



Review

The molecular physiology of nuclear retinoic acid receptors. From health to disease[☆]

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ABSTRACT

The nuclear retinoic acid (RA) receptors (RAR α , β and γ) are transcriptional transregulators, which control the expression of specific gene subsets subsequently to ligand binding and to strictly controlled phosphorylation processes. Consequently RARs maintain homeostasis through the control of cell proliferation and differentiation. Today, it is admitted that, analogous to the paradigm established by the hematopoietic system, most adult tissues depict a differentiation hierarchy starting from rare stem cells. Here we highlight that the integrity of RARs is absolutely required for homeostasis in adults. Indeed, strictly controlled levels of RARs are necessary for the correct balance between self-renewal and differentiation of tissue stem cells. In addition, loss, accumulation, mutations or aberrant modifications of a specific RAR lead to uncontrolled proliferation and/or to differentiation block and thereby to cancer. This article is part of a Special Issue entitled: Translating nuclear receptors from health to disease.

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1. Introduction

Nuclear retinoic acid (RA) receptors (RARs) consist of three subtypes, α (NR1B1), β (NR1B2) and γ (NR1B3), encoded by separate genes. For each subtype, there are at least 2 isoforms, which are generated by differential promoter usage and alternative splicing and differ only in their N-terminal regions. RARs are activated by retinoids, which cover natural vitamin A metabolites exemplified by retinoic acid and synthetic active analogs [1].

RARs function as ligand-dependent transcriptional regulators, heterodimerized with retinoid X receptors (RXRs), which also consist of three types, α NR2B1, β (NR2B2) and γ (NR2B3) [2–4]. As such they regulate the expression of subsets of target genes involved in cellular differentiation, proliferation and apoptosis [5–8]. Consequently, RARs play critical roles in a variety of biological processes, including development, reproduction, immunity, organogenesis and homeostasis, as assessed by vitamin A-deficiency (VAD), pharmacological and genetic studies conducted in the mouse [9–13].

As a general trend, the RAR α subtype has ubiquitous or quite widespread expression patterns. In contrast, the other RARs (RAR β and RAR γ) show complex tissue specific expression [11]. The generation of germline mutations of the RAR and RXR coding genes in mice by homologous recombination [12] provided powerful tools to study the role of specific RAR/RXR heterodimers during embryonic patterning and organogenesis. However the determination of their functions in a defined adult cell type/tissue or at a given time of the adult life by this strategy was limited, due to *in utero* lethality or arrest of the

development of a given organ at an early stage. To overcome these limitations, strategies for spatio-temporally controlled somatic mutagenesis in mice have been designed [14] and provided invaluable models to elucidate some physiological functions of RARs and RXRs in adults. Such a strategy has been already applied to skin and liver [15,16], which have the ability to proliferate during the whole adult life. It has been also applied to hematopoietic stem cells (HSCs) [17], which serve as a paradigm for understanding non-only stem cell biology, but also tissue homeostasis due to their ability to supply blood cells for the entire life of individuals.

In addition to physiological roles, RARs are also involved in diseases and cancers subsequently to mutations, fusions to other proteins, altered expression levels or aberrant post-translational modifications, resulting into altered functions and disruption of homeostasis.

Here we will survey the state of our current knowledge in the role of the different RAR subtypes in healthy adults, focusing on cells or tissues with constant renewal capacity such as tissue stem cells (hematopoietic, mesenchymal and neural stem cells). We will also review the recent insights concerning the deregulation of RAR activities in some diseases or cancers.

2. RAR structure and canonical RAR-mediated regulation of transcription

RARs have a well-defined domain organization and structure (Fig. 1A), consisting mainly of an unstructured and non-conserved N-terminal domain (NTD) and two highly conserved and structured domains, a central DNA-binding domain (DBD) linked to a C-terminal ligand-binding domain (LBD) [1,2,18]. Briefly, the DBD contains two typical cysteine-rich zinc-binding motifs and two α helices, which cross at right angles, folding into a globular conformation to form the

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core of the DBD. By contrast, the LBD is more complex because it contains not only the ligand-binding pocket, but also the main dimerization domain and a hydrophobic cleft involved in coregulator binding. The LBD shows a common fold comprising 12 conserved alpha helices and a short beta turn, arranged in three layers to form an antiparallel «alpha-helical sandwich» (Fig. 1A).

In the past 20 years, it has been established that the basic mechanism for transcriptional regulation by RARs relies on DNA binding to specific sequence elements located in the promoters of target genes and on ligand-induced conformational changes in the LBD that direct the dissociation/association of several coregulator complexes [1,2,19]. RARs heterodimerized with RXRs bind to specific DNA sequences named RA response elements (RAREs), composed typically of two direct repeats of a core hexameric motif, PuG [G/T] TCA [18,20]. The classical RARE is a 5 bp-spaced direct repeat (referred to as DR5). However, the heterodimers also bind to direct repeats separated by 1 bp (DR1) or 2 bp (DR2). Recently the development of high throughput technologies such as chromatin immunoprecipitation (ChIP) coupled with array hybridization or flow cell sequencing expanded the repertoire of potential high affinity response elements [5,21].

When bound to RAREs in the absence of ligand, RARs are associated with large protein complexes endowed with enzymatic activities, which maintain chromatin in a condensed and repressed state [22]. Ligand binding, which is facilitated by RAR association with cellular RA binding protein (CRABP) [23,24], induces conformational changes in the LBD [1,25], the most striking one being the repositionment of helix 12. This structural switch provokes corepressors release and initiates an ordered and coordinated recruitment of a series of coregulator complexes with different enzymatic activities including histone acetyl and methyl transferases, and DNA-dependent ATPases [26,27]. At the end, these events alter the chromatin structure surrounding the promoter of target genes and pave the way for the recruitment of the transcription machinery including RNA Polymerase II and the general transcription factors.

3. New pictures of RAR signaling: RARs are phosphoproteins and cross-talk with kinase cascades

Today it is becoming increasingly evident that, in addition to these classical genomic effects, RARs also induce non-genomic responses such as the rapid and transient activation of several kinase cascades. Indeed, in response to RA, RARs activate p38MAPK [28–30] and a downstream mitogen and stress-activated kinase, MSK1 [31]. These non-genomic effects are mediated by a subpopulation of RARs anchored at the cytoplasmic side of the cell membrane [32,33] and our unpublished observations), suggesting a new paradigm by which RARs would integrate membrane/cytoplasm events that would orchestrate several post-translational modifications in order to fine-tune transcription [34].

In line with this, it emerged that RARs are phosphorylated in response to RA [1,31] at two serine residues located in unstructured domains that provide the flexibility needed for modifications by enzymes such as kinases. One serine is located in a loop between helices 9 and 10 (L9-10) within the LBD and belongs to an arginine-lysine-rich motif that corresponds to a consensus phosphorylation motif for several kinases including MSK1 (Fig. 1A). In contrast, the second serine residue is located in a proline-rich motif within the NTD (Fig. 1A) and is phosphorylated by cdk7 [35,36], which forms with cyclin H and MAT1 the CAK subcomplex of the general transcription factor TFIIF. Most interestingly, the correct positioning of cdk7 and thereby the efficiency of the NTD phosphorylation rely on the docking of cyclin H at a specific site of the LBD located in loop L8-9 and the N-terminal part of H9 (Fig. 1A) [37].

Bruck et al. [31] demonstrated that in the case of the RAR α subtype, the phosphorylation of these two residues results from a coordinated phosphorylation cascade starting with the phosphorylation of the serine in L9-10 (Fig. 1B). Phosphorylation of this residue increases the

dynamics/flexibility of the nearby loop L8-9 (Samarut et al., manuscript submitted), which corresponds to the docking site of cyclin H [37]. Consequently, the binding efficiency of cyclin H is increased, allowing the right positioning of cdk7 and the phosphorylation of the serine located in the NTD by this kinase [38]. Finally phosphorylation of this N-terminal residue allows the recruitment of RAR α to promoters [31]. Note that the RAR γ subtype can be phosphorylated at the same residues and that phosphorylation of the NTD signals the dissociation of vinexin β , an actin-binding protein, which represses RAR γ activity [39].

4. RAR functions in hematopoiesis

One of the best example of RAR functions in adults and for the entire life of individuals is hematopoiesis, which refers to the formation and development of the blood cells [40–42]. In adults, the major hematopoietic organ is the bone marrow where hematopoietic stem cells (HSCs) reside as rare cells in individual niches, and must continuously supply blood cells to host circulation and tissues (homeostasis) [43]. Because mature blood cells are predominantly short lived, multipotent HSCs are required throughout life to replenish multipotent progenitors, committed precursors and mature blood cells (red blood cells, megakaryocytic, lymphocytes and myeloid cells that include monocytes/macrophages, neutrophils and eosinophils) through differentiation processes (Fig. 2). These processes are orchestrated by numerous factors such as cytokines, growth factors and several transcription factors including RARs. Simultaneously, the HSC pool has to be maintained via cell fate decisions of self-renewal, apoptosis/senescence or differentiation. Thus hematopoiesis requires a balance between the self-renewal of the hematopoietic stem cells and their differentiation into the different lineages.

Easy access to hematopoietic cells has enabled a vast number of studies over the last several decades and increased the understanding about the role of RARs in the self-renewal and the differentiation of HSCs. The hematopoietic system expresses significantly the RAR α and RAR γ subtypes (RAR β being generally poorly expressed), and thus most studies concentrated on these two nuclear receptors and compared the behavior of bone marrow HSCs from WT or knockout mice [17,44–46]. As mice deficient for RAR α or RAR γ are viable, bone marrow cells isolated from such mice provided interesting tools to study the role of the individual receptors. Strategies for spatio-temporally controlled somatic mutagenesis of RARs and for RAR overexpression were also designed. These genetic studies were completed by the analysis of VAD animals, and by pharmacological studies based on the use of selective RAR agonist or antagonists. Overall, they provided solid evidence that RAR α and RAR γ are dispensable for hematopoiesis but that each RAR has specific and distinct modulatory roles (Fig. 2).

