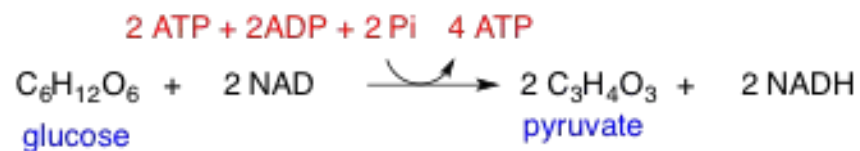


# Whole-cell models; Digital evolution

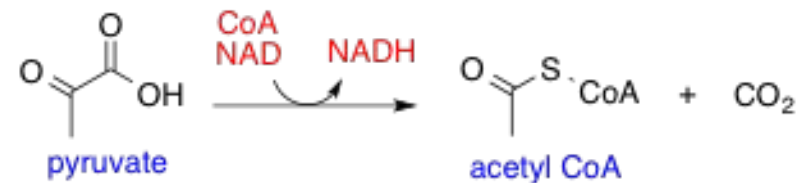
- Flux balance analysis
- Artificial life

# Metabolic networks

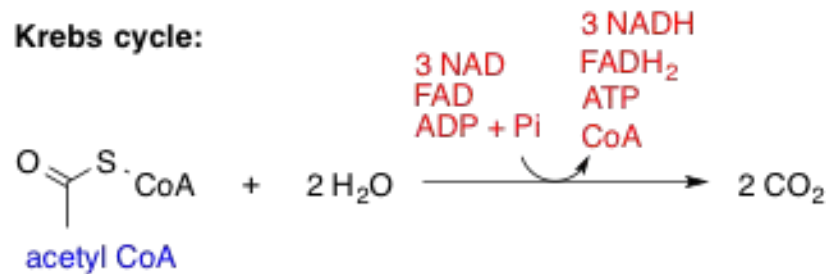
## Glycolysis:



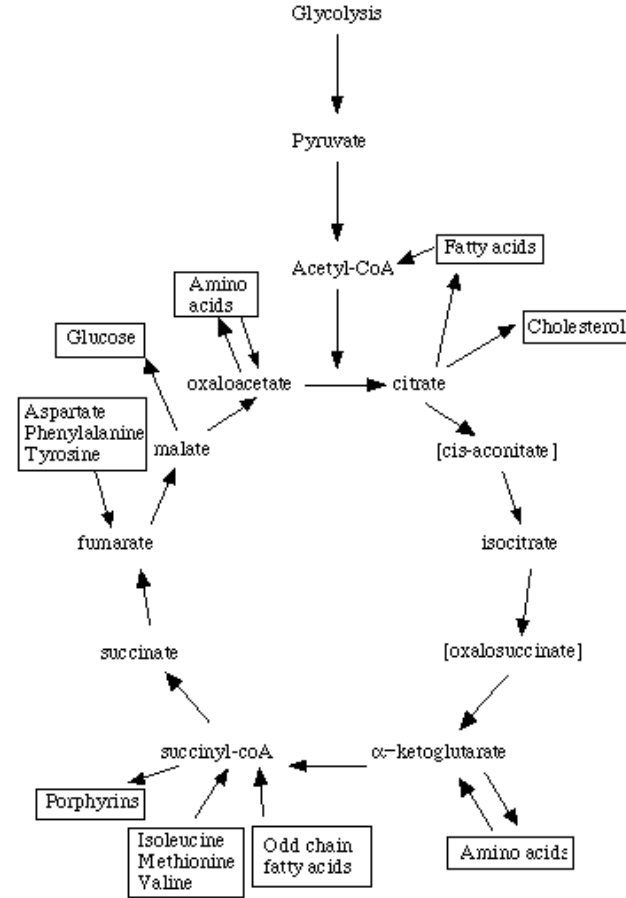
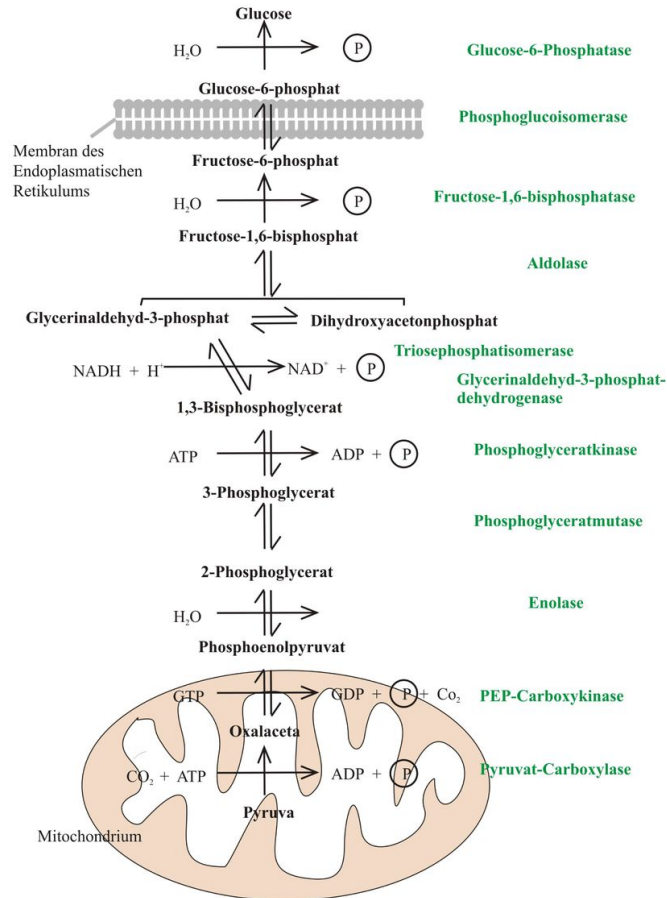
## Oxidative decarboxylation:



