



The key role of oncopharmacology in therapeutic management, from common to rare cancers: A literature review

Baptiste Louveau, Fanélie Jouenne, Florentia Kaguelidou, Alexandra Landras, Lauriane Goldwirt, Samia Mourah

► To cite this version:

Baptiste Louveau, Fanélie Jouenne, Florentia Kaguelidou, Alexandra Landras, Lauriane Goldwirt, et al.. The key role of oncopharmacology in therapeutic management, from common to rare cancers: A literature review. *Thérapie*, EDP Sciences - Depuis 2016, la revue *Thérapie* n'est plus publiée par EDP Sciences.> *Thérapies* (Elsevier), 2020, 75, pp.183 - 193. 10.1016/j.therap.2020.02.010 . hal-03490262

HAL Id: hal-03490262

<https://hal.archives-ouvertes.fr/hal-03490262>

Submitted on 20 May 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial| 4.0 International License

THERAPIES special issue number 2 march april 2020

HEADING: DRUG AND ORPHAN DISEASES

The key role of oncopharmacology in therapeutic management, from common to rare cancers: a literature review

Key role of oncopharmacology in therapeutic management, from common to rare cancers

**Baptiste Louveau^{a,b}, Fanélie Jouenne^{a,b}, Florentia Kaguelidou^{c,d}, Alexandra Landras^{a,b},
Lauriane Goldwirt^{a,b}, Samia Mourah^{a,b,*}**

^a *Department of pharmacology and tumor genomics, Assistance Publique-Hôpitaux de Paris, Saint Louis hospital, 75010 Paris, France*

^b *Université de Paris, INSERM UMRS 976, 75010 Paris, France*

^c *Department of pediatric pharmacology and pharmacogenetics, Assistance Publique-Hôpitaux de Paris, Robert Debré Hospital, 75019, Paris, France*

^d *INSERM, CIC 1426, 75019, Paris, France*

Received September 16, 2019; accepted November 15, 2019

***Corresponding author.** Department of pharmacology and tumor genomics, Saint Louis Hospital AP-HP, 1 Avenue Claude Vellefaux, 75475 Paris cedex 10, France.

Tel: 0033 1 42 49 48 85

E-mail adress: samia.mourah@aphp.fr (S. Mourah)

Summary

The therapeutic management of cancers has undergone considerable changes due to the emergence of genomics tools and tumor molecular deciphering. In this context, a dual pharmacological approach based on pharmacogenomic analyses and therapeutic drug monitoring is now part of the routine care in cancer management for personalized therapies. First, molecular and immune profiling of tumors allows the emergence of new pharmacological targets in common as well as in rare cancers. Second, pharmacogenomic analyses coupled to therapeutic drug monitoring guide the prescription by adjusting regimen and managing drug resistance.

KEYWORDS

Oncopharmacology; Pharmacogenomics; Therapeutic drug monitoring; Targeted therapies; Immunotherapies; Resistance

Abbreviations

ALCL: anaplastic large-cell lymphoma

ALL: acute lymphoblastic leukemia

AML: acute myeloid leukemia

AEs: adverse events

AUC: area under the curve

BRAFi: BRAF inhibitor CAR: chimeric antigen receptor

CML: chronic myeloid leukemia

CTLA4: cytotoxic T-lymphocyte-associated antigen 4

ECD: Erdheim–Chester disease

EGFR: epidermal growth factor receptor

HL: Hodgkin lymphoma

ICIs: immune checkpoint inhibitors

IFN- α : interferon alfa

LCH: Langerhans cell histiocytosis

MAPK: mitogen-activated protein kinase

MEKi: MEK inhibitor

NHLs: non Hodgkin lymphoma

NSCLC: non-small cell lung cancer

PD-1: programmed cell death I

PD-L1: programmed cell death ligand I

PI3K/AKT: phosphoinositide 3-kinase/ protein kinase B

RTK: receptor tyrosine kinases

SCLC: small cell lung cancer

TDM: therapeutic drug monitoring

TKI: tyrosine kinase inhibitor

Introduction

Recent advances in cancer patients have required a new multidisciplinary pharmacological approach in terms of treatment choice, optimization and post marketing authorization surveillance [1]. Genomic tools have considerably changed the treatment of cancers these last years. Based on the characterization of molecular alterations and tumor immune features, targeted therapies and then immunotherapies have greatly improved the clinical course of patients and enlarged the role of pharmacology in cancer therapeutic management. Thus, a dual pharmacological approach relying on therapeutic drug monitoring and pharmacogenomic analyses is now part of the routine care. Indeed, identifying tumor molecular alterations is a key point to guide therapeutic strategies and provide new insights regarding the understanding of drug resistance and the development of response biomarkers.

In this review, we present the contribution of oncoparmacology in patient's management, from common to rare tumor types.

From tumor molecular alterations to pharmacological targets

Genomic and transcriptomic approaches allowed a molecular deciphering of tumors and uncovered alterations in signaling pathways underlining tumor pathogenesis processes. This leads to a better molecular characterization of a wide range of cancers, from common to rare, paving the way for innovative targeted therapies development.

Lung cancer

Lung cancer is a major cause of mortality and the most commonly diagnosed cancer worldwide with 11.6% of the total cases [2]. Comprehensive molecular profiling, notably from the Cancer Genome Atlas research network identified alterations frequently retrieved in the different histological lung subtypes [3].

In adenocarcinoma, the most frequent form of non-small cell lung cancer (NSCLC), the RTK/RAS/RAF signaling pathway is altered in 75% of cases. The major factors leading to this activation are *NF1*, *KRAS*, *EGFR* and *BRAF* mutations with *EGFR* being the most common driver mutation described in approximately 15% of cases [4]. Based on this molecular characterization,

tyrosine kinase inhibitors (TKIs) targeting EGFR such as erlotinib and gefitinib have greatly improve clinical outcome with an overall response rate of 75% in patients with *EGFR* sensitizing mutations (5). Following the same logic, a combination of dabrafenib (BRAF inhibitor, BRAFi) and trametinib (MEK inhibitor, MEKi) has been approved and showed good clinical benefit in *BRAFV600* mutated (*BRAFV600^{mut}*) patients. Besides the mitogen-activated protein kinase (MAPK) pathway, rearrangements of the *ALK* gene were also uncovered in 3-5% of NSCLC patients prompting to develop successive generations of ALK kinase inhibitors (crizotinib, ceritinib, osimertinib...), which reached an overall response rate of 50 to 60% [6]. Finally, *ROS1* rearrangements uncovered in 2% of patients constitute a good pharmacological target [7,8].

Beyond targeting the driver mutations, efforts regarding the characterization of the tumoral immunologic features and microenvironment have allowed NSCLC to benefit from immunotherapy [9]. Immune checkpoints inhibitors (ICIs) targeting the PD1/PD-L1 interaction (nivolumab, pembrolizumab, durvalumab) or the CTLA4 receptor (ipilimumab) have thus improved the survival compared to molecular targeted therapies and provide innovative therapeutic options for patients not harboring previous molecular alterations [10].

Squamous cell carcinomas which represent approximately 30% of lung cancers are defined by multiple and complex genomic mutations in several pathways, including actionable oncogenic alterations in *FGFR1*, *PIK3CA*, *DDR2*, *MET* and *BRAF* genes [11] constituting opportunities for molecular targeted therapies. Although clinical trials targeting these pathways have failed, ICIs have led to a meaningful increase in response rate and survival [12].

Small cell lung cancer (SCLC) accounts for approximately 15% of lung cancers and is characterized by rapid disease progression. Facing a lack in early detection and a limited amount of tumor tissue for translational research, few is known about its molecular characterization and no targeted therapies are approved for its treatment [13,14]. Currently, mRNA profiling distinguishes two groups according to expression of *CHGA*, *GRP*, *ASCL1* and *DLK1* and co-inactivation of *TP53* and *RBI* are frequent [15]. Besides, several trials evaluating ICIs are ongoing.

