



# LECTURE III: DENOISING & CLUSTERING

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# ILLUMINA AMPLICON READS

- Illumina MiSeq sequencing is the current state of the art technology for amplicon sequencing because of its high yield relative to cost.
- The error rate ( $\sim 1\%$ ) of Illumina sequencing is so high that the chance of capturing true reads is small (about 1% for a 450 base fragment, not adjusted for improvements due to overlapping reads).
- This problem is not new, and has been discussed ever since high-throughput sequencing was first applied to taxonomical markers.



# PROPOSED SOLUTIONS

- For many years now, the common practice was to solve this by UPGMA- style clustering at a fixed sequence distance (97% similarity).
- Sequences were clustered into operational taxonomic units based on sequence similarity or dissimilarity cutoffs.

# UPMG

Single linkage clustering.

Advantages:

- reduces the impact of clustering parameters on the resulting OTUs by avoiding arbitrary global clustering thresholds and input sequence ordering dependence.

# UPARSE

UPARSE implements a greedy algorithm that performs OTU clustering

## Advantages:

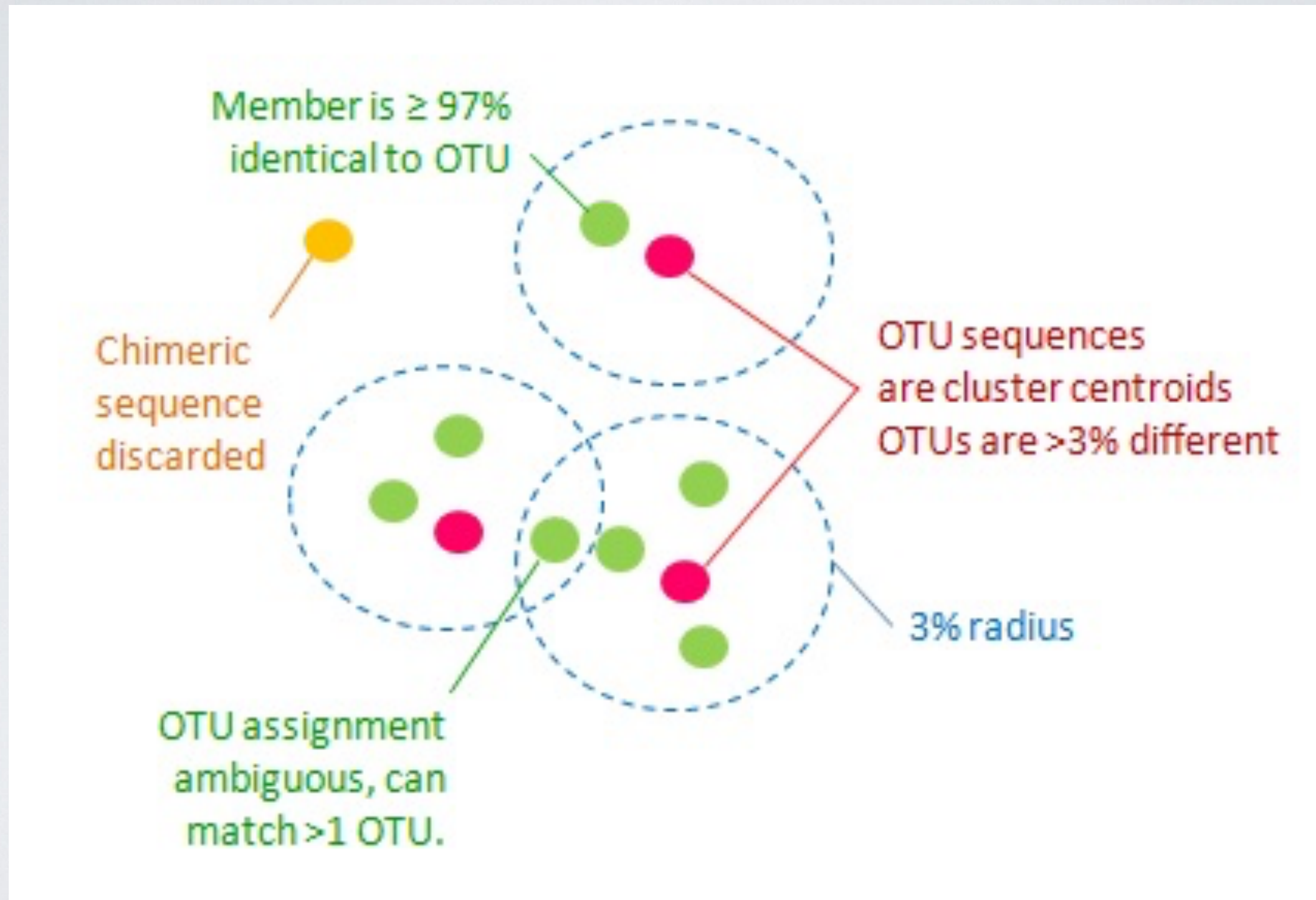
- Fast and greedy.

## Disadvantages:

- Greedy clustering methods suffer from two fundamental problems.
  1. They use an arbitrary fixed global clustering threshold. As lineages evolve at variable rates, no single cut-off value can accommodate the entire tree of life.
  2. The input order of amplicons strongly influences the clustering results. Previous centroid selections are not re-evaluated as clustering progresses, which can generate inaccurately formed OTUs



# UPARSE



[https://drive5.com/usearch/manual/uparseotu\\_algo.html](https://drive5.com/usearch/manual/uparseotu_algo.html)

# SWARM

Swarm is a de novo clustering algorithm based on an unsupervised single-linkage-clustering method.