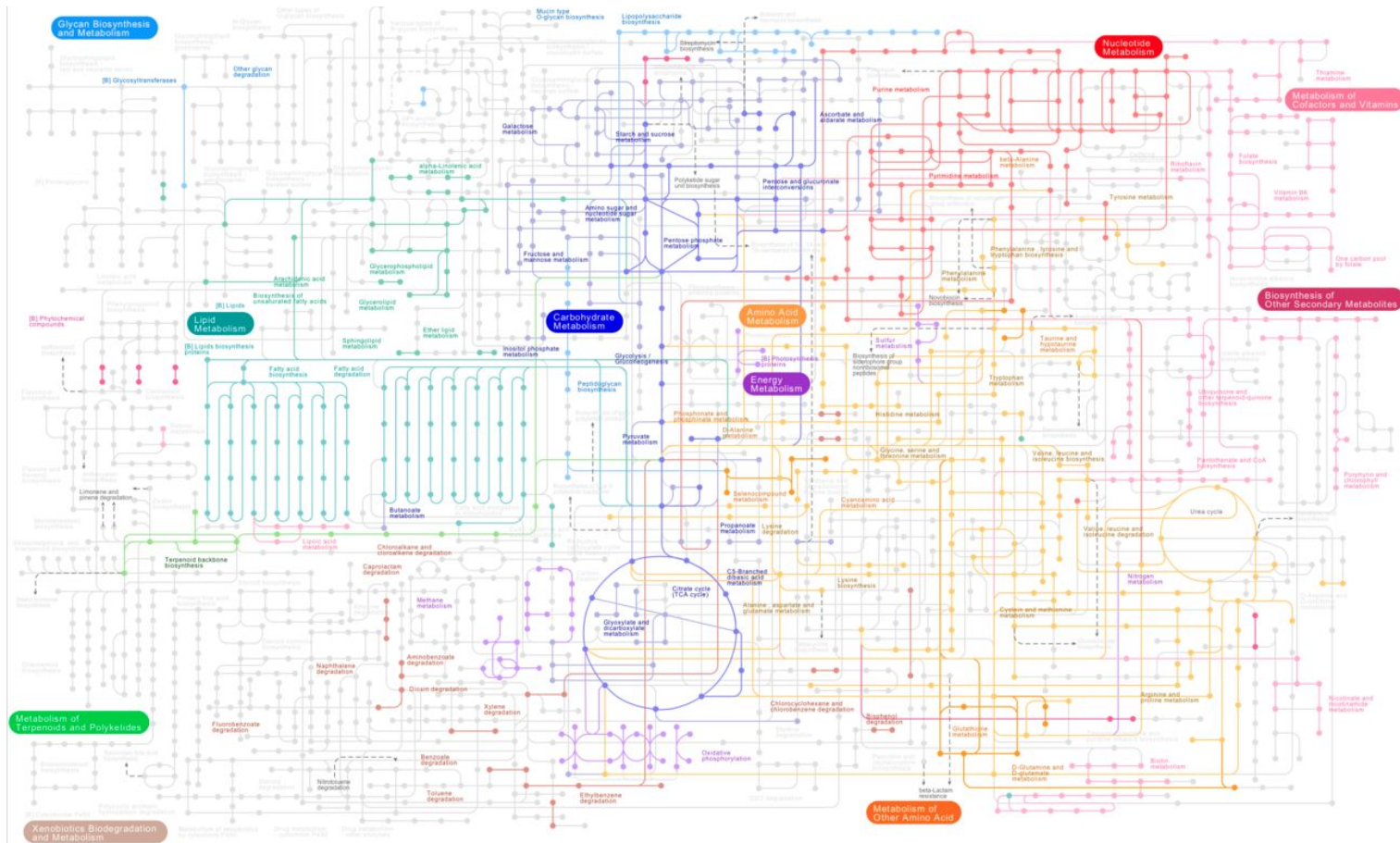
## Krebs cycle:



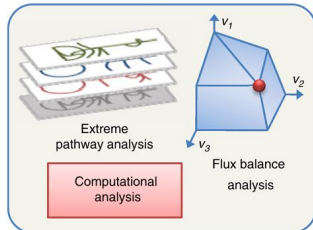
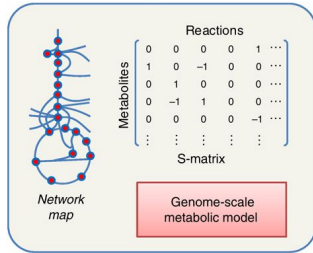
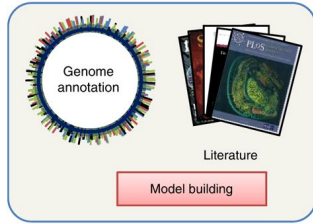
# Metabolic networks



# Metabolic networks



# Genome-scale metabolic network reconstruction & model



## Genome-scale metabolic network reconstruction:

- A collection of biochemical transformation derived from the genome annotation and the literature of a particular organism.
- Formed based on an organism-specific knowledge base.
- A network reconstruction is unique to an organism.

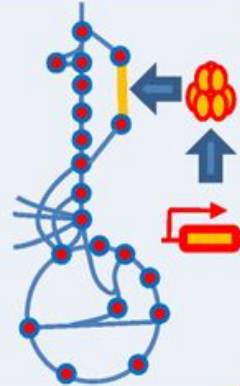
## Genome-scale metabolic network model:

- Derived from a *reconstruction* by converting it into a mathematical form (i.e., an in silico model) and by assessing its phenotypic properties computationally.

# Genome-scale metabolic network reconstruction & model

## 3. Directing hypothesis-driven discovery

A Metabolic GENRE aided in determining pathway usage and discovering a novel citramalate synthase gene in *G. sulfurreducens*. GENREs have also helped study the effects of transposons on downstream genes, and identify transcriptional timing patterns in *S. cerevisiae*.



## 2. Guidance of metabolic engineering

Metabolic GENREs guided efforts to engineer malate and succinate producing strains of *S. cerevisiae* and *M. succiniciproducens*. GENREs have also helped determine ways to increase the respiration rate of *G. sulfurreducens* and scale-up vaccine production against *N. meningitides*.



# Flux balance analysis (FBA)

FBA: metabolic network  $\rightarrow$  linear programming optimization problem.

