, which is the number of cells theoretically recordable by a single tetrode (Henze et al., 2000); these estimates vary by neuron size, type and brain region. Yet, in practice, only 5–15 neurons can be reliably separated in limited-duration recordings. The low firing-rates of many neurons (Buzsáki and Mizuseki, 2014; Schwindel et al., 2014; Shoham et al., 2006), as well as low amplitudes of the majority of detected spikes (Figure 2;  $<40 \mu\text{V}$ ; Schomburg et al., 2012) preclude a reliable neuron isolation with currently available clustering algorithms (Harris et al., 2000). Furthermore, mechanical damages associated with probes movements may damage neuronal processes, or tear small vessels (Claverol-Tinture and Nadasdy, 2004; Koza et al., 2010; Seymour and Kipke, 2007; Tsai et al., 2009).

Electrical recording from neurons is invasive. Therefore, the desire for large-scale monitoring of neurons with many recording sites in a small volume and the desire to minimize the tissue damage inflicted by the electrodes compete with each other (Scott et al., 2012). Micro-Electro-Mechanical System (MEMS)-based recording devices can reduce the technical limitations inherent in wire electrodes because with the same amount of tissue displacement, the number of monitoring sites can be substantially increased (Kipke et al., 2008; Najafi and Wise, 1986), and a tapered tip is readily achieved. Most micromachined electrode



**Figure 2. Electric Signals in the Extracellular Space**

(A) Simultaneous intracellular and extracellular recordings of action potential in a hippocampal CA1 neuron *in vivo*. Note that the trough and peak of the extracellular unit spike (0.1 Hz–5 kHz) correspond to the maximum rate of rise and maximum rate of fall of the intracellular action potential.

(B) Extracellular contribution of an action potential ("spike") to the LFP in the vicinity of the spiking pyramidal cell. The magnitude of the spike is normalized. The peak-to-peak voltage range is indicated by the color of the traces. Note that the spike amplitude decreases rapidly with distance from the soma. The distance dependence of the spike amplitude within the pyramidal layer is shown (bottom left panel) with voltages drawn to scale, using the same color identity as the traces in the boxed area. The same traces are shown normalized to the negative peak (bottom right panel).

(C) Multisite electrodes can estimate the position of the recorded neurons by triangulation of extracellular voltage measurements. Distance of the electrode tips from a single pyramidal cell is indicated by arrows. The spike amplitude of neurons ( $>60 \mu\text{V}$ ) within the gray cylinder (50  $\mu\text{m}$  radius), containing  $\sim 100$  neurons, can be used for effective separation by current clustering methods.

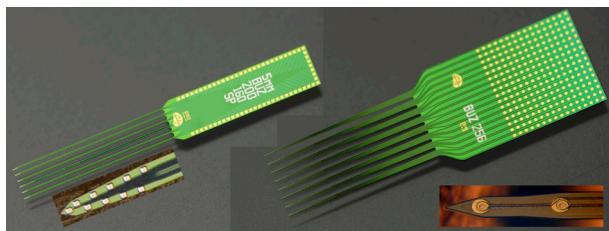
(D) Coherence maps of gamma activity (30–90 Hz) in the hippocampus of a freely behaving rat. The ten example sites (black dots) served as reference sites, and coherence was calculated between the reference site and the remaining 255 locations of an 8-shank (32-site vertical linear spacing at 50  $\mu\text{m}$ ) probe.

(A) Reproduced from (Buzsáki et al., 1996). (B) Reproduced from (Buzsáki et al., 2012). (C) Reproduced from (Buzsáki, 2004). (D) Reproduced from (Berényi et al., 2014).

arrays ("silicon probes"; Figure 3) are formed by photolithographic patterning of thin films of conductors and insulators on silicon substrate. Such a fabrication process allows tailoring the size, shape and the arrangement of the electrodes according to the neural density and local circuit architecture of a specific brain region. Advances in lithography, metal deposition procedures and quality control, an ever-growing number of probe architectures have been developed over the years to meet experiment-guided specific requirements (Bartels et al., 2008; Hofmann et al., 2006; Jamille and David, 2002; Ludwig et al., 2006; McCreery et al., 2006; Motta and Judy, 2005; Musallam et al., 2007; Neves and Ruther, 2007; Nordhausen et al., 1996; Rennaker et al., 2005; Seidl et al., 2011; Wise et al., 2004). The resulting silicon devices enable high-density LFP recordings and large-scale single unit recordings in a variety of brain structures and species (Figure 4; Agarwal et al., 2014; Barthó et al., 2004; Blanche et al., 2005; Broome et al., 2006; Csicsvari et al., 2003; Du et al., 2011; Fujisawa et al., 2008; Lin et al., 2014; Montgomery et al., 2008; Pouzat et al., 2002).

Further advances in the electrode design and arrangement require to push beyond existing technology capabilities to achieve higher recording densities. One of these is the need to reduce probe dimensions to enable probe designs of about the

same size, but having significantly more electrode sites. Such a goal is limited mainly by the width of the embedded conductive leads (also called traces or interconnects) that are needed to connect each electrode site to an extracranial bond pad for connection to the external electronics interface. Experience in rodents suggests that when the width of the probe shanks is wider than  $>60 \mu\text{m}$  (assuming a shank thickness less than 20  $\mu\text{m}$ ), the unit recording quality of the probe decreases rapidly, perhaps due to compression or damage to the dendrites of the surrounding neurons (Claverol-Tinture and Nadasdy, 2004). There is a viable technology roadmap for developing next-generation fine-featured probes for high-resolution recording. One approach involves incremental advances in current MEMS microfabrication processes that result in smaller minimum feature sizes. A related approach involves using alternative MEMS technologies that intrinsically support finer features, e.g., electron-beam lithography (Du et al., 2011). A third MEMS-based approach is to integrate active electronic components in the probe shanks to provide electronic switching that reduces the lead count, and thereby reduces the required shank dimensions (Seidl et al., 2011). Efforts are under way to manufacture a single-shank (50  $\mu\text{m}$  wide) probe with up to 960 closely spaced (20  $\mu\text{m}$ ) contacts, of which 384 can be selected for simultaneous



**Figure 3. Silicon Probes Designed for Unit Sampling and High Spatial Density Monitoring of LFP**

(A) Six-shank “decatrodes” (Buzsaki64sp probe from NeuroNexus) for recording and effective separation of single neurons spikes. Vertical separation of the recording sites ( $160 \mu\text{m}^2$ ):  $20 \mu\text{m}$ . Shanks are  $15 \mu\text{m}$  thick and  $52 \mu\text{m}$  wide.