4.1. Stem cells maintenance: Critical role for RAR γ

Hematopoiesis requires not only the continuous production of mature blood cells from HSCs but also the maintenance of the HSC pool and RAR γ has been found to be a critical regulator of this balance [46]. Indeed bone marrow cells from RAR γ null mice only, and not from RAR α —/— mice, exhibited markedly reduced numbers of HSCs associated with increased numbers of more mature progenitor cells. The observation that primitive precursors overexpressing RAR γ exhibited a more undifferentiated phenotype corroborated the role of RAR γ in HSCs maintenance.

4.2. Lymphocytes (B and T) and macrophages: A role for RAR γ in inflammatory cytokine production

B and T lymphocytes are derived from the multipotent common lymphoid progenitors (CLP, Fig. 2) and are involved in the immune response. Subsequent to the observation that RAR γ expression is up regulated in developing T lymphocytes [17], one suggested that this RAR subtype would play a role in lymphocyte function. However the analysis

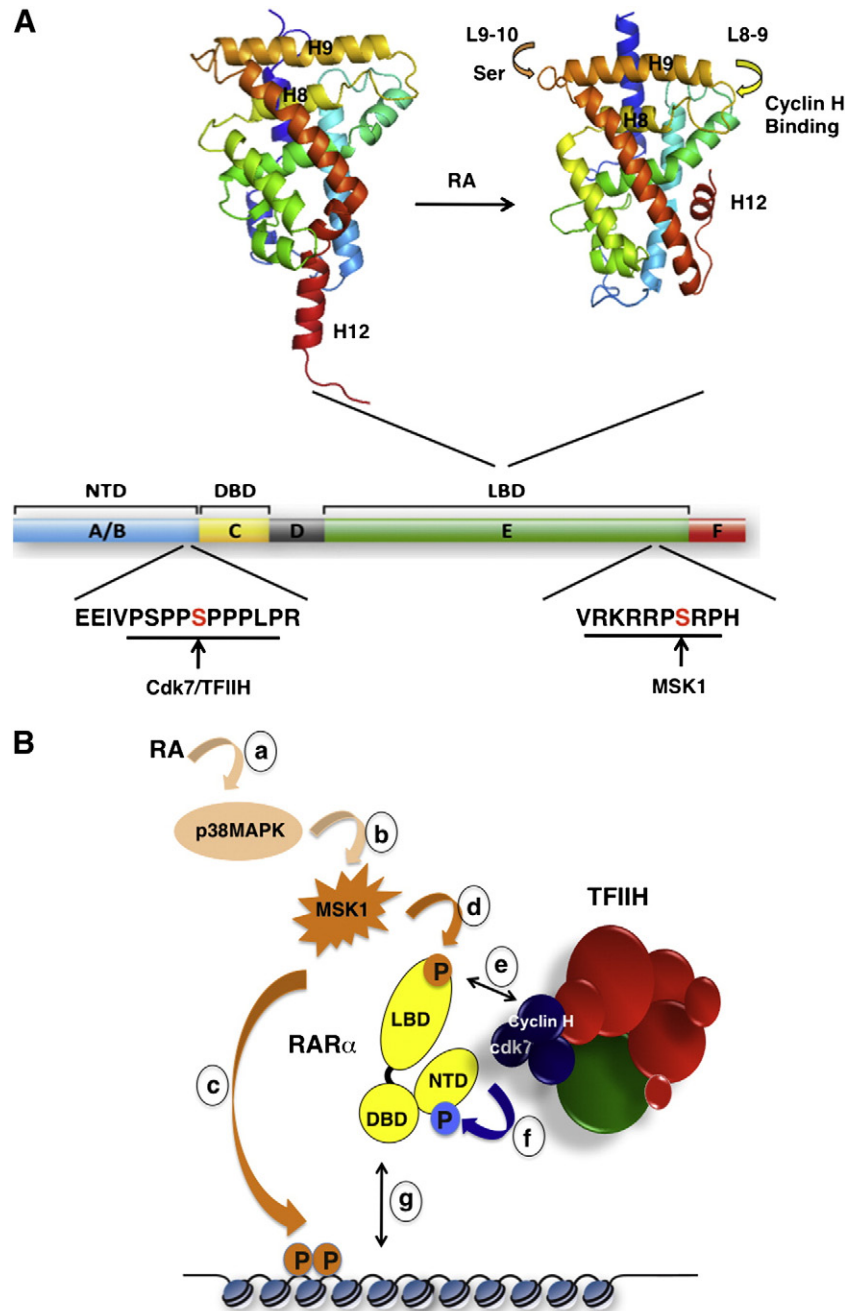


Fig. 1. A. Schematic representation (not to scale) of the different domains of RARs with the main phosphorylation sites. RARs consist mainly of an N-terminal domain (NTD), and a central DNA-binding domain (DBD) linked to a C-terminal ligand-binding domain (LBD). The LBD is composed of 12 conserved α helices and a β -turn separated by disordered loops. The structures in the absence (PDB1LBD) and in the presence (PDB2LBD) of ligand are shown with the swing of helix 12. The serine located in loop L9-10 of RAR α is also shown as well as the cyclin H binding site, which encompasses loop L8-9. B. Recapitulation of the phosphorylation cascade induced by RA. In response to RA, p38MAPK (a) and the downstream protein kinase MSK1 (b) are activated. MSK1 phosphorylates histones at H3S10 (c) as well as RAR α at a serine residue located in the LBD (d). Subsequently, the cyclin H subunit of the CAK subcomplex of TFIIH is recruited to an adjacent domain (e), allowing the formation of a RAR α /TFIIH complex and the phosphorylation of the NTD by the cdk7 kinase (f). Finally, RAR α phosphorylated and associated with TFIIH is recruited to response elements located in the promoter of target genes (g).

of bone marrow cells from mice that conditionally lacked RAR γ in all hematopoietic cells indicated that RAR γ is not essential for T and B lymphocyte proliferation and differentiation [17]. RAR γ was also dispensable for antibody production by B cells. However, RAR γ was required for CD8+ T cell response and cytotoxicity, i.e., the production of interferon gamma (IFN- γ), tumor necrosis factor alpha (TNF α) and cytokines, which remove pathogens and induce inflammation. Most interestingly, macrophages, which are derived from the multipotent common myeloid progenitors (CMP) and which are also involved in the

immune response, also require RAR γ for the production of inflammatory cytokines [17,47]. It has been suggested that RAR γ would regulate the transcription of the cytokines genes by direct binding to promoter regulatory elements. However the promoter regions of IL-6, IL-12, IFN- γ and TNF- α do not contain any RARE, suggesting an indirect role of RAR γ that remains to be elucidated.

Paradoxically, RA and its derivatives suppress inflammation and cytokine production in clinical treatments. Given that this effect was not impaired in RAR γ -deficient macrophages and T cells [17], it has been

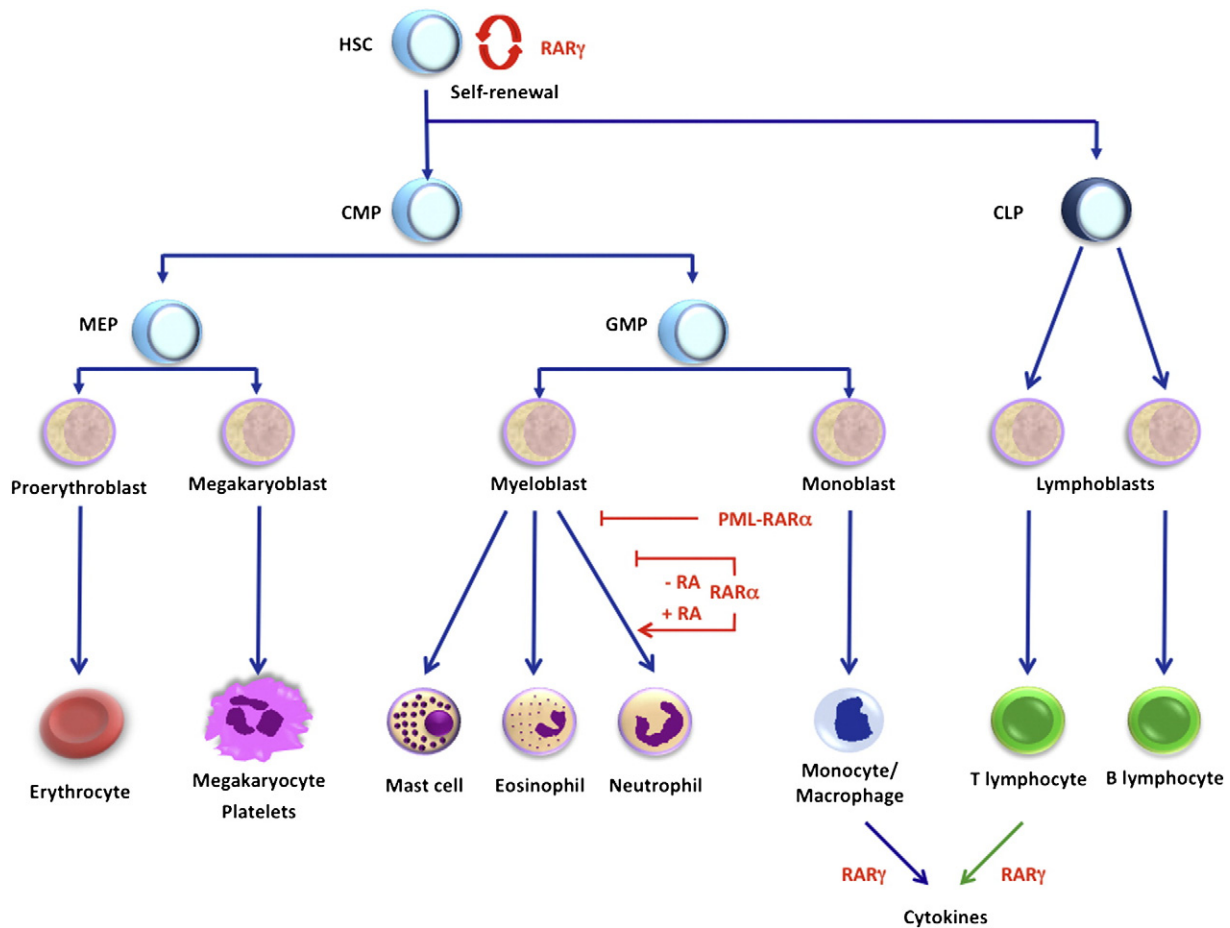


Fig. 2. Recapitulation of the different hematopoietic stages. The stages, which are regulated by RARs, are indicated. HSC, hematopoietic stem cells; CMP, common myeloid progenitor; CLP, common lymphoid progenitor; MEP, megakaryocyte/erythroid progenitor; GMP, granulocyte/macrophage progenitor.