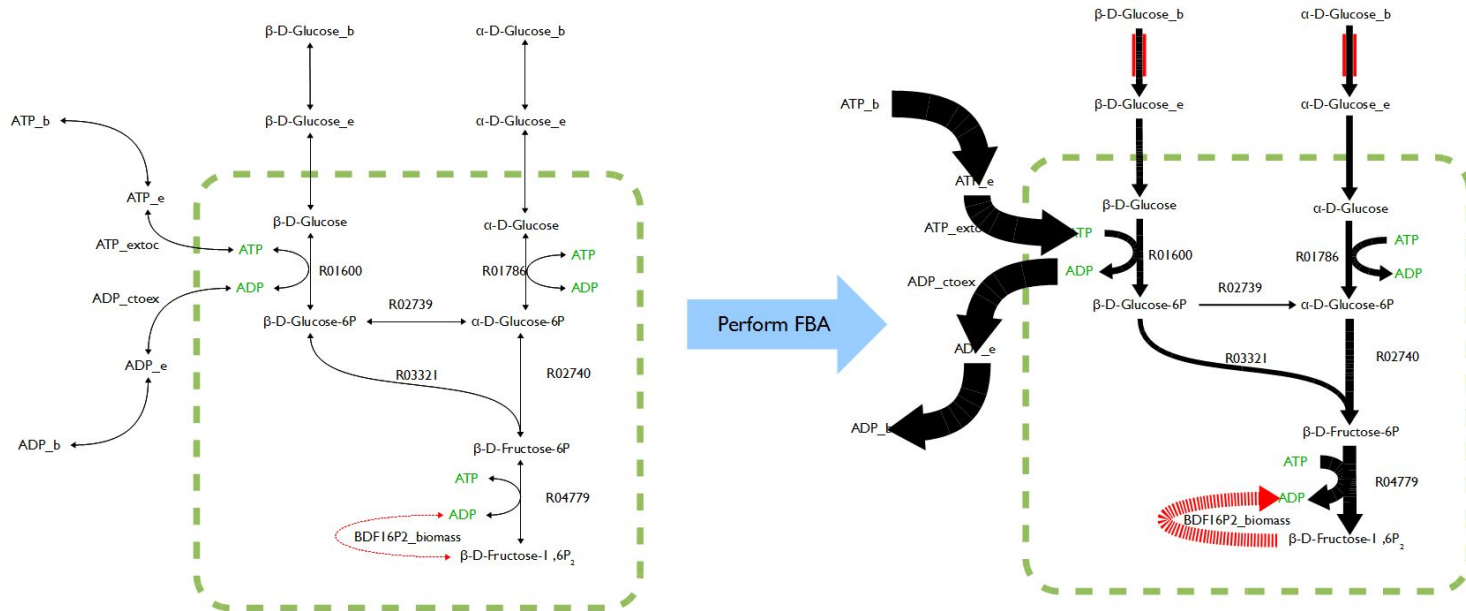
The main constraints in FBA: steady-state mass conservation of metabolites.

- Relies on balancing metabolic fluxes
- Is based on the fundamental law of mass conservation
- Is performed under steady-state conditions (an example of constraint...)
- Requires information only about:
  - a. the stoichiometry of metabolic pathways,
  - b. metabolic demands, and
  - c. a few strain specific parameters
- Does NOT require enzymatic kinetic data

# Flux balance analysis

The results of FBA on a metabolic network of the top six reactions of glycolysis.

- The predicted flux through each reaction is proportional to the width of the line.
- **Red springy arrow:** Objective function; **Red bars:** Constraints on  $\alpha$ -D-glucose and  $\beta$ -D-glucose import.

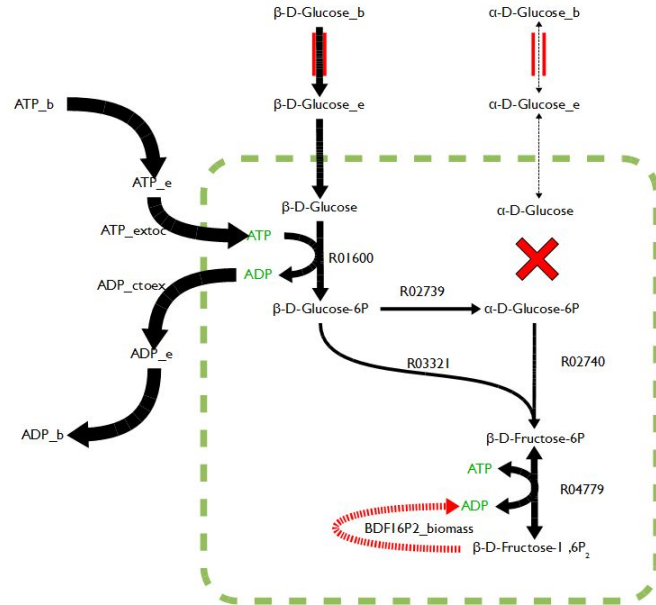




# Flux balance analysis

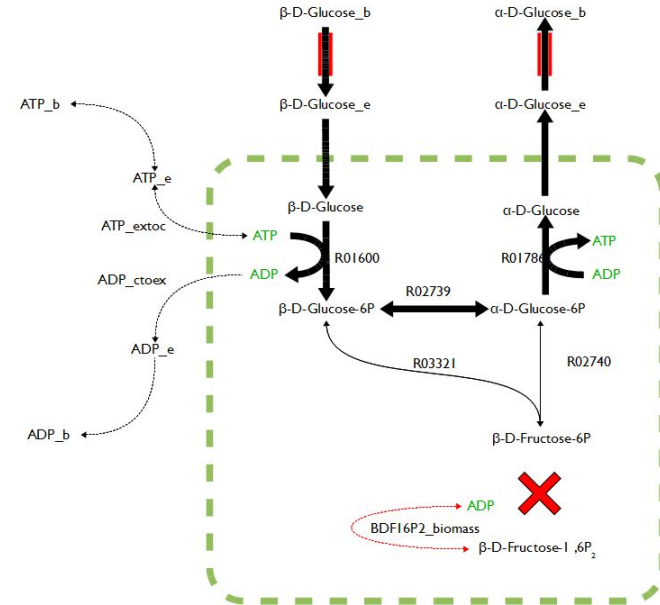
*Non-lethal gene deletion* in a metabolic network.

- Flux through the objective function is halved but is still present.



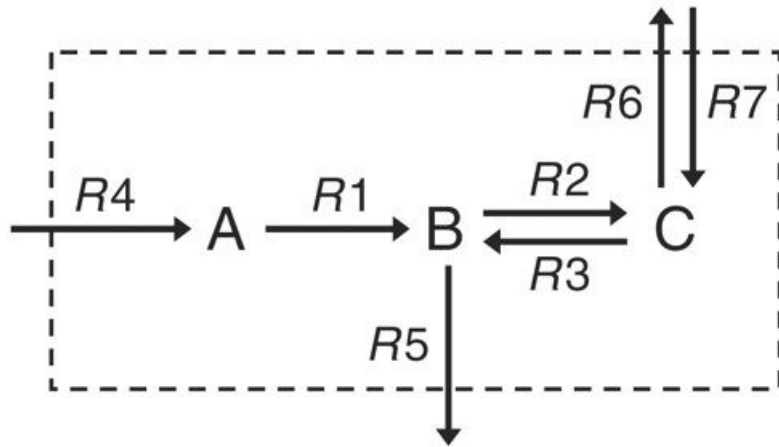
*Lethal gene deletion* in a metabolic network.