(B) Eight-shank probe for large spatial coverage ( $2.1 \text{ mm} \times 1.6 \text{ mm}$ ; Buzsaki256 probe from NeuroNexus). Recording sites ( $160 \mu\text{m}^2$ ) are separated by  $50 \mu\text{m}$ . Shanks taper from  $96 \mu\text{m}$  to  $13 \mu\text{m}$  at the tip.

recording at a given time (HHMI-Allen Institute-Gatsby Foundation-Wellcome Trust Consortium), thus enabling more powerful experimental approaches to understanding the interaction of neurons and neuronal pools. A non-MEMS approach is to use small diameter (less than 7 microns), parylene coated carbon fibers to create fine-featured microelectrode arrays having large numbers of ultrasmall microelectrodes (Kozai et al., 2012).

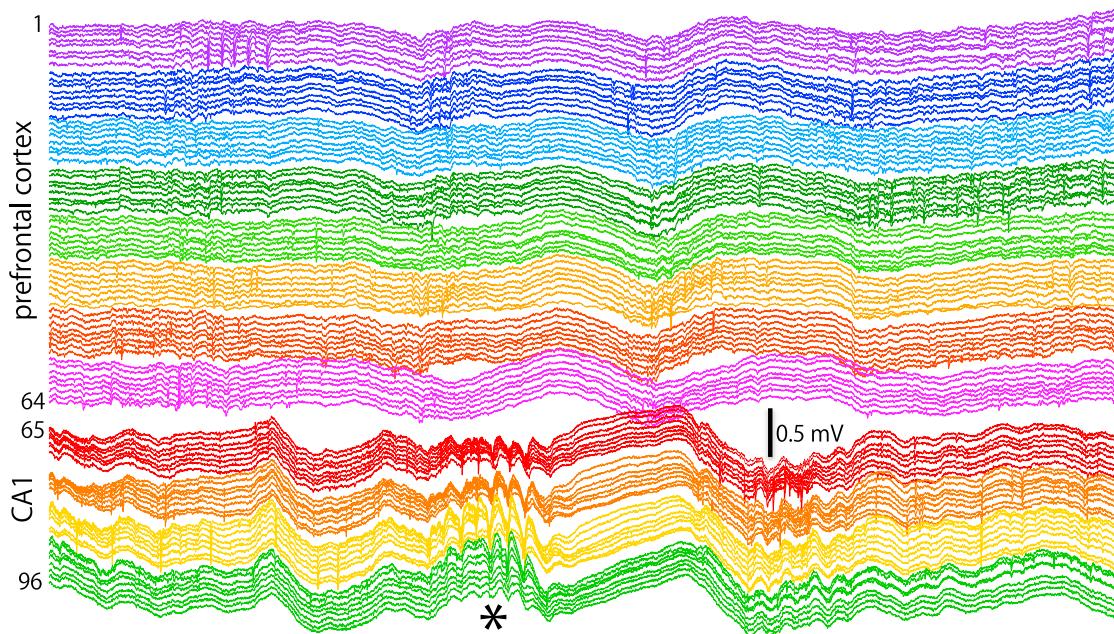
A second advance in probe design space is to optimize shank geometry and recording site positions for maximal dense local sampling of neurons, which is important for capturing the dynamics of neuronal ensembles with complex connectivity patterns. In particular, one goal is to minimize the “backside shielding” effect of typical planar silicon probes that have electrode sites all on one side (the “topside”; Moffitt and McIntyre, 2005). One design approach for minimizing backside shielding is to locate electrode sites at or near the edge of the shank, perhaps in concert with additional sites located off the edge (Seymour et al., 2011). An alternative approach is to extend planar architectures to manufacture double-sided electrodes (Du et al., 2009; Perlin and Wise, 2004). A third design space opportunity is three-dimensional microelectrode arrays, which, in principle, offer high-resolution functional mapping of local circuits (Wise et al., 2008). Experiments indicate that with three-dimensional triangulation the source localization of neurons can precise as  $17 \mu\text{m}$  (Du et al., 2009). Three-dimensional electrode arrays present difficult technical challenges involving advanced hermetic packaging having micron-level precision, complex lead transfers for hundreds of leads, and sophisticated surgical techniques, combined with important practical needs for reliability and manufacturability (Bai et al., 2000; Hoogerwerf and Wise, 1994; Neves and Ruther, 2007; Pang et al., 2005; Perlin and Wise, 2010; Yao et al., 2007). Furthermore, inserting three-dimensional probes into the vessel-rich neocortex (Tsai et al., 2009) without compromising the neuropil will remain a serious challenge.

