

The Norwegian Insect Monitoring Program

Lessons in metabarcoding-based biomonitoring

Marie Louise Davey, Senior Researcher



Global Insect Decline



RESEARCH ARTICLE

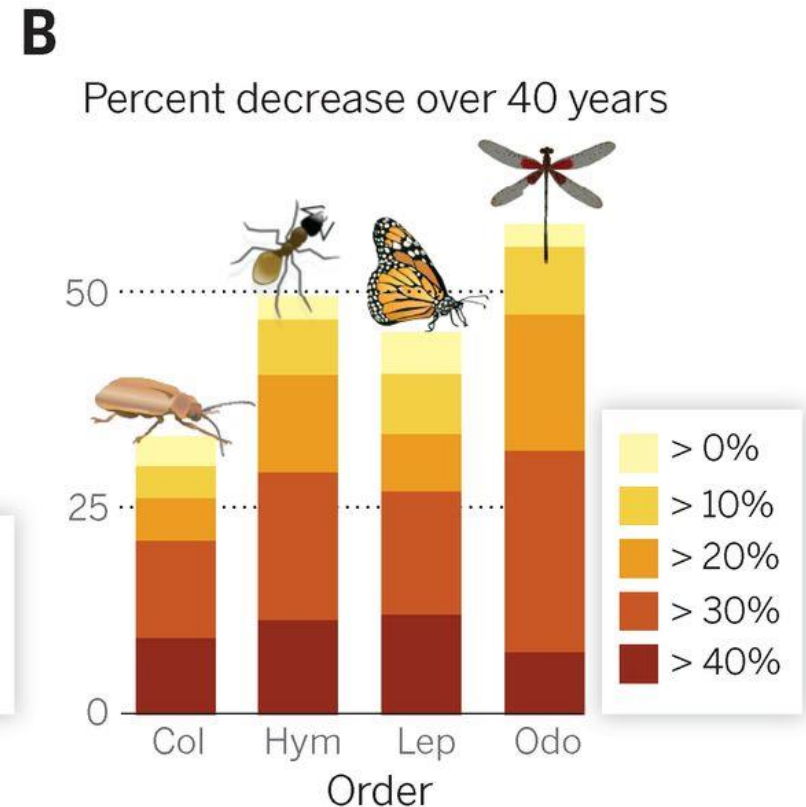
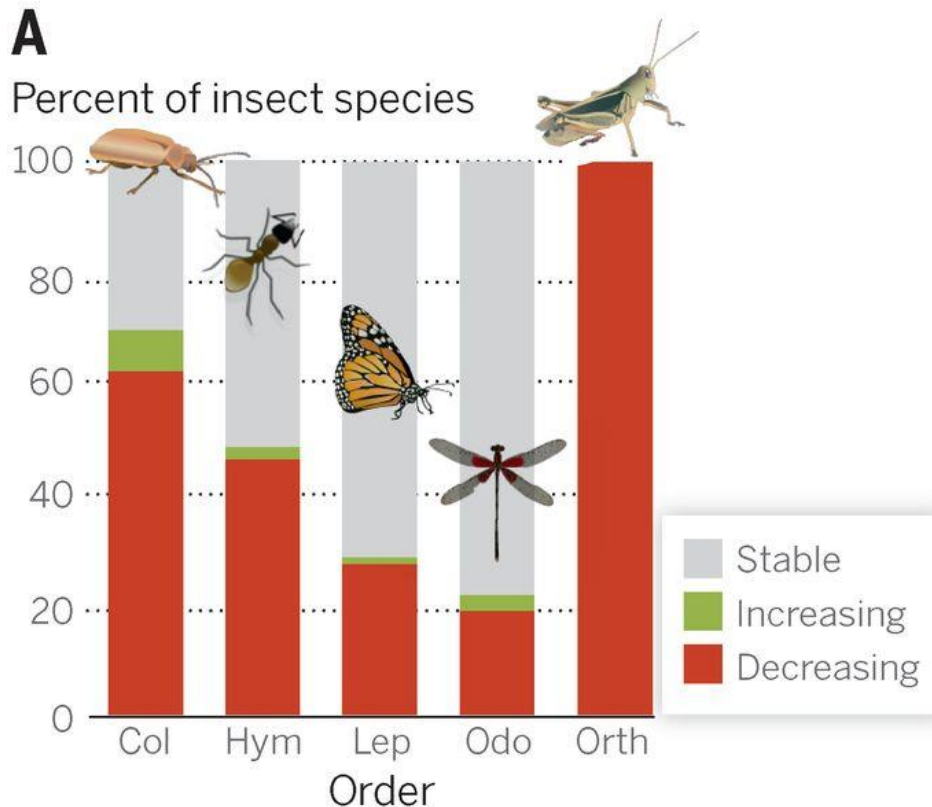
More than 75 percent decline over 27 years in total flying insect biomass in protected areas

Caspar A. Hallmann^{1*}, Martin Sorg², Eelke Jongejans¹, Henk Siepel¹, Nick Hofland¹, Heinz Schwan², Werner Stenmans², Andreas Müller², Hubert Sumser², Thomas Hörren², Dave Goulson³, Hans de Kroon¹

1 Radboud University, Institute for Water and Wetland Research, Animal Ecology and Physiology & Experimental Plant Ecology, PO Box 9100, 6500 GL Nijmegen, The Netherlands, **2** Entomological Society Krefeld e.V., Entomological Collections Krefeld, Marktstrasse 159, 47798 Krefeld, Germany, **3** University of Sussex, School of Life Sciences, Falmer, Brighton BN1 9QG, United Kingdom

* c.hallmann@science.ru.nl

Global Insect Decline



Defaunation in the Anthropocene

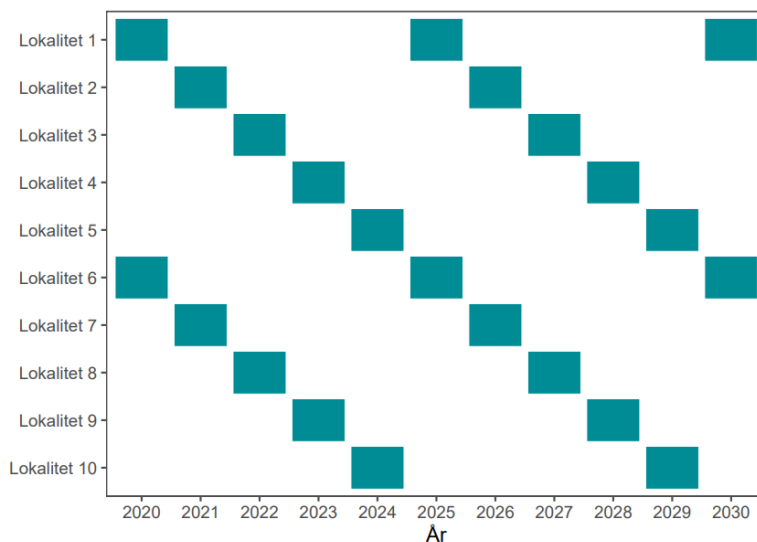
RODOLFO DIRZO, HILLARY S. YOUNG, MAURO GALETTI, GERARDO CEBALLOS, [...], AND BEN COLLEN