Melanoma

Melanoma accounts for approximately 10% of cutaneous cancers and presents a very bad prognosis when diagnosed at an advanced or metastatic stage [16]. Until 2010, cytotoxic chemotherapies such as dacarbazine were the only available treatments for metastatic forms and displayed a poor clinical benefit. Applying whole exome sequencing revealed that melanoma tumors harbor a high rate of

genomic alterations [17] and led to the exhaustive identification of significantly recurrently altered genes. It allowed to establish a molecular classification defining 4 subtypes harboring the following genotypes: (i) *BRAF* kinase mutations; (ii) *NRAS* small G protein mutations; (iii) *NFI* mutations and (iv) triple wild-type, (heterogeneous group with no *BRAF*, *RAS* or *NFI* mutations but alterations of *GNAQ*, *GNA11*, *KIT* or *CTNNB1* can be retrieved) [18].

Regarding therapeutic management, two major altered signaling pathways were highlighted raising high hopes for the use of molecular therapies: (i) the MAPK pathway activated in more than 75% of melanomas due to *BRAF* (50% of melanomas), *NRAS* or *cKIT* somatic mutations; (ii) The PI3K/AKT pathway with alterations on genes such as *PTEN* or *AKT* [19–21].

Two BRAFi, vemurafenib and dabrafenib, were thus approved to target the constitutive activation of the MAPK pathway successive to *BRAFV600* mutations. Firstly used as a monotherapy, these drugs improve survival in metastatic melanoma patients and a clinical response was observed in 50% of patients [22,23]. Facing multiple treatment escape, BRAFi are now used in combination with MEKi (cobimetinib, trametinib) which is the standard of care in *BRAFV600^{mut}* metastatic melanoma patients [24,25]. This clinical benefit achieved with BRAFi+MEKi compared to BRAFi alone points out the interest of targeting an altered pathway at different levels to delay escape to therapy. In that extent, trials evaluating the clinical benefit of combining PI3K inhibitors with MAPK inhibitors (MAPKi) are ongoing with promising efficacy data albeit restrained by severe adverse events (AEs) such as stomatitis, creatine kinase increase or cutaneous rash [26,27]. Moreover, alterations of the apoptosis or cell cycle pathways were identified in melanoma, supporting the evaluation of new targeted approach such as CDK4/6 inhibitors [28].

As described in lung cancers subtypes, the characterization of the immune tumoral features has led to the approval of ICIs in metastatic melanoma. Anti-PD1/PD-L1 and anti-CTLA4 immunotherapies thus provide long-term clinical response and are now used as first or second line therapy (after BRAFi+MEKi) in metastatic melanoma patients according to the existence of *BRAFV600* mutation [29–32].

Hematological malignancies

Hematological malignancies concern both adults and children. Tumor mutational profiling of hematological malignancies is significantly different with multiple genomic alterations in elderly patients whereas few targetable mutations have been identified in pediatric patients so far, restraining the development of targeted therapies in this subpopulation [33]. In addition, emerging

evidence shows that immunotherapies are effective against chemo resistant cancer cells when administered alone or in combination with chemotherapy [34].

In chronic myeloid leukemia (CML), TKIs such as imatinib, dasatinib and nilotinib are used and target the oncogenic pathway of Philadelphia (Ph) chromosome (translocation t(9,12)), the main genetic alteration in CML, which results in the BCR-ABL fusion gene. TKIs decrease the induction of proliferation through the ABL-activation [35]. In B-cell acute lymphoblastic leukemia (ALL), blinatumomab, a bi-specific monoclonal antibody mediating the interaction between CD19+ on leukemic blasts and CD3+ on T-cells, is now marketed for the treatment of Ph- recurrent or refractory patients [36]. The induction treatment for children with ALL is a combinatory chemotherapy and the addition of a TKIs, such as imatinib, may be beneficial for Ph+ ALL patients [36]. In acute myeloid leukemia (AML), 85% of children go into remission after chemotherapy induction. However, for relapsing or refractory cases, a promising option besides stem cell transplant or a second cycle of chemotherapy, is the off-label use of gemtuzumab ozogamicin, a monoclonal antibody targeting CD33 [37].

Currently, treatment of Hodgkin lymphoma (HL) is mainly based on the use of chemotherapy and radiation therapy. However, immunotherapy is increasingly used [34,36] and brentuximab vedotin, a conjugate between an anti-CD30 antibody and a cytotoxic agent, is currently approved for use in adults only. ICIs and notably anti-PD1 immunotherapy are labeled for use in adults with refractory or relapsed HL even after stem cell transplant and/or use of brentuximab vedotin. Clinical trials in childhood HL are ongoing.

Treatment of non-Hodgkin lymphomas (NHLs) is mainly based on multi-agent chemotherapy combined or not with local treatments such as surgery and radiation. In patients with CD20+ diffuse large B-cell NHL, rituximab, an anti-CD20 antibody, is indicated in combination with chemotherapies [38].

Treatment with brentuximab vedotin has also been approved to treat relapsed or refractory systemic anaplastic large-cell lymphoma (ALCL) in adults and trials are ongoing in children [39]. Also, the use of ALK inhibitors (crizotinib, ceritinib), alone or in combination with CD4 inhibitors may be more effective in treating children with relapsed or refractory ALCL [40].

Adaptive T-cell therapy has been a breakthrough for the treatment of hematological cancers, especially the development of chimeric antigen receptor (CARs) transgenic T-cells targeting CD19. Use of this therapy in patients with refractory disease to standard chemotherapy has resulted in approximately 66% complete response rate despite short-term toxicity [34]. In children, recent trials demonstrate a 81% overall remission rate within 3 months and no minimal residual disease in responders [41]. Chimeric antigen receptors (CARs) are engineered receptors

that graft a defined specificity onto an immune cell, typically a T-cell, and enhance T-cells function [42]. CARs consist of a T-cell activating domain and an extracellular immunoglobulin-derived heavy and light chains to direct specificity, and second generation CARs also include co-stimulatory domains [43]. Their activation leads to T-cell proliferation, cytokines secretion, and cytotoxicity. Transgenic CAR T-cells such as CTL019-tisagenlecleucel target the antigen CD19 which can be found on the cell surface of most B-cell derived ALL. Clinical trials evaluating the use of these CAR T-cells have recently showed substantially improved outcomes in patients with refractory B-cell cancers, including NHL, CML and ALL [41,44,45]. Nevertheless, some challenges are inherent to this treatment like short-term AEs, the low levels of long-run persistence of CAR T-cells or the risk of an immune escape [42].

Histiocytic neoplasms

Histiocytic neoplasms are a heterogeneous group of rare clonal hematopoietic diseases with a therapeutic management greatly improved by the genomic approach. For instance, recurrent molecular alterations have been identified in Langerhans cell histiocytosis (LCH) and Erdheim–Chester disease (ECD).

LCH is a rare disease with varying clinical presentations, from localized to severe multivisceral forms [46]. The pulmonary form of the adult occurs selectively in young smokers [47] and tobacco smoking is the only risk factor clearly identified to date [47]. LCH treatment is highly dependent from the clinical presentation [48,49] and surgery, radiation therapy, or chemotherapies have been historically used [49]. In adults, the association vinblastine/prednisone as well as cytarabine or cladribine are usually prescribed [48,50]. In pulmonary LCH, although cladribine may improve lung function [51], vinblastine is ineffective and there is currently no effective treatment [52]. Moreover, these chemotherapies are highly toxic with variable response rate requiring the development of novel therapeutic strategies.

In 2010, the tumoral aspect of the LCH was confirmed and *BRAFV600E* oncogene mutation was identified in approximately half of LCH tissue lesions [53,54]. Recently, wide exome sequencing of LCH lesions revealed other gene alterations in the MAPK pathway, including *BRAF* deletions [55], *MAP2K1* mutations/deletions in about 25% of cases [56–58], as well as *NRAS* [54], *ARAF* [59], *MAP3K1* [58], and *KRAS* mutations [60,61]. These findings highlight that a somatic activating mutation in the MAPK pathway is carried by most LCH patients and support the clinical use of MAPKi. In LCH adults harboring a *BRAFV600* mutation, an overall response rate of 43%

and no disease progression were observed under vemurafenib [62–64]. In children, clinical trials evaluating the safety, the tolerability and the pharmacokinetics of BRAFi are ongoing. MEKi have shown good results *in vitro* [61] and trametinib was effective and well-tolerated in a patient with progressive pulmonary LCH [65]. Overall, MEKi is a relevant therapeutic option as it showed good response in patients with histiocytic neoplasm harboring mutations in *RAF*, *RAS* and *MEK* genes [66]. Moreover, TKIs have emerged as a relevant pharmacological approach and studies have shown the effectiveness of imatinib in the treatment of LCH patients [67,68]. In an open-label multicenter trial, the pan-AKT inhibitor afuresertib provided a clinical stabilization in LCH patients [69]. Despite not targetable, other altered genes such as *ASXL1*, *DNMT3A*, *IDH1*, *TET2* [61,70], *PTPN11* [71] *TP53* and *MET* [53] have also been identified.