**Advantages:**

- reduces the impact of clustering parameters on the resulting OTUs by avoiding arbitrary global clustering thresholds and input sequence ordering dependence.



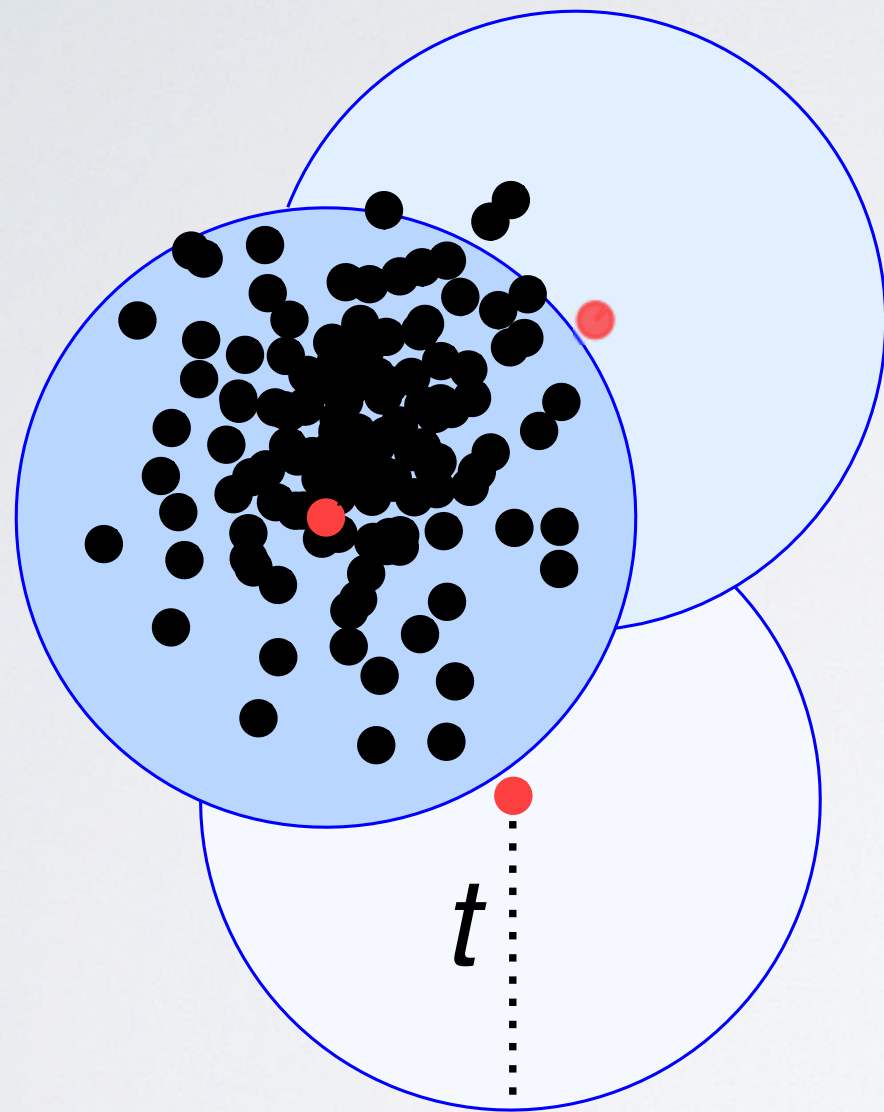
# SWARM

**Swarm builds OTUs in two steps:**

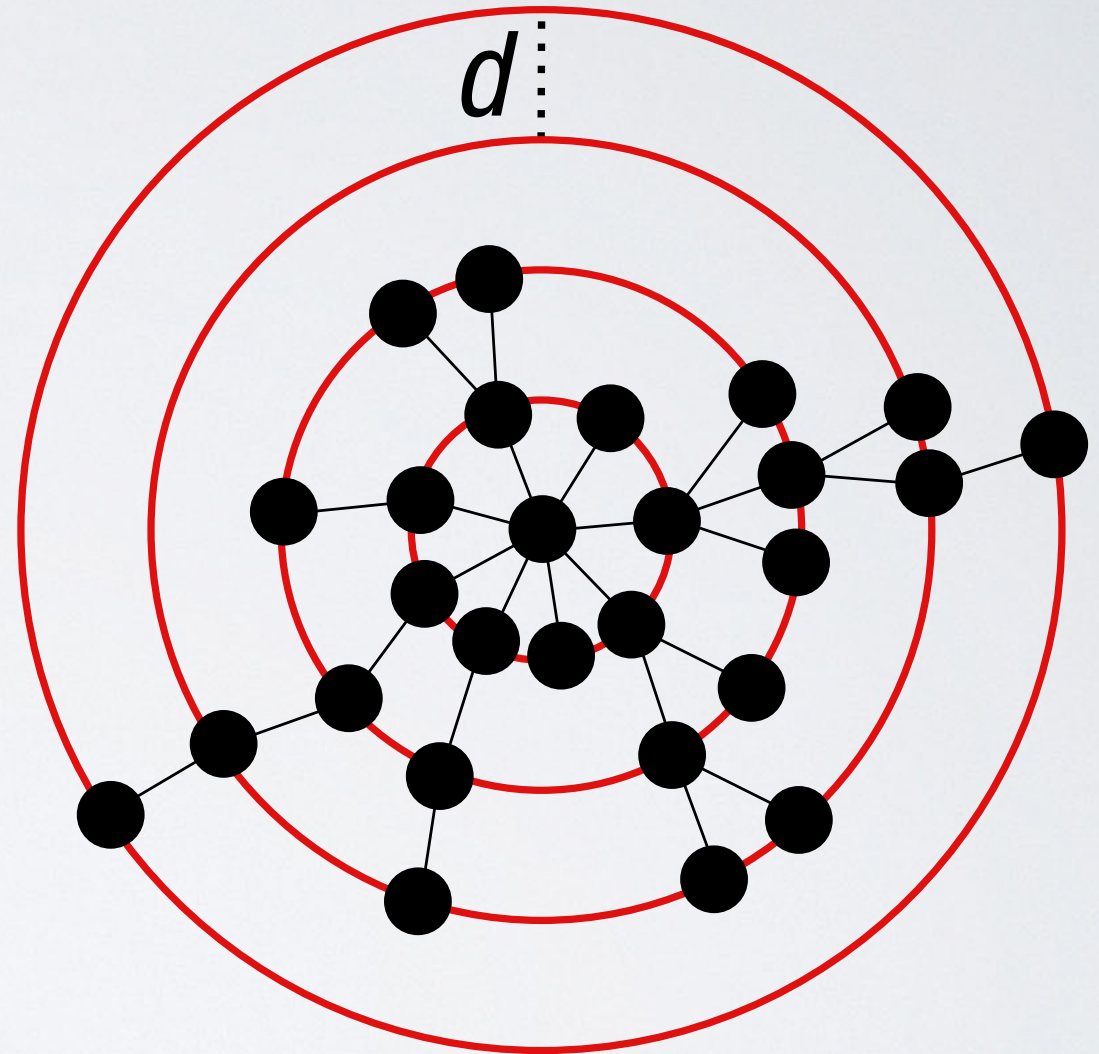
1. an initial set of OTUs is constructed by iteratively agglomerating similar amplicons
2. amplicon abundance values are used to reveal OTUs' internal structures and to break them into sub-OTUs, if necessary.