SCIENCE • 25 Jul 2014 • Vol 345, Issue 6195 • pp. 401-406 • DOI: 10.1126/science.1251817

www.nina.no

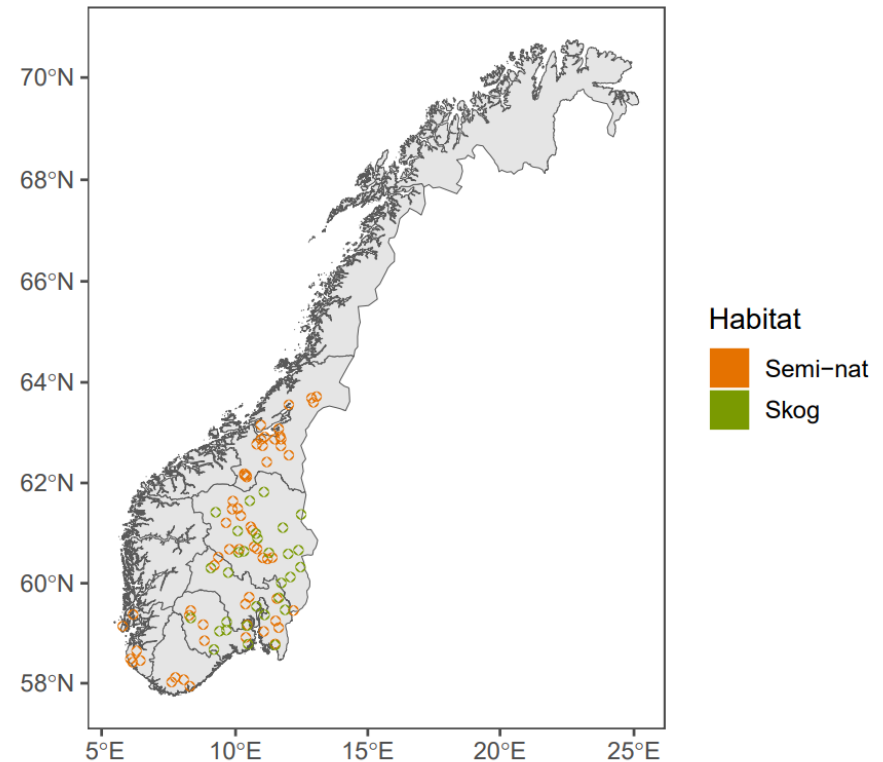
Norwegian Insect Monitoring Program

- Funded by the Norwegian Environment Agency
 - ▶ Initiated in 2019 with a pilot project in Trøndelag for method development
 - ▶ Uses a field design with 50 monitoring sites per habitat type visited on a 5-year rolling basis.
 - ▶ Based primarily on malaise traps for flying insects, with supplemental window traps for beetles



Norwegian Insect Monitoring Program

- ▶ Launched in 2020 in semi-natural grasslands and forests in eastern Norway
- ▶ Program is scaling up to national coverage
 - expanded to semi-natural grasslands in central Norway (2021), southern Norway (2022), and northern Norway (2023)



Norwegian Insect Monitoring Program

- A variety of bioclimatic data is recorded at each locality
- Insect biomass caught is monitored at 2 week intervals from April to October
- Monitoring at the «species» level is by metabarcoding-based identification of the insect biomass caught at each locality

Why metabarcoding for biomonitoring?

- High throughput
- Cost effective
- Standardized for comparable results
- Accurate and precise
- Relevant data for relevant organisms

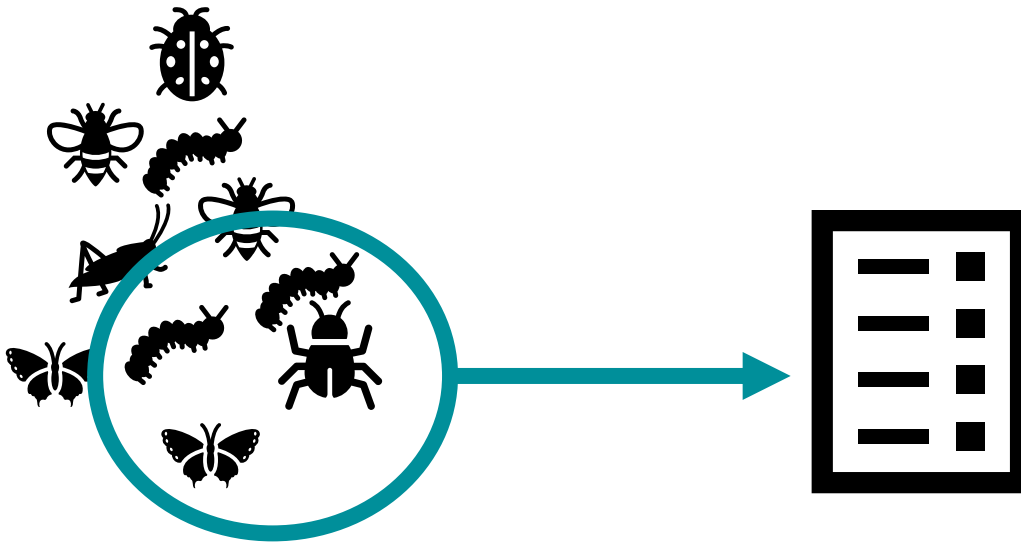
Why metabarcoding for biomonitoring?

	Metabarcoding	Morphological
Traps/Sites:	100/100	73/55
Sampling Duration:	1 year	3 years
Processing Time:	3 months	15 years
Completion	100%	1%
Species Recovered:	16 000	4 000
New to Science:	?	700
Cots:	2 million USD	3 million USD*