Similarly, ECD is a rare non-Langerhans cell histiocytosis (approximately 500 patients diagnosed worldwide since 1930) and no therapeutic trials have been performed so far. Interferon alfa (IFN- α), has long been the first-line therapy [72,73], but the occurrence of severe AEs and the development of secondary resistance to IFN- α high doses [74,75] motivated the investigation of alternative pharmacological strategies. Second-line treatments such as anakinra, cladribine, TKIs and infliximab have been proposed [76] but the small numbers of patients did not allow to define the optimal strategy. Since more than 50% of patients harbor a *BRAFV600E* mutation [77,78], MAPKi may be a relevant therapeutic option and positive outcomes were reported in refractory *BRAFV600E* patients treated with vemurafenib [62]. Other cases reported the efficacy of BRAFi but also MEKi therapy [79,80].

Pharmacogenomic in the management of cancers

Tumor molecular genotyping as part of routine care in France

Drug development and approval for molecularly stratified tumor subgroups have rendered molecular testing mandatory and require that molecular analyses be performed nationwide. To this end, the French National Cancer Institute (INCa) and the French Ministry of Health have set up since 2007, a national network of 28 regional molecular genetics centers working in a multidisciplinary collaboration, involving clinicians, pathologists and molecular biologists. Selective molecular tests are performed in these facilities. They are free of charge for all patients in their region, irrespective of the type of establishment in which they are receiving treatment. An evolutive catalogue of validated molecular predictive biomarkers performed in the INCa platforms

and guiding the prescription of targeted therapies is available. Thus, the genotyping of tumors has become a theranostic tool, which conditions the prescription of targeted therapies. For instance, *EGFR* genotype status will determine the prescription of anti-EGFR targeted therapies in NSCLC and the presence of *BCR-ABL* fusion transcript will condition the use of imatinib, dasatinib or nilotinib in CML. In metastatic melanoma, *BRAF* genotyping is performed for all patients who may benefit from BRAFi. In acral lentiginous and mucosal melanomas c-KIT genotyping will guide a potential treatment with TKIs.

A specific program has also been implemented by INCa to anticipate the launch of new targeted therapies and to accelerate the time-to-access to new drugs and experimental therapies. This initiative has been operational for few years now and has been successful in meeting its initial aims of uniform nationwide test provision and fast implementation of molecular tests for new tumor biomarkers.

Besides these initiatives, INCa implements recommendations regarding molecular tests prescription, genotyping, results and activity reporting.

Management of resistance to therapies

Resistance mechanisms

Multiple resistance mechanisms have emerged and have restrained the clinical benefit of targeted therapies and immunotherapies. In that extent, pharmacogenomic analyses at baseline and under treatment are a key point in therapeutic management allowing the understanding of preexisting and acquired genomic alterations involved in resistance.

Resistance to targeted therapies was firstly documented for imatinib in CML based on mutations in the targeted kinases [81]. In NSCLC, a similar direct oncogene reactivation, due to the activating *EGFR*T790M mutation, was involved in resistance to first-generation TKIs. Successive generations of EGFR inhibitors were hence developed [82]. In metastatic melanoma, multiple resistance mechanisms can bypass BRAF inhibition and acquired resistance occurs in 50% of patients [22,23]. A MAPK pathway reactivation is retrieved in more than 75% of cases [83–87]. Resistance may also be driven by alterations in other oncogenic pathways promoting cell survival and proliferation as well. In NSCLC, resistance to gefitinib through amplification of an alternative

tyrosine kinase receptors such as *MET* was highlighted [88]. In melanoma, the PI3K/AKT pathway may thus be activated through *PTEN* and *AKT* mutations and alterations of cell cycle genes have been associated to resistance [89,90]. Recent studies revealed baseline genomic features predictive of clinical response under MAPKi. For instance, in melanoma, Wongchenko et al. [91] and Yan et al. [92] highlighted improved clinical response in patients carrying baseline *NF1* alterations and higher expression of immune response-related genes. A MAPK pathway activity score predictive of vemurafenib response has been proposed as well [93]. In NSCLC, sensitizing *EGFR* mutations, and *ALK* or *ROS1* rearrangements are known predictive biomarkers of response to targeted therapies and several gene alterations (*KRAS*, *MET*) are candidates currently studied [94].

Deciphering the efficacy of MAPKi newly prescribed in histiocytic neoplasms, data on resistance mechanisms in this rare disease are beginning to emerge. Recently, a novel *MAP2K1* mutation was related to resistance in a patient treated with trametinib [95]. Similarly, *RASA1* loss was highlighted as a mechanism of resistance to BRAFi treatment which was overcome by adding a MEKi [96].

Resistance to ICIs is reported in 50% of treated patients. As pharmacological activity of immunotherapies relies on T-cells, dysfunction of tumor-specific T-cells drives resistance [97]. *JAK1/JAK2* mutations or alterations of the β -catenin pathway may thus decrease T-cells activity and lead to resistance [98]. Moreover, Hugo et al. identified genes overexpressed in resistance cases and proposed a transcriptomic innate anti-PD1 resistance signature [99]. Other resistance mechanisms described to date include expression of alternative checkpoints (*LAG-3*, *TIM-3*...) or down-regulation of major histocompatibility complex I [100,101].

A higher TMB was significantly associated with favorable outcome in those patients [102]. Regarding anti-PD1/PD-L1 immunotherapies, PD-L1 expression was also depicted as a promising biomarker of response to anti-PD1 therapy but faced important limitations [103].

Identifying non-invasive biomarkers has raised a lot of interest these last few years and circulating tumor DNA (ctDNA) has been increasingly studied in the field of both molecular therapies and immunotherapies. Many recent data suggest that undetectable or low level of ctDNA at therapy initiation are predictive of increased clinical benefit [104,105].

Overcoming resistance

Many efforts are undertaken to overcome resistance, and several trials are ongoing to evaluate the benefit of combining immunotherapies and targeted therapies to overcome the loss of clinical

benefit. In melanoma, translational data have shown that MAPKi may improve the efficacy of immunotherapy by impacting the tumoral microenvironment. This approach is of great interest as it would allow to synergize the high response rate observed under MAPKi and the long-term clinical benefit provided by ICIs [106,107]. Preliminary results from the phase II TRIDeNT and phase III COMBI-I trials (evaluating respectively dabrafenib+trametinib+nivolumab and dabrafenib+trametinib +spartalizumab) revealed improved response rate [108,109]. Combination of MEKi and anti-PD-L1 is also currently under evaluation in *BRAF* wild-type advanced melanoma but shows less conclusive efficacy.

In NSCLC, the same strategy is experimented with clinical trials testing the combined administration of TKIs and ICIs. For instance, the TATTON phase I trial evaluated the combination of osimertinib and durvalumab. Although efficacy results seemed promising, major AEs occurred [110]. Overall, these data emphasize the therapeutic potential of combining immunotherapies and targeted therapies. Nevertheless, as the effect of targeted therapies on the microenvironment is time-dependent, further studies must be addressed to establish the optimal timing and dosing in this context of combinatory treatments.

Therapeutic drug monitoring in the management of cancers

Besides the genomic approach inherent to these innovative therapies, cancer patient's management requires a careful therapeutic monitoring. Oral administration of molecular targeted therapies has improved the patient's quality of life and treatment efficacy but has also raised concerns regarding fixed doses regimen, drug adherence, and inter-individual pharmacokinetics variability.

Therapeutic drug monitoring (TDM) relies on the quantification and interpretation of drug concentrations in biological fluids considering their metabolic profile, their large inter-individual variability, and the efficacy or AEs exposure relationship. The objectives are to minimize underdosing, prevent from drug resistance, determine the etiology of AEs and monitor dose reductions or interactions in high-risks patients. However, different levels of evidence are retrieved according to the cancer type: it is strongly recommended for common cancers such as blood cancer, recommended in lung cancer or melanoma and remains to be evaluated for rare cancers due to the few pharmacokinetic-pharmacodynamic studies.