- No flux through the objective function  $\rightarrow$  pathway is no longer functional.



# Flux balance analysis

## 1. Reaction network formalism



### Chemical reactions

#### Internal

$R1: -1 A \rightarrow 1 B$

$R2: -1 B \rightarrow 1 C$

$R3: -1 C \rightarrow 1 B$

#### Exchange

$R4: 1 A$

$R5: -1 B$

$R6: -1 C$

$R7: 1 C$

$S =$

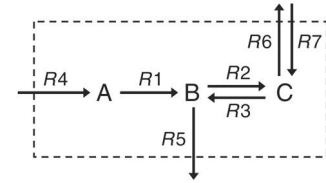
	$R1$	$R2$	$R3$	$R4$	$R5$	$R6$	$R7$
<b>A</b>	-1	0	0	1	0	0	0
<b>B</b>	1	-1	1	0	-1	0	0
<b>C</b>	0	1	-1	0	0	-1	1

# Flux balance analysis

## 1. Reaction network formalism

$S =$

	$R1$	$R2$	$R3$	$R4$	$R5$	$R6$	$R7$
<b>A</b>	-1	0	0	1	0	0	0
<b>B</b>	1	-1	1	0	-1	0	0
<b>C</b>	0	1	-1	0	0	-1	1



$$\begin{matrix} & R1 & R2 & R3 & R4 & R5 & R6 & R7 \\ \begin{matrix} A \\ B \\ C \end{matrix} & \begin{bmatrix} -1 & 0 & 0 & 1 & 0 & 0 & 0 \\ 1 & -1 & 1 & 0 & -1 & 0 & 0 \\ 0 & 1 & -1 & 0 & 0 & -1 & 1 \end{bmatrix} & * & \begin{bmatrix} v_1 \\ \vdots \\ v_7 \end{bmatrix} & = & ?
 \end{matrix}$$

# Flux balance analysis

## 2. FBA formulation

### Dynamic mass balance

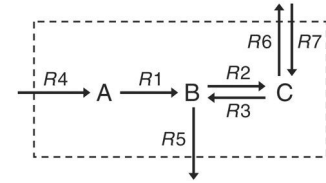
$$\frac{dC}{dt} = \mathbf{S}\mathbf{v}$$

$C$  : Concentration

$t$  : Time

$\mathbf{S}$  : Stoichiometric matrix

$\mathbf{v}$  : Flux vector



### Steady-state assumption

$$\mathbf{S}\mathbf{v} = \mathbf{0}$$

### LP formulation

Objective: max  $Z = \mathbf{c} \cdot \mathbf{v}$

Constraints:

$$\begin{array}{c} \text{A} \\ \text{B} \\ \text{C} \end{array} \begin{array}{c} R1 \\ R2 \\ R3 \\ R4 \\ R5 \\ R6 \\ R7 \end{array} \begin{bmatrix} -1 & 0 & 0 & 1 & 0 & 0 & 0 \\ 1 & -1 & 1 & 0 & -1 & 0 & 0 \\ 0 & 1 & -1 & 0 & 0 & -1 & 1 \end{bmatrix} \begin{bmatrix} v_1 \\ \vdots \\ v_7 \end{bmatrix} = \mathbf{0}$$

$$0 \leq v_1, \dots, v_7 \leq 10$$

# Flux balance analysis: Objective function

**Objective function:** Physiologically-meaningful or design-based objective for the interrogation or exploitation of a given system.

Examples:

- Maximizing...
  - biomass or cell growth
  - maximizing ATP production
  - maximizing the rate of synthesis of a particular product
- Minimizing...
  - ATP production
  - nutrient uptake (both to determine conditions of optimal metabolic energy efficiency)

# Flux balance analysis: Constraints

No. of equations (one per reactant)  $\ll$  no. of unknown variables (reaction fluxes).

- An *under-determined* set of linear equations.
- Therefore, optimize fluxes given cellular objective given a bunch of constraints.
- **Principal constraint:** mass balance
- **Additional constraints:**
  - physico-chemical constraints
  - spatial or topological constraints
  - condition dependent environmental constraints
  - regulatory constraints
- All constraints together represent a set of linear equations.

# Flux balance analysis

## II. FBA formulation

### Dynamic mass balance

$$\frac{dC}{dt} = Sv$$

$C$  : Concentration

$t$  : Time

$S$  : Stoichiometric matrix

$v$  : Flux vector

### Steady-state assumption

$$Sv = 0$$

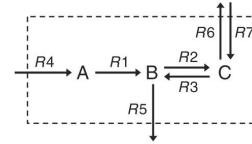
### LP formulation

Objective: max  $Z = c \cdot v$

Constraints:

$$\begin{matrix} & R1 & R2 & R3 & R4 & R5 & R6 & R7 \\ \begin{matrix} A \\ B \\ C \end{matrix} & \begin{bmatrix} -1 & 0 & 0 & 1 & 0 & 0 & 0 \\ 1 & -1 & 1 & 0 & -1 & 0 & 0 \\ 0 & 1 & -1 & 0 & 0 & -1 & 1 \end{bmatrix} & \begin{bmatrix} v_1 \\ \vdots \\ v_7 \end{bmatrix} \end{matrix} = 0 \quad 0 \leq v_1, \dots, v_7 \leq 10$$

### I. Reaction network formalism

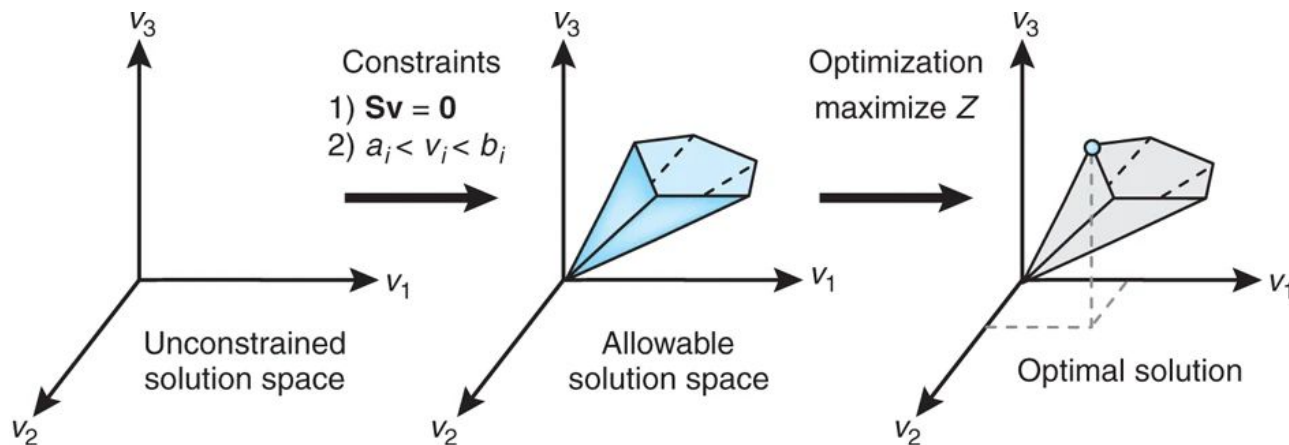


Chemical reactions	
Internal	Exchange
R1: -1 A → 1 B	R4: 1 A
R2: -1 B → 1 C	R5: -1 B
R3: -1 C → 1 B	R6: -1 C
	R7: 1 C