*20% voluntary labour

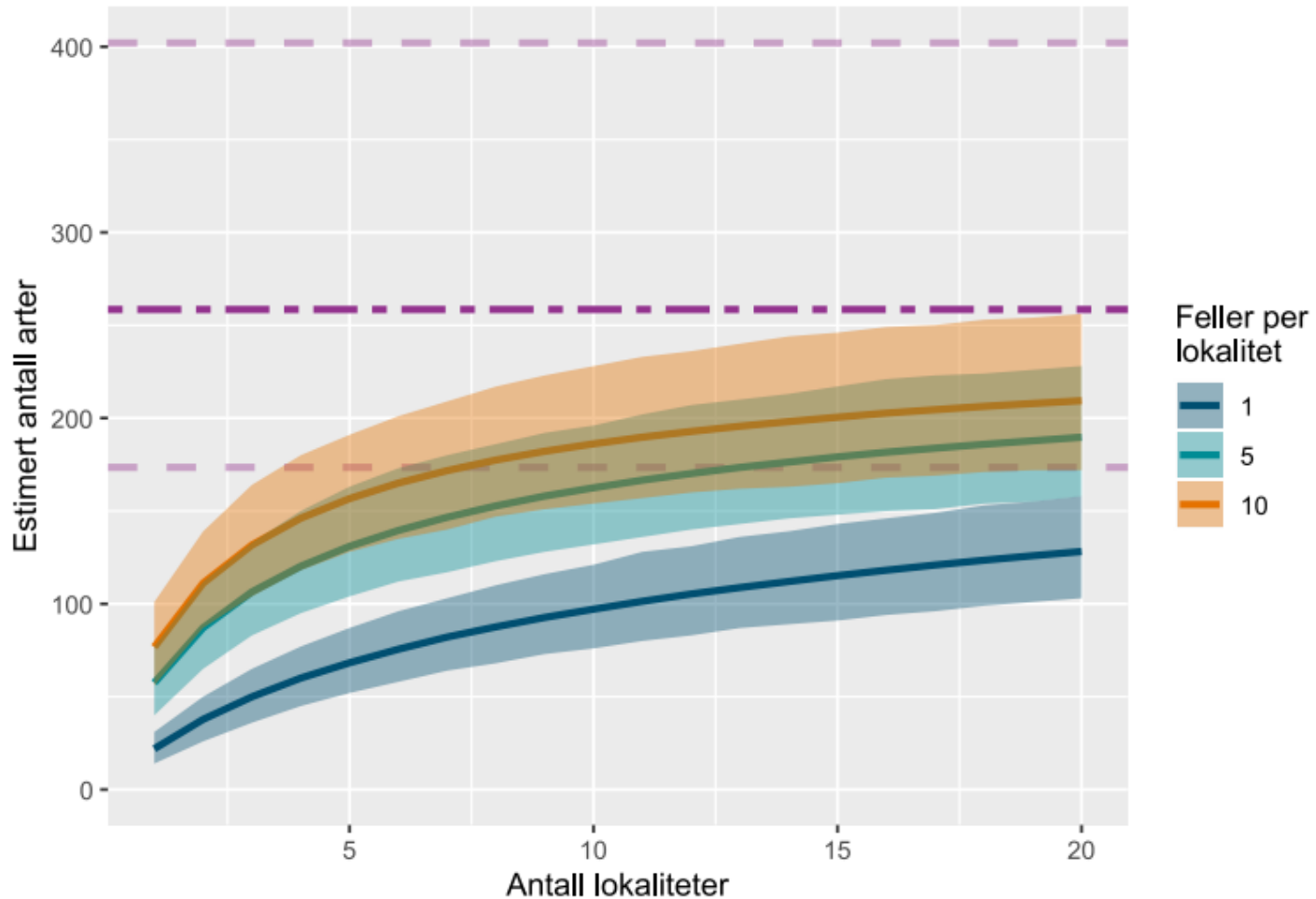
Designing a monitoring program

- How do we effectively sample insects on a national scale?

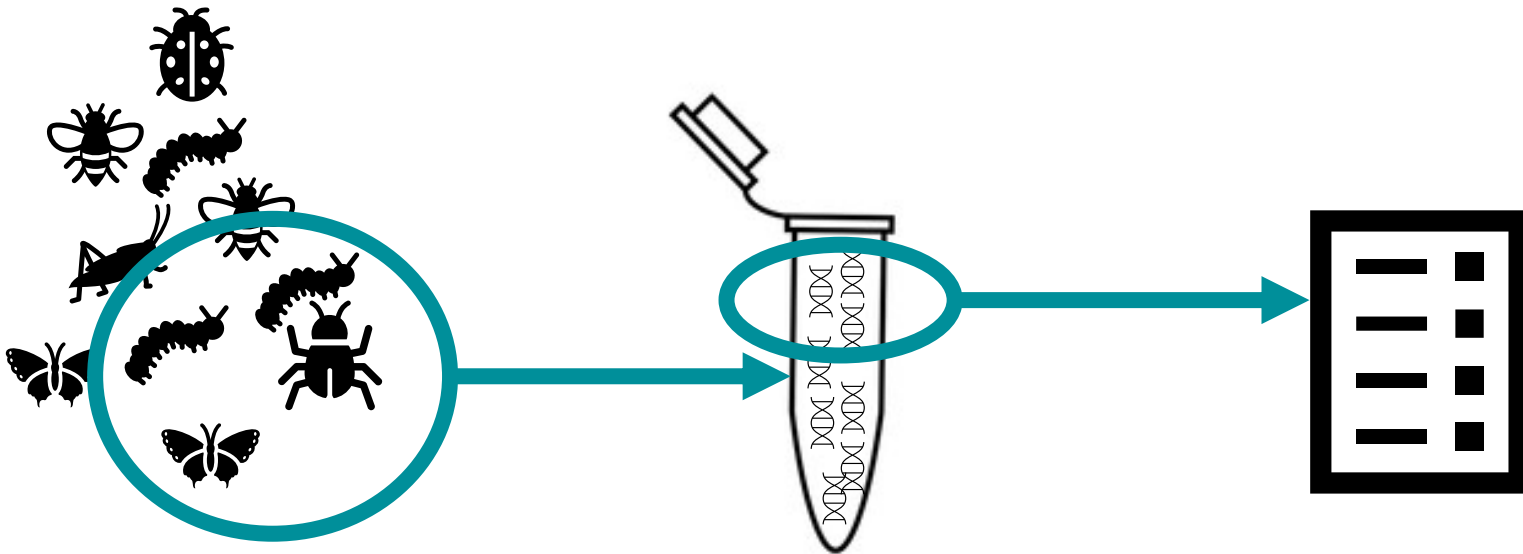




Designing a monitoring program



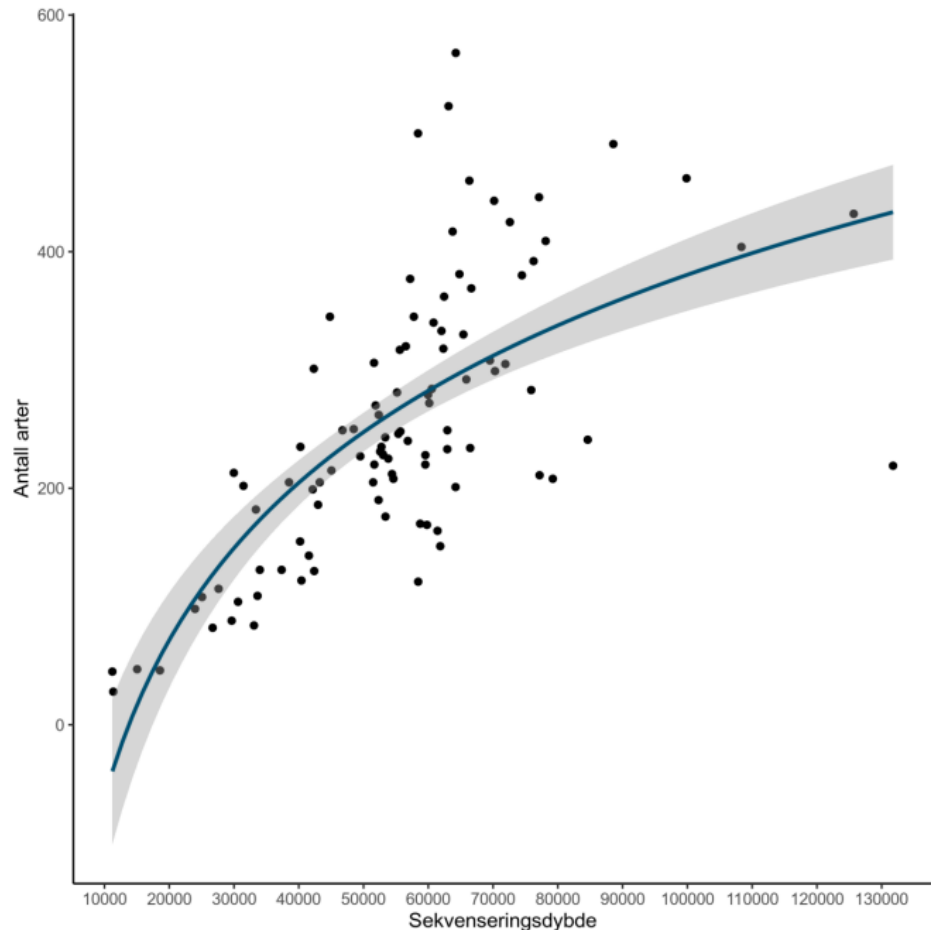
Designing a monitoring program



- Metabarcoding effectively introduces an additional sampling event
 1. sampling of organisms
 2. sampling of DNA molecules