General pharmacokinetic characteristics

The main pharmacokinetic characteristics of targeted therapies are summarized in Table 1. The absorption phase of these oral treatments is generally rapid with a plasma peak obtained in 3-6 hours. Food intake significantly increases the bioavailability of lapatinib, nilotinib, bosutinib and vemurafenib, slightly increases the bioavailability of erlotinib and significantly decreases the bioavailability of crizotinib, dabrafenib, dasatinib and sorafenib. Diet has no significant effect on the bioavailability of cobimetinib, and ponatinib. These drugs are widely distributed in tissues and are highly bound to proteins. They are substrates and modulators of efflux transporters (including P-glycoprotein and BCRP) and can therefore interact with the drugs substrates [111]. Moreover, their metabolism is essentially hepatic and mainly involves CYP3A4 cytochrome, which is a source of drug and food interactions. All TKIs are mainly excreted in the stool with a minor fraction excreted in the urine. Renal function therefore has little influence on their pharmacokinetics.

Relationship between exposure and efficacy or adverse events

Lung cancer

Despite no association with response was highlighted [112], higher trough concentrations of erlotinib are significantly associated with an improved progression-free survival and overall survival [113–115]. In addition, cutaneous toxicity of erlotinib was correlated with plasma exposure [115].

Melanoma

A study conducted in 27 melanoma patients indicated that patients with sorafenib plasma exposure (AUC_{ss}) greater than 100 µg.h/mL had a better tumor response and greater progression-free survival [116]. Patients with severe AEs (grade 3-4) had a higher plasma exposure (61.9 versus 53 µg.h/mL) [117].

A relationship between tumor progression and vemurafenib plasma exposure was also described with a suggested steady-state pharmacokinetic target of above 42 µg/mL [118–120]. Kramkimel *et al.* also showed a correlation between the increase in vemurafenib plasma concentration and the occurrence of skin AEs of grade >2 in the first 3 months of treatment [120].

Similarly, a study conducted in 27 melanoma patients showed a growing risk of AEs occurrence with dabrafenib trough plasma concentrations [121].

Hematological malignancies

Results from studies in CML patients showed that imatinib trough concentration were significantly higher in patients with a major molecular response and have suggested an imatinib minimum concentration of 1000 ng/mL to achieve a good molecular response [122–124].

A further study conducted in 542 CML patients treated with nilotinib suggested that obtaining the major molecular response after 12 months of follow-up was significantly associated with trough concentration. A correlation between increased plasma exposure (AUC) and corrected QT interval elongation as well as bilirubin elevation was highlighted [125].

Moreover, additional data in CML patients treated with nilotinib uncovered a significant shorter time to reach a major molecular response with residual concentrations >500 ng/mL. This study also correlated plasma exposure with the occurrence of known AEs [126].

Regarding dasatinib, Wang *et al.* showed that the major cytogenetic response was significantly correlated with the residual concentration. Low concentrations were associated with a decreased risk of pleural effusion [127,128].

The use of bosutinib trough concentrations to predict treatment efficacy and occurrence of AEs was raised in a study by Hsyu *et al.* The probability of achieving a complete cytogenetic response at 1 year, a major molecular response and a complete hematologic response increased with bosutinib trough concentrations [129].

Taken together, these data demonstrate the clinical interest of TDM as a complement to genomic analyses in oral molecular targeted therapy management in order to ensure treatment efficacy and safety. Few data are currently available in the field of immunotherapy and future pharmacokinetic-pharmacodynamic evidences would be of great interest in therapeutic management and notably in prediction of severe AEs. In addition, studies conducted on rare cancers such as LCH are missing to date and will be of major importance to improve patient's follow-up.

Conclusion

Tumor molecular deciphering has paved the way for targeted therapy development positioning the pharmacology contribution at the forefront of cancer patients' management. Pharmacogenomics is now complementary to TDM, which provides crucial data for the patient follow-up. As improving therapeutic strategy stratification and overcoming therapeutic resistance will be of major concern in the future, pharmacogenomic analyses will certainly play a pivotal role in treating both common and rare tumor types.

Disclosure of interest

Authors have no competing interest to declare.

References

- [1] Conte C, Vaysse C, Bosco P, Noize P, Fourrier-Reglat A, Despas F, et al. The value of a health insurance database to conduct pharmacoepidemiological studies in oncology. *Therapies* 2019;74(2):279-88.
- [2] Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries - Bray - 2018 - CA: A Cancer Journal for Clinicians - Wiley Online Library. <https://onlinelibrary-wiley-com.gate2.inist.fr/doi/full/10.3322/caac.21492>. [Accessed December 27, 2019].
- [3] Cancer Genome Atlas Research Network, Weinstein JN, Collisson EA, Mills GB, Shaw KRM, Ozenberger BA, et al. The Cancer Genome Atlas Pan-Cancer analysis project. *Nat Genet* 2013;45(10):1113-20.
- [4] Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature* 2014;511(7511):543-50.
- [5] Shepherd FA, Rodrigues Pereira J, Ciuleanu T, Tan EH, Hirsh V, Thongprasert S, et al. Erlotinib in previously treated non-small-cell lung cancer. *N Engl J Med* 2005;353(2):123-32.
- [6] Shaw AT, Engelman JA. ALK in lung cancer: past, present, and future. *J Clin Oncol Off J Am Soc Clin Oncol*. 10 mars 2013;31(8):1105-11.

- [7] Shaw AT, Ou SH, Bang YJ, Camidge DR, Solomon BJ, Salgia R, et al. Crizotinib in ROS1-rearranged non-small-cell lung cancer. *N Engl J Med* 2014;371(21):1963-71.
- [8] Lin JJ, Shaw AT. Recent advances in targeting ROS1 in lung cancer. *J Thorac Oncol* 2017;12(11):1611-25.
- [9] Beatty GL, Gladney WL. Immune escape mechanisms as a guide for cancer immunotherapy. *Clin Cancer Res* 2015;21(4):687-92.
- [10] Ellis PM, Vella ET, Ung YC. Immune checkpoint inhibitors for patients with advanced non-small-cell lung cancer: a systematic review. *Clin Lung Cancer* 2017;18(5):444-459.e1.
- [11] Cancer Genome Atlas Research Network. Comprehensive genomic characterization of squamous cell lung cancers. *Nature* 2012;489(7417):519-25.
- [12] Paik PK, Pillai RN, Lathan CS, Velasco SA, Papadimitrakopoulou V. New treatment options in advanced squamous cell lung cancer. *Am Soc Clin Oncol Educ Book*. 2019 Jan;39:e198-e206
- [13] Semanova EA, Nagel R, Berns A. Origins, genetic landscape, and emerging therapies of small cell lung cancer. *Genes Dev* 15 juill 2015;29(14):1447-62.
- [14] Byers LA, Rudin CM. Small cell lung cancer: where do we go from here? *Cancer* 2015;121(5):664-72.
- [15] Inamura K. Lung cancer: understanding its molecular pathology and the 2015 WHO classification. *Front Oncol* 2017;7:193.
- [16] Institut national du cancer. Les cancers en France. 2016. <http://www.e-cancer.fr/Actualites-et-evenements/Actualites/Les-cancers-en-France-2016-une-edition-100-interactive>. [Accessed December 27, 2019].
- [17] Reddy BY, Miller DM, Tsao H. Somatic driver mutations in melanoma. *Cancer* 2017;123(S11):2104-17.