The above discussion of probe geometry can help estimate the optimal configuration of future probes and the maximum yield of simultaneously recordable neurons. For the purpose of recording and separating the maximum number of neurons, the recording sites should tile the entire shank surface of the

probe, which should, in turn, have minimal dimensions. Putting aside the problem of interconnects for a moment, and assuming the need for at least  $8 \times 8 \mu\text{m}^2$  recording sites with  $10 \mu\text{m}$  center-to-center separation, 200 sites per shank on a  $20 \mu\text{m}$ -wide probe can cover the entire cortical gray matter (<2 mm). Adding recording sites on both sides would double the number of monitoring sites and increase the unit yield (Du et al., 2009). Placing three shanks in  $\leq 100 \mu\text{m}$  triangle would allow effective three-dimensional triangulation of cortical neurons in all cortical layers. Such a probe could, in principle, record from between 1,000 and 5,000 neurons and determine the three-dimensional position of both cell bodies and the main dendrites of each neuron. These technical improvements are within reach but will require high-density off-chip lead transfers, probably in conjunction with on-probe circuitry (Al-Ashmouny et al., 2012; Bai and Wise, 2001; Perlin and Wise, 2010; Wise et al., 2004). We should emphasize that even slim-shank probes come at the expense of tissue damage, fractional displacement volume and associated disruption of physiological activity that needs to be carefully investigated (Claverol-Tinture and Nadasdy, 2004; Polikov et al., 2005; Tsai et al., 2009).

Unfortunately, there is not a “one-fit-for-all” solution for probe design. Various neuron types have different sizes, dendritic configurations and densities and require “custom-fit” probes for effective neuron isolation. This area of research could benefit tremendously from appropriate modeling of the extracellular space populated with realistic neuron geometries (Reimann et al., 2013), glia and vessels and physiological spiking patterns. In turn, virtual electrodes could be embedded into such a “neuron cube” to identify the most appropriate probe geometry, recording site configurations and neuron shielding effects. Such a neuron cube model could also offer “ground truth” for the neuron-spike identity problem and be exploited for perfecting unit clustering algorithms.

Major questions in neuroscience address circuit changes and stability over extended time periods, and dealing with such problems presents important challenges for long-term, high-quality unit recordings. Although several laboratories have reported extended recordings for weeks and even months with diverse types of microelectrodes in various species (Chestek et al., 2011; Jackson and Fetz, 2007; Kipke et al., 2003; Nicolelis et al., 1997; Rousche and Normann, 1998; Ruther et al., 2011; Sumer et al., 2005), chronic performance of electrodes, to date, is unpredictable and highly variable. A major cause of probe failure over time is mechanical instability and neuron damage. Ultrathin, and flexible probes can alleviate the mechanical damage associated with probe penetration by reducing the mechanical mismatch between probe and brain tissue, and allow the probe to move with the adjacent tissue and neurons (Bjornsson et al., 2006). Biodegradable materials, such as silk fibroin, patterned on Parylene substrate, can provide sufficient mechanical support for the probes to be inserted in the brain, while allowing free-flotation of the flexible probe after the silk layer is absorbed (Kim et al., 2010; Tien et al., 2013; Wu et al., 2015). There is a tremendous need to improve the electrode/tissue interface to enhance the signal-to-noise ratio (SNR) and increase the yield of neurons per recording session. Conducting polymers such as PEDOT:PSS are used to decrease the electrochemical impedance



**Figure 4. High-Quality Unit Recordings**

High-quality unit recordings from the superficial layers of the prefrontal cortex (traces 1–64) and the hippocampal CA1 pyramidal layer (traces 65–96). During sleep, 400 msec epoch. Asterisk indicates sharp wave ripple. Data are reproduced from Fujisawa et al. (2008).

mismatch between electrode and brain tissue interface and further reduce the chronic reactive tissue response (Abidian and Martin, 2008; Ludwig et al., 2006; Shain et al., 2003; Spataro et al., 2005). Combining electrical recordings with local delivery of electrical stimulation, chemical substances, such as neurotransmitters, and light for optogenetics (see below) will play an growing role in the exploration of neural circuit mechanisms (Chen et al., 1997; Cheung et al., 2003; Li et al., 2007; Seidl et al., 2010, 2011).

Penetrating probes can isolate multiple single-neurons from any structure. However, increasing the number of shanks per probe can cause irreversible damage to brain tissue limiting the monitoring of large-scale neural dynamics occurring over contiguous areas of cortex. An emerging approach is to use surface electrodes. The NeuroGrid is an organic material-based, ultraconformable, biocompatible and scalable neural interface device that can acquire both LFPs and spikes from superficial cortical neurons without penetrating the brain surface (Khodagholy et al., 2015). The device is fabricated from “soft organic” material to allow conformation of the probe on the curvilinear surface of the brain, as well as improving the efficiency of ion to electron conversion (Khodagholy et al., 2011). In concert with advances in microelectronics and data processing such as CMOS technologies used in Multi-Electrode Arrays (MEAs) that can yield several thousands recordings sites, LFP and spike samples recorded by the NeuroGrid can be substantially increased (Hutzel and Fromherz 2004; Lei et al., 2011; Muller et al., 2012; Chichilnisky and Baylor, 1999; Meister et al., 1994).

#### Microdrives and Probe-Preamplifier Interconnect

Implanted and “fixed”-depth electrodes have good mechanical stability but their unit yield is low. During surgical anesthesia the brain expands and shrinks, and therefore even precisely placed

electrodes move their position relative to neuronal bodies. An effective way of increasing unit yield is to place the electrodes several hundred  $\mu\text{m}$  above the target area and move them slowly ( $<100 \mu\text{m}$  steps a day) by a microdrive. The unproven assumption of this practice is that moving the shank tips so slowly, the neurons, glia and vasculature have time to move away and rebuild (Fujisawa et al., 2008). Advancement of the electrode can be achieved by microdrives (Fee and Leonardo, 2001; Vandecasteele et al., 2012; Wilson and McNaughton, 1993; Yamamoto and Wilson, 2008). The key requirements of a microdrive are precision, mechanical stability and minimal size and weight. Drives add additional weight and volume to the head-gear and are seriously limiting factors in experiments that require free movement of small rodents. In our experience, a head implant totaling 10% of the preoperative body weight does not impede free behavior. Assuming an adult weight of a 30 g mouse, the package weight budget should be  $<3 \text{ g}$ , although for many experiments the weight should be even less.