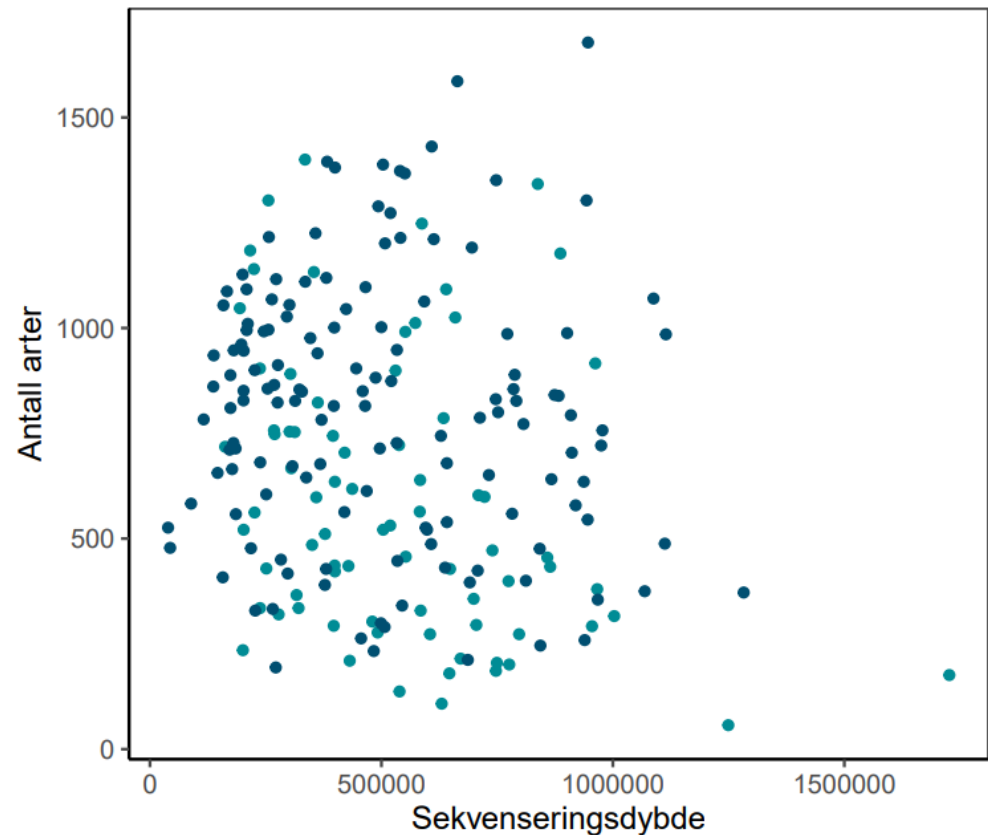
Designing a monitoring program

- Sufficient sequencing depth is vital in order to rival detection capacity of other methods



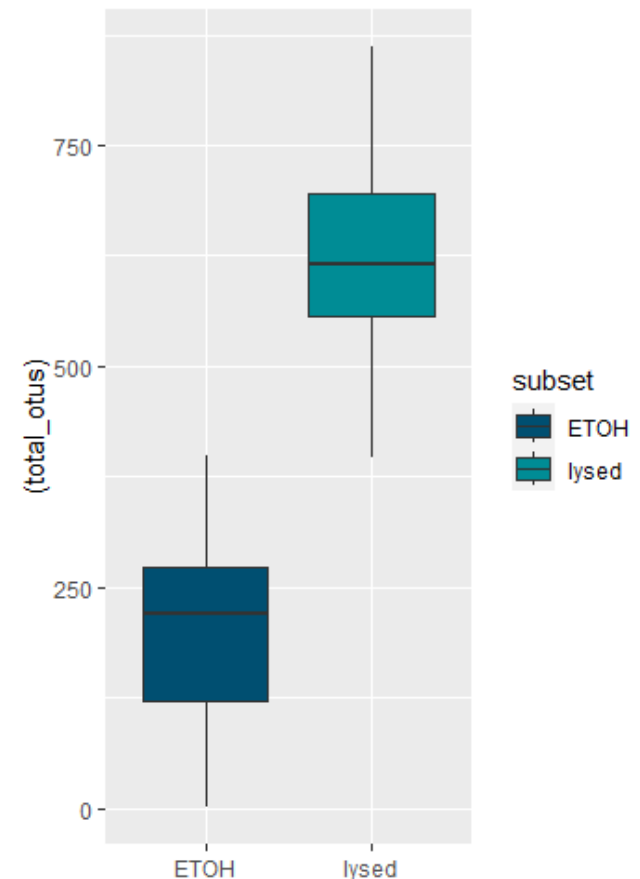
Designing a monitoring program

- Sufficient sequencing depth is vital in order to rival detection capacity of other methods
- NorIns relies on NovaSeq sequencing that generates 0.5 to 1 million sequences passing quality control per sample



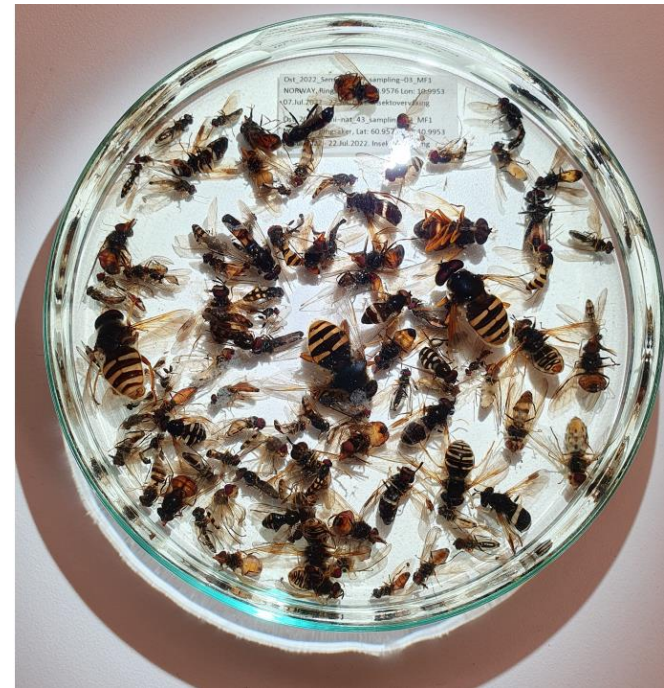
Designing a monitoring program

- DNA isolation methods can impact species recovery
- NorIns uses a 3 hour, minimally destructive soft lysis protocol that preserves sufficient morphological characters in hard bodied specimens to allow for later identification