proposed that RA would interfere with signaling pathways involved in the production of inflammatory cytokines such as the nuclear factor kappa B (NF κ B) pathway, which is critical for the expression of various pro-inflammatory genes, including IL-6, IL-12, TNF α , and IFN- γ [48,49]. It has been proposed that the heterodimeric partner RXR α might bind and inhibit NF κ B [50]. However recent evidence rather suggests that RA would suppress the expression of the inflammatory genes by inhibiting kinase pathways independently of RAR activation [51–53]. Whatever the mechanism is, it might be also at the basis of the beneficial effects of RA in sustaining the stability and functionality of regulatory T cells (nTregs), which maintain immunological homeostasis and self-tolerance and thus play a crucial role in controlling autoimmune diseases [54].

4.3. Neutrophil differentiation: Bidirectional role of RAR α

According to several studies, RA has the capacity to drive pluripotent hematopoietic progenitors into the neutrophil lineage. RA can also reprogram hematopoietic progenitors already engaged in another lineage into the neutrophil fate, indicating that RA plays an important role in controlling granulopoiesis. According to genetic and pharmacological studies, neutrophil differentiation is regulated essentially by RAR α , which plays a bidirectional role, inhibiting differentiation when unliganded or bound to antagonists and promoting differentiation when bound to RA or specific RAR α agonists [44,45]. In line with this, RAR α antagonists promoted the accumulation of immature neutrophils and overexpression of RAR α inhibited neutrophil differentiation in the absence of RA, but increased the sensitivity of the transduced cells to the

antiproliferative and differentiative activity of RA and selective RAR α agonists. Moreover, progenitors from RAR α –/– cells differentiated faster into neutrophils, compared to cells from wild type mice [44].

Several RA target genes exemplified by C/EBP ϵ are activated during the RA-induced differentiation of HSCs into neutrophils [40,44]. However, high throughput analysis should improve our understanding of how RAR α controls the expression of lineage-specific myeloid genes. Nevertheless recent studies highlighted a novel level of regulation of neutrophil differentiation by RAR α via microRNAs. Indeed, specific microRNAs are expressed in HSCs [55] and their expression is dynamically regulated during early hematopoiesis and lineage commitment [56]. Most interestingly, some of them are induced by RA [57] and the promoters of the corresponding genes depict DR elements, which bind RAR α [58]. Further investigations should reveal how these microRNAs orchestrate the transcription of specific genes and thereby granulopoiesis.

Finally, given that MAPKs and other kinase signaling pathways play critical roles in the maintenance of hematopoietic homeostasis [59,60], one can suggest that RAR α phosphorylation would also control neutrophil differentiation. Results from our laboratory indicate that indeed RAR α is phosphorylated in myeloid cells (our unpublished results). However in these cells, a loss of RAR α phosphorylation by CAK would be required for the expression of target genes and granulopoiesis [61,62].

Of note is that cells of the neutrophil lineage express both the RAR α 1 and RAR α 2 isoforms. It has been proposed that RAR α 2 would be a more potent inhibitor of differentiation than RAR α 1 [44,63] but

the role of each RAR α isoforms in granulopoiesis is still elusive and debated [64].

5. RAR γ in skin proliferation and differentiation. Role in psoriasis?

In contrast to blood cells, the skin cells can renew but not fully regenerate, making poorly satisfactory the prognosis of healing after wound or deep burn. However, RA has long been known to influence epidermal proliferation and to have a benefit action during wound healing. Indeed topical treatment of skin with retinoids induces hyperproliferation of keratinocytes located in the basal layer, leading to the thickening of the differentiated suprabasal layers and an epidermal hyperplasia [15,65].

In human and mouse epidermis, RAR γ represents 90% of the RARs and thus is the predominant RAR, with the remaining 10% being RAR α and with RAR β being not expressed [66]. The analysis of mice selectively lacking both RAR α and RAR γ in adult epidermis by inducible conditional somatic mutagenesis [15] demonstrated that the complete absence of RARs in basal keratinocytes does not alter their homeostatic proliferation. Thus the self-renewal of epidermal keratinocytes does not seem to require an epidermal RAR-dependent signaling pathway. However, RAR γ present in suprabasal keratinocytes was found to be involved in the RA-induced hyperproliferation, which takes place in the basal layer, through a paracrine signal. Therefore the beneficial actions of retinoids during wound healing have been attributed to this mechanism. It must be noted that the injection of mesenchymal stem cells (MSCs) in skin also has benefits in skin repair and regeneration as they transdifferentiate into keratinocytes [67]. However, the question whether RAR γ plays a role in this process still remains.

Retinoids are highly effective in the treatment of skin disorders such as psoriasis and acne [68,69]. Psoriatic skin is characterized by hyperproliferation of the basal epidermal layers and hyperkeratosis but no clear-cut alterations in the expression levels and distribution of RARs have been reported. In fact, psoriasis is essentially an inflammatory disease in which skin contains large numbers of immune T cells and macrophages that produce cytokines and other inflammatory molecules (interleukins, TNF α and IFN- γ) [70]. In line with this, the effects of retinoids on this disease have been correlated to their ability to suppress inflammation through inhibition of cytokine production (see above 4.2). Accordingly, microarray experiments indicated that RA downregulates a great proportion of the psoriasis-related genes [71].

6. New unconventional roles for RAR α in the nervous system

It has long been thought that the adult mammalian central nervous system does not undergo notable cell turnover. However, recent evidence has shown that neural stem cells are present in two specialized niches of the adult brain, the subventricular zone and the subgranular zone of the hippocampus. They maintain neurogenesis and gliogenesis throughout adult life [72]. Several studies indicate that RA induces the differentiation of these stem cells into neurons [73,74], as described for multipotent embryonic stem cells [75–77]. However no genetic studies have been performed yet in the field and thus the question whether RARs are involved in neuronal stem cell proliferation and differentiation still remains.

In fact, recent work highlighted an unexpected role for RAR α in neuron homeostatic plasticity. Indeed, the function of the normal nervous system requires that, in the hippocampus, neurons maintain a constant overall activity level that is achieved by homeostatic synaptic plasticity [78]. This type of plasticity, manifested as increased synaptic transmission in response to reduced neural activity, requires the translation of new proteins such as the GluR1 receptors in dendrites. Recently Aoto et al. [79] reported that RAR α plays a role during homeostatic plasticity by increasing the size of the postsynaptic glutamate receptor response. RAR α is expressed in the hippocampus, but in addition to its classical presence in the nucleus,

it is also present in neuronal dendrites [80]. During homeostatic plasticity and in response to RA, RAR α accumulates in dendrite granules where it plays a non-genomic role as an mRNA binding protein and stimulates GluR1 translation [81,82].

Finally, one must note that synaptic plasticity underlies hippocampus-dependent spatial learning and memory [78]. Given that deficiencies in retinoid metabolism and signaling cause impaired synaptic plasticity and learning [83], it has been suggested that such deficiencies might result in neurological diseases [84]. One example is Alzheimer's disease, which is associated with a variety of pathophysiological features including amyloid plaque, inflammation, neurodegeneration, altered neurotransmission and age-related changes [85]. In that context, retinoids are candidate's drugs for treatment of this disease. [85,86]. This expands the scope of the biological function of RARs and retinoids beyond their role as regulators of gene transcription. Note however that RAR α can also regulate neurotransmission through the direct transcriptional regulation of the dopamine D2 receptor [87].

7. RARs in cancers

In keeping with their ability to regulate growth and differentiation throughout life, it is evident that abnormal expression and/or functions of RARs might be involved in several cancers. Accordingly, RAR loss, overexpression, mutations or aberrant post-translational modifications have been correlated with tumoral growth.

7.1. RAR α rearrangements and mutations in myeloid leukemia (APL)

The blood system serves as a paradigm for understanding not only stem cell biology and organogenesis but also oncogenesis. Indeed, in human myeloid leukemia, the RAR α gene is the target of chromosomal rearrangements leading to the production of fusion proteins that allow the cells to continue to proliferate and/or prevent the terminal differentiation seen in normal myelocytes [88].

The best example is acute promyelocytic leukemia (APL or AML-M3 according to the FAB classification) in which maturation arrest occurs at the level of the promyelocyte (Fig. 2). In APLs the rearrangements lead to the production of fusion proteins containing a fixed C-terminal portion corresponding to most of the RAR α polypeptide (including the DBD and the LBD) and a variable N-terminal portion originating from the PML, PLZF, NuMA, nucleophosmin or STAT5b proteins (for review, see reference [89]). Recently, novel rearrangements have been identified involving the regulatory type I alpha subunit of PKA [90] and FIP1L1 [91]. A number of laboratories analyzed how these rearrangements disturb the functions of normal RAR α in controlling the proliferation of promyelocytes and their differentiation into neutrophils.