Any small drift ( $<10 \mu\text{m}$ ) between the electrode tip and the neuron(s) can affect recording stability. Such drifts are typically brought about by headshakes, intense grooming, banging the head stage against cage walls and, importantly, during plugging and unplugging the head connectors. The latter problem can be reduced significantly if the microdrive, containing the electrodes, is physically separated from the head connector by a flexible interconnect cable. Polyimide cables have been used successfully for such purpose because of their flexibility and integrated design with silicon probes (Berényi et al., 2014; Stieglitz et al., 2005) (NeuroNexus Inc.). Cables can also be fabricated as an integral part of the probe so that bonds are required only at the connector end (Yao et al., 2007). With high site-count probes,

however, there are still remaining challenges. Higher-density bonding technologies are needed to permit lead transfers on <20  $\mu\text{m}$  centers, and traditional connectors with >100 sites and thick cables are not practical for freely behaving small rodents. On-site multiplexing is one solution but size, weight and cost factors must be considered.

#### On-Head Signal Amplification and Multiplexing

A critical aspect of miniaturization is the deployment of signal multiplexers. Earlier versions of multiplexers had either low channel counts or limited high- or low-pass frequency characteristics for simultaneous recordings of both unit and LFP signals in the physiological range and required de-multiplexing and digitization of the transmitted multiplexed analog signals (Berényi et al., 2014; Du et al., 2011; Harrison, 2008; Olsson et al., 2005; Szuts et al., 2011; Viventi et al., 2011). The recent availability of small size 32- and 64-channel digital multiplexers (RHD-2132 or RHD-2164 chip or die; Intan, Inc.) has accelerated the spread of high-channel-count recordings in physiological laboratories. The digital output of the signal multiplexers (Harrison, 2008) allows direct streaming of the neurophysiological data to the computer. Despite important advances in miniaturization, high-channel-count recordings (>100) in behaving mice and other small animals still remain a challenge because routing the probe sites to the inputs of the signal multiplexers still requires large connectors. Placing headstages permanently on the head of the animal (Berényi et al., 2014) is an alternative but expensive solution and they add unwanted volume and weight. Given that only 15% of the weight of the functional 64-site electrode system comes from the digitizing headstage integrated circuit (the bulk coming from the printed-circuit board, connector, and sealing epoxy), the most significant development step in packaging and electronics toward a 1,000 site recording system usable in a behaving mouse would be a next-generation integrated circuits that would support larger numbers of channels and that could be intimately packaged with a high-density neural probe (Lei et al., 2011).

An ideal solution to size reduction is integrating the signal amplifiers and multiplexers to the probe end (Csicsvari et al., 2003; Olsson et al., 2005) or into the flexible interconnect cable. The current limitation of such improvement is the high cost of design and fabrication, which can exceed several million dollars. In principle, the active circuits can be placed on the probe shanks, effectively reducing the number of interconnects between the recording sites and electronics outside the brain. However, active circuits also occupy volume and limit the number of recording sites. Experience dictates that any structure >60  $\mu\text{m}$  wide deteriorates the recording ability of probes. Furthermore, such brain-embedded circuits must operate at low enough power levels so that significant heating of the probe is avoided since heating can impact local neurons. Active circuits must also be electrically shielded or located such that the electric fields generated by the circuit components do not influence the spiking activity of nearby neurons (Jefferys, 1995).

#### Data Transmission: Cable, Telemetry, and Data Loggers

Cables interconnecting the head stage and the recording system can be completely eliminated by telemetry or data loggers. While up to 128-channel telemetry systems have been successfully used in small animals (Greenwald et al., 2011; Harrison et al.,

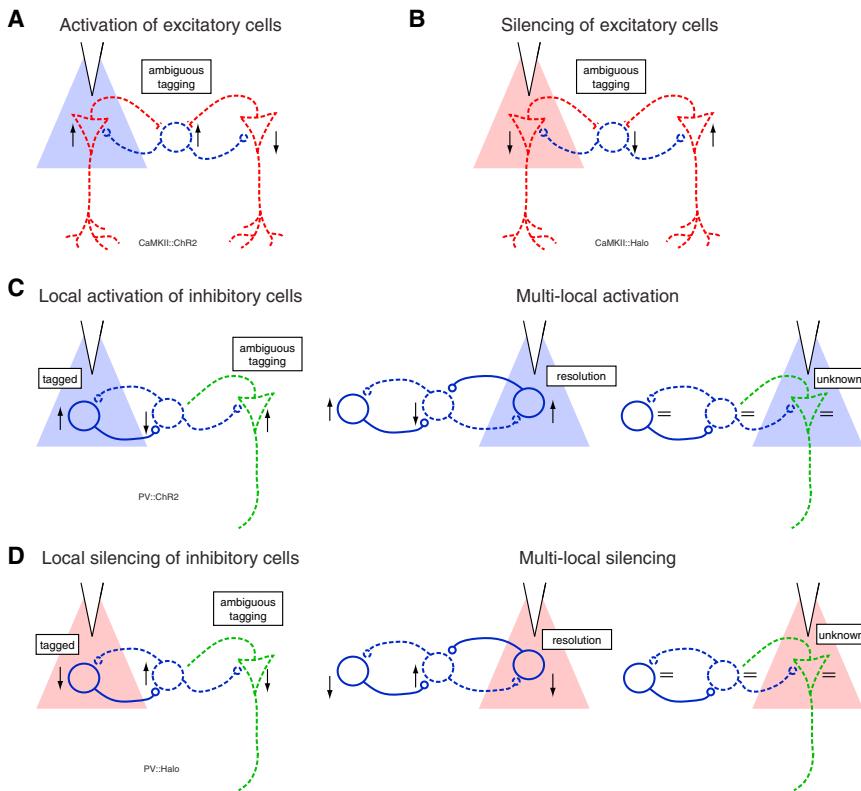
2011; Sodagar et al., 2007; Szuts et al., 2011), cable connections allows much higher bandwidth, higher channel counts and lower noise. On-probe multiplexing followed by analog-to-digital conversion, thresholding, and data compression can make use of the relatively sparse nature of neural recordings to boost the channel capacity by well over an order of magnitude in some situations (Olsson et al., 2005). Data loggers present another solution for specific applications, such as studying underground, underwater behaviors, social interactions or flying (Yartsev and Ulanovsky, 2013). Unfortunately, the power source required for telemetry adds additional weight and limits the duration of the experiments, while less power-intensive data loggers do not allow the experimenter to monitor the recorded data during the experiments. A combination of these two approaches may optimize their cost-benefit by storing all data on a data logger, and transmitting a few user-selected channels, when needed, via a telemeter and allowing bidirectional communication perturbation of circuits and recording (see below). Externally powered devices (Wentz et al., 2011) may provide solutions for long-term wireless interfacing.