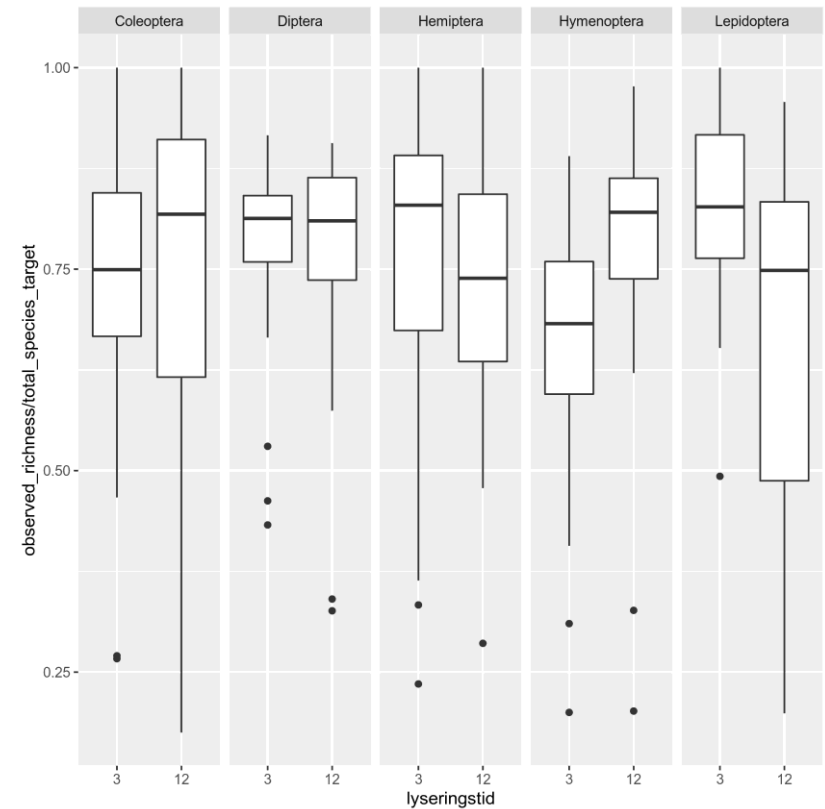
Designing a monitoring program

- DNA isolation methods can impact species recovery
- NorIns uses a 3 hour, minimally destructive soft lysis protocol that preserves sufficient morphological characters in hard bodied specimens to allow for later identification



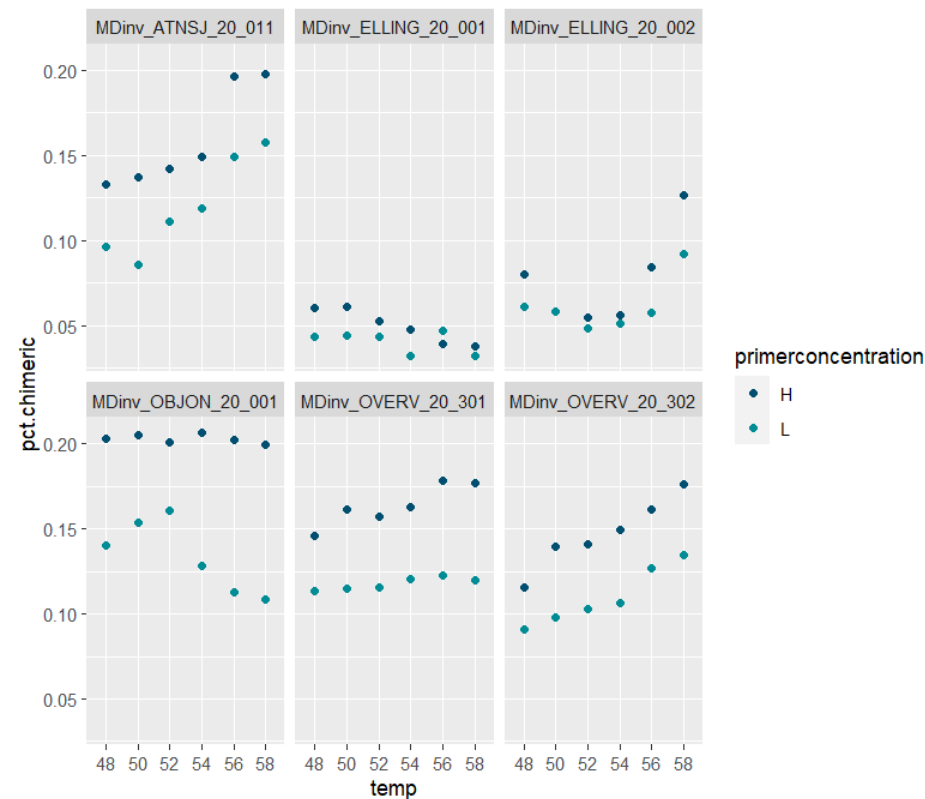
Designing a monitoring program

- DNA isolation method can impact species recovery
- Lysis time can impact recovery of hard vs. soft bodied taxa and large vs. small taxa