- [18] Cancer Genome Atlas Network. Genomic classification of cutaneous melanoma. *Cell* 2015;161(7):1681-96.
- [19] Reddy BY, Miller DM, Tsao H. Somatic driver mutations in melanoma. *Cancer* 2017;123(S11):2104-17.
- [20] Lim SY, Menzies AM, Rizos H. Mechanisms and strategies to overcome resistance to molecularly targeted therapy for melanoma. *Cancer* 2017;123(S11):2118-29.
- [21] Hodis E, Watson IR, Kryukov GV, Arold ST, Imielinski M, Theurillat JP, et al. A landscape of driver mutations in melanoma. *Cell* 2012;150(2):251-63.
- [22] Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 2011;364(26):2507-16.
- [23] Hauschild A, Grob JJ, Demidov LV, Jouary T, Gutzmer R, Millward M, et al. Dabrafenib in BRAF-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial. *Lancet* 2012;380(9839):358-65.
- [24] Ascierto PA, McArthur GA, Dréno B, Atkinson V, Liskay G, Di Giacomo AM, et al. Cobimetinib combined with vemurafenib in advanced BRAF(V600)-mutant melanoma (coBRIM): updated efficacy results from a randomised, double-blind, phase 3 trial. *Lancet Oncol* 2016;17(9):1248-60.
- [25] Long GV, Stroyakovskiy D, Gogas H, Levchenko E, de Braud F, Larkin J, et al. Combined BRAF and MEK inhibition versus BRAF inhibition alone in melanoma. *N Engl J Med* 2014;371(20):1877-88.
- [26] Bedard PL, Tabernero J, Janku F, Wainberg ZA, Paz-Ares L, Vansteenkiste J, et al. A phase Ib dose-escalation study of the oral pan-PI3K inhibitor buparlisib (BKM120) in combination with the oral MEK1/2 inhibitor trametinib (GSK1120212) in patients with selected advanced solid tumors. *Clin Cancer Res* 2015;21(4):730-8.

- [27] Grilley-Olson JE, Bedard PL, Fasolo A, Cornfeld M, Cartee L, Razak ARA, et al. A phase Ib dose-escalation study of the MEK inhibitor trametinib in combination with the PI3K/mTOR inhibitor GSK2126458 in patients with advanced solid tumors. *Invest New Drugs* 2016;34(6):740-9.
- [28] Hamilton E, Infante JR. Targeting CDK4/6 in patients with cancer. *Cancer Treat Rev* 2016;45:129-38.
- [29] Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. *N Engl J Med* 2011;364(26):2517-26.
- [30] Robert C, Long GV, Brady B, Dutriaux C, Maio M, Mortier L, et al. Nivolumab in previously untreated melanoma without BRAF mutation. *N Engl J Med* 2015;372(4):320-30.
- [31] Larkin J, Hodi FS, Wolchok JD. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N Engl J Med* 2015;373(13):1270-1.
- [32] Wolchok JD, Chiarion-Sileni V, Gonzalez R, Rutkowski P, Grob JJ, Cowey CL, et al. Overall survival with combined nivolumab and ipilimumab in advanced melanoma. *N Engl J Med* 2017;377(14):1345-56.
- [33] Gröbner SN, Worst BC, Weischenfeldt J, Buchhalter I, Kleinheinz K, Rudneva VA, et al. The landscape of genomic alterations across childhood cancers. *Nature* 2018 15;555(7696):321-7.
- [34] Mackall CL, Merchant MS, Fry TJ. Immune-based therapies for childhood cancer. *Nat Rev Clin Oncol* 2014;11(12):693-703.
- [35] Holyoake TL, Vetrie D. The chronic myeloid leukemia stem cell: stemming the tide of persistence. *Blood* 2017;129(12):1595-606.
- [36] Burdach SEG, Westhoff MA, Steinhäuser MF, Debatin KM. Precision medicine in pediatric oncology. *Mol Cell Pediatr* 2018;5(1):6.

- [37] Wayne AS, Fitzgerald DJ, Kreitman RJ, Pastan I. Immunotoxins for leukemia. *Blood* 2014;123(16):2470-7.
- [38] Samochatova EV, Maschan AA, Shelikhova LN, Myakova NV, Belogurova MB, Khlebnikova OP, et al. Therapy of advanced-stage mature B-cell lymphoma and leukemia in children and adolescents with rituximab and reduced intensity induction chemotherapy (B-NHL 2004M protocol): the results of a multicenter study. *J Pediatr Hematol Oncol* 2014;36(5):395-401.
- [39] Palanca-Wessels MC, Press OW. Advances in the treatment of hematologic malignancies using immunoconjugates. *Blood* 2014;123(15):2293-301.
- [40] Mossé YP, Lim MS, Voss SD, Wilner K, Ruffner K, Laliberte J, et al. Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: a Children's Oncology Group phase 1 consortium study. *Lancet Oncol* 2013;14(6):472-80.
- [41] Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H, et al. Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N Engl J Med* 2018;378(5):439-48.
- [42] June CH, Sadelain M. Chimeric antigen receptor therapy. *N Engl J Med* 2018;379(1):64-73.
- [43] Vairy S, Garcia JL, Teira P, Bittencourt H. CTL019 (tisagenlecleucel): CAR-T therapy for relapsed and refractory B-cell acute lymphoblastic leukemia. *Drug Des Devel Ther* 2018;12:3885-98.
- [44] Brudno JN, Kochenderfer JN. Chimeric antigen receptor T-cell therapies for lymphoma. *Nat Rev Clin Oncol* 2018;15(1):31-46.
- [45] Park JH, Rivière I, Gonen M, Wang X, Sénéchal B, Curran KJ, et al. Long-term follow-up of CD19 CAR therapy in acute lymphoblastic leukemia. *N Engl J Med* 2018;378(5):449-59.

- [46] Guyot-Goubin A, Donadieu J, Barkaoui M, Bellec S, Thomas C, Clavel J. Descriptive epidemiology of childhood Langerhans cell histiocytosis in France, 2000–2004. *Pediatr Blood Cancer* 2008;51(1):71-5.
- [47] Vassallo R, Harari S, Tazi A. Current understanding and management of pulmonary Langerhans cell histiocytosis. *Thorax* 2017;72(10):937-45.
- [48] Girschikofsky M, Arico M, Castillo D, Chu A, Doberauer C, Fichter J, et al. Management of adult patients with Langerhans cell histiocytosis: recommendations from an expert panel on behalf of Euro-Histio-Net. *Orphanet J Rare Dis* 2013;8:72.
- [49] Haupt R, Minkov M, Astigarraga I, Schäfer E, Nanduri V, Jubran R, et al. Langerhans cell histiocytosis (LCH): guidelines for diagnosis, clinical work-up, and treatment for patients till the age of 18 years. *Pediatr Blood Cancer* 2013;60(2):175-84.
- [50] Grobost V, Khouatra C, Lazor R, Cordier JF, Cottin V. Effectiveness of cladribine therapy in patients with pulmonary Langerhans cell histiocytosis. *Orphanet J Rare Dis* 2014;9. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4268858/>. [Accessed December 27, 2019].
- [51] Lorillon G, Bergeron A, Detournignies L, Jouneau S, Wallaert B, Frija J, et al. Cladribine is effective against cystic pulmonary Langerhans cell histiocytosis. *Am J Respir Crit Care Med* 2012;186(9):930-2.
- [52] Tazi A, Lorillon G, Haroche J, Neel A, Dominique S, Aouba A, et al. Vinblastine chemotherapy in adult patients with langerhans cell histiocytosis: a multicenter retrospective study. *Orphanet J Rare Dis* 2017; 12: 95. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5441059/>. [Accessed December 27, 2019].
- [53] Badalian-Very G, Vergilio JA, Degar BA, MacConaill LE, Brandner B, Calicchio ML, et al. Recurrent BRAF mutations in Langerhans cell histiocytosis. *Blood* 2010;116(11):1919-23.
- [54] Mourah S, How-Kit A, Meignin V, Gossot D, Lorillon G, Bugnet E, et al. Recurrent NRAS mutations in pulmonary Langerhans cell histiocytosis. *Eur Respir J* 2016;47(6):1785-96.