Most of the studies have been performed on PML-RAR α , which results from the reciprocal translocation t (15; 17) [92] and which is involved in >95% of cases. At the molecular level, PML-RAR α behaves as an altered RAR with stronger repressive activities. Indeed, in contrast to RAR α , PML-RAR α has the ability to homodimerize or oligomerize with or without union to RXR α [93,94]. Such a property expands the recruitment of corepressor complexes encompassing a wide spectrum of epigenetic enzymes with strong repressive properties towards target genes [95]. Moreover, according to recent high throughput studies [21], the oligomeric PML-RAR α complexes also gain an expanded DNA-binding capacity away from the classical DR5, DR2 and DR1 motifs towards more widely spaced DRs. Finally, PML-RAR α was also found to repress the expression of microRNA genes and thus to increase the expression of key cancer proteins [58].

Another important property is that, in contrast to RAR α , the PML-RAR α fusion protein is insensitive to physiological concentrations of RA and requires higher pharmacological concentrations to overcome the transcription and differentiation block [96]. Such pharmacological doses of RA induce the release of microRNA expression [58], as well as large-scale epigenetic changes (H3 acetylation, H3K27 methylation,

H3K9 methylation etc.) at thousand of gene regions [21]. Most interestingly, the global changes in repressive marks are not triggered by the release of the chromatin-modifying enzymes from the PML-RAR α complex itself but by the rapid loss of the fusion protein at the DNA-binding sites ([21] and Cassinat et al., manuscript submitted). Such changes in the epigenetic landscape of APL cells have been correlated to the reversal of the differentiation block and to the cessation of proliferation and thus are at the basis of therapies combining RA and epi-drugs such as HDACs [96–98].

It must be noted that some APL patients depict mutations in the ligand-binding domain (LBD) of the RAR α portion of the PML-RAR α fusion protein [99,100]. Some of these patients had point mutations in the ligand-binding pocket that disrupted RA binding. Others had mutations that produced extensive deletion of the entire LBD. All these mutations have been associated with poor clinical outcome, relapse and RA resistance. However, signaling pathways such as G-CSF were recently found to achieve the differentiation of such RA-resistant leukemic cells [101]. Though the molecular mechanism of the release of the RA resistance by G-CSF still remains ill defined, recent studies highlighted a key role for MAPK. Indeed, when combined with RA, G-CSF restores the expression of RAR α -target genes through the activation of the MAPK/ERK pathway [102] and the subsequent relief of aberrant epigenetic modifications of histones (Cassinat et al., manuscript submitted).

Finally, it must be noted that, due to their quiescent state, several leukemic stem cells resist to RA treatment, resulting in disease relapse. Therefore new therapeutic approaches based on the proteasomal degradation of the PML-RAR α oncoprotein have been developed in order to trigger the eradication of the leukemia-initiating cells and thus long-term remission [103–105]. Such therapies combine RA to arsenic and/or agents that activate cyclic AMP signaling [106,107].

7.2. Loss of RAR β and RAR γ in cancers: Are RARs tumor suppressors?

In several tissues and organs, the loss of a RAR subtype has been associated with tumorigenesis. Consequently, RARs have been considered as tumor suppressors. In keratinocytes, it is the RAR γ subtype that has been considered as a tumor suppressor. Indeed, in KO mice, the loss of RAR γ predisposed skin to tumors [108] and RAR γ was found to be absent in oral keratinocytes from head and neck cancers [109]. In addition, premature skin aging and skin cancer induced by UV radiation from the sun have been correlated to a dramatic decrease in RAR γ levels subsequently to the UV-induced proteasomal degradation of the receptor [110,111].

In contrast, in other tissues and organs such as lung, breast and cervix, it is the loss of RAR β expression that has been associated with tumorigenesis. Indeed, in such cancers, the RAR β gene is frequently deleted or the RAR β promoter silenced due to aberrant DNA methylation or repressive histone modifications [96,112–115]. Accordingly, reexpression of RAR β [116] or restoration of the RAR β promoter activity by demethylation of DNA or HDAC inhibition [117,118] decreased the tumorigenicity of the cancer cells.

The mechanism of the tumor suppressive effect of RARs is still debated but according to several studies, RARs would block tumor promotion through inhibiting the activity of the AP-1 complex (heterodimers of the proto-oncogene products c-Fos and c-Jun), which is known to regulate the expression of several genes involved in oncogenesis and cell transformation and which is controlled by a complex network of phosphorylations by stress-kinases ([1,96] and references therein). Though several models have been proposed, the molecular basis of the anti-AP-1 activity of RARs is still debated [1,96]. Nevertheless, AP-1 inhibition generally correlates with the growth inhibitory effect of retinoids on tumors. Therefore, the capacity of RARs to inhibit AP-1-responsive genes seems to be the basis for the chemopreventive and chemotherapeutic effects of RA in the treatment of hyperproliferative diseases [96].

Note that in one leukemic patient, a RAR α mutant named RAR α 1delBC (deleted for the DBD and the N-terminal domain) has been isolated [119]. However no specific correlations with the growth of the leukemic cells have been found yet.

7.3. Non-classic roles of RARs in tumor cells growth

Unexpectedly, in some cases, RARs have been correlated with survival instead of growth arrest. As an example, in hepatocellular carcinoma, RAR γ has been found to be overexpressed [120] and to contribute to cell growth through its altered subcellular localization and strong cytoplasmic accumulation. Indeed, in the cytosol, RAR γ interacts with the p85 α regulatory subunit of PI3K leading to the activation of the PI3K/Akt pathway, which is one of the major survival pathways in cancer cells [121]. Another example, still in the liver, is RAR α , which behaves as an oncogene in the absence of the corepressor TRIM24 (TIF1 α) [122].

One must note that in breast cancer cells, RAR α expression can be induced by estrogens. Moreover RAR α has been shown to share with the estrogen receptor ER α , a subset of binding regions within the genome [123,124] and to be part of the ER transcriptional complexes [124]. Through this mechanism, which is distinct from the classical one, it has been proposed that RAR α would participate to estrogen-mediated proliferation. Note however that this model occurs only in the presence of estrogens and is not incompatible with a shift towards the classical antiproliferative activity of RAR α upon RA addition.

Finally, in some cancers, RA does not inhibit cell growth but promotes cell survival. Unexpectedly, studies from Noy's laboratory demonstrated that this effect is not mediated by RARs but by the activation of the orphan peroxisome proliferation-activated receptor β/δ (PPAR β/δ), which in turn induces the expression of survival genes [125]. This shift in RA signaling would be regulated by the ratio between the CRABP II and FABP5 proteins, which specifically deliver RA to RARs and PPAR β/δ respectively.

7.4. Aberrant RAR phosphorylations

Today, new roles for phosphorylation in the regulation of RAR functions are being discovered at an accelerated pace. Accordingly, it is now increasingly clear that cancers are characterized by amplified or deregulated cytosolic kinase cascades [126], ending at Akt or at different MAPKs (Erks, JNKs, p38MAPK), which enter the nucleus. Breast cancers characterized by the overexpression of the receptor tyrosine kinase ErbB-2 [127–129] are a typical example of such cancers, which are generally resistant to the antiproliferative action of RA [130]. In these cancers (Duong et al., manuscript in preparation) and others [131,132], RAR α is aberrantly phosphorylated. Subsequently, RAR α is degraded and/or its transcriptional activity is abrogated.

Note that in xeroderma pigmentosum patients, who are characterized by mutations affecting subunits of the core of TFIIH, RAR α is not efficiently phosphorylated by the cdk7/cyclinH/MAT1 subcomplex of the general transcription factor TFIIH. This deficient phosphorylation has characteristic downstream consequences on the expression of RAR target genes [133]. This has been correlated at least in part to the clinical abnormalities of the patients but also to their high risk of skin cancer in response to UV.

8. Conclusion and future directions

Analogous to the paradigm established by the hematopoietic system, there is now increasing evidence for the existence of stem cells and a differentiation hierarchy in most adult tissues, including brain [72], the mammary gland [134] and dermis [67]. In this review we have highlighted that strictly controlled levels of RARs are necessary for the correct balance between self-renewal and differentiation of tissue stem cells (Table 1). Thus, the integrity of RARs is absolutely required for

homeostasis in adults. Consequently, it is evident that loss, accumulation, mutations or aberrant modifications of a specific RAR predispose to cancer and lead to uncontrolled proliferation and/or to differentiation block (Table 1). Therefore, deciphering how RARs and retinoids control the expression of subsets of target genes is a continuous challenge in homeostasis, oncogenesis and therapy. In line with this, microRNAs appear to be novel and promising RARs targets, not only during homeostatic cell fate determination but also in cancer therapy [55,58].

It is important to note that in addition to its ability to control cell growth and differentiation, RA has been recently shown to regulate lipid homeostasis [135]. However this effect is mediated not only by RARs but also by the peroxisome proliferation-activated receptor β/δ (PPAR β/δ), introducing an additional level of complexity. Finally, it is getting increasingly evident that RARs are involved in unconventional non-genomic effects such as neuron plasticity [79], opening new avenues of investigations.

Here we did not cover RXRs, the RAR partners, but it is evident that they also play a role in homeostasis [16,136] and that their expression levels [112,137] as well as their post-translational modifications [138] are affected in several diseases and cancers.