# SWARM

a



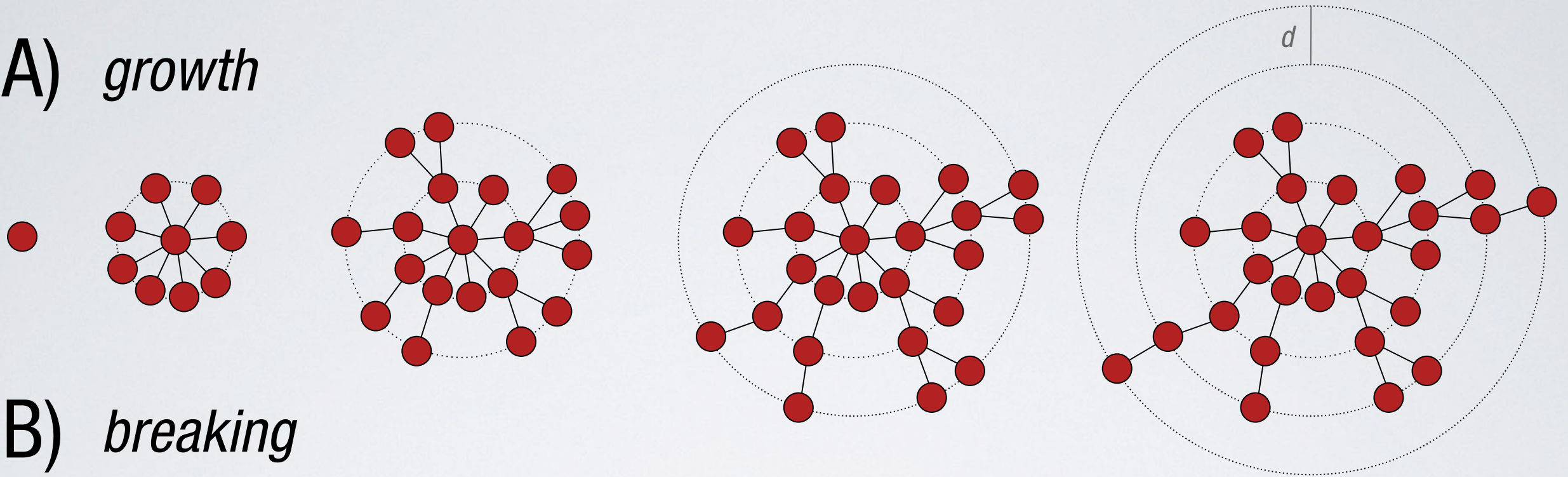
b



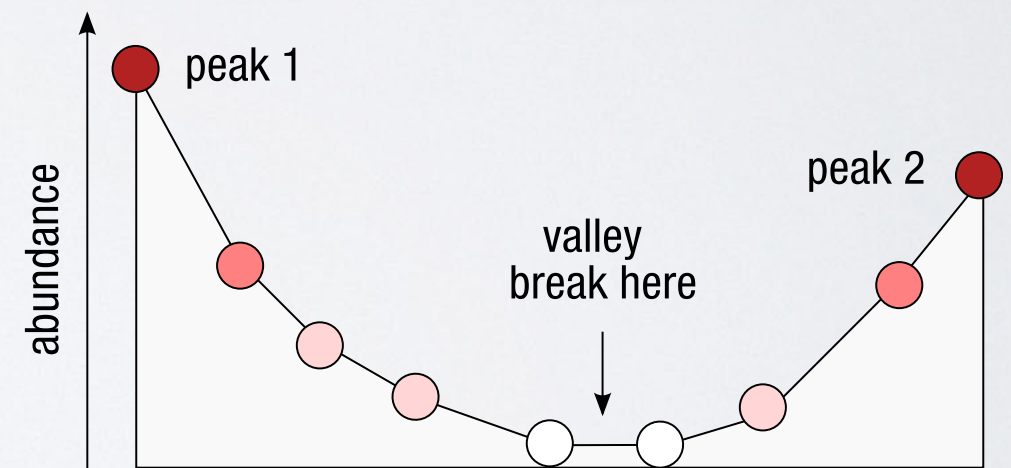
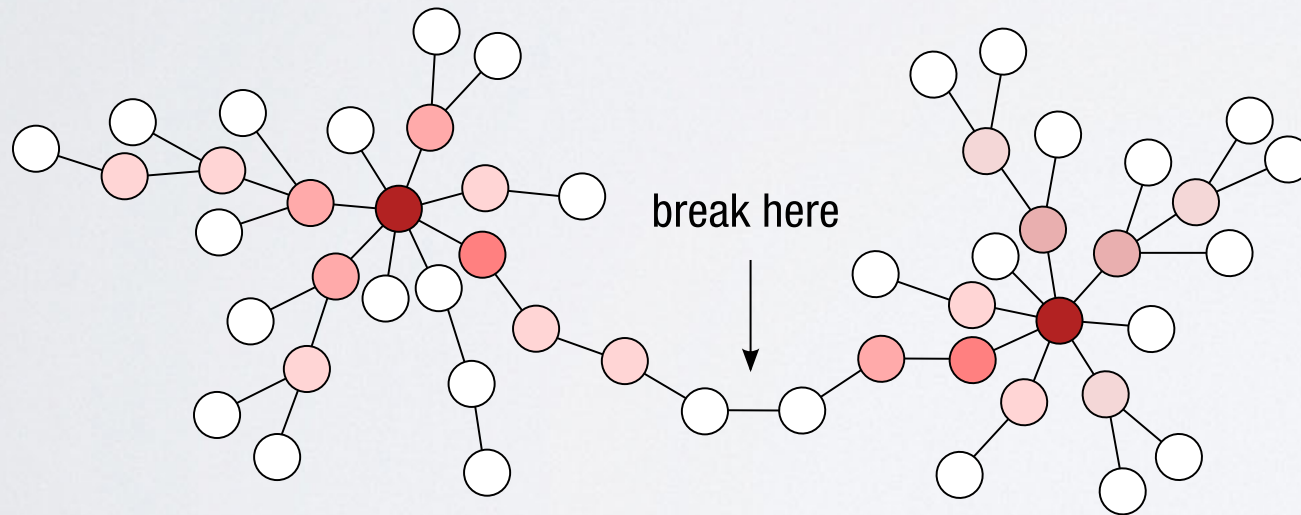