- [55] Chakraborty R, Burke TM, Hampton OA, Zinn DJ, Lim KPH, Abhyankar H, et al. Alternative genetic mechanisms of BRAF activation in Langerhans cell histiocytosis. *Blood* 2016;128(21):2533.
- [56] Brown NA, Furtado LV, Betz BL, Kiel MJ, Weigelin HC, Lim MS, et al. High prevalence of somatic MAP2K1 mutations in BRAF V600E–negative Langerhans cell histiocytosis. *Blood* 2014;124(10):1655-8.
- [57] Chakraborty R, Hampton OA, Shen X, Simko SJ, Shih A, Abhyankar H, et al. Mutually exclusive recurrent somatic mutations in MAP2K1 and BRAF support a central role for ERK activation in LCH pathogenesis. *Blood* 2014;124(19):3007-15.
- [58] Nelson DS, van Halteren A, Quispel WT, van den Bos C, Bovée JVMG, Patel B, et al. MAP2K1 and MAP3K1 mutations in langerhans cell histiocytosis. *Genes Chromosomes Cancer* 2015;54(6):361-8.
- [59] Nelson DS, Quispel W, Badalian-Very G, Van Halteren AG, Bos C van den, Bovée JV, et al. Somatic activating ARAF mutations in Langerhans cell histiocytosis. *Blood* 2014;123(20):3152-5.
- [60] Jouenne F, Lorillon G, Laurent-Issartel C, Sadoux A, Meignin V, Leschi C, et al. Genetic landscape of pulmonary langerhans cell histiocytosis. In 2018. p. OA3781.
- [61] Lee LH, Gasilina A, Roychoudhury J, Clark J, McCormack FX, Pressey J, et al. Real-time genomic profiling of histiocytoses identifies early-kinase domain BRAF alterations while improving treatment outcomes. *JCI Insight* 2017 Feb 9;2(3):e89473.
- [62] Haroche J, Cohen-Aubart F, Emile JF, Arnaud L, Maksud P, Charlotte F, et al. Dramatic efficacy of vemurafenib in both multisystemic and refractory Erdheim-Chester disease and Langerhans cell histiocytosis harboring the BRAF V600E mutation. *Blood* 2013;121(9):1495-500.
- [63] Diamond EL, Subbiah V, Lockhart AC, Blay JY, Puzanov I, Chau I, et al. Vemurafenib for BRAF V600–mutant Erdheim-Chester disease and Langerhans cell histiocytosis. *JAMA Oncol* 2018;4(3):384-8.

- [64] Hyman DM, Puzanov I, Subbiah V, Faris JE, Chau I, Blay JY, et al. Vemurafenib in multiple nonmelanoma cancers with BRAF V600 mutations. *N Engl J Med* 2015;373(8):726-36.
- [65] Lorillon G, Jouenne F, Baroudjian B, de Margerie-Mellon C, Vercellino L, Meignin V, et al. Response to trametinib of a pulmonary Langerhans cell histiocytosis harboring a MAP2K1 deletion. *Am J Respir Crit Care Med* 2018 Sep 1;198(5):675-8.
- [66] Diamond EL, Durham BH, Ulaner GA, Drill E, Buthorn J, Ki M, et al. Efficacy of MEK inhibition in patients with histiocytic neoplasms. *Nature* 2019;567(7749):521-4.
- [67] Montella L, Insabato L, Palmieri G. Imatinib mesylate for cerebral Langerhans'-cell histiocytosis. *N Engl J Med* 2004;351(10):1034-5.
- [68] Janku F, Amin HM, Yang D, Garrido-Laguna I, Trent JC, Kurzrock R. Response of histiocytoses to imatinib mesylate: fire to ashes. *J Clin Oncol* 2010;28(31):e633-6.
- [69] Arceci RJ, Allen CE, Dunkel IJ, Jacobsen E, Whitlock J, Vassallo R, et al. A phase IIa study of afuresertib, an oral pan-AKT inhibitor, in patients with Langerhans cell histiocytosis. *Pediatr Blood Cancer* 2017;64(5):e26325.
- [70] Diamond EL, Durham BH, Haroche J, Yao Z, Ma J, Parikh SA, et al. Diverse and targetable kinase alterations drive histiocytic neoplasms. *Cancer Discov* 2016;6(2):154.
- [71] Farnault L, Hélias-Rodzewicz Z, Venton G, Fanciullino R, Gabriel S, Mescam L, et al. Response to trametinib of histiocytosis with an activating PTPN11 mutation. *Leuk Lymphoma* 2020 Jan;61(1):194-197.
- [72] Braitheh F, Boxrud C, Esmaeli B, Kurzrock R. Successful treatment of Erdheim-Chester disease, a non-Langerhans-cell histiocytosis, with interferon- α . *Blood* 2005;106(9):2992-4.
- [73] Haroche J1, Amoura Z, Trad SG, Wechsler B, Cluzel P, Grenier PA, et al. Variability in the efficacy of interferon- α in Erdheim-Chester disease by patient and site of involvement: results in eight patients. *Arthritis Rheum* 2006 Oct;54(10):3330-6.

- [74] Arnaud L, Hervier B, Neel A, Hamidou MA, Kahn JE, Wechsler B, et al. CNS involvement and treatment with interferon- are independent prognostic factors in Erdheim-Chester disease: a multicenter survival analysis of 53 patients. *Blood* 2011;117(10):2778-82.
- [75] Haroche J, Amoura Z, Dion E, Wechsler B, Costedoat-Chalumeau N, Cacoub P, et al. Cardiovascular involvement, an overlooked feature of Erdheim-Chester disease: report of 6 new cases and a literature review. *Medicine (Baltimore)* 2004;83(6):371-92.
- [76] Tzoulis C, Schwarzmüller T, Gjerde IO, Sjøfteland E, Neckelmann G, Biermann M, et al. Excellent response of intramedullary Erdheim-Chester disease to vemurafenib: a case report. *BMC Res Notes* 2015 Apr 30;8:171.
- [77] Emile JF, Charlotte F, Amoura Z, Haroche J. BRAF mutations in Erdheim-Chester disease. *J Clin Oncol* 2012;31(3):398.
- [78] Cangi MG, Biavasco R, Cavalli G, Grassini G, Dal-Cin E, Campochiaro C, et al. BRAFV600E-mutation is invariably present and associated to oncogene-induced senescence in Erdheim-Chester disease. *Ann Rheum Dis* 2015;74(8):1596-602.
- [79] Haroche J, Cohen-Aubart F, Emile JF, Maksud P, Drier A, Tolédano D, et al. Reproducible and sustained efficacy of targeted therapy with vemurafenib in patients with BRAFV600E-mutated Erdheim-Chester disease. *J Clin Oncol* 2014;33(5):411-8.
- [80] Nordmann TM, Juengling FD, Recher M, Berger CT, Kalbermatten D, Wicki A, et al. Trametinib after disease reactivation under dabrafenib in Erdheim-Chester disease with both BRAF and KRAS mutations. *Blood* 2017;129(7):879-82.
- [81] Gorre ME, Sawyers CL. Molecular mechanisms of resistance to STI571 in chronic myeloid leukemia. *Curr Opin Hematol* 2002;9(4):303-7.
- [82] Yu HA, Arcila ME, Rekhtman N, Sima CS, Zakowski MF, Pao W, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clin Cancer Res* 2013;19(8):2240-7.

- [83] Shi H, Moriceau G, Kong X, Lee MK, Lee H, Koya RC, et al. Melanoma whole-exome sequencing identifies (V600E)B-RAF amplification-mediated acquired B-RAF inhibitor resistance. *Nat Commun* 2012;3:724.
- [84] Poulikakos PI, Persaud Y, Janakiraman M, Kong X, Ng C, Moriceau G, et al. RAF inhibitor resistance is mediated by dimerization of aberrantly spliced BRAF(V600E). *Nature* 2011;480(7377):387-90.
- [85] Rizos H, Menzies AM, Pupo GM, Carlino MS, Fung C, Hyman J, et al. BRAF inhibitor resistance mechanisms in metastatic melanoma: spectrum and clinical impact. *Clin Cancer Res* 2014;20(7):1965-77.
- [86] Van Allen EM, Wagle N, Sucker A, Treacy DJ, Johannessen CM, Goetz EM, et al. The genetic landscape of clinical resistance to RAF inhibition in metastatic melanoma. *Cancer Discov* 2014;4(1):94-109.
- [87] Johnson DB, Menzies AM, Zimmer L, Eroglu Z, Ye F, Zhao S, et al. Acquired BRAF inhibitor resistance: A multicenter meta-analysis of the spectrum and frequencies, clinical behaviour, and phenotypic associations of resistance mechanisms. *Eur J Cancer Oxf* 2015;51(18):2792-9.
- [88] Bean J, Brennan C, Shih JY, Riely G, Viale A, Wang L, et al. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. *Proc Natl Acad Sci U S A* 2007;104(52):20932-7.
- [89] Rebecca VW, Alicea GM, Paraiso KHT, Lawrence H, Gibney GT, Smalley KSM. Vertical inhibition of the MAPK pathway enhances therapeutic responses in NRAS-mutant melanoma. *Pigment Cell Melanoma Res* 2014;27(6):1154-8.
- [90] Shi H, Hugo W, Kong X, Hong A, Koya RC, Moriceau G, et al. Acquired resistance and clonal evolution in melanoma during BRAF inhibitor therapy. *Cancer Discov* 2014;4(1):80-93.