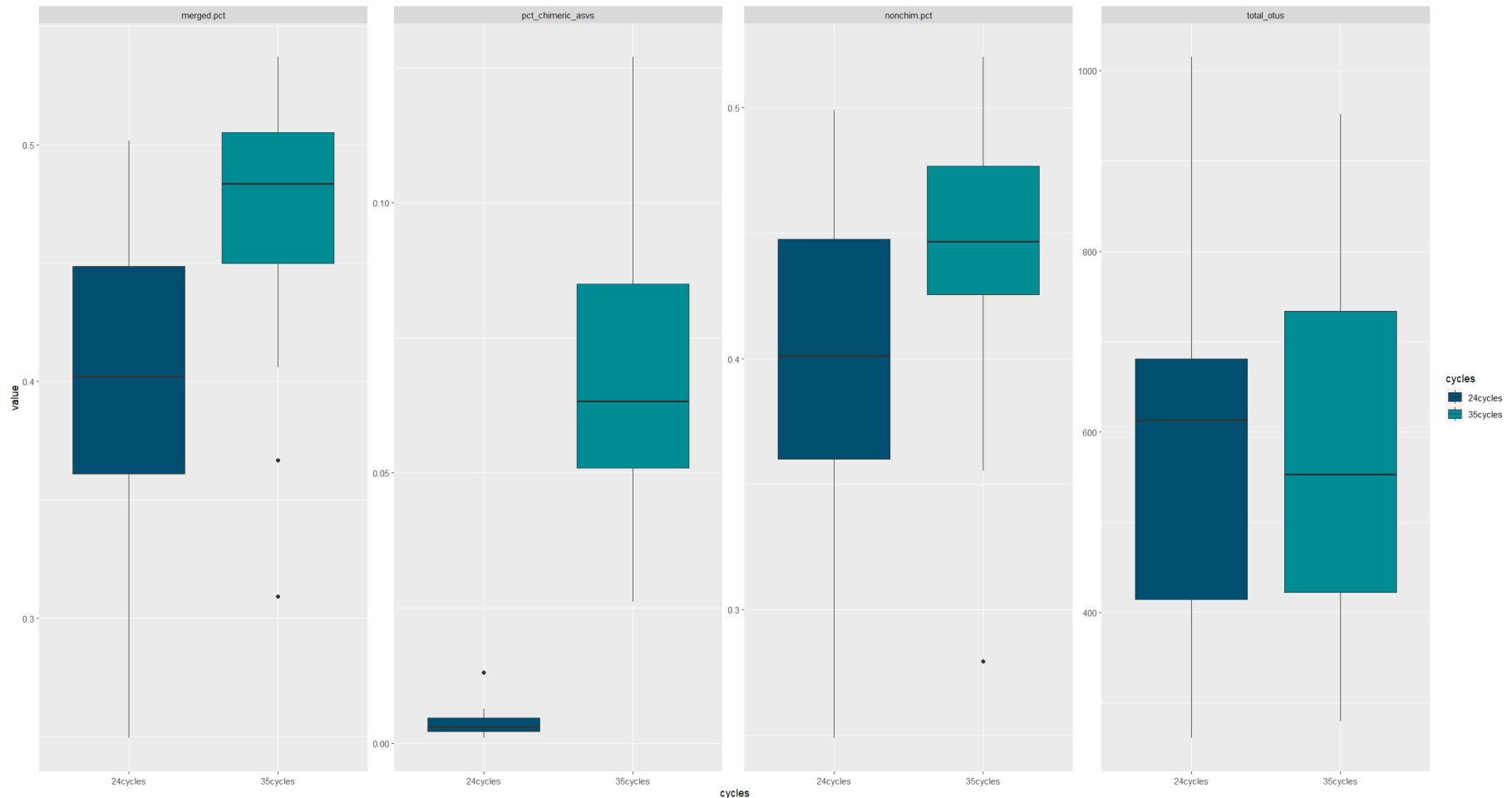


Designing a monitoring program

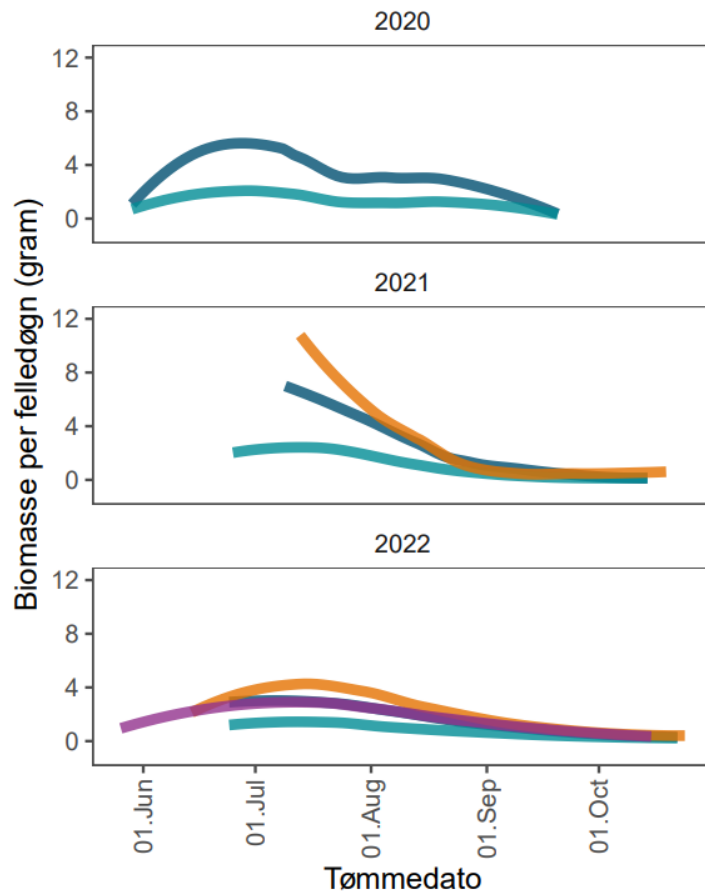
- PCR conditions need to be optimized, particularly for chimera formation
- Incomplete chimera removal likely leads to inflated alpha diversity estimates
- The solution is to avoid chimera formation in the first place



Designing a monitoring program

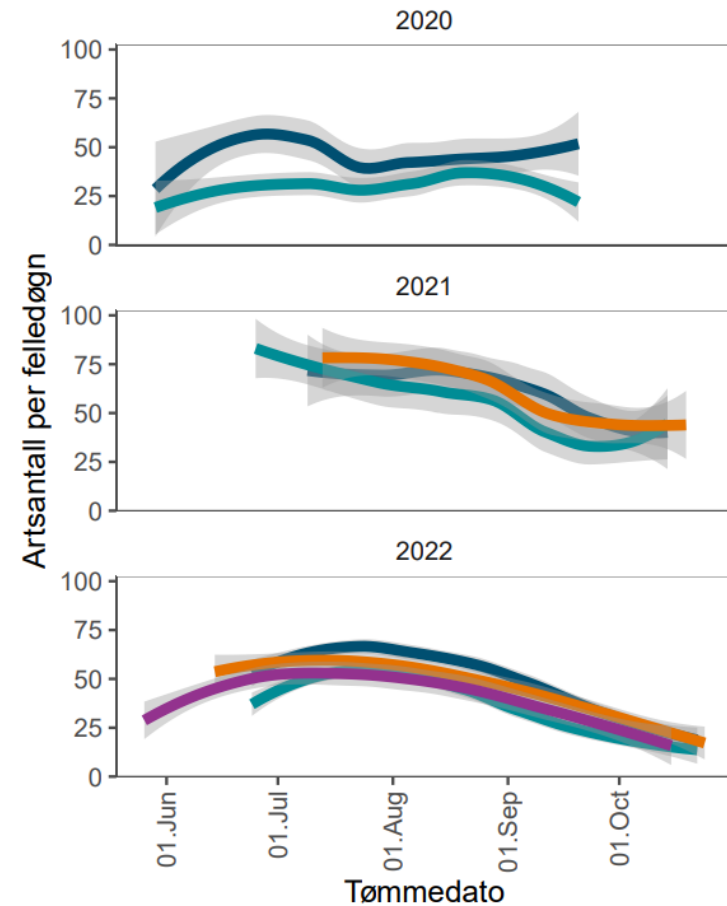


Three years of monitoring



Tidsserie

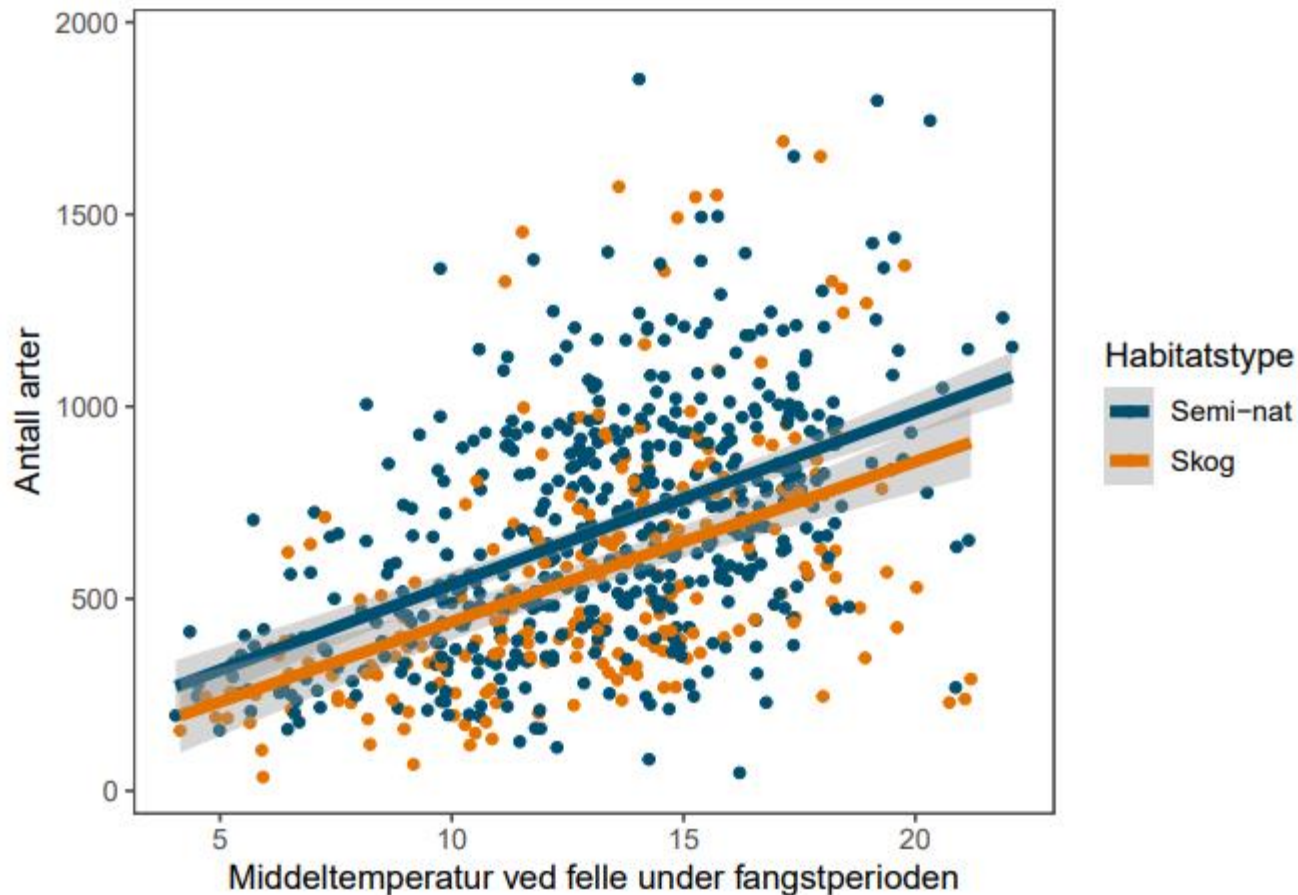
- Semi-nat_Østlandet
- Skog_Østlandet
- Semi-nat_Trøndelag
- Semi-nat_Sørlandet