→ S =

	R1	R2	R3	R4	R5	R6	R7
A	-1	0	0	1	0	0	0
B	1	-1	1	0	-1	0	0
C	0	1	-1	0	0	-1	1

# Constraint-based modeling



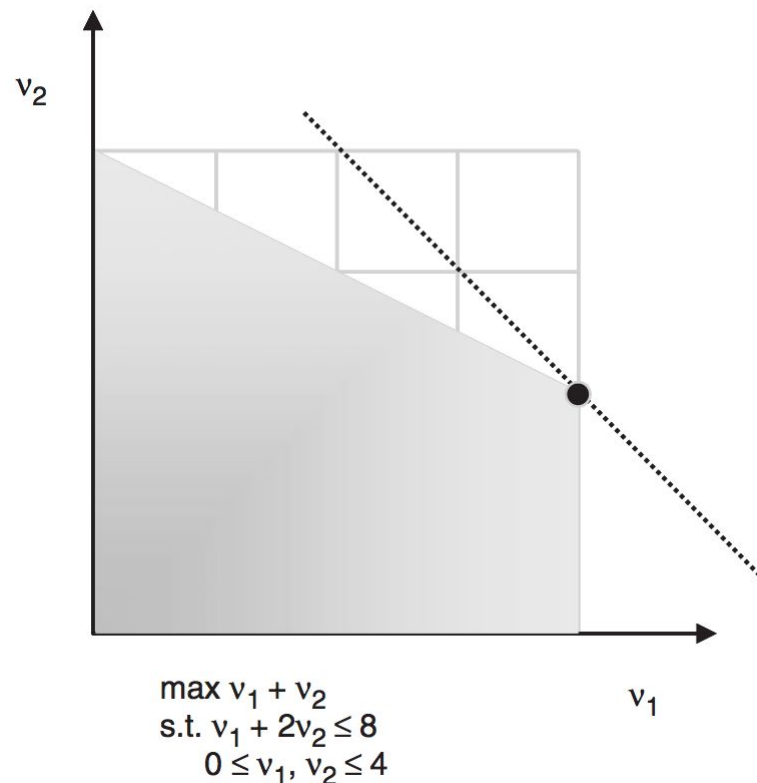
1. No constraints: flux may lie at any point in solution space.
2. Mass balance constraints (imposed by the stoichiometry) and capacity constraints (imposed by the lower and upper bounds:  $a_i$  &  $b_i$ ): defines allowable solution space.
  - a. Any flux distribution within this space is allowable; Points outside this space are denied
3. Optimization of an objective function: A single optimal flux distribution that lies on the edge of the allowable solution space.



# Constraint-based modeling

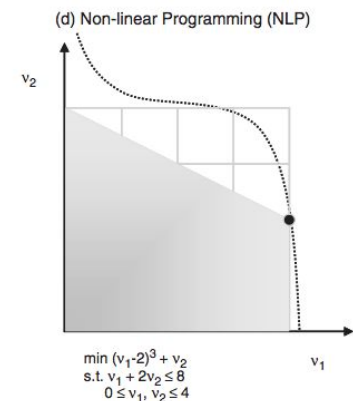
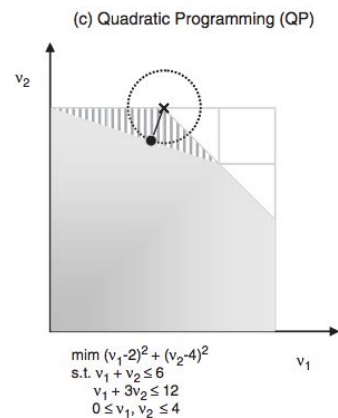
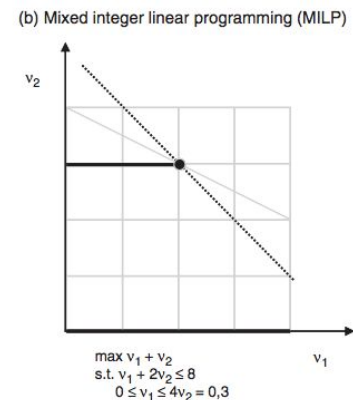
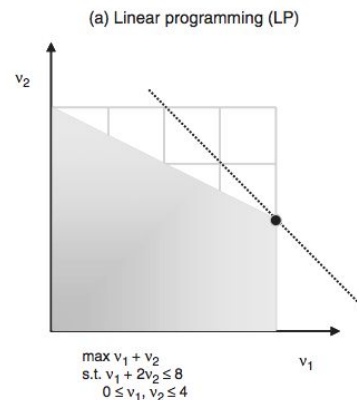
## Linear programming

- Feasible solution space:
  - shaded area and solid lines
  - defined by:
    - flux capacities,
    - stoichiometric relationships, and
    - design specification (e.g. gene deletions).
- Objective function: dotted line
- Optimal solution: circular dot



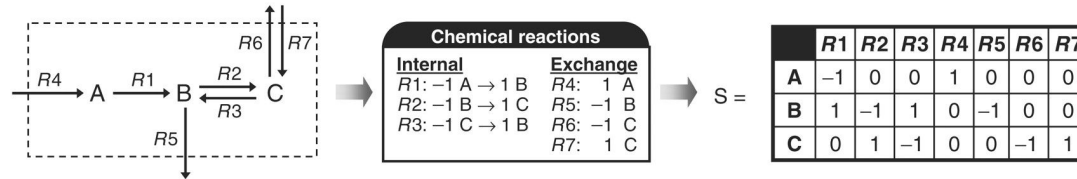
# Constraint-based modeling

- Mixed Integer LP (MILP):
  - Integer variables are involved in a linear programming problem (e.g. binary variable formulation for gene deletion).
- Quadratic programming (QP):
  - Quadratic objective function subject to linear constraints.
  - This technique is generally used for finding the closest point to a specified point.
- Nonlinear programming (NLP):
  - Nonlinear objectives or constraints.
  - Generally difficult to solve for global optimal solution because of its non-convexity.