# SWARM

## A) *growth*



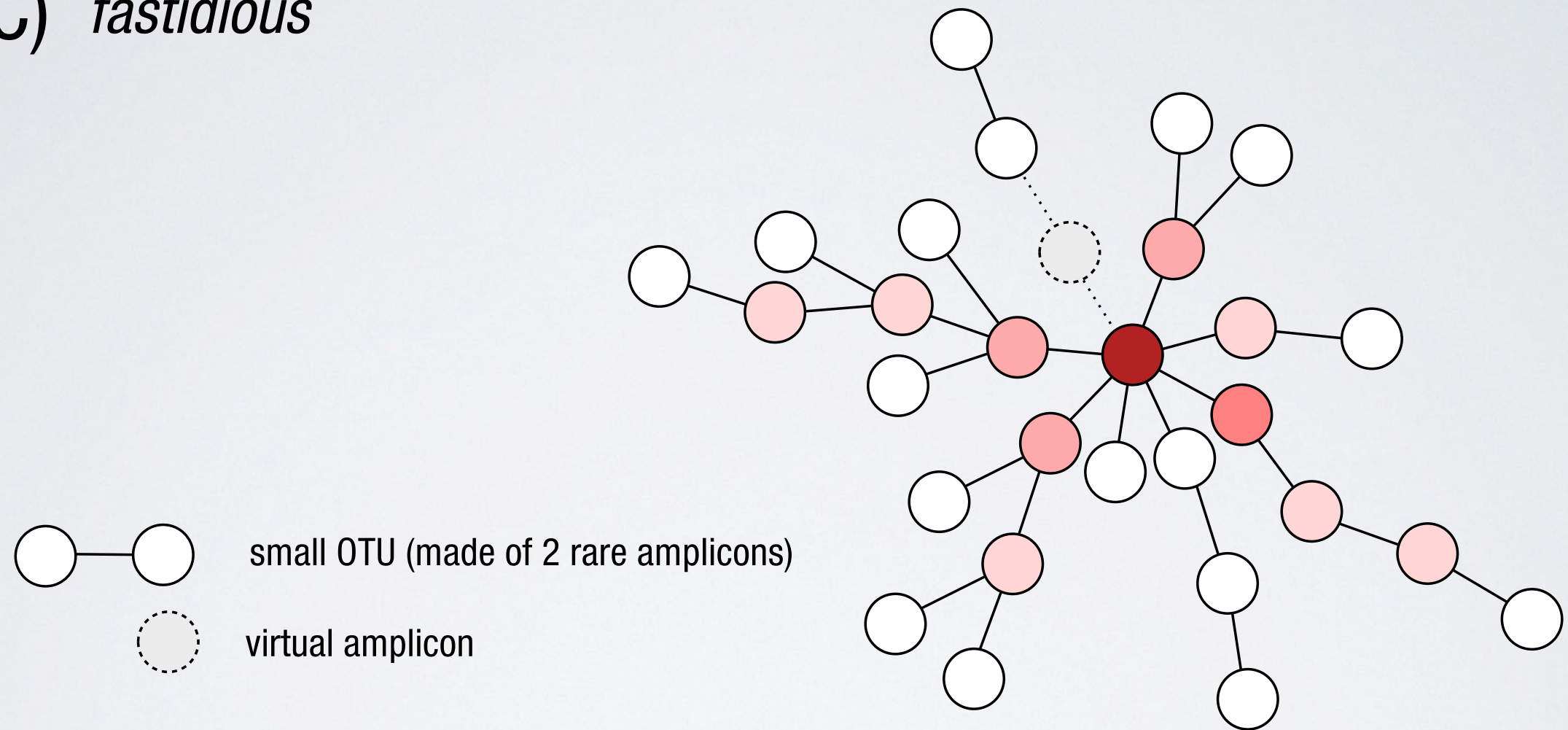
## B) *breaking*





# SWARM

## C) *fastidious*



# LINGERING PROBLEMS WITH “OTU”

imagine sequencing reads  
streaming from a single  