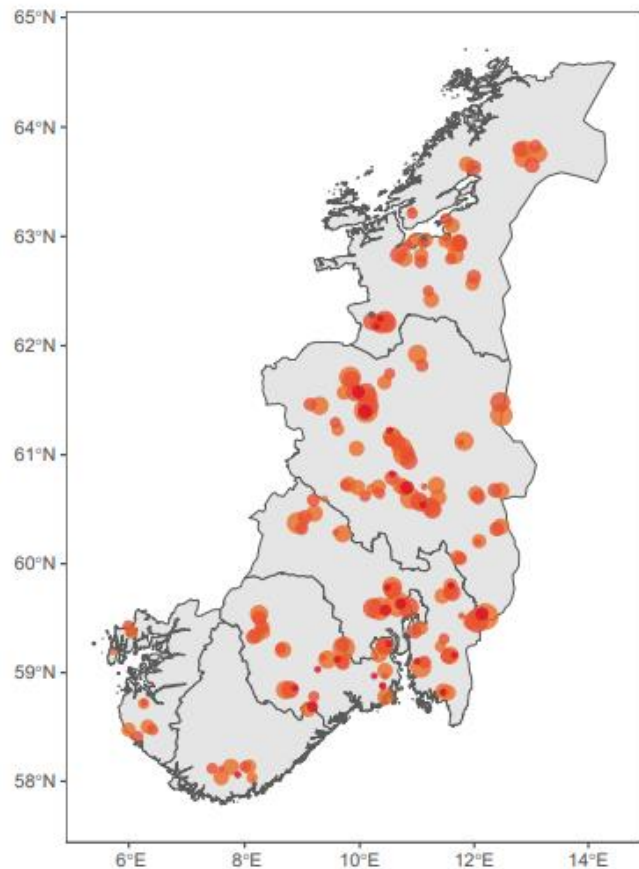
Tidsserie

- Semi-nat_Østlandet
- Skog_Østlandet
- Semi-nat_Trøndelag
- Semi-nat_Sørlandet

Three years of monitoring



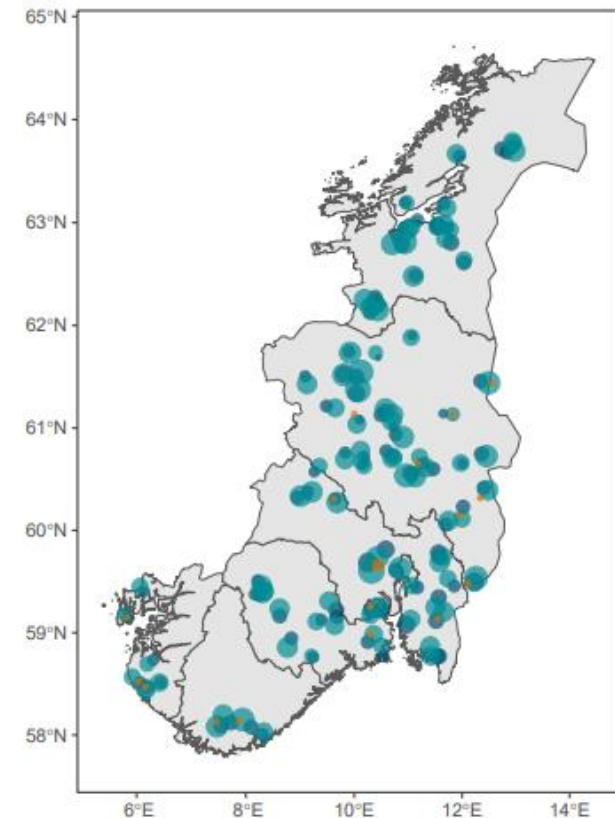
Three years of monitoring



Antall arter per kategori og lokalitet

• 1 • 5 • 10 • 15 • 20

Rødlistekategori NT (83 stk.) VU (56 stk.) EN (14 stk.) RE (1 stk.)



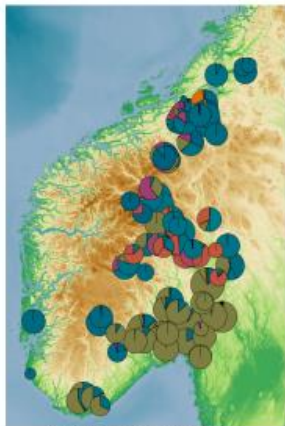
Antall arter per kategori og lokalitet

• 1 • 10 • 20 • 30 • 40 • 42

Kategori Fennoskandisk forek. (81 stk.) Potensielt fremmede arter (176 stk.) På fremmedartslista (10 stk.)