- [91] Wongchenko MJ, McArthur GA, Dréno B, Larkin J, Ascierto PA, Sosman J, et al. Gene expression profiling in BRAF-mutated melanoma reveals patient subgroups with poor outcomes to vemurafenib that may be overcome by cobimetinib plus vemurafenib. *Clin Cancer Res* 2017 Sep 1;23(17):5238-5245.
- [92] Yan Y, Wongchenko MJ, Robert C, Larkin J, Ascierto PA, Dréno B, et al. Genomic features of exceptional response in vemurafenib ± cobimetinib-treated patients with BRAFV600-mutated metastatic melanoma. *Clin Cancer Res* 2019. DOI: 10.1158/1078-0432.CCR-18-0720. <https://clincancerres.aacrjournals.org/content/early/2019/03/01/1078-0432.CCR-18-0720>. [Accessed December 27, 2019].
- [93] Wagle MC, Kirouac D, Klijn C, Liu B, Mahajan S, Junttila M, et al. A transcriptional MAPK pathway activity score (MPAS) is a clinically relevant biomarker in multiple cancer types. *NPJ Precis Oncol* 2018;2(1):7.
- [94] Ahmadzada T, Kao S, Reid G, Boyer M, Mahar A, Cooper WA. An Update on predictive biomarkers for treatment selection in non-small cell lung cancer. *J Clin Med* 2018;7(6).
- [95] Azorsa DO, Lee DW, Wai DH, Bista R, Patel AR, Aleem E, et al. Clinical resistance associated with a novel MAP2K1 mutation in a patient with Langerhans cell histiocytosis. *Pediatr Blood Cancer* 2018;65(9):e27237.
- [96] Jouenne F, Reger de Moura C, Lorillon G, Meignin V, Dumaz N, Lebbe C, et al. RASA1 loss in a BRAF-mutated Langerhans cell sarcoma: a mechanism of resistance to BRAF inhibitor. *Ann Oncol* 2019;30(7):1170-2.
- [97] Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, Adaptive, and Acquired Resistance to Cancer Immunotherapy. *Cell* 2017;168(4):707-23.
- [98] Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinas H, Hugo W, Hu-Lieskovan S, et al. Mutations associated with acquired resistance to PD-1 blockade in melanoma. *N Engl J Med* 2016;375(9):819-29.

- [99] Hugo W, Zaretsky JM, Sun L, Song C, Moreno BH, Hu-Lieskovan S, et al. Genomic and transcriptomic features of response to anti-PD-1 therapy in metastatic melanoma. *Cell* 2016;165(1):35-44.
- [100] McGranahan N, Furness AJS, Rosenthal R, Ramskov S, Lyngaa R, Saini SK, et al. Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. *Science* 2016;351(6280):1463-9.
- [101] Thommen DS, Schreiner J, Müller P, Herzig P, Roller A, Belousov A, et al. Progression of lung cancer is associated with increased dysfunction of T cells defined by coexpression of multiple inhibitory receptors. *Cancer Immunol Res* 2015;3(12):1344-55.
- [102] Goodman AM, Kato S, Bazhenova L, Patel SP, Frampton GM, Miller V, et al. Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. *Mol Cancer Ther* 2017;16(11):2598-608.
- [103] Otoshi T, Nagano T, Tachihara M, Nishimura Y. Possible biomarkers for cancer immunotherapy. *Cancers (Basel)* 2019 Jul 3;11(7). pii: E935.
- [104] Gray ES, Rizos H, Reid AL, Boyd SC, Pereira MR, Lo J, et al. Circulating tumor DNA to monitor treatment response and detect acquired resistance in patients with metastatic melanoma. *Oncotarget* 2015;6(39):42008-18.
- [105] Cabel L, Riva F, Servois V, Livartowski A, Daniel C, Rampanou A, et al. Circulating tumor DNA changes for early monitoring of anti-PD1 immunotherapy: a proof-of-concept study. *Ann Oncol* 2017;28(8):1996-2001.
- [106] Pelster MS, Amaria RN. Combined targeted therapy and immunotherapy in melanoma: a review of the impact on the tumor microenvironment and outcomes of early clinical trials. *Ther Adv Med Oncol* 2019;11:1758835919830826.
- [107] Moya-Horno I, Viteri S, Karachaliou N, Rosell R. Combination of immunotherapy with targeted therapies in advanced non-small cell lung cancer (NSCLC). *Ther Adv Med Oncol* 2018;10:1758834017745012.

- [108] Tawbi HA-H, Amaria RN, Glitza IC, Milton D, Hwu W-J, Patel SP, et al. Safety and preliminary activity data from a single center phase II study of triplet combination of nivolumab (N) with dabrafenib (D) and trametinib (T) [trident] in patients (Pts) with BRAF-mutated metastatic melanoma (MM). *J Clin Oncol* 2018;36(15_Suppl 15):9560. https://ascopubs.org/doi/abs/10.1200/JCO.2018.36.15_suppl.9560. [Accessed December 27, 2019].
- [109] Dummer R, Arance Fernández AM, Hansson J, Larkin JMG, Long GV, Gasal E, et al. Preliminary findings from part 1 of COMBI-i: A phase III study of anti-PD-1 antibody PDR001 combined with dabrafenib (D) and trametinib (T) in previously untreated patients (pts) with advanced BRAF V600-mutant melanoma. *J Clin Oncol* 2018;36(5_suppl):189.
- [110] Chih-Hsin Yang J, Shepherd FA, Kim DW, Lee GW, Lee JS, Chang GC, et al. Osimertinib plus durvalumab versus osimertinib monotherapy in EGFR T790M-positive NSCLC following previous EGFR TKI therapy: CAURAL brief report. *J Thorac Oncol Off Publ Int Assoc Study Lung Cancer*. mai 2019;14(5):933-9.
- [111] Deng J, Shao J, Markowitz JS, An G. ABC transporters in multi-drug resistance and ADME-Tox of small molecule tyrosine kinase inhibitors. *Pharm Res* 2014;31(9):2237-55.
- [112] Tiseo M, Andreoli R, Gelsomino F, Mozzoni P, Azzoni C, Bartolotti M, et al. Correlation between erlotinib pharmacokinetics, cutaneous toxicity and clinical outcomes in patients with advanced non-small cell lung cancer (NSCLC). *Lung Cancer Amst Neth* 2014;83(2):265-71.
- [113] Motoshima K, Nakamura Y, Sano K, Ikegami Y, Ikeda T, Mizoguchi K, et al. Phase II trial of erlotinib in patients with advanced non-small-cell lung cancer harboring epidermal growth factor receptor mutations: additive analysis of pharmacokinetics. *Cancer Chemother Pharmacol* 2013;72(6):1299-304.
- [114] Calvo E, Malik SN, Siu LL, Baillargeon GM, Irish J, Chin SF, et al. Assessment of erlotinib pharmacodynamics in tumors and skin of patients with head and neck cancer. *Ann Oncol* 2007;18(4):761-7.
- [115] Soulieres D, Senzer NN, Vokes EE, Hidalgo M, Agarwala SS, Siu LL. Multicenter phase II study of erlotinib, an oral epidermal growth factor receptor tyrosine kinase inhibitor, in patients

with recurrent or metastatic squamous cell cancer of the head and neck. *J Clin Oncol* 2004;22(1):77-85.

[116] Pécuchet N, Lebbe C, Mir O, Billemonet B, Blanchet B, Franck N, et al. Sorafenib in advanced melanoma: a critical role for pharmacokinetics? *Br J Cancer* 2012;107(3):455-61.

[117] Boudou-Rouquette P, Ropert S, Mir O, Coriat R, Billemonet B, Tod M, et al. Variability of sorafenib toxicity and exposure over time: a pharmacokinetic/pharmacodynamic analysis. *Oncologist* 2012;17(9):1204-12.