true sequence...



# LINGERING PROBLEMS WITH “OTU”

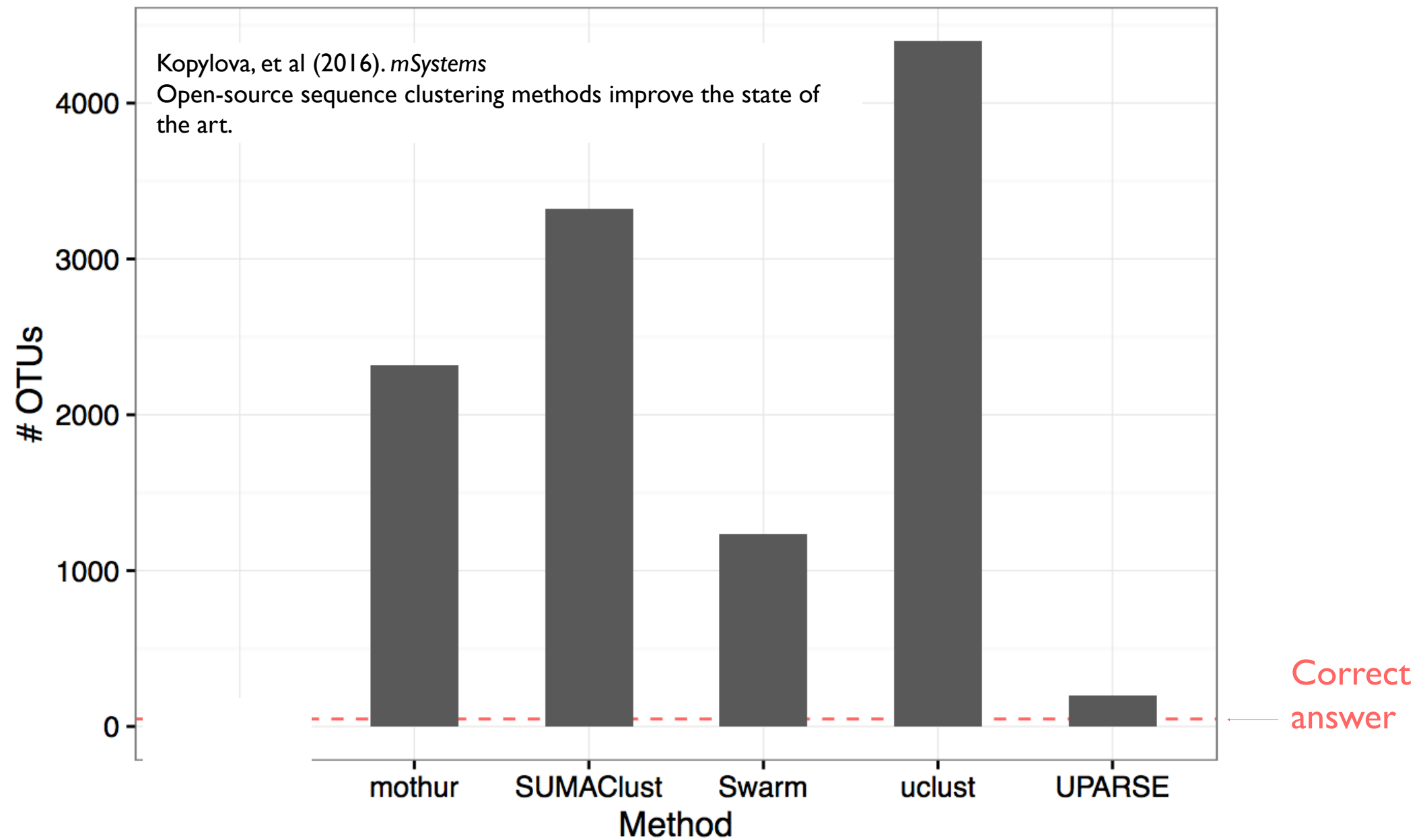
The deeper you sequence, the more you expect to find reads outside the radius by chance.