#### Data Acquisition and Online Processing

Recent rapid progress with on-head digital multiplexing (Harrison, 2008) and direct streaming of the neurophysiological data to the computer has made high-density recordings affordable even for small laboratories. Community-driven improvement of shared software (e.g., [klustaviewas@googlegroups.com](mailto:klustaviewas@googlegroups.com); <http://neuroscope.sourceforge.net/>; <http://klusta-team.github.io/klustakwik/>; <http://www.open-ephys.org/>) is expected to provide new solutions for flexible data display and fast processing of the recorded physiological and behavioral data for closed-loop interactions with brain circuits. These include on-line detection of particular LFP and unit firing patterns of isolated single spikes, bursts or arbitrary combinations of multiple site spike timing relationships associated with any chosen aspect of behavior or spatial localization of the animal to drive instantaneous feedback perturbation of multiple single neurons or sites (Stark et al., 2012). On-line or real-time processing of even complex neural data is becoming feasible by microprocessors and field-programmable gate arrays (FPGAs; Muller et al., 2012).

#### Combination of Large-Scale Unit Recordings with Optogenetics

The recent advent of optogenetics (Boyden et al., 2005; Deisseroth, 2011; Zemelman et al., 2002) provides a solution for identifying specific, genetically defined neuronal subtypes in blind extracellular recordings by expressing light-sensitive opsins in a given neuronal population. Both activation and silencing strategies can be used for this purpose. Several laboratories have offered solutions for optical stimulation of local circuits combined with simultaneous neuronal recordings (Cardin et al., 2010; Halassa et al., 2011; Han et al., 2009; Kim et al., 2013; Kravitz et al., 2010; Lu et al., 2012; Rubehn et al., 2013; Voigts et al., 2013; Zhang et al., 2009). However, a number of technical issues must be addressed to exploit the full potential of combined large-scale recording and optogenetics, including local delivery of low-intensity light, application of appropriate stimulus waveforms, and replacement of large benchtop lasers with small head-mounted LEDs or laser diodes (see below).



**Figure 5. Ambiguity and Disambiguation of Neuron Identity by Optical Tagging**

(A) Identification of ChR2-expressing excitatory cells (red) by light pulses is often ambiguous because they can drive nearby interneurons (blue) at a short latency. Since opsin expression level and spiking threshold may vary among principal cells, and since synaptic transmission from principal cells to interneurons may be rapid and strong, some principal cells may show decreased firing rates.

(B) Similarly, silencing of several excitatory cells can bring about a short-latency rate decrease of interneurons and circuit-mediated rate increase of excitatory cells.

(C) Strong optogenetic activation of inhibitory cells may also be ambiguous, since disynaptic disinhibition of opsin-free cells (dashed green; recorded on another shank) may also occur at a short latency. The ambiguity may be resolved or reduced if the neuron responds at a shorter latency upon direct illumination of the neighboring shank (central panel) but not if no change in firing rate is detected (right panel).

(D) Silencing of inhibitory cells results in decreased spiking of illuminated opsin-expressing cells (direct effect), increased spiking of opsin-free cells (synaptic effect), and perhaps interneuron-mediated spiking decrease of other cells (disynaptic effect; green). The direct and disynaptic effects are particularly hard to differentiate upon prolonged global illumination, but may be disambiguated using sequential multisite illumination.

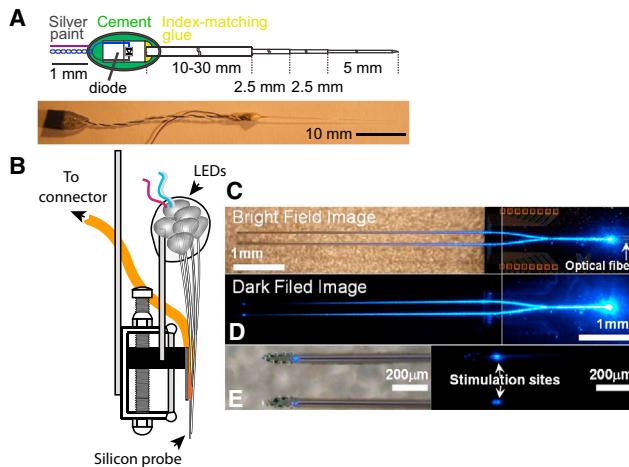
Integrated recording-optogenetic methods can be used to accomplish at least three goals: (1) identification of genetically labeled neurons (optical “tagging”), (2) physiological characterization and classification of neuron types and (3) testing the causal roles of the identified neurons in the performance of local circuits.