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References

- [1] C. Rochette-Egly, P. Germain, Dynamic and combinatorial control of gene expression by nuclear retinoic acid receptors, *Nucl. Recept. Signal.* 7 (2009) e005.
- [2] P. Germain, B. Staels, C. Dacquet, M. Spedding, V. Laudet, Overview of nomenclature of nuclear receptors, *Pharmacol. Rev.* 58 (2006) 685–704.
- [3] P. Germain, P. Chambon, G. Eichele, R.M. Evans, M.A. Lazar, M. Leid, A.R. De Lera, R. Lotan, D.J. Mangelsdorf, H. Gronemeyer, International Union of Pharmacology. LXIII. Retinoid X receptors, *Pharmacol. Rev.* 58 (2006) 760–772.
- [4] P. Germain, P. Chambon, G. Eichele, R.M. Evans, M.A. Lazar, M. Leid, A.R. De Lera, R. Lotan, D.J. Mangelsdorf, H. Gronemeyer, International Union of Pharmacology. LX. Retinoic acid receptors, *Pharmacol. Rev.* 58 (2006) 712–725.
- [5] L. Delacroix, E. Moutier, G. Altobelli, S. Legras, O. Poch, M.A. Choukrallah, I. Bertin, B. Jost, I. Davidson, Cell-specific interaction of retinoic acid receptors with target genes in mouse embryonic fibroblasts and embryonic stem cells, *Mol. Cell. Biol.* 30 (2010) 231–244.
- [6] G. Bour, R. Taneja, C. Rochette-Egly, Mouse embryocarcinoma F9 cells and retinoic acid. A model to study the molecular mechanisms of endodermal differentiation, in: R. Raneja (Ed.), *Nuclear Receptors in Development*, vol. 16, Elsevier Press Inc., 2006, pp. 211–253.
- [7] C. Eifert, N. Sangster-Guilty, L.M. Yu, S.V. Chittur, A.V. Perez, J.A. Tine, P.J. McCormick, Global gene expression profiles associated with retinoic acid-induced differentiation of embryonal carcinoma cells, *Mol. Reprod. Dev.* 73 (2006) 796–824.
- [8] L.J. Gudas, J.A. Wagner, Retinoids regulate stem cell differentiation, *J. Cell. Physiol.* (2010).
- [9] M. Mark, N.B. Ghyselinck, P. Chambon, Function of retinoid nuclear receptors: lessons from genetic and pharmacological dissections of the retinoic acid signaling pathway during mouse embryogenesis, *Annu. Rev. Pharmacol. Toxicol.* 46 (2006) 451–480.
- [10] K. Niederreither, P. Dolle, Retinoic acid in development: towards an integrated view, *Nat. Rev. Genet.* 9 (2008) 541–553.
- [11] P. Dolle, Developmental expression of retinoic acid receptors (RARs), *Nucl. Recept. Signal.* 7 (2009) e006.
- [12] M. Mark, N.B. Ghyselinck, P. Chambon, Function of retinoic acid receptors during embryonic development, *Nucl. Recept. Signal.* 7 (2009) e002.
- [13] D. Su, L.J. Gudas, Gene expression profiling elucidates a specific role for RARgamma in the retinoic acid-induced differentiation of F9 teratocarcinoma stem cells, *Biochem. Pharmacol.* 75 (2008) 1129–1160.
- [14] D. Metzger, A.K. Indra, M. Li, B. Chapellier, C. Calleja, N.B. Ghyselinck, P. Chambon, Targeted conditional somatic mutagenesis in the mouse: temporally-controlled knock out of retinoid receptors in epidermal keratinocytes, *Meth. Enzymol.* 364 (2003) 379–408.
- [15] B. Chapellier, M. Mark, N. Messaddeq, C. Calleja, X. Warot, J. Brocard, C. Gerard, M. Li, D. Metzger, N.B. Ghyselinck, P. Chambon, Physiological and retinoid-induced proliferations of epidermis basal keratinocytes are differently controlled, *EMBO J.* 21 (2002) 3402–3413.

Table 1

Recapitulation of RAR functions during homeostasis and of their deregulation in cancers or diseases.

Tissue	RAR	Physiological role	Associated disease	RAR alterations
Bone marrow	RAR α	Neutrophil differentiation [44,45]	Leukemia (APL)	RAR α chromosomal translocations [88,89] Mutations in RAR α LBD [99,100]
	RAR γ	HSC renewal [46] T cell CD8+ cytotoxicity [17]	Inflammation	
Fibroblasts	RAR α	Growth control	Xeroderma pigmentosum	RAR α hypophosphorylation [133]
Liver	RAR γ /RAR α	Growth control	Cancer	RAR γ overexpression/cytosolic localization [120] Loss of TRIM24 [122]
Lung	RAR β /RAR α	Growth control	Cancer	RAR β deletion/silencing [96,112,113] RAR α aberrant phosphorylation [131,132]
Mammary gland	RAR β /RAR α	Growth control	Cancer	RAR β deletion/silencing [115] RAR α aberrant phosphorylation [131,132]
Neurons	RAR α	Plasticity [79]	Alzheimer? [85,86]	
Skin	RAR γ	Keratinocyte proliferation [15]	Aging/cancer	RAR γ loss [109–111]

- [16] T. Imai, M. Jiang, P. Kastner, P. Chambon, D. Metzger, Selective ablation of retinoid X receptor alpha in hepatocytes impairs their lifespan and regenerative capacity, *Proc. Natl. Acad. Sci. USA* 98 (2001) 4581–4586.
- [17] I. Dzhalalov, P. Chambon, Y.W. He, Regulation of CD8+ T lymphocyte effector function and macrophage inflammatory cytokine production by retinoic acid receptor gamma, *J. Immunol.* 178 (2007) 2113–2121.
- [18] P. Germain, L. Altucci, W. Bourguet, C. Rochette-Egly, H. Gronemeyer, Nuclear receptor superfamily: principles of signaling, in: J. Miyamoto, J. Burger (Eds.), *Pure Appl. Chem.* 75 (2003) 1619–1664.
- [19] C. Rochette-Egly, Dynamic combinatorial networks in nuclear receptor-mediated transcription, *J. Biol. Chem.* 280 (2005) 32565–32568.
- [20] V. Laudet, H. Gronemeyer, *Nuclear receptor factsbook*, Academic Press, London, 2001.
- [21] J.H. Martens, A.B. Brinkman, F. Simmer, K.J. Francoijs, A. Nebbioso, F. Ferrara, L. Altucci, H.G. Stunnenberg, PML-RARalpha/RXR alters the epigenetic landscape in acute promyelocytic leukemia, *Cancer Cell* 17 (2010) 173–185.
- [22] V. Perissi, K. Jepsen, C.K. Glass, M.G. Rosenfeld, Deconstructing repression: evolving models of co-repressor action, *Nat. Rev. Genet.* 11 (2010) 109–123.
- [23] L. Delva, J.N. Bastie, C. Rochette-Egly, R. Kraiba, N. Balitrand, G. Despouy, P. Chambon, C. Chomienne, Physical and functional interactions between cellular retinoic acid binding protein II and the retinoic acid-dependent nuclear complex, *Mol. Cell. Biol.* 19 (1999) 7158–7167.
- [24] A.S. Budhu, N. Noy, Direct channeling of retinoic acid between cellular retinoic acid-binding protein II and retinoic acid receptor sensitizes mammary carcinoma cells to retinoic acid-induced growth arrest, *Mol. Cell. Biol.* 22 (2002) 2632–2641.
- [25] A. le Maire, C. Teyssier, C. Erb, M. Grimaldi, S. Alvarez, A.R. de Lera, P. Balaguer, H. Gronemeyer, C.A. Royer, P. Germain, W. Bourguet, A unique secondary-structure switch controls constitutive gene repression by retinoic acid receptor, *Nat. Struct. Mol. Biol.* 17 (2010) 801–807.
- [26] P. Lefebvre, P.J. Martin, S. Flajollet, S. Dedieu, X. Billaut, B. Lefebvre, Transcriptional activities of retinoic acid receptors, *Vitam. Horm.* 70 (2005) 199–264.
- [27] M.G. Rosenfeld, V.V. Lunyak, C.K. Glass, Sensors and signals: a coactivator/corepressor/epigenetic code for integrating signal-dependent programs of transcriptional response, *Genes Dev.* 20 (2006) 1405–1428.
- [28] Y. Alsayed, S. Uddin, N. Mahmud, F. Lekmine, D.V. Kalvakolanu, S. Minucci, G. Bokoch, L.C. Platanias, Activation of Rac1 and the p38 mitogen-activated protein kinase pathway in response to all-trans-retinoic acid, *J. Biol. Chem.* 276 (2001) 4012–4019.
- [29] M. Gianni, E. Parrella, I. Raska, E. Gaillard, E.A. Nigro, C. Gaudon, E. Garattini, C. Rochette-Egly, P38MAPK-dependent phosphorylation and degradation of SRC-3/AIB1 and RARalpha-mediated transcription, *EMBO J.* 25 (2006) 739–751.
- [30] M. Gianni, A. Bauer, E. Garattini, P. Chambon, C. Rochette-Egly, Phosphorylation by p38MAPK and recruitment of SUG-1 are required for RA-induced RAR γ degradation and transactivation, *EMBO J.* 21 (2002) 3760–3769.
- [31] N. Bruck, D. Vitoux, C. Ferry, V. Duong, A. Bauer, H. de The, C. Rochette-Egly, A coordinated phosphorylation cascade initiated by p38MAPK/MSK1 directs RARalpha to target promoters, *EMBO J.* 28 (2009) 34–47.