# Flux balance analysis

## I. Reaction network formalism



## II. FBA formulation

### Dynamic mass balance

$$\frac{dC}{dt} = Sv$$

**C** : Concentration  
**t** : Time  
**S** : Stoichiometric matrix  
**v** : Flux vector

### Steady-state assumption

$$Sv = 0$$

### LP formulation

Objective: max  $Z = v_5$

Constraints:

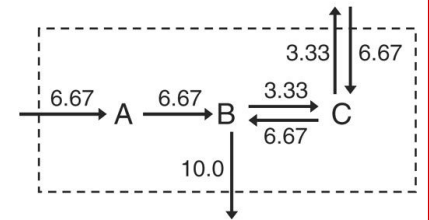
$$\begin{matrix} A \\ B \\ C \end{matrix}
 \begin{bmatrix} R1 & R2 & R3 & R4 & R5 & R6 & R7 \\ -1 & 0 & 0 & 1 & 0 & 0 & 0 \\ 1 & -1 & 1 & 0 & -1 & 0 & 0 \\ 0 & 1 & -1 & 0 & 0 & -1 & 1 \end{bmatrix}
 \begin{bmatrix} v_1 \\ \vdots \\ v_7 \end{bmatrix} = 0$$

$0 \leq v_1, \dots, v_7 \leq 10$

## III. Hypothetical flux distribution at steady-state

$$Z = 10$$

$$v = [6.67 \ 3.33 \ 6.67 \ 6.67 \ 10.0 \ 3.33 \ 6.67]^T$$



# Genome-scale metabolic network reconstruction & model

Organism	Strain	Genes	Version	GR	Mets	Rxns	Comp
<i>Bacillus subtilis</i>		4,225	model_v3	844	988	1,020	2 (c,e)
<i>Escherichia coli</i>	K12 MG1655	4,405	iAF1260	1,260	1,039	2,077	3 (c,e,p)
<i>Helicobacter pylori</i>	26695	1,632	iIT341	341	485	476	2 (c,e)
<i>Pseudomonas putida</i>	KT2440	5,350	iNJ746	746	911	950	3 (c,p,e)
<i>Pseudomonas putida</i>	KT2440	5,350	iJP815	815	886	877	2 (c,e)
<i>Pseudomonas aeruginosa</i>	PA01	5,640	iM01056	1,056	760	883	2 (c,e)
<i>Mycoplasma genitalium</i>	G-37	521	iPS189	189	274	262	2 (c,e)
<i>Lactobacillus plantarum</i>	WCFS1	3,009		721	531	643	2 (c,e)
<i>Streptomyces coelicolor</i>	A3(2)	8,042		700	500	700	2 (c,e)
<i>Leishmania major</i>	Friedlin	8,370	iAC560	560	1,101	1,112	8 (a,f,y,c,e,m,r,n)
<i>Saccharomyces cerevisiae</i>	Sc288	6,183	iMM904	904	713	1,412	8 (c,e,m,x,n,r,v,g)
<i>Homo sapiens</i>		28,783	Recon 1	1,496	2,766	3,311	8 (c,e,m,x,n,r,v,g)

# Constructing a genome-scale metabolic model

Conversion of reconstruction into a model

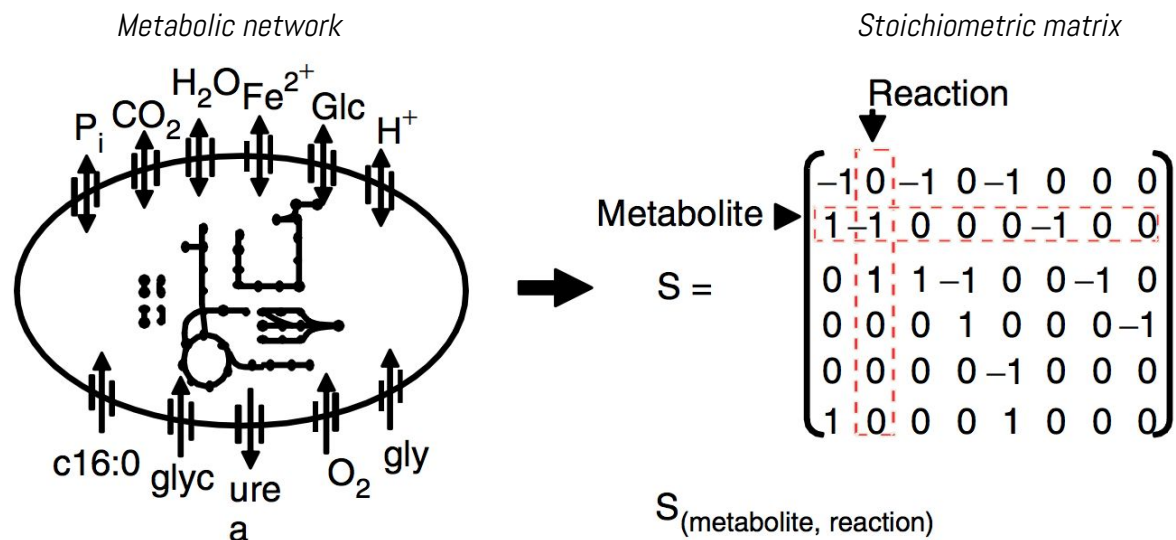
## 3. Conversion of reconstruction into computable format

- 38| Initialize the COBRA toolbox.
- 39| Load reconstruction into Matlab.
- 40| Verify S matrix.
- 41| Set objective function.
- 42| Set simulation constraints.