#### Strategies for Optogenetic Tagging of Single Neurons

The optogenetic method can, in principle, be used to distinguish cortical principal cells from interneurons, identify neurons with specific neuromodulators (Madisen et al., 2012) and differentiate physiologically similar neurons projecting to different targets (Packer et al., 2013). The rapidly growing Cre mouse (Madisen et al., 2012; Taniguchi et al., 2011) and rat (Witten et al., 2011) lines, intersection virus methods (Gradinaru et al., 2010) and other novel ways of marking neurons pave the way to ever more precise definitions of neuronal identity (Packer et al., 2013; Scanziani and Häusser, 2009). However, because neurons are both embedded in circuits and contribute to circuit function, their perturbation can bring about secondary changes, which need to be separated from the primary action of optical stimulation (Figure 5). On the one hand, variable expression levels of opsins can yield false negative effects. On the other hand, indirect inactivation of neurons can arise from an undesirable recruitment of nontagged neurons by *trans*-synaptic effects. Optogenetic activation of principal neurons by light pulses can discharge their partner interneurons (Csicsvari et al., 1998; Miles, 1990) and, potentially other principal cells at short latency and high fidelity. Separation of directly and indirectly driven neurons by latency criteria is highly unreliable (Kvitsiani et al., 2013). Silencing principal cells suffer from the same technical problems

since the connected partners can also decrease their rates. Optogenetic activation of inhibitory interneurons can induce a robust rate increase in the directly activated interneurons and suppressed spiking or no-change in the remaining neurons. Yet strong inhibition, brought about by strong light and synchronously discharging interneurons, may bring about rebound spiking in principal cells and possibly other interneurons (Figure 5C; Stark et al., 2013). Tagging of inhibitory interneurons by optogenetic silencing is more reliable than by their activation, although disynaptic disinhibition can mediate rate decrease in opsin-free cells. Several complementary strategies can be used to increase the precision of optogenetic tagging of neurons, including the application of different waveform patterns, but perhaps more important is using minimum light intensities of focally applied light and monitoring the induced effects both locally and more distally from the light source (Figure 5).

Many of current technical problems of neuron identification and local circuit analysis arises from the use of large-diameter optical fibers (Cardin et al., 2010; Halassa et al., 2011; Han et al., 2009; Kravitz et al., 2010) or brain surface illumination (Huber et al., 2008; Lee et al., 2012; Moore and Wehr, 2013) at high light power. To activate neurons at a distance from a light source, high intensities (several mW or higher) are often used (Cardin et al., 2010; Halassa et al., 2011; Han et al., 2009; Huber et al., 2008; Kravitz et al., 2010; Moore and Wehr, 2013). High-power photo-stimulation can generate photoelectric artifacts, activate/silence numerous neurons both directly and indirectly (Figure 5) and induce synchronous discharges in multiple neurons, making “spike sorting” difficult or impossible. Such technical problems



**Figure 6. Diode Probes for Focal In/Activation of Neurons**

(A) Schematic of a single LED-fiber assembly. The LED is coupled to a  $50\text{ }\mu\text{m}$  multimode fiber, etched to a point at the distal (brain) end. (B) Schematic of a drive equipped with a six-shank diode probe with LED-fibers mounted on each shank. Etched optical fibers are attached  $\sim 40\text{ }\mu\text{m}$  above the recordings sites on the silicon probe shanks. (C) Recording silicon probe integrated with a waveguide. Light transmission through the optical splitter waveguides integrated on the fabricated neural probe: (C and E) bright-field microscope images; (D and F) dark-field images. Light can be delivered to multiple shanks from a single fiber source via an optical splitter or different wave-length lights can be delivered to the same shank through an optical mixer. (A) and (B) are reproduced after Stark et al. (2012) and (C)–(E) after Im et al. (2011).

can be significantly reduced or eliminated by etching small-core ( $\leq 50\text{ }\mu\text{m}$ ) optical fibers to a point ( $\leq 10\text{ }\mu\text{m}$ ) and mounting them close ( $< 40\text{--}50\text{ }\mu\text{m}$ ) to the recording sites (Figure 6; Royer et al., 2010; Stark et al., 2012). Experiments have shown that less than  $1\text{ }\mu\text{W}$  of light ( $< 0.1\text{ mW/mm}^2$ ) is sufficient to activate ChR2-expressing neurons *in vivo* (Stark et al., 2012; Stark et al., 2014), and with the development of more sensitive opsins (Chuong et al., 2014), light requirements are expected to decrease. Such hybrid devices can eliminate photoelectric artifacts and reduce artificially induced overlapping spikes.

The low light intensity requirement can eliminate the need for bench top lasers and optical cables that restrain the freedom of the animal's movement. Instead, miniature light emitting diodes (LED) and/or laser diodes can be coupled to short, small-diameter ( $50\text{ }\mu\text{m}$ ) multimode fibers and attached directly to the shanks of a silicon probe or tetrode (Figure 6). The small size and weight of these integrated "diode probes" allow fast, multi-site and multicolor optogenetic manipulations in freely moving animals with concurrent monitoring of the manipulated neurons (Royer et al., 2010; Royer et al., 2012; Stark et al., 2012). To replace the labor-intensive manual process, at least two approaches are being tested. The first is the monolithic integration of all optical and electrical components on the shanks of the silicon probe (Wu et al., 2013; Zorzos et al., 2010). Various waveguide configurations can be designed with photolithography and the distances between optical stimulation sites and recording electrodes can be precisely determined (Zorzos et al., 2010; Zorzos et al., 2012; Wu et al., 2013). At a given stimulation site, different wavelengths can be selected by switching