- [32] S. Masia, S. Alvarez, A.R. de Lera, D. Barettoni, Rapid, nongenomic actions of retinoic acid on phosphatidylinositol-3-kinase signaling pathway mediated by the retinoic acid receptor, *Mol. Endocrinol.* 21 (2007) 2391–2402.
- [33] N. Dey, P.K. De, M. Wang, H. Zhang, E.A. Dobrotka, K.A. Robertson, D.L. Durden, CSK controls retinoic acid receptor (RAR) signaling: a RAR-c-SRC signaling axis is required for neurogenic differentiation, *Mol. Cell. Biol.* 27 (2007) 4179–4197.
- [34] S. Lalevee, C. Ferry, C. Rochette-Egly, Phosphorylation control of nuclear receptors, *Meth. Mol. Biol.* 647 (2010) 251–266.
- [35] J. Bastien, S. Adam-Stitah, T. Riedl, J.M. Egly, P. Chambon, C. Rochette-Egly, TFIH interacts with the retinoic acid receptor gamma and phosphorylates its AF-1-activating domain through cdk7, *J. Biol. Chem.* 275 (2000) 21896–21904.
- [36] C. Rochette-Egly, S. Adam, M. Rossignol, J.M. Egly, P. Chambon, Stimulation of RAR alpha activation function AF-1 through binding to the general transcription factor TFIH and phosphorylation by CDK7, *Cell* 90 (1997) 97–107.
- [37] G. Bour, E. Gaillard, N. Bruck, S. Lalevee, J.L. Plassat, D. Busso, J.P. Samama, C. Rochette-Egly, Cyclin H binding to the RAR(alpha) activation function (AF)-2 domain directs phosphorylation of the AF-1 domain by cyclin-dependent kinase 7, *Proc. Natl Acad. Sci. USA* 102 (2005) 16608–16613.
- [38] E. Gaillard, N. Bruck, Y. Brelivet, G. Bour, S. Lalevee, A. Bauer, O. Poch, D. Moras, C. Rochette-Egly, Phosphorylation by protein kinase A potentiates retinoic acid receptor A activity by means of increasing interaction with and phosphorylation by cyclin H/cdk7, *Proc. Natl Acad. Sci. USA* 103 (2006) 9548–9553.
- [39] S. Lalevee, G. Bour, M. Quinternet, E. Samarut, P. Kessler, M. Vitorino, N. Bruck, M.A. Delsuc, J.L. Vonesch, B. Kieffer, C. Rochette-Egly, Vinexin(beta), an atypical "sensor" of retinoic acid receptor gamma signaling: union and sequestration, separation, and phosphorylation, *FASEB J.* (2010).
- [40] S.H. Orkin, L.I. Zon, Hematopoiesis: an evolving paradigm for stem cell biology, *Cell* 132 (2008) 631–644.
- [41] S.H. Orkin, L.I. Zon, SnapShot: hematopoiesis, *Cell* 132 (2008) 712.
- [42] H. Ueno, I.L. Weissman, The origin and fate of yolk sac hematopoiesis: application of chimera analyses to developmental studies, *Int. J. Dev. Biol.* 54 (2010) 1019–1031.
- [43] A. Wilson, E. Laurenti, A. Trumpp, Balancing dormant and self-renewing hematopoietic stem cells, *Curr. Opin. Genet. Dev.* 19 (2009) 461–468.
- [44] P. Kastner, S. Chan, Function of RARalpha during the maturation of neutrophils, *Oncogene* 20 (2001) 7178–7185.
- [45] S.J. Collins, The role of retinoids and retinoic acid receptors in normal hematopoiesis, *Leukemia* 16 (2002) 1896–1905.
- [46] L.E. Purton, S. Dworkin, G.H. Olsen, C.R. Walkley, S.A. Fabb, S.J. Collins, P. Chambon, RARgamma is critical for maintaining a balance between hematopoietic stem cell self-renewal and differentiation, *J. Exp. Med.* 203 (2006) 1283–1293.
- [47] C. Gordy, I. Dzhalgalov, Y.W. He, Regulation of CD8(+) T cell functions by RARgamma, *Semin. Immunol.* 21 (2009) 2–7.
- [48] S. Wang, H. Uchi, S. Hayashida, K. Urabe, Y. Moroi, M. Furue, Differential expression of phosphorylated extracellular signal-regulated kinase 1/2, phosphorylated p38 mitogen-activated protein kinase and nuclear factor-kappaB p105/p50 in chronic inflammatory skin diseases, *J. Dermatol.* 36 (2009) 534–540.
- [49] L. Vermeulen, W.V. Berghe, I.M. Beck, K. De Bosscher, G. Haegeman, The versatile role of MSKs in transcriptional regulation, *Trends Biochem. Sci.* 34 (2009) 311–318.
- [50] S.Y. Na, B.Y. Kang, S.W. Chung, S.J. Han, X. Ma, G. Trinchieri, S.Y. Im, J.W. Lee, T.S. Kim, Retinoids inhibit interleukin-12 production in macrophages through physical associations of retinoic X receptor and NFkappaB, *J. Biol. Chem.* 274 (1999) 7674–7680.
- [51] M. Kirchmeyer, M. Koufany, S. Sebillaud, P. Netter, J.Y. Jouzeau, A. Bianchi, All-trans retinoic acid suppresses interleukin-6 expression in interleukin-1-stimulated synovial fibroblasts by inhibition of ERK1/2 pathway independently of RAR activation, *Arthritis Res. Ther.* 10 (2008) R141.
- [52] I.M. Beck, W.V. Berghe, S. Gerlo, N. Bougarne, L. Vermeulen, K. De Bosscher, G. Haegeman, Glucocorticoids and mitogen- and stress-activated protein kinase 1 inhibitors: possible partners in the combat against inflammation, *Biochem. Pharmacol.* 77 (2009) 1194–1205.
- [53] I.M. Beck, W. Vanden Berghe, L. Vermeulen, K.R. Yamamoto, G. Haegeman, K. De Bosscher, Crosstalk in inflammation: the interplay of glucocorticoid receptor-based mechanisms and kinases and phosphatases, *Endocr. Rev.* 30 (2009) 830–882.
- [54] X. Zhou, N. Kong, J. Wang, H. Fan, H. Zou, D. Horwitz, D. Brand, Z. Liu, S.G. Zheng, Cutting edge: all-trans retinoic acid sustains the stability and function of natural regulatory T cells in an inflammatory milieu, *J. Immunol.* 185 (2010) 2675–2679.
- [55] A. Fatica, A. Rosa, M. Ballarino, M.L. De Marchis, K.D. Rasmussen, I. Bozzoni, Role of microRNAs in myeloid differentiation, *Biochem. Soc. Trans.* 36 (2008) 1201–1205.
- [56] C.Z. Chen, L. Li, H.F. Lodish, D.P. Bartel, MicroRNAs modulate hematopoietic lineage differentiation, *Science* 303 (2004) 83–86.
- [57] F. Fazi, A. Rosa, A. Fatica, V. Gelmetti, M.L. De Marchis, C. Nervi, I. Bozzoni, A microcircuitry comprised of microRNA-223 and transcription factors NF1-A and C/EBPalpha regulates human granulopoiesis, *Cell* 123 (2005) 819–831.
- [58] A. Saumet, G. Vetter, M. Bouttier, E. Portales-Casamar, W.W. Wasserman, T. Maurin, B. Mari, P. Barbry, L. Vallar, E. Friederich, K. Arar, B. Cassinat, C. Chomienne, C.H. Lecellier, Transcriptional repression of microRNA genes by PML-RARA increases expression of key cancer proteins in acute promyelocytic leukemia, *Blood* 113 (2009) 412–421.
- [59] C.R. Geest, M. Buitenhuis, A.G. Laarhoven, M.B. Bierings, M.C. Bruin, E. Vellenga, P.J. Coffer, p38 MAP kinase inhibits neutrophil development through phosphorylation of C/EBPalpha on serine 21, *Stem Cells* 27 (2009) 2271–2282.