[118] Funck-Brentano E, Alvarez JC, Longvert C, Abe E, Beauchet A, Funck-Brentano C, et al. Plasma vemurafenib concentrations in advanced BRAFV600mut melanoma patients: impact on tumour response and tolerance. *Ann Oncol* 2015;26(7):1470-5.

[119] Goldwirt L, Chami I, Feugeas J-P, Pages C, Brunet-Possenti F, Allayous C, et al. Reply to « Plasma vemurafenib concentrations in advanced BRAFV600mut melanoma patients: impact on tumour response and tolerance » by Funck-Brentano et al. *Ann Oncol* 2016;27(2):363-4.

[120] Kramkimel N, Thomas-Schoemann A, Sakji L, Golmard J, Noe G, Regnier-Rosencher E, et al. Vemurafenib pharmacokinetics and its correlation with efficacy and safety in outpatients with advanced BRAF-mutated melanoma. *Target Oncol* 2016;11(1):59-69.

[121] Rousset M, Dutriaux C, Bosco-Lévy P, Prey S, Pham-Ledard A, Dousset L, et al. Trough dabrafenib plasma concentrations can predict occurrence of adverse events requiring dose reduction in metastatic melanoma. *Clin Chim Acta* 2017;472:26-9.

[122] Takahashi N, Wakita H, Miura M, Scott SA, Nishii K, Masuko M, et al. Correlation between imatinib pharmacokinetics and clinical response in Japanese patients with chronic-phase chronic myeloid leukemia. *Clin Pharmacol Ther* 2010;88(6):809-13.

[123] Picard S, Titier K, Etienne G, Teilhet E, Ducint D, Bernard MA, et al. Trough imatinib plasma levels are associated with both cytogenetic and molecular responses to standard-dose imatinib in chronic myeloid leukemia. *Blood* 2007;109(8):3496-9.

- [124] Marin D, Bazeos A, Mahon FX, Eliasson L, Milojkovic D, Bua M, et al. Adherence is the critical factor for achieving molecular responses in patients with chronic myeloid leukemia who achieve complete cytogenetic responses on imatinib. *J Clin Oncol* 2010;28(14):2381-8.
- [125] Larson RA, Yin OQP, Hochhaus A, Saglio G, Clark RE, Nakamae H, et al. Population pharmacokinetic and exposure-response analysis of nilotinib in patients with newly diagnosed Ph+ chronic myeloid leukemia in chronic phase. *Eur J Clin Pharmacol* 2012;68(5):723-33.
- [126] Giles FJ, Yin OQP, Sallas WM, le Coutre PD, Woodman RC, Ottmann OG, et al. Nilotinib population pharmacokinetics and exposure-response analysis in patients with imatinib-resistant or -intolerant chronic myeloid leukemia. *Eur J Clin Pharmacol* 2013;69(4):813-23.
- [127] Wang X, Roy A, Hochhaus A, Kantarjian HM, Chen TT, Shah NP. Differential effects of dosing regimen on the safety and efficacy of dasatinib: retrospective exposure-response analysis of a Phase III study. *Clin Pharmacol* 2013;5:85-97.
- [128] Yu H, Steeghs N, Nijenhuis CM, Schellens JHM, Beijnen JH, Huitema ADR. Practical guidelines for therapeutic drug monitoring of anticancer tyrosine kinase inhibitors: focus on the pharmacokinetic targets. *Clin Pharmacokinet* 2014;53(4):305-25.
- [129] Hsyu PH, Mould DR, Upton RN, Amantea M. Pharmacokinetic-pharmacodynamic relationship of bosutinib in patients with chronic phase chronic myeloid leukemia. *Cancer Chemother Pharmacol* 2013;71(1):209-18.

Table 1. Main pharmacokinetic characteristics of targeted therapies.

| International non-proprietary name | Metabolic pathway | P-gp | | BCRP | | T _{max} | T _{1/2} | Food effect | CYP3A4 inhibitor effect | CYP3A4 inducer effect |
|------------------------------------|---------------------------------|-----------|-----------|-----------|-----------|------------------|------------------|----------------------------|---------------------------------|-----------------------------|
| | | Substrate | Inhibitor | Substrate | Inhibitor | [h] | [h] | | | |
| Lung Cancer | | | | | | | | | | |
| Ceritinib | CYP3A4 | +++ | ++ | x | ++ | 4-6 | 31-41 | Slight increase | +290% AUC | -70% AUC |
| | | | | | | | | | +120% C _{max} | -44% C _{max} |
| Crizotinib | CYP3A4, CYP3A5 | +++ | +++ | x | x | 4-6 | 42 | -14% AUC | +3,2 x AUC | -84% AUC |
| | | | | | | | | -14% C _{max} | +1,4 x C _{max} | -79% C _{max} |
| Erlotinib | CYP3A4 | +++ | +++ | +++ | +++ | 4 | 36,2 | Slight increase | +86% AUC | -69% AUC |
| | | | | | | | | | +69% C _{max} | |
| Gefitinib | CYP3A4, CYP2D6 | +++ | NA | +++ | +++ | | | -83% AUC | +80% AUC | |
| Melanoma | | | | | | | | | | |
| Cobimetinib | CYP3A4, CYP3A5 | +++ | x | x | ++ | 2,4 | 43,6 | Negligible | Increased AUC, C _{max} | lower AUC, C _{max} |
| Dabrafenib | CYP2C8, CYP3A4 | +++ | NA | +++ | NA | 2 | 8 | -31% AUC | +71% AUC | NA |
| | | | | | | | | -51% C _{max} | +33% C _{max} | |
| Sorafenib | CYP3A4, UGT1A9 | x | NA | +++ | NA | 3 | 25-48 | -30% AUC | Negligible | -37% AUC |
| Trametinib | Carboxylesterase | +++ | + | x | + | 1,5 | 127 | -70% AUC | Negligible | Negligible |
| | | | | | | | | -10% C _{max} | | |
| Vemurafenib | CYP3A4 | +++ | + | +++ | +++ | 4 | 51,6 | +4,6 x AUC | NA | -40% AUC |
| | | | | | | | | +2,5 x C _{max} | | |
| Blood cancers | | | | | | | | | | |
| Bosutinib | CYP3A4 | x | +++ | x | +++ | 6 | 34 | +170% AUC | + 200% AUC | -6% AUC |
| | | | | | | | | +180% C _{max} | +150% C _{max} | -14% C _{max} |
| Dasatinib | CYP3A4 | +++ | + | +++ | ++ | 0,5-4 | 5-6 | -14% AUC | | -82% AUC |
| Ibrutinib | CYP3A4 | x | ++ | x | ++ | 1-2 | 4-13 | Increase AUC | +24 x AUC | -90% AUC |
| | | | | | | | | | +29 x C _{max} | -92% C _{max} |
| Idelalisib | Aldehyde oxydase, CYP3A, UGT1A4 | ++ | + | ++ | + | 1-2 | 8,2 | +36% AUC | +79% AUC | -75% AUC |
| | | | | | | | | Unchanged C _{max} | +26% C _{max} | |
| Imatinib | CYP2A4 | +++ | +++ | +++ | +++ | | 18 | -11% C _{max} | +40% AUC | -74% AUC |
| | | | | | | | | | +26% C _{max} | -54% C _{max} |
| Nilotinib | CYP3A4, CYP2C8 | + | +++ | +++ | +++ | | 17 | +82% AUC | +300% AUC | -80% AUC |

| | | | | | | | | | | |
|-----------|--------|-----|----|-----|-----|---|----|---------------------------|--------------------------|-----------------------|
| | | | | | | | | +112% C _{max} | | -64% C _{max} |
| Ponatinib | CYP3A4 | +++ | ++ | +++ | +++ | 4 | 22 | Negligible | +78% AUC | -64% AUC |
| | | | | | | | | | +47% C _{max} | -42% C _{max} |

+++ : strong substrate or inhibitor; ++: moderate substrate or inhibitor; + : weak substrate or inhibitor; x : neither substrate or inhibitor.

AUC: Area under the curve; BCRP: breast cancer resistance protein; CYP: cytochrome P; h: hours; NA: not available;

P-gp: P-glycoprotein; UGT: glucuronosyltransferase.