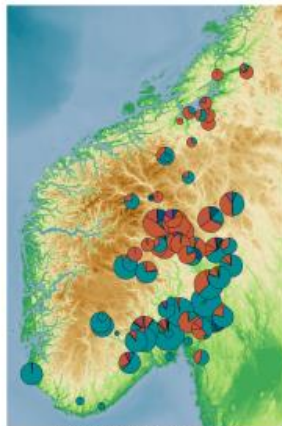
Beyond species lists: Genetic diversity

Sommerfugler



Celypha lacunana

Biller



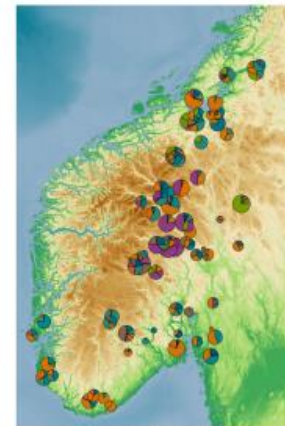
Malthodes fuscus

Tovinger

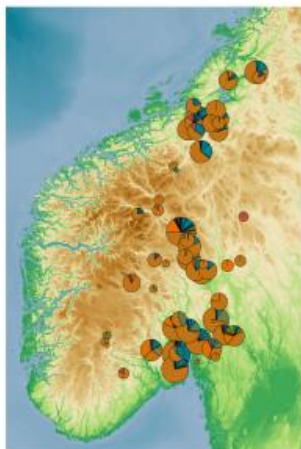


Psychoda phalaenoides

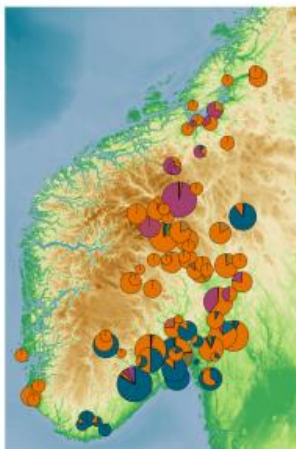
Vepser



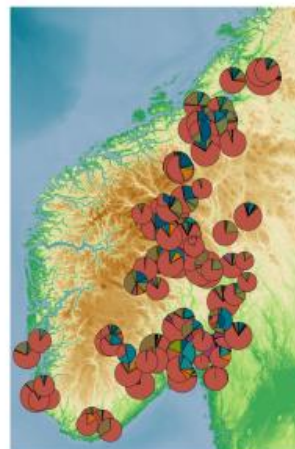
Aclastus minutus



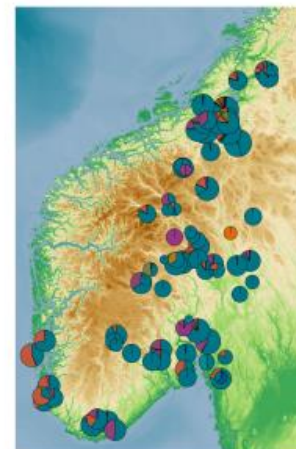
Phyllocnistis labyrinthella



Malthodes mysticus

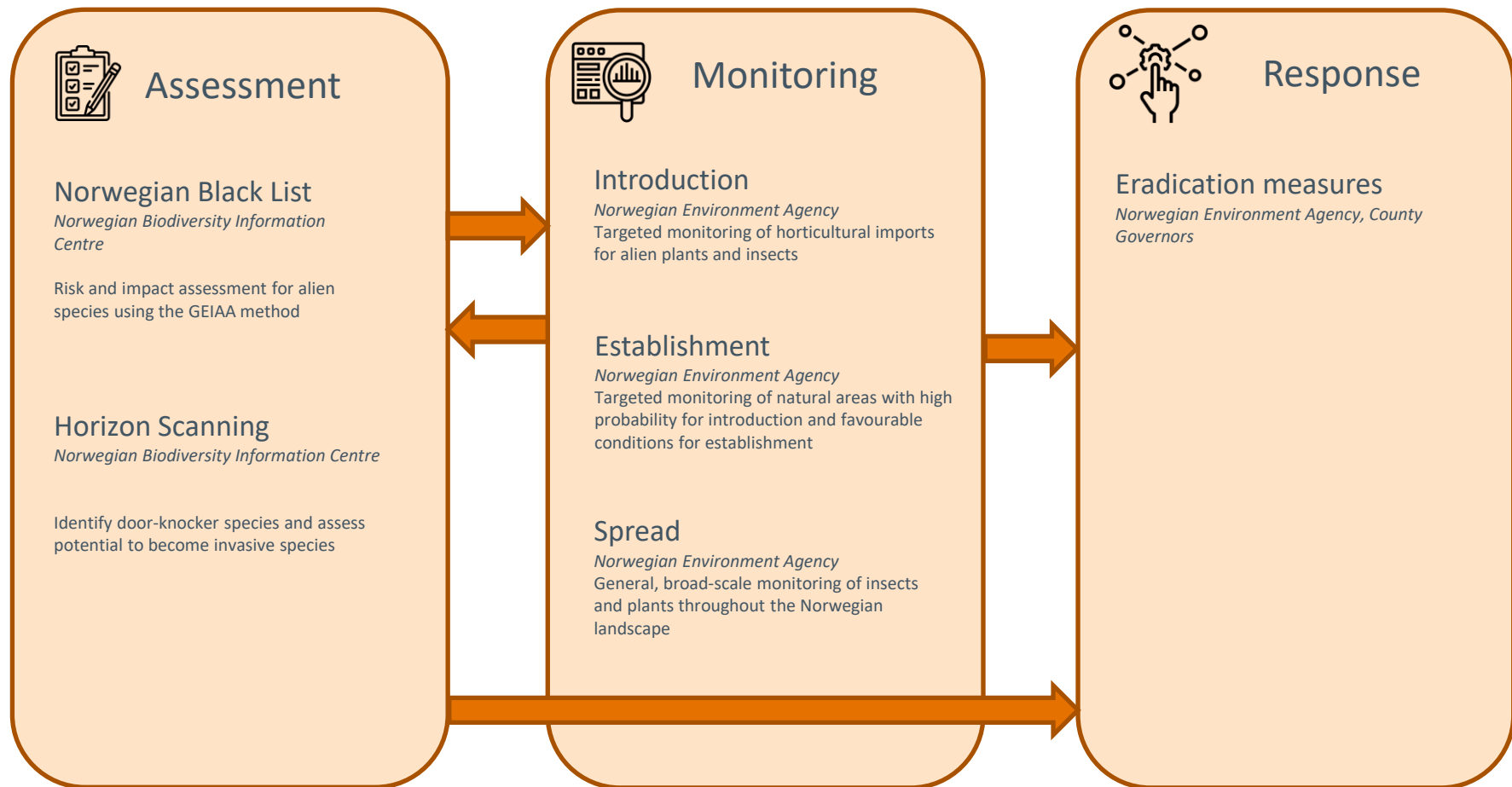


Scatopsiara atomaria

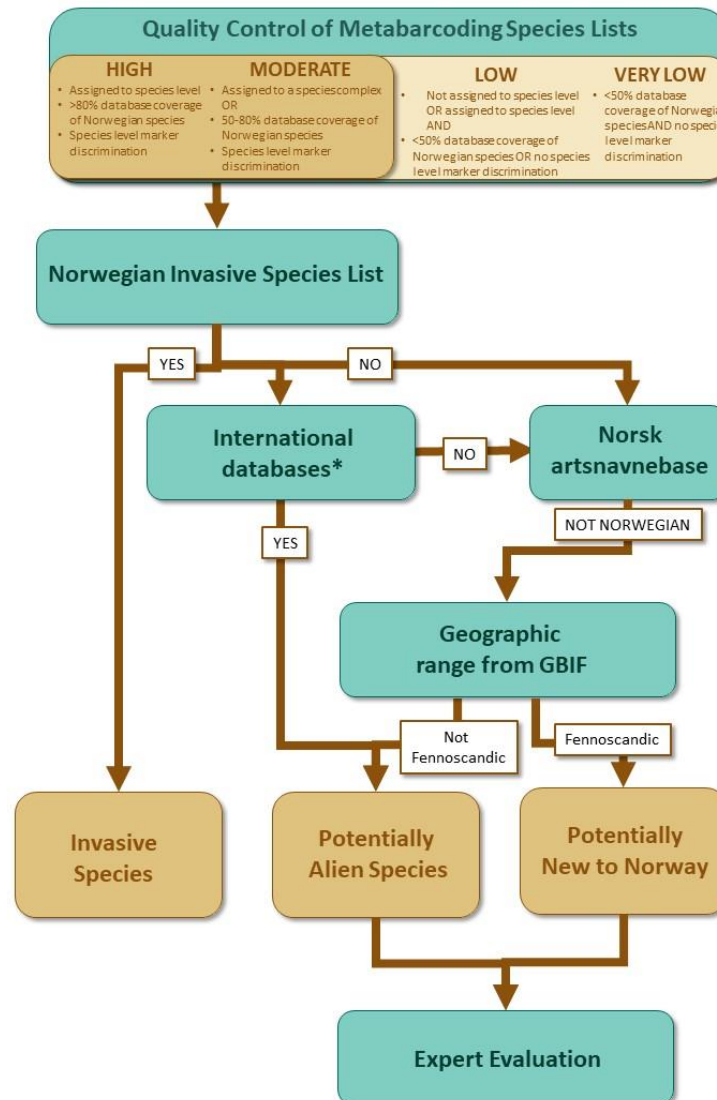


Sussaba cognata

Beyond species lists: Integrated Monitoring



Beyond species lists: Horizon scanning



Questions?

