

Portable Proteomics Pipeline (P3) Labelled (iTRAQ4) Quantification

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Portable Proteomics Pipeline (P3) is a series of protein mass spectrometry data pre-processing pipelines in Docker containers. The packages includes protein identification, filtering, and quantification for both labeled and label-free mass spectrometry data.

This document discuss the `kristiyanto/p3:itraquant` container, a spectrum count quantification pipeline for label-free mass spectrometry protein quantification. The pipeline is based on MSNbase pipeline [4].

Input

The container takes the following files:

```
p3.config # Configuration file
*.mzid    # mzIdentML files resulted from p3:msgf container or other identification tools.
*.mzML    # Mass Spectrometer output files
```

`p3.config` may contain various parameters for p3 related containers. To run spectrum count quantification, `p3.config` must contain the following information:

```
[itraq4]
evaluate_treshold = 1e-10 # Evaluate treshold. Features with evaluate higher than
                           # this value will be discarded.
                           # Check MSNbase: removeNoID documentation for more detailed information.
pNA      = 0              # 0 to 1. Ratio of NA allowed for a feature.
                           # 0 indicates that features with any missing value will be discarded.
                           # Check MSNbase: filterNA documentation for more detailed information.
quant_method = sum        # Quantification method.
                           # Check MSNbase: quantify documentation for more detailed information.
combine_by = mean         # Function used for feature aggregation.
                           # Check MSNbase: combineFeatures for more detailed documentation for more deta.
```

Running the Container

to run the container, a docker engine must be installed. A more information about installing Docker engine is available at <https://docs.docker.com/engine/installation/>. Input files must be mounted to

/root/data within the container. This can be done by using `-v` switch. For MacOS and Windows users, the folder should be located under `C:\Users` or `/Users/`. More information about volumes in Docker containers is available at <http://container-solutions.com/understanding-volumes-docker/>

```
# Download/update the container from DockerHub
docker pull kristiyanto/p3:itraquant
# Run the container
docker run --rm -v /Users/path/files:/root/data kristiyanto/p3:itraquant
```

Output

Once the quantification process is completed, `LabelledQuant.txt`, `evaluate.txt`, and `msnset.rda` are generated. `LabelledQuant.txt` is a tab delimited file with the quantification results, with each column represented from each of the `mzML` file provided. `evaluate.txt` is the raw data of the identification and evaluate for each features. `msnset.rda` is an `msnset` object for the final result that can be easily imported to R for additional analysis.

Pipeline

`kristiyanto/p3:scquant` is based on R, and it uses `MSNbase` [4] package and pipeline. A more detailed information about the pipeline is available at http://bioconductor.org/packages/release/bioc/vignettes/MSnID/inst/doc/msnid_vignette.pdf.

For this documentation, three paired of experiment files are processed.

```
# Files
print(mzid.files)

## [1] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f01.mzid"
## [2] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid"
## [3] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f03.mzid"
```

```
print(mzml.files)

## [1] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f01.mzML"
## [2] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzML"
## [3] "TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f03.mzML"
```

Identification

Identification is performed by using `addIdentificationData()` function from `MSnBase` package.

idSummary(msexp.id)

```
##                                spectrumFile
## 1 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f01.mzML
## 2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzML
## 3 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f03.mzML
##                                idFile coverage
## 1 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f01.mzid    0.977
## 2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid    0.976
## 3 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f03.mzid    0.978
```

fData(msexp.id)[1:3,]

```
##      spectrum scan number(s) passthreshold rank calculatedmasstocharge
## X1.1      1      48      TRUE      1      329.2057
## X1.2      2      48      TRUE      1      323.6872
## X1.3      3      48      TRUE      1      379.2012
##      experimentalmasstocharge chargestate ms-gf:denovoscore ms-gf:evaluate
## X1.1      329.4545      4      117      5.595577
## X1.2      323.9427      4      118      1.038507
## X1.3      379.4494      4      93      4.714499
##      ms-gf:rawscore ms-gf:specvalue assumedissociationmethod
## X1.1      33      5.147427e-07      HCD
## X1.2      48      9.608537e-08      HCD
## X1.3      9      4.315219e-07      HCD
##      isotopeerror isdecoy post pre end start      accession length
## X1.1      1      FALSE      R      R      44      34 ref|NP_775799.2      301
## X1.2      1      FALSE      H      R      1120      1111 ref|NP_075463.2      3013
## X1.3      1      FALSE      S      R      223      212 ref|NP_057257.1      338
##                                description
## X1.1      hypothetical protein LOC161502, gi|148747373 [Homo sapiens]
## X1.2      protein furry homolog, gi|117606355 [Homo sapiens]
## X1.3      hemK methyltransferase family member 1, gi|7705409 [Homo sapiens]
##      pepseq modified modification
## X1.1      DKGKLLIQRSR      FALSE      <NA>
## X1.2      RFLFPQQSLR      FALSE      <NA>
## X1.3      IWIIHLDMTSEK      FALSE      <NA>
##                                idFile
## X1.1 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f01.mzid
## X1.2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid
## X1.3 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f03.mzid
##      databaseFile nprot npes.prot npsm.prot npsm.pep
## X1.1 ID_003632_9011437E.fasta      1      1      2      1
## X1.2 ID_003632_9011437E.fasta      1      2      7      2
## X1.3 ID_003632_9011437E.fasta      1      1      2      1
```

