## Class 17

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First lets import the transcript count estimates into R

```
library(tximport)
  # setup the folder and filenames to read
  folders <- dir(pattern="SRR21568*")</pre>
  samples <- sub("_quant", "", folders)</pre>
  files <- file.path( folders, "abundance.h5" )</pre>
  names(files) <- samples</pre>
  txi.kallisto <- tximport(files, type = "kallisto", txOut = TRUE)</pre>
1 2 3 4
  head(txi.kallisto$counts)
                 SRR2156848 SRR2156849 SRR2156850 SRR2156851
ENST00000539570
                                            0.00000
                                      0
                                                              0
ENST00000576455
                           0
                                      0
                                            2.62037
                                                              0
ENST00000510508
                          0
                                           0.00000
                                                              0
ENST00000474471
                          0
                                      1
                                           1.00000
                                                              0
ENST00000381700
                           0
                                           0.00000
                                                              0
                           0
                                                              0
ENST00000445946
                                            0.00000
```

These are our estimated transcript counts for each sample. Lets see how many transcripts are in each sample

```
colSums(txi.kallisto$counts)
```

```
SRR2156848 SRR2156849 SRR2156850 SRR2156851
2563611 2600800 2372309 2111474
```

In at least one sample, lets see how many transcripts are detected

```
sum(rowSums(txi.kallisto$counts)>0)
```

## [1] 94561

Lets next filter out annotated transcripts with no reads and those with no change over the samples

```
#Filter out the no reads
to.keep <- rowSums(txi.kallisto$counts) > 0
kset.nonzero <- txi.kallisto$counts[to.keep,]

#Filter out the no change
keep2 <- apply(kset.nonzero,1,sd)>0
x <- kset.nonzero[keep2,]</pre>
```

##Principal Component Analysis

Lets perform PCA on the transcriptomic profiles of these samples. We will make it so that each feature contributes equally to the PCA

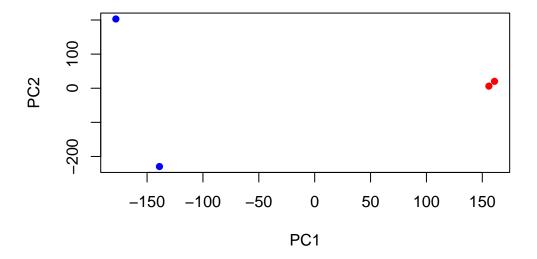
```
pca <- prcomp(t(x), scale=TRUE)
summary(pca)</pre>
```

Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 183.6379 177.3605 171.3020 1e+00
Proportion of Variance 0.3568 0.3328 0.3104 1e-05
Cumulative Proportion 0.3568 0.6895 1.0000 1e+00
```

Now lets use the first two PCs as a coordinate system to visualize the summarized transcriptomic profiles in each sample

```
xlab="PC1", ylab="PC2", pch=16)
```



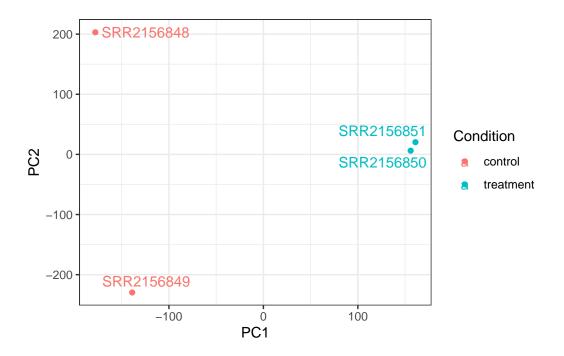
Use ggplot to make a similar figure of PC1 vs PC2 and a seperate figure PC1 vs PC3 and PC2 vs PC3.

```
library(ggrepel)

# Make metadata object for the samples
colData <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(colData) <- colnames(txi.kallisto$counts)

# Make the data.frame for ggplot
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)

ggplot(y) +
   aes(PC1, PC2, col=Condition) +
   geom_point() +
   geom_text_repel(label=rownames(y)) +
   theme_bw()</pre>
```

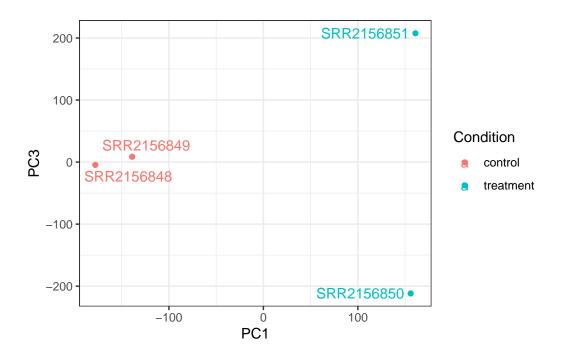


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rownames(colData) <- colnames(txi.kallisto$counts)

# Make the data.frame for ggplot
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)

ggplot(y) +
   aes(PC2, PC3, col=Condition) +
   geom_point() +
   geom_text_repel(label=rownames(y)) +
   theme_bw()</pre>
```