# LINGERING PROBLEMS WITH “OTU”

## Typical “OTU” performance on validation data (“mock community”)



<http://benjjneb.github.io/dada2/R/SotA.html>

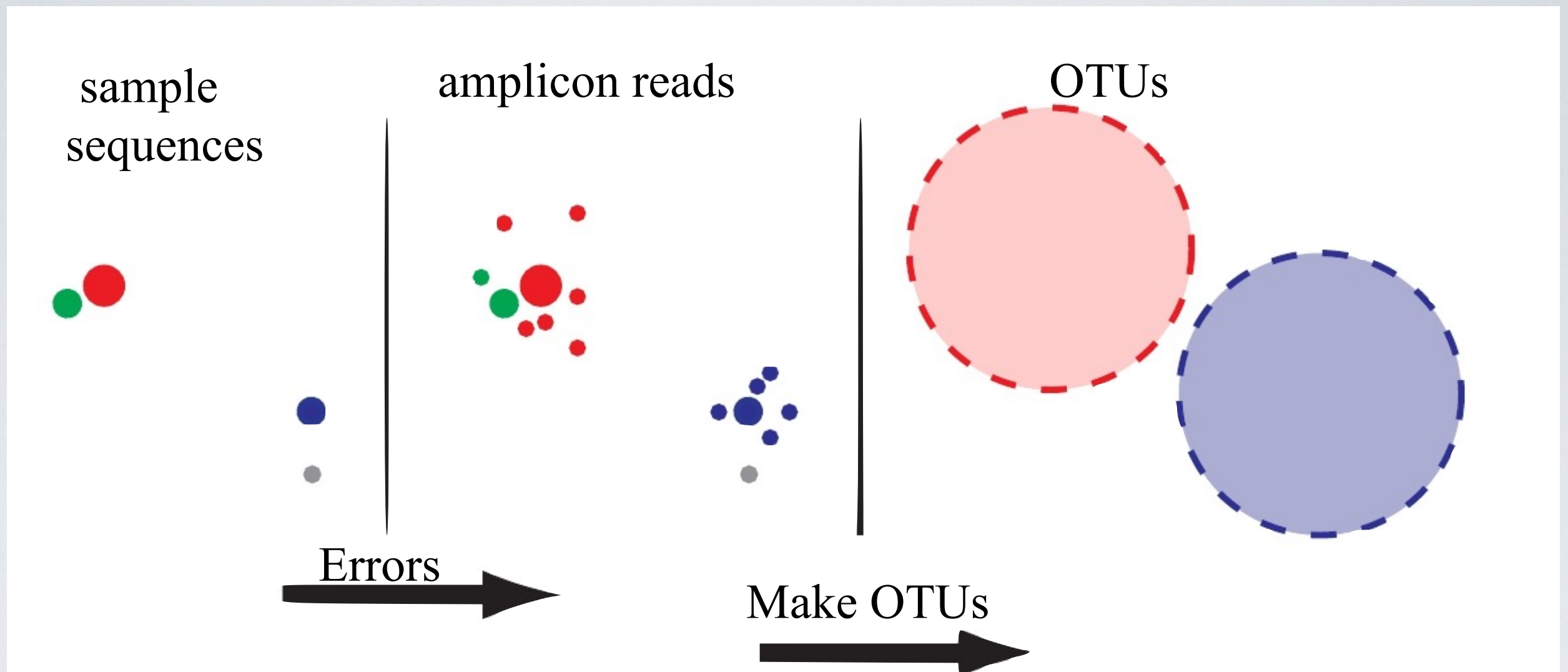
sample  
sequences

amplicon reads

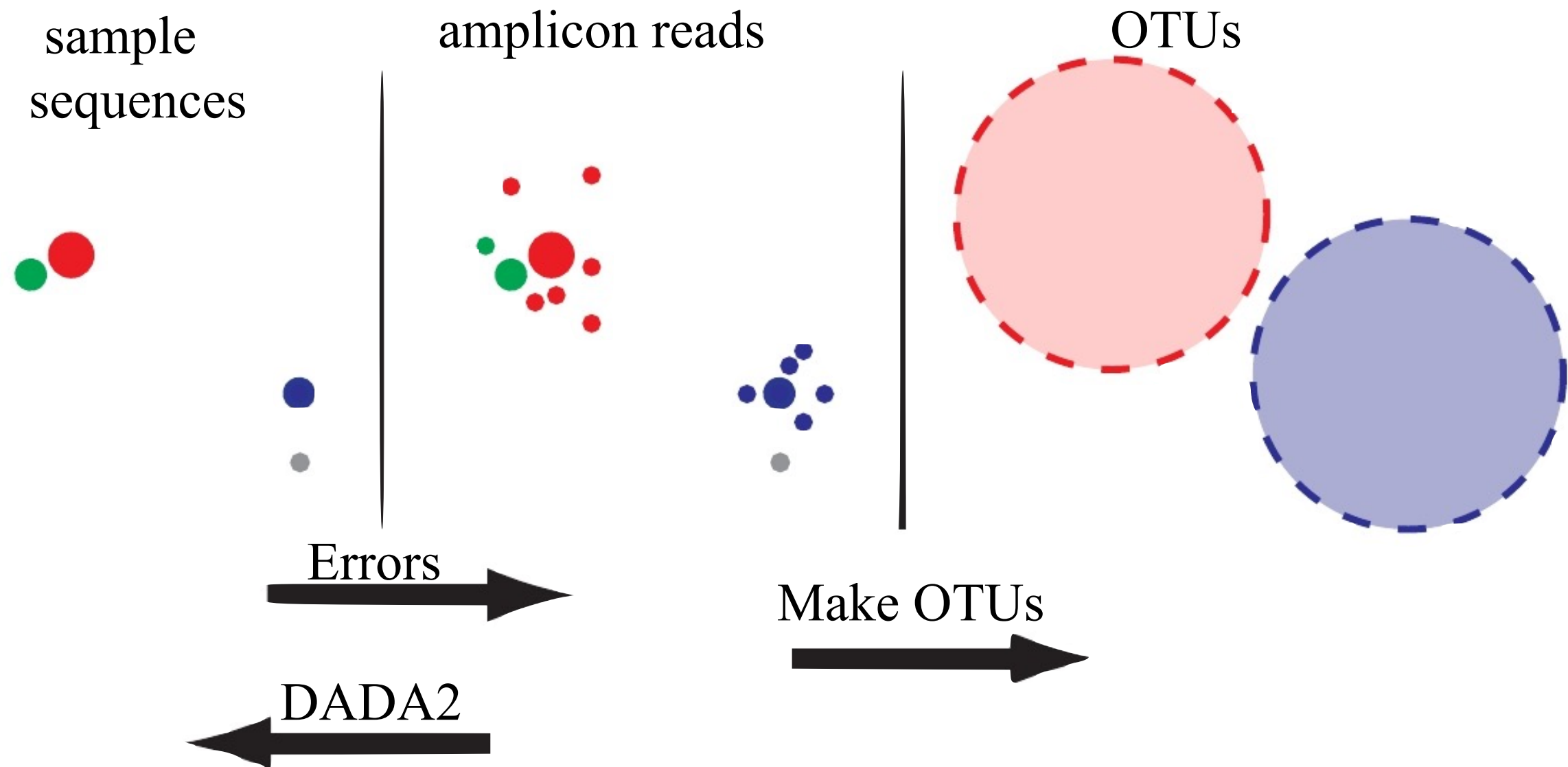


Errors



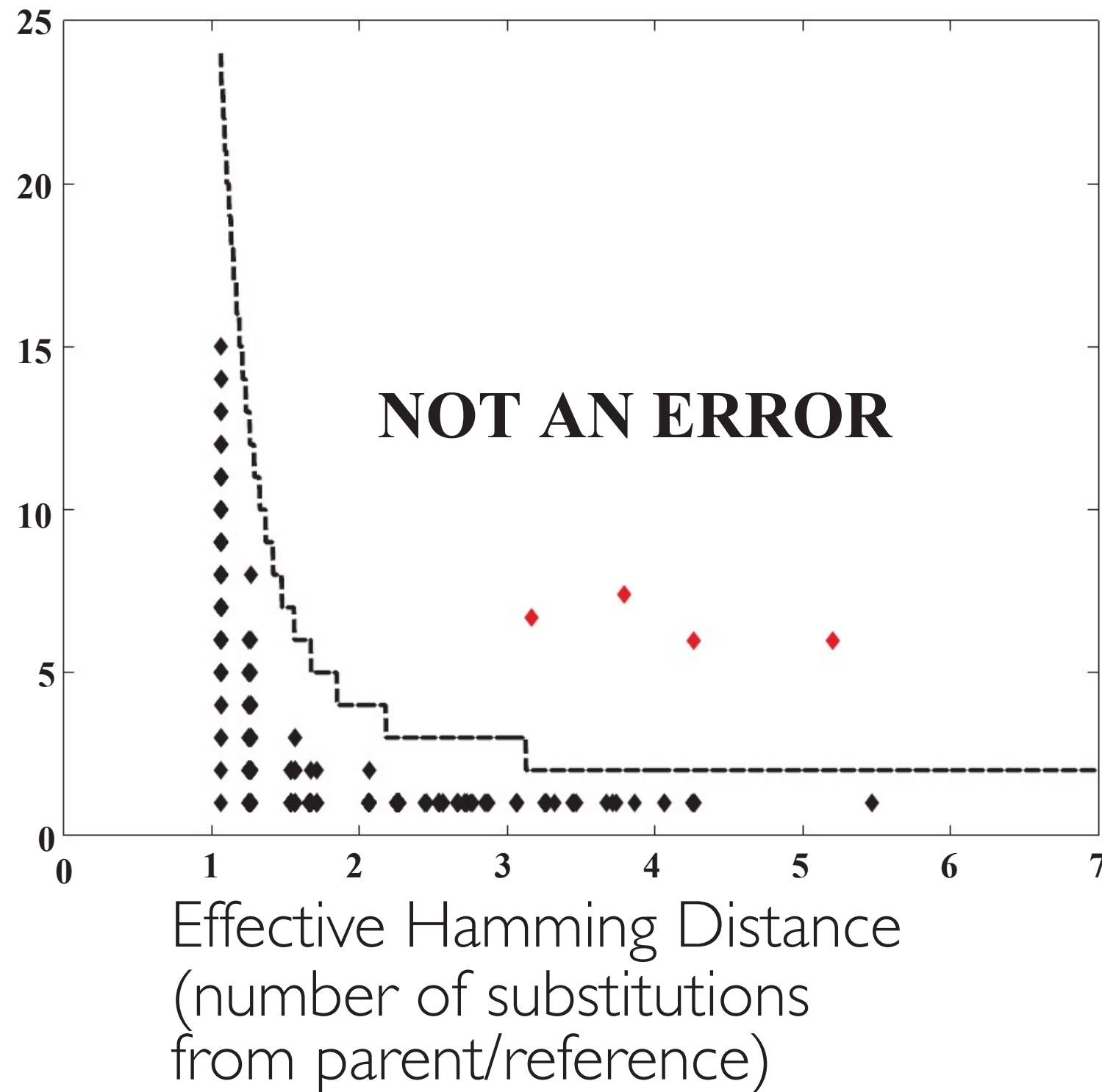






# The shape of amplicon sequencing errors

counts,  
unique  
sequence



Slide adapted from Benjamin Callahan

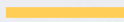

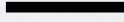

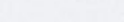
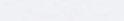
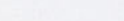
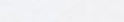
# DADA2