- [60] C.R. Geest, P.J. Coffer, MAPK signaling pathways in the regulation of hematopoiesis, *J. Leukoc. Biol.* 86 (2009) 237–250.
- [61] P. Luo, A. Wang, K.J. Payne, H. Peng, J.G. Wang, Y.K. Parrish, J.W. Rogerio, T.J. Triche, Q. He, L. Wu, Intrinsic retinoic acid receptor alpha-cyclin-dependent kinase-activating kinase signaling involves coordination of the restricted proliferation and granulocytic differentiation of human hematopoietic stem cells, *Stem. Cells* 25 (2007) 2628–2637.
- [62] J. Wang, L.W. Barsky, C.H. Shum, A. Jong, K.I. Weinberg, S.J. Collins, T.J. Triche, L. Wu, Retinoid-induced G1 arrest and differentiation activation are associated with a switch to cyclin-dependent kinase-activating kinase hypophosphorylation of retinoic acid receptor alpha, *J. Biol. Chem.* 277 (2002) 43369–43376.
- [63] J. Labrecque, D. Allan, P. Chambon, N.N. Iscove, D. Lohnes, T. Hoang, Impaired granulocytic differentiation in vitro in hematopoietic cells lacking retinoic acid receptors alpha1 and gamma, *Blood* 92 (1998) 607–615.
- [64] S. Wang, G. Tricot, L. Shi, W. Xiong, Z. Zeng, H. Xu, M. Zangari, B. Barlogie, J.D. Shaughnessy Jr., F. Zhan, RARalpha2 expression is associated with disease progression and plays a crucial role in efficacy of ATRA treatment in myeloma, *Blood* 114 (2009) 600–607.
- [65] G.J. Fisher, J.J. Voorhees, Molecular mechanisms of retinoid actions in skin, *FASEB J.* 10 (1996) 1002–1013.
- [66] G.J. Fisher, H.S. Talwar, J.H. Xiao, S.C. Datta, A.P. Reddy, M.P. Gaub, C. Rochette-Egly, P. Chambon, J.J. Voorhees, Immunological identification and functional quantitation of retinoic acid and retinoid X receptor proteins in human skin, *J. Biol. Chem.* 269 (1994) 20629–20635.
- [67] X. Fu, H. Li, Mesenchymal stem cells and skin wound repair and regeneration: possibilities and questions, *Cell Tissue Res.* 335 (2009) 317–321.
- [68] P.C. van de Kerkhof, Update on retinoid therapy of psoriasis in: an update on the use of retinoids in dermatology, *Dermatol. Ther.* 19 (2006) 252–263.
- [69] A. Menter, N.J. Korman, C.A. Elmets, S.R. Feldman, J.M. Gelfand, K.B. Gordon, A.B. Gottlieb, J.Y. Koo, M. Lebwohl, H.W. Lim, A.S. Van Voorhees, K.R. Beutner, R. Bhushan, Guidelines of care for the management of psoriasis and psoriatic arthritis: section 4. Guidelines of care for the management and treatment of psoriasis with traditional systemic agents, *J. Am. Acad. Dermatol.* 61 (2009) 451–485.
- [70] E.D. Roberson, A.M. Bowcock, Psoriasis genetics: breaking the barrier, *Trends Genet.* 26 (2010) 415–423.
- [71] D.D. Lee, O. Stojadinovic, A. Krzyzanowska, C. Vouthounis, M. Blumenberg, M. Tomic-Canic, Retinoid-responsive transcriptional changes in epidermal keratinocytes, *J. Cell. Physiol.* 220 (2009) 427–439.
- [72] L. Conti, E. Cattaneo, Neural stem cell systems: physiological players or in vitro entities? *Nat. Rev. Neurosci.* 11 (2010) 176–187.
- [73] M.B. Gonçalves, J. Boyle, D.J. Webber, S. Hall, S.L. Minger, J.P. Corcoran, Timing of the retinoid-signalling pathway determines the expression of neuronal markers in neural progenitor cells, *Dev. Biol.* 278 (2005) 60–70.
- [74] M.B. Gonçalves, M. Agudo, S. Connor, S. McMahon, S.L. Minger, M. Maden, J.P. Corcoran, Sequential RARbeta and alpha signalling in vivo can induce adult forebrain neural progenitor cells to differentiate into neurons through Shh and FGF signalling pathways, *Dev. Biol.* 326 (2009) 305–313.
- [75] M. Kim, A. Habiba, J.M. Doherty, J.C. Mills, R.W. Mercer, J.E. Huettner, Regulation of mouse embryonic stem cell neural differentiation by retinoic acid, *Dev. Biol.* 328 (2009) 456–471.
- [76] M. Bibel, J. Richter, K. Schrenk, K.L. Tucker, V. Staiger, M. Korte, M. Goetz, Y.A. Barde, Differentiation of mouse embryonic stem cells into a defined neuronal lineage, *Nat. Neurosci.* 7 (2004) 1003–1009.
- [77] M.P. Stavridis, B.J. Collins, K.G. Storey, Retinoic acid orchestrates fibroblast growth factor signalling to drive embryonic stem cell differentiation, *Development* 137 (2010) 881–890.
- [78] M.A. Sutton, E.M. Schuman, Dendritic protein synthesis, synaptic plasticity, and memory, *Cell* 127 (2006) 49–58.
- [79] J. Aoto, C.I. Nam, M.M. Poon, P. Ting, L. Chen, Synaptic signaling by all-trans retinoic acid in homeostatic synaptic plasticity, *Neuron* 60 (2008) 308–320.
- [80] N. Chen, B. Onisko, J.L. Napoli, The nuclear transcription factor RARalpha associates with neuronal RNA granules and suppresses translation, *J. Biol. Chem.* 283 (2008) 20841–20847.
- [81] B. Maghsoodi, M.M. Poon, C.I. Nam, J. Aoto, P. Ting, L. Chen, Retinoic acid regulates RARalpha-mediated control of translation in dendritic RNA granules during homeostatic synaptic plasticity, *Proc. Natl Acad. Sci. USA* 105 (2008) 16015–16020.
- [82] M.M. Poon, L. Chen, Retinoic acid-gated sequence-specific translational control by RARalpha, *Proc. Natl Acad. Sci. USA* 105 (2008) 20303–20308.
- [83] D.L. Misner, S. Jacobs, Y. Shimizu, A.M. de Urquiza, L. Solomin, T. Perlmann, L.M. De Luca, C.F. Stevens, R.M. Evans, Vitamin A deprivation results in reversible loss of hippocampal long-term synaptic plasticity, *Proc. Natl Acad. Sci. USA* 98 (2001) 11714–11719.
- [84] M.A. Lane, S.J. Bailey, Role of retinoid signalling in the adult brain, *Prog. Neurobiol.* 75 (2005) 275–293.
- [85] K. Shudo, H. Fukasawa, M. Nakagomi, N. Yamagata, Towards retinoid therapy for Alzheimer's disease, *Curr. Alzheimer Res.* 6 (2009) 302–311.
- [86] F. Fahrenholz, F. Tippmann, K. Endres, Retinoids as a perspective in treatment of Alzheimer's disease, *Neurodegener. Dis.* 7 (2010) 190–192.
- [87] T.A. Samad, W. Krezel, P. Chambon, E. Borrelli, Regulation of dopaminergic pathways by retinoids: activation of the D2 receptor promoter by members of the retinoic acid receptor-retinoid X receptor family, *Proc. Natl Acad. Sci. USA* 94 (1997) 14349–14354.
- [88] S. Sell, Leukemia: stem cells, maturation arrest, and differentiation therapy, *Stem Cell Rev.* 1 (2005) 197–205.