# Constructing a genome-scale metabolic model

## Conversion of reconstruction into a model

### Mathematical representation



By definition:

- Substrates have negative coefficients (i.e., they are consumed)
- Products have positive coefficients (i.e., they are produced)

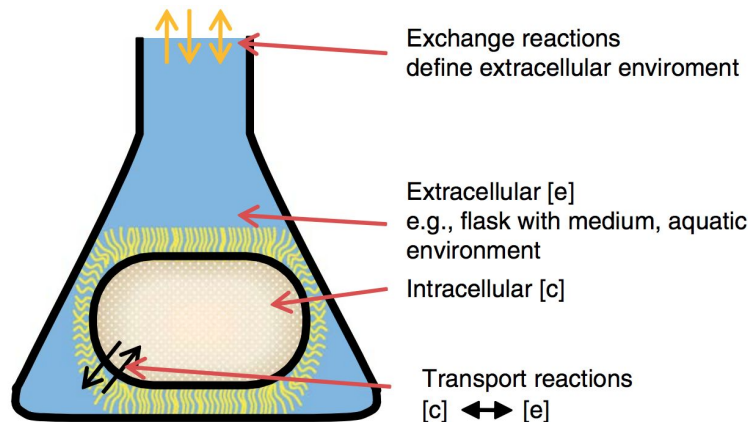
$\mathbf{v}$  is a vector of reaction fluxes

Conservation of mass: All steady states can be described by  $\mathbf{S}\mathbf{v} = 0$

# Constructing a genome-scale metabolic model

## Conversion of reconstruction into a model

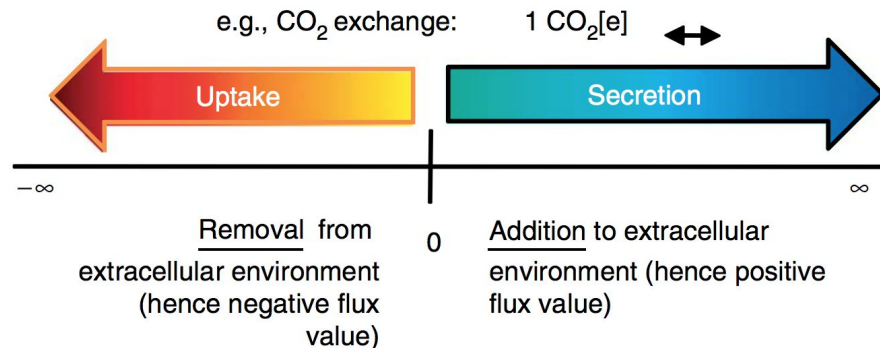
### Definition of system boundaries



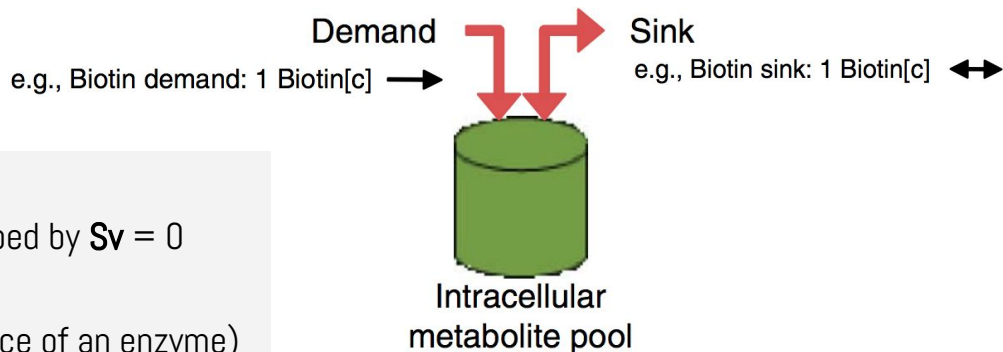
### Constraints:

- Mass conservation: all steady states can be described by  $\mathbf{Sv} = 0$
- Thermodynamics (reaction directionality)
- Enzyme capacity or regulation (i.e., presence/absence of an enzyme)

### Exchange reactions



### Demand/sink reactions



# Constructing a genome-scale metabolic model

Network evaluation  $\approx$  Debugging

## 4. Network evaluation

- 43–44| Test if network is mass-and charge balanced.
- 45| Identify metabolic dead-ends.
- 46–48| Perform gap analysis.
- 49| Add missing exchange reactions to model.
- 50| Set exchange constraints for a simulation condition.
- 51–58| Test for stoichiometrically balanced cycles.
- 59| Re-compute gap list.
- 60–65| Test if biomass precursors can be produced in standard medium.
- 66| Test if biomass precursors can be produced in other growth media.
- 67–75| Test if the model can produce known secretion products.
- 76–78| Check for blocked reactions.
- 79–80| Compute single gene deletion phenotypes.
- 81–82| Test for known incapacilities of the organism.
- 83| Compare predicted physiological properties with known properties.
- 84–87| Test if the model can grow fast enough.
- 88–94| Test if the model grows too fast.



# Constructing a genome-scale metabolic model

## Network evaluation $\approx$ Debugging

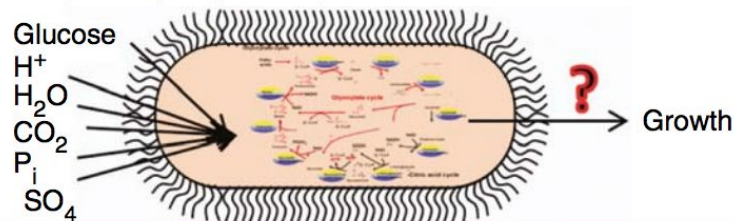
### Analysis of biomass precursors synthesis

- Biomass precursors = cellular growth requirements
- Pathways to synthesize precursors must be complete (i.e., functional) for the network to simulate growth
- Testing synthesis of each separate biomass precursor is part of the debugging process



### Analysis of growth in minimal medium

- Minimal medium is defined for many organisms and can be found in primary literature
- Contains at least 1 C-, N-, S- and P-source
- Auxotrophs may require the presence of addition metabolites



### Test for growth on known carbon sources

- Exchange reactions define medium and environment
- Transport reactions allow network to consume carbon sources
- Biodegradative pathways that are required for carbon utilization

### Secretion capability

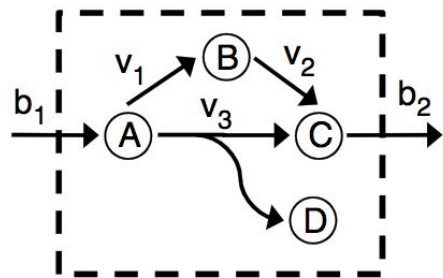
- Transport and exchange reactions are required in reconstruction to enable secretion
- Secretion may only occur under certain circumstances (e.g., D-lactic acid formation under anoxic conditions)
- Comparison with known secretion pattern of multiple metabolites (e.g., secretion of a certain ratio of  $CO_2$  and acetate)