“raw” reads



dereplicate

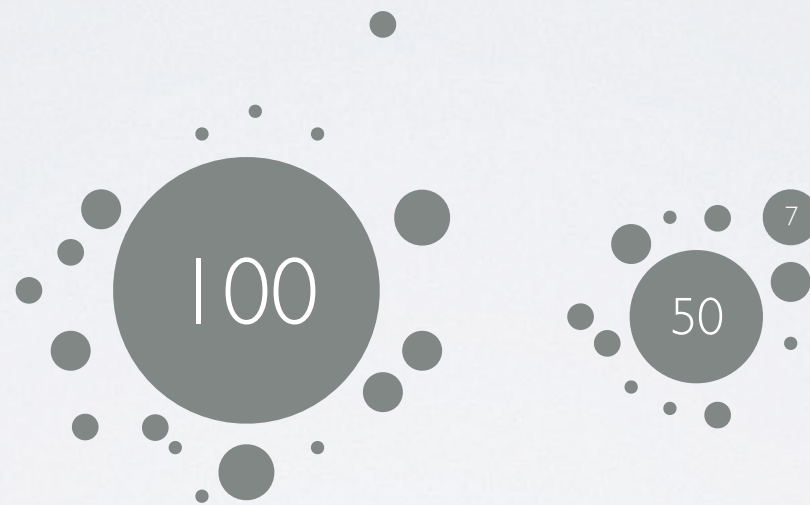
Input:

unique sequences	abundance	mean-Q
	100	32
	50	32
	7	20
	5	...
	4	...
	3	...
	2	...
	2	...



# DADA2

Initial guess: one real sequence + errors



# DADA2

Infer initial error model under this assumption.

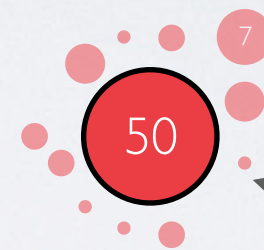


$\text{Pr}(i \rightarrow j) =$

	A	C	G	T
A	0.97	$10^{-2}$	$10^{-2}$	$10^{-2}$
C	$10^{-2}$	0.97	$10^{-2}$	$10^{-2}$
G	$10^{-2}$	$10^{-2}$	0.97	$10^{-2}$
T	$10^{-2}$	$10^{-2}$	$10^{-2}$	0.97

# DADA2

Update the model.



not an error

$\text{Pr}(i \rightarrow j) =$

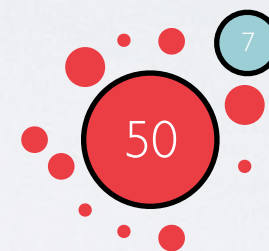
	A	C	G	T
A	0.997	$10^{-3}$	$10^{-3}$	$10^{-3}$
C	$10^{-3}$	0.997	$10^{-3}$	$10^{-3}$
G	$10^{-3}$	$10^{-3}$	0.997	$10^{-3}$
T	$10^{-3}$	$10^{-3}$	$10^{-3}$	0.997



# DADA2

Update model again

not an error → •



not an error

$\text{Pr}(i \rightarrow j) =$

	A	C	G	T
A	0.998	$1 \times 10^{-4}$	$2 \times 10^{-3}$	$2 \times 10^{-4}$
C	$6 \times 10^{-5}$	0.998	$3 \times 10^{-4}$	$1 \times 10^{-3}$
G	$1 \times 10^{-4}$	$1 \times 10^{-4}$	0.998	$6 \times 10^{-5}$
T	$2 \times 10^{-4}$	$2 \times 10^{-3}$	$1 \times 10^{-4}$	0.998

# DADA2 ASSUMPTIONS

## DADA2 Error Model:

- Errors independent b/w different sequences
- Errors independent b/w sites within a sequence
- Sequence  $i$  is produced from parent sequence  $j$  with probability equal to the product of site-wise substitution probabilities:

$$\lambda_{j \rightarrow i} = \prod_{l=0}^L p(j(l) \rightarrow i(l), q(l)))$$

- Each substitution probability depends on original nt, substituting nt, and quality score at position in  $i$



# DADA2 ASSUMPTIONS

## DADA2 Abundance Model:

- Errors are independent across reads
- Abundance of reads w/ sequence  $i$  produced from more-abundant parent sequence  $j$  is poisson distributed
- Expectation of abundance equals error rate,  $\lambda_{j \rightarrow i}$ , multiplied by the abundance of sequence  $j$
- $i$  has count greater than or equal to one
- “Abundance p-value” for sequence  $i$  is thus:

$$p_A(j \rightarrow i) = \sum_{a=a_1}^{\infty} \rho_{pois}(n_j \lambda_{j \rightarrow i}, a) / (1 - \rho_{pois}(n_j \lambda_{j \rightarrow i}, 0))$$

- “Probability of seeing an abundance of sequence  $i$  that is equal to or greater than observed value, by chance, given sequence  $j$ .” (Bonferroni-corrected)
- A low  $p_A$  indicates there are more reads of sequence  $i$  than can



# APPLICATIONS

- Any amplicon-seq data, not just 16S rRNA or even microbiome
- Sequence variants unique to an individual host
- Sequence variants associated with a clinical outcome
- Improved meta-genomic inference (e.g. PiCRUST)
- Mitigate ambiguity of representative genome(s) to use
- Detecting pathogens (special cases)