- [89] P.P. Pandolfi, Oncogenes and tumor suppressors in the molecular pathogenesis of acute promyelocytic leukemia, *Hum. Mol. Genet.* 10 (2001) 769–775.
- [90] A. Catalano, M.A. Dawson, K. Somana, S. Opat, A. Schwarzer, L.J. Campbell, H. Iland, The PRKAR1A gene is fused to RARA in a new variant acute promyelocytic leukemia, *Blood* 110 (2007) 4073–4076.
- [91] A. Buijs, M. Bruin, Fusion of FIP1L1 and RARA as a result of a novel t(4;17)(q12;q21) in a case of juvenile myelomonocytic leukemia, *Leukemia* 21 (2007) 1104–1108.
- [92] H. de The, C. Chomienne, M. Lanotte, L. Degos, A. Dejean, The t(15;17) translocation of acute promyelocytic leukaemia fuses the retinoic acid receptor alpha gene to a novel transcribed locus, *Nature* 347 (1990) 558–561.
- [93] B.B. Zeisig, C. Kwok, A. Zelent, P. Shankaranarayanan, H. Gronemeyer, S. Dong, C. W. So, Recruitment of RXR by homotetrameric RARalpha fusion proteins is essential for transformation, *Cancer Cell* 12 (2007) 36–51.
- [94] J. Zhu, R. Nasr, L. Peres, F. Riaucoux-Lormiere, N. Honore, C. Berthier, D. Kamashev, J. Zhou, D. Vitoux, C. Lavau, H. de The, RXR is an essential component of the oncogenic PML/RARA complex in vivo, *Cancer Cell* 12 (2007) 23–35.
- [95] L. Di Croce, V.A. Raker, M. Corsaro, F. Fazi, M. Fanelli, M. Faretta, F. Fuks, F. Lo Coco, T. Kouzarides, C. Nervi, S. Minucci, P.G. Pelicci, Methyltransferase recruitment and DNA hypermethylation of target promoters by an oncogenic transcription factor, *Science* 295 (2002) 1079–1082.
- [96] L. Altucci, H. Gronemeyer, The promise of retinoids to fight against cancer, *Nat. Rev. Cancer* 1 (2001) 181–193.
- [97] S. Minucci, C. Nervi, F. Lo Coco, P.G. Pelicci, Histone deacetylases: a common molecular target for differentiation treatment of acute myeloid leukemias? *Oncogene* 20 (2001) 3110–3115.
- [98] K. Petrie, A. Zelent, S. Waxman, Differentiation therapy of acute myeloid leukemia: past, present and future, *Curr. Opin. Hematol.* 16 (2009) 84–91.
- [99] S. Cote, A. Rosenauer, A. Bianchini, K. Seiter, J. Vandewiele, C. Nervi, W.H. Miller Jr., Response to histone deacetylase inhibition of novel PML/RARalpha mutants detected in retinoic acid-resistant APL cells, *Blood* 100 (2002) 2586–2596.
- [100] E. Schachter-Tokarz, C. Kelaidi, B. Cassinat, C. Chomienne, C. Gardin, E. Raffoux, H. Dombret, P. Fenaux, R. Gallagher, PML-RARalpha ligand-binding domain deletion mutations associated with reduced disease control and outcome after first relapse of APL, *Leukemia* 24 (2010) 473–476.
- [101] T. Higuchi, M. Kizaki, M. Omine, Induction of differentiation of retinoic acid-resistant acute promyelocytic leukemia cells by the combination of all-trans retinoic acid and granulocyte colony-stimulating factor, *Leuk. Res.* 28 (2004) 525–532.
- [102] A. Glasow, N. Prodromou, K. Xu, M. von Lindern, A. Zelent, Retinoids and myelomonocytic growth factors cooperatively activate RARA and induce human myeloid leukemia cell differentiation via MAP kinase pathways, *Blood* 105 (2005) 341–349.
- [103] K. Ito, R. Bernardi, A. Morotti, S. Matsuo, G. Saglio, Y. Ikeda, J. Rosenblatt, D.E. Avigan, J. Teruya-Feldstein, P.P. Pandolfi, PML targeting eradicates quiescent leukaemia-initiating cells, *Nature* 453 (2008) 1072–1078.
- [104] M.H. Tatham, M.C. Geoffroy, L. Shen, A. Plechanovova, N. Hattersley, E.G. Jaffray, J. J. Palvimo, R.T. Hay, RNF4 is a poly-SUMO-specific E3 ubiquitin ligase required for arsenic-induced PML degradation, *Nat. Cell Biol.* 10 (2008) 538–546.
- [105] V. Lallemand-Breitenbach, M. Jeanne, S. Benhenda, R. Nasr, M. Lei, L. Peres, J. Zhou, J. Zhu, B. Raught, H. de The, Arsenic degrades PML or PML-RARalpha through a SUMO-triggered RNF4/ubiquitin-mediated pathway, *Nat. Cell Biol.* 10 (2008) 547–555.
- [106] R. Nasr, M.C. Guillemin, O. Ferhi, H. Soilihi, L. Peres, C. Berthier, P. Rousselot, M. Robledo-Sarmiento, V. Lallemand-Breitenbach, B. Gourmel, D. Vitoux, P.P. Pandolfi, C. Rochette-Egly, J. Zhu, H. de The, Eradication of acute promyelocytic leukemia-initiating cells through PML-RARA degradation, *Nat. Med.* 14 (2008) 1333–1342.
- [107] R. Nasr, V. Lallemand-Breitenbach, J. Zhu, M.C. Guillemin, H. de The, Therapy-induced PML/RARA proteolysis and acute promyelocytic leukemia cure, *Clin. Cancer Res.* 15 (2009) 6321–6326.
- [108] C.F. Chen, P. Goyette, D. Lohnes, RARgamma acts as a tumor suppressor in mouse keratinocytes, *Oncogene* 23 (2004) 5350–5359.
- [109] I. Klaassen, R.H. Brakenhoff, S.J. Smeets, G.B. Snow, B.J. Braakhuis, Expression of retinoic acid receptor gamma correlates with retinoic acid sensitivity and metabolism in head and neck squamous cell carcinoma cell lines, *Int. J. Cancer* 92 (2001) 661–665.
- [110] Z. Wang, M. Boudjelal, S. Kang, J.J. Voorhees, G.J. Fisher, Ultraviolet irradiation of human skin causes functional vitamin A deficiency, preventable by all-trans retinoic acid pre-treatment, *Nat. Med.* 5 (1999) 418–422.
- [111] M. Boudjelal, Z. Wang, J.J. Voorhees, G.J. Fisher, Ubiquitin/proteasome pathway regulates levels of retinoic acid receptor gamma and retinoid X receptor alpha in human keratinocytes, *Cancer Res.* 60 (2000) 2247–2252.
- [112] E. Picard, C. Seguin, N. Monhoven, C. Rochette-Egly, J. Siat, J. Borrelly, Y. Martinet, N. Martinet, J.M. Vignaud, Expression of retinoid receptor genes and proteins in non-small-cell lung cancer, *J. Natl Cancer Inst.* 91 (1999) 1059–1066.
- [113] Y.A. Suh, H.Y. Lee, A. Virmani, J. Wong, K.K. Mann, W.H. Miller Jr., A. Gazdar, J.M. Kurie, Loss of retinoic acid receptor beta gene expression is linked to aberrant histone H3 acetylation in lung cancer cell lines, *Cancer Res.* 62 (2002) 3945–3949.
- [114] Z. Zhang, K. Joh, H. Yatsuki, W. Zhao, H. Soejima, K. Higashimoto, M. Noguchi, M. Yokoyama, T. Iwasaka, T. Mukai, Retinoic acid receptor beta2 is epigenetically silenced either by DNA methylation or repressive histone modifications at the promoter in cervical cancer cells, *Cancer Lett.* 247 (2007) 318–327.
- [115] M. Widschwendter, J. Berger, M. Hermann, H.M. Muller, A. Amberger, M. Zeschigk, A. Widschwendter, B. Abendstein, A.G. Zeimet, G. Daxenbichler, C. Marth, Methylation and silencing of the retinoic acid receptor-beta2 gene in breast cancer, *J. Natl Cancer Inst.* 92 (2000) 826–832.
- [116] B. Houle, C. Rochette-Egly, W.E. Bradley, Tumor-suppressive effect of the retinoic acid receptor beta in human epidermoid lung cancer cells, *Proc. Natl Acad. Sci. USA* 90 (1993) 985–989.
- [117] E.F. Farias, K. Petrie, B. Leibovitch, J. Murtagh, M.B. Chornet, T. Schenk, A. Zelent, S. Waxman, Interference with Sin3 function induces epigenetic reprogramming and differentiation in breast cancer cells, *Proc. Natl Acad. Sci. USA* 107 (2010) 11811–11816.
- [118] A. Cras, D. Darsin-Bettinger, N. Balitrand, B. Cassinat, A. Soulie, M.E. Toubert, L. Delva, C. Chomienne, Epigenetic patterns of the retinoic acid receptor beta2 promoter in retinoic acid-resistant thyroid cancer cells, *Oncogene* 26 (2007) 4018–4024.
- [119] A. Parrado, G. Despouy, R. Kraiba, C. Le Pogam, S. Dupas, M. Choquette, M. Robledo, J. Larghero, H. Bui, I. Le Gall, C. Rochette-Egly, C. Chomienne, R.A. Padua, Retinoic acid receptor alpha1 variants, RARalpha1DeltaB and RARalpha1DeltaBC, define a new class of nuclear receptor isoforms, *Nucleic Acids Res.* 29 (2001) 4901–4908.
- [120] T.D. Yan, H. Wu, H.P. Zhang, N. Lu, P. Ye, F.H. Yu, H. Zhou, W.G. Li, X. Cao, Y.Y. Lin, J. Y. He, W.W. Gao, Y. Zhao, L. Xie, J.B. Chen, X.K. Zhang, J.Z. Zeng, Oncogenic potential of retinoic acid receptor-gamma in hepatocellular carcinoma, *Cancer Res.* 70 (2010) 2285–2295.
- [121] T.F. Franke, PI3K/Akt: getting it right matters, *Oncogene* 27 (2008) 6473–6488.
- [122] K. Khetchoumian, M. Teletin, J. Tisserand, M. Mark, B. Herquel, M. Ignat, J. Zucman-Rossi, F. Cammas, T. Lerouge, C. Thibault, D. Metzger, P. Chambon, R. Losson, Loss of Trim24 (Tif1alpha) gene function confers oncogenic activity to retinoic acid receptor alpha, *Nat. Genet.* 39 (2007) 1500–1506.
- [123] S. Hua, R. Kittler, K.P. White, Genomic antagonism between retinoic acid and estrogen signaling in breast cancer, *Cell* 137 (2009) 1259–1271.
- [124] C.S. Ross-Innes, R. Stark, K.A. Holmes, D. Schmidt, C. Spyrou, R. Russell, C.E. Massie, S.L. Vowler, M. Eldridge, J.S. Carroll, Cooperative interaction between retinoic acid receptor-alpha and estrogen receptor in breast cancer, *Genes Dev.* 24 (2010) 171–182.
- [125] T.T. Schug, D.C. Berry, N.S. Shaw, S.N. Travis, N. Noy, Opposing effects of retinoic acid on cell growth result from alternate activation of two different nuclear receptors, *Cell* 129 (2007) 723–733.
- [126] P. Blume-Jensen, T. Hunter, Oncogenic kinase signalling, *Nature* 411 (2001) 355–365.
- [127] D.J. Slamon, G.M. Clark, S.G. Wong, W.J. Levin, A. Ullrich, W.L. McGuire, Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene, *Science* 235 (1987) 177–182.
- [128] M.M. Moasser, The oncogene HER2: its signaling and transforming functions and its role in human cancer pathogenesis, *Oncogene* 26 (2007) 6469–6487.
- [129] N.E. Hynes, G. MacDonald, ErbB receptors and signaling pathways in cancer, *Curr. Opin. Cell Biol.* 21 (2009) 177–184.
- [130] A.M. Tari, S.J. Lim, M.C. Hung, F.J. Esteve, G. Lopez-Berestein, Her2/neu induces all-trans retinoic acid (ATRA) resistance in breast cancer cells, *Oncogene* 21 (2002) 5224–5232.
- [131] H. Srinivas, D.M. Juroske, S. Kalyankrishna, D.D. Cody, R.E. Price, X.C. Xu, R. Narayanan, N.L. Weigel, J.M. Kurie, c-Jun N-terminal kinase contributes to aberrant retinoid signaling in lung cancer cells by phosphorylating and inducing proteasomal degradation of retinoic acid receptor alpha, *Mol. Cell Biol.* 25 (2005) 1054–1069.
- [132] H. Srinivas, D. Xia, N.L. Moore, I.P. Uray, H. Kim, L. Ma, N.L. Weigel, P.H. Brown, J. M. Kurie, Akt phosphorylates and suppresses the transactivation of retinoic acid receptor alpha, *Biochem. J.* 395 (2006) 653–662.
- [133] A. Keriell, A. Stary, A. Sarasin, C. Rochette-Egly, J.M. Egly, XPD mutations prevent TFIIH-dependent transactivation by nuclear receptors and phosphorylation of RARalpha, *Cell* 109 (2002) 125–135.
- [134] J.E. Visvader, Keeping abreast of the mammary epithelial hierarchy and breast tumorigenesis, *Genes Dev.* 23 (2009) 2563–2577.
- [135] D.C. Berry, N. Noy, All-trans-retinoic acid represses obesity and insulin resistance by activating both peroxisome proliferation-activated receptor beta/delta and retinoic acid receptor, *Mol. Cell Biol.* 29 (2009) 3286–3296.
- [136] A.K. Indra, E. Castaneda, M.C. Antal, M. Jiang, N. Messaddeq, X. Meng, C.V. Loehr, P. Grigolio, S. Kato, W. Wahli, B. Desvergne, D. Metzger, P. Chambon, Malignant transformation of DMBA/TPA-induced papillomas and nevi in the skin of mice selectively lacking retinoid-X-receptor alpha in epidermal keratinocytes, *J. Invest. Dermatol.* 127 (2007) 1250–1260.
- [137] N. Martinet, F. Alla, G. Farre, T. Labib, H. Drouot, R. Vidili, E. Picard, M.P. Gaube, D. Le Faou, J. Siat, J. Borelly, P. Vermeylen, T. Bazarbachi, J.M. Vignaud, Y. Martinet, Retinoic acid receptor and retinoid X receptor alterations in lung cancer precursor lesions, *Cancer Res.* 60 (2000) 2869–2875.
- [138] R. Matsushima-Nishiwaki, M. Okuno, S. Adachi, T. Sano, K. Akita, H. Moriwaki, S.L. Friedman, S. Kojima, Phosphorylation of retinoid X receptor alpha at serine 260 impairs its metabolism and function in human hepatocellular carcinoma, *Cancer Res.* 61 (2001) 7675–7682.