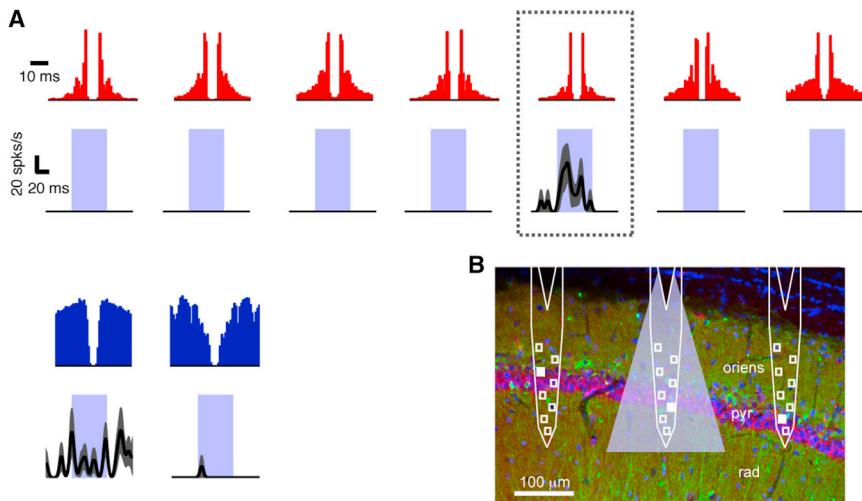
between the light sources and allowing fast and complex manipulation of neural activity by exciting and silencing of the same neurons (Stark et al., 2012). Another approach is placing an assembly of micro-light-emitting diodes ( $\mu$ LEDs) in the vicinity of the recording sites (Kim et al., 2013). Either configuration will offer unmatched spatial precision and capability of targeted perturbation and recording from specified neuron types. Dramatic reduction of light intensity combined with dense recording of the surrounding neurons allows single neuron stimulation (Figure 7) and estimation of the absolute numbers of directly driven neurons in a small illuminated volume. Being able to assess the fraction of neurons that can affect a particular physiological pattern, perception, memory or overt behavior is an important goal of neuronal circuit analysis.

#### Characterization of Neuron Subtypes

To be able to identify diverse components of circuits, an iterative refinement of a library of physiological parameters is needed so that subsequently the various neurons can be recognized reliably by using purely physiological criteria without the need for optogenetics (Kvitsiani et al., 2013; Madisen et al., 2012; Pi et al., 2013; Royer et al., 2012; Stark et al., 2013). We are only beginning to understand how different methods of light delivery impact different neuron types. A hitherto unexploited path for characterizing the physiological properties of optically tagged neurons is their input-output analysis. By analogy to neuron characterization by intracellular current injections (Ascoli et al., 2008) localized optical stimulation can be used to activate neurons and observe their characteristic response properties to pulses, sinusoid, white noise stimuli or more complex patterns. Although the photo-bleaching and light adaptation properties of the various opsins (Lin et al., 2009) are currently a potential obstacle with bright light sources, the availability of faster reacting and highly sensitive opsins (Berndt et al., 2011; Chuong et al., 2014; Gunaydin et al., 2010; Klapproetke et al., 2014; Zhang et al., 2011) will reduce such concerns. Comparison of pyramidal cells and interneurons has already demonstrated that ChR2 expressing parvalbumin interneurons follow optical responses much more efficiently than neighboring pyramidal neurons (Stark et al., 2013). Further experiments may validate the utility of such waveform characterization methods of neuron identity. Overall, combining optogenetic manipulation with high-density electrophysiological monitoring can offer high fidelity identification of circuit components, a prerequisite for a rational perturbation of identified neuron types and quantities for understanding their impact in circuit function (Roux et al., 2014; Roux and Buzsáki, 2015).

#### Studying Circuits with Correlational and Perturbation Methods

Perhaps the most critical goal of combining optogenetics with large-scale extracellular recordings is to identify the causal role of specific neuron classes in local circuit operations. Perturbation strategies can probe the involvement of particular neuron types in any defined computation, such as gain control, plasticity, oscillations, circuit stability and spike transfer modes. Closed-loop optogenetic activation or silencing of targeted neurons, combination of predetermined spike patterns in multiple cells contingent upon behavioral parameters, and/or selected features of the LFP can alter the timing of action potentials (Stark et al., 2013; Stark et al., 2012; Stark et al., 2014) and induce or suppress



**Figure 7. Optogenetic Activation of a Single Neuron in the Behaving Mouse**

Autocorrelograms and optogenetically evoked responses (light blue rectangles) of pyramidal cells (red) and putative interneurons (blue) in a freely moving CaMKII::ChR2 mouse during weak 50 ms light pulses (0.01 mW/mm<sup>2</sup> at the center of the CA1 pyramidal layer). All neurons were recorded from the middle shank of the diode-probe (inset). Note robust response of a single pyramidal cell (boxed). Ten repetitions. No neurons were activated by light on the adjacent shanks. At higher intensities (>0.05 mW/mm<sup>2</sup>) multiple other neurons also increased their firing (data not shown).

correlated firing between cells during movement, perception, learning, memory, sleep or any other aspects of brain activity. Simultaneous monitoring of neuronal responses to optogenetic perturbation is critical for uncovering the mechanisms how each neuron type contributes to circuit performance. Optogenetics combined with large-scale extracellular recordings has already proven to be effective in studying the functional roles of specific GABAergic interneuron classes in both hippocampus and neocortex, as well as other brain regions (Alonso et al., 2012; Brown et al., 2012; Piet et al., 2013; Royer et al., 2012). Strong optogenetic activation of parvalbumin-expressing interneurons at gamma frequency was shown to coordinate the timing of sensory inputs relative to a gamma cycle and enhance signal transmission (Cardin et al., 2009; Carlén et al., 2012; Sohal et al., 2009), whereas at weaker activation produced theta resonance and excess spiking in nearby pyramidal cells (Stark et al., 2013). Local stimulation of pyramidal cells induced high frequency oscillations, typical of hippocampal ripples (Stark et al., 2014). Channelrhodopsin-2-assisted circuit analysis in the amygdala identified neurons critical in gating conditioned fear (Habensak et al., 2010). Optogenetic activation of orexin neurons in the hypothalamus could change sleep choreography (de Lecea and Huerta, 2014). Periodic light stimulation of neurons of the thalamic reticular nucleus induced state-dependent neocortical spindles (Halassa et al., 2011) or evoked generalized spike and wave discharges (Berényi et al., 2012). On the other hand, tonic photoactivation of reticular neurons reduced focal seizures in the neocortex (Paz et al., 2013). Similarly, spontaneous seizures could be suppressed by optogenetic activation of parvalbumin interneurons in the hippocampus (Krook-Magnuson et al., 2013). Optogenetic activation of dentate gyrus neurons could induce false memories, making mice acting accordingly in a fear conditioning paradigm (Ramirez et al., 2013). This nonexhaustive list of experiments demonstrate the power of combined closed-loop recording-optogenetic methods for understanding complex neuronal interactions in intact and altered networks (Roux et al., 2014), and foretell how such methods could one day be harnessed for clinical applications.

neurons, the appropriate combination of which can provide single neuron, single spike monitoring and control in behaving animals, not possible by other methods. Although several aspects of the combined recording/optogenetics technique are highly scalable and one can imagine exponential growth of the technology (Alivisatos et al., 2013; Marblestone et al., 2013), our aim was to discuss techniques that can be deployed currently or in the near future for the ultradense sampling of neuronal circuits. Combination of electrical recording and optogenetics with imaging (Grienberger and Konnerth, 2012; Yuste and Katz, 1991) can further enhance the power of these methods.