# Constructing a genome-scale metabolic model

# Network evaluation $\asymp$ Debugging

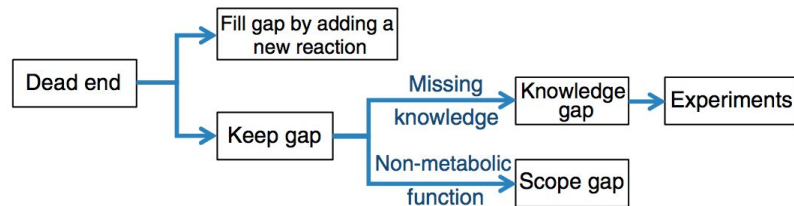
## Identifying gaps

### Connectivity based (topology):

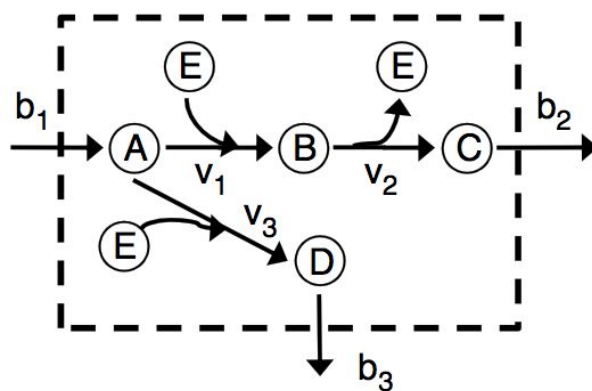


	$v_1$	$v_2$	$v_3$	$b_1$	$b_2$
A	-1	0	-1	1	0
B	1	-1	0	0	0
C	0	1	1	0	-1
D	0	0	1	0	0

## Dead end



## Functionality based (computation)

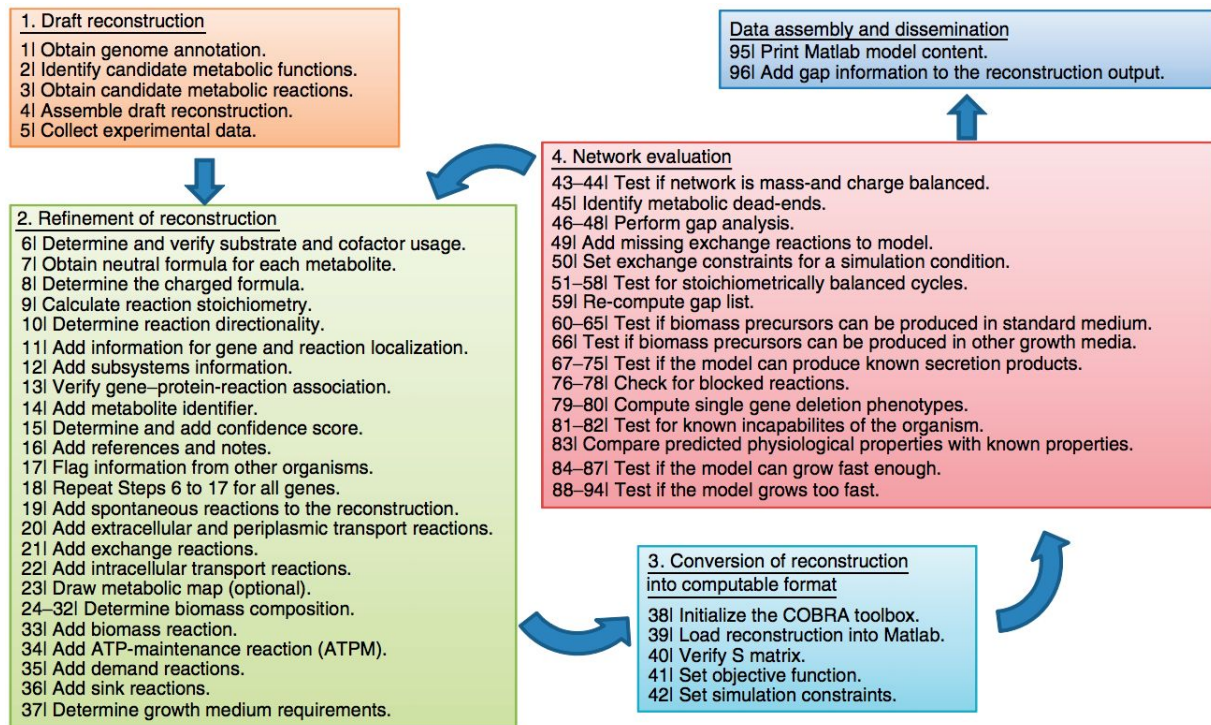


	$v_1$	$v_2$	$v_3$	$b_1$	$b_2$	$b_3$
A	-1	0	-1	1	0	0
B	1	-1	0	0	0	0
C	0	1	0	0	-1	0
D	0	0	1	0	0	-1
E	-1	1	-1	0	0	0

# Constructing a genome-scale metabolic model

Procedure to iteratively  
reconstruct metabolic networks.

- Iterate stages 2–4 are continuously...
- ...until model predictions are similar to the phenotypic characteristics of the target organism and/or all experimental data for comparison are exhausted.





# Genome-scale metabolic network reconstruction & model

## 1. Contextualization of HT data

Several studies have overlaid gene microarray data on a metabolic GENRE to determine condition-dependent cell phenotypes. Metabolic GENREs have also been used to interpret metabolomic data,  $^{13}\text{C}$  flux data, and to link multiple data types.



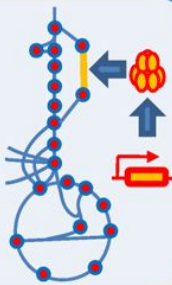
## 2. Guidance of metabolic engineering

Metabolic GENREs guided efforts to engineer malate and succinate producing strains of *S. cerevisiae* and *M. succiniciproducens*. GENREs have also helped determine ways to increase the respiration rate of *G. sulfurreducens* and scale-up vaccine production against *N. meningitidis*.



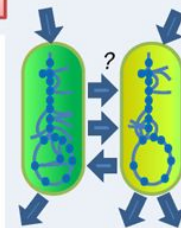
## 3. Directing hypothesis-driven discovery

A Metabolic GENRE aided in determining pathway usage and discovering a novel citramalate synthase gene in *G. sulfurreducens*. GENREs have also helped study the effects of transposons on downstream genes, and identify transcriptional timing patterns in *S. cerevisiae*.



## 4. Interrogation of multi-species relationships

A dual-species metabolic model was built to study interactions between the syntrophic bacteria, *D. vulgaris* and *M. maripaludis*. Metabolic models have also been used in comparisons of multiple species, such as an analysis of pathway differences between four halophilic bacteria.



## 5. Network property discovery

Metabolic GENREs have been used to study metabolite connectivity, and pathway redundancy *in silico*. Pathway-analysis tools have also spawned techniques such as flux coupling analysis, which has helped identify novel drug targets in *M. tuberculosis*.