Spectacular progress has been made over the past few years in signal multiplexing, moving from expensive and bulky modular amplifier-digitizer systems to affordable, high-channel count signal multiplexed systems (Berényi et al., 2014; <http://www.intantech.com/>; <http://www.open-ephys.org/>). There are no longer financial constraints even for small labs to record from a few hundred channels. It is expected that channel count will greatly increase in the coming years, paralleled with a substantial decrease in the size of signal multiplexing devices. Silicon probes with several dozens and even hundreds of sites are currently commercially available and probes with even higher channel counts in both two- and three-dimensional configurations are being tested experimentally (Du et al., 2009; Riera et al., 2012). To make these techniques fruitful for experiments on mice and other small size animals will require orders of magnitude improvement in several key parameters, particularly on-probe signal multiplexing (Olsson et al., 2005). Substantially improved power efficiency of telemetry and data loggers or their external powering may eventually eliminate the need for cable connections between the animal and recording equipment. A foreseeable challenge with brain embedded electronics is that the electromagnetic fields needed for external powering may inappropriately stimulate neurons. Miniaturization and improved stability of micromanipulators to allow long-term recordings from multiple brain regions simultaneously are equally important steps.

### Looking into the Future

Good research practice stands on the foundation of accurate measurement. Our review has focused on the large-scale recording of neuronal spike data and the optogenetic manipulation of

Obtaining high-quality, high-density data from multiple interconnected structures is only the first necessary step. The second big challenge is to develop fast automatic or semi-automatic and validated algorithms for spike clustering with common standards to facilitate standardization and within- and across-laboratory replication of neuronal identification (Buzsáki, 2004; Einevoll et al., 2012). Finally, making sense of the large-scale and properly preprocessed data requires a further concerted effort (Packer et al., 2013). Effective dissemination of the newly developed methods and algorithms and sharing as well as maintaining well-curated data with common standards and seamless user accessibility (Mizuseki et al., 2014; CRCNS.org) are among the key tasks ahead of us for advancing rapid progress.

Successful analysis of neuronal circuits requires not only large-scale recording of neurons at high speed but also their interactive control at the single-neuron, single-spike level. While optogenetics has emerged as an appropriate method for manipulating identified neurons at sufficient speed, substantial improvement is needed to achieve the desired spatial resolution at the single neuron level. Such development will require the integration of silicon recording technology with the local and programmable delivery of light, using either waveguides or neuron-size LEDs mixed with the recording sites (Kim et al., 2013; Wu et al., 2013, 2015). Neuron identity can be obtained online by optogenetics. Multicolor techniques can be deployed for activating and silencing the same neurons in the behaving animal (Stark et al., 2012) and for selective perturbation of excitatory and inhibitory neurons.

Optogenetic techniques are often cited as a solution for the long-awaited mechanistic understanding of causal effects of neurons in their native environment (Boyden et al., 2005; Packer et al., 2013; Scanziani and Häusser, 2009). However, these aspects of optogenetics are often overstated since identifying the “cause” by local perturbations in complex systems can be a daunting task. While tracking cause-effect relationships in linear systems (such as A causes B) is relatively straightforward, in complex systems with multiple re-entrant loops and emergent properties such as the brain, circuit perturbations can bring about multiple and hard-to-interpret consequences since A and B can have bidirectional or circular causal relationships. Because brain circuits are perpetually active and their components display qualitatively different interactions in different brain states, imposition of synthetic patterns has to compete with an ongoing brain-based program and can induce outcomes different from the native functions. These and other unexpected surprises of the perturbation technique (Miesenböck, 2009) emphasize the need for the focal delivery of light limited to the volume of recorded neurons and the application of control theory (Zhou et al., 1996). Despite these expected difficulties, multisite-multineuron close-loop control experiments offer unprecedented opportunities for the exploration of neuronal circuits. They can be used to induce, modify or annihilate network oscillations, test hypothesized rules of *in vivo* plasticity, examine the critical role of timing (e.g., by introducing controlled jitter of spike timing) and, perhaps most importantly, determine how synthetic spatiotemporal patterns of activity of identified neurons induce specific behaviors. This exercise amounts to understanding the neuronal syntax or brain coding.

## ACKNOWLEDGMENTS

This work was supported by National Institute of Health Grants NS34994, MH54671, NIH 1U01NS090526, 1R21EB019221, and NS074015; NSF ECCS 1407977 the Human Frontier Science Program; and the National Science Foundation (Temporal Dynamics of Learning Center Grant SBE 0542013). A.B. was supported by a Marie Curie FP7-PEOPLE-2009-IOF grant (number 254780), EU-FP7-ERC-2013-Starting grant (number 337075), and the “Lendület” program of the Hungarian Academy of Sciences. E.S. was supported by the Rothschild Foundation, the Human Frontiers in Science Project (LT-000346/2009-L), and the Machiah Foundation (20090098). A.B. is the founder and owner of Amplipex Ltd., Szeged, Hungary, which manufactures signal-multiplexed head stages and demultiplexing systems. D.R.K. is the founder and current executive director of NeuroNexus Technologies, Inc., a subsidiary of Greatbatch, Inc.

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