Filter

Filtering is done by: 1) Removing unidentified features, and features with e-value higher than `evaluate_treshold` value described in `p3.config`. 2) features that are assigned by multiple protein groups.

```
# Prior Filtering
length(msexp.id)
```

```
## [1] 47545
```

```
print(evaluate_treshold)
```

```
## [1] 1e-10
```

```
k          <- (fData(msexp.id)$'ms-gf:value'< evaluate_treshold)
k[is.na(k)] <- FALSE
print(sum(k)) # Number of kept features
```

```
## [1] 48
```

```
msexp.filter1 <- removeNoId(msexp.id, keep=k)
# Subsequent value filter
length(msexp.filter1)
```

```
## [1] 48
```

```
msexp.filter2 <- removeMultipleAssignment(msexp.filter1)
# Subsequent multiple assignment filter
length(msexp.filter2)
```

```
## [1] 28
```

```
fData(msexp.filter2)[1:3,]
```

```
##          spectrum scan number(s) passthreshold rank calculatedmasstocharge
## X10257.2      866          11331          TRUE    1          886.9222
## X11009.2     3374          12159          TRUE    1          855.1025
## X11424.2     4757          12615          TRUE    1          914.4767
##          experimentalmasstocharge chargestate ms-gf:denovoscore
## X10257.2          887.4300          2          182
## X11009.2          855.4400          3          176
## X11424.2          914.4795          2          137
```

```
##          ms-gf:evaluate ms-gf:rawscore ms-gf:specevalue
## X10257.2 5.779398e-12          177      5.227042e-19
## X11009.2 9.363158e-17          159      8.292638e-24
## X11424.2 2.077475e-13          129      1.872784e-20
##          assumedissociationmethod isotopeerror isdecoy post pre end start
## X10257.2          HCD              1  FALSE    E  R 326   312
## X11009.2          HCD              1  FALSE    N  K 125   103
## X11424.2          HCD              0  FALSE    G  K 145   130
##          accession length
## X10257.2 ref|NP_006816.2    602
## X11009.2 ref|NP_005902.1    395
## X11424.2 ref|NP_000916.2    359
##
## X10257.2          cytoskeleton-associated protein
## X11009.2          S-adenosylmethionine synthase isoform typ
## X11424.2 pyruvate dehydrogenase E1 component subunit beta, mitochondrial isoform 1 precursor
##          pepseq modified      modification
## X10257.2 STLQTMESDIYTEVR    FALSE      <NA>
## X11009.2 TCNVLVALEQQSPDIAQGVHLDR    TRUE 57.021463735 (2)
## X11424.2 TYYMSGGLQPPIVFR    FALSE      <NA>
##          idFile
## X10257.2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid
## X11009.2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid
## X11424.2 TCGA_13-1489_42-2590_36-2529_117C_W_PNNL_B2S5_f02.mzid
##          databaseFile nprot npep.prot npsm.prot npsm.pep
## X10257.2 ID_003632_9011437E.fasta    1      1      3      1
## X11009.2 ID_003632_9011437E.fasta    1      1      1      1
## X11424.2 ID_003632_9011437E.fasta    1      1      1      1
```

Quantification

Quantification is done by using the MSnBase package [4], with method specified on the `p3.config` file.

```
head(exprs(qnt))
```

```
##          iTRAQ4.114 iTRAQ4.115 iTRAQ4.116 iTRAQ4.117
## X10257.2          NA          NA    97508.10          NA
## X11009.2    25410.31    25410.31    25410.31    25410.31
## X11424.2    27149.91    27149.91    27149.91    27149.91
## X12133.1    43594.16    43594.16    43594.16    43594.16
## X12133.3    24688.00    24688.00    24688.00    24688.00
## X12294.2    12488.43    12488.43    12488.43    12488.43
```

```
print(pNA)
```

```
## [1] 0
```

```
qnt.filtered      <- filterNA(qnt, pNA = pNA)
head(exprs(qnt.filtered))
```

```
##           iTRAQ4.114 iTRAQ4.115 iTRAQ4.116 iTRAQ4.117
## X11009.2    25410.31   25410.31   25410.31   25410.31
## X11424.2    27149.91   27149.91   27149.91   27149.91
## X12133.1    43594.16   43594.16   43594.16   43594.16
## X12133.3    24688.00   24688.00   24688.00   24688.00
## X12294.2    12488.43   12488.43   12488.43   12488.43
## X12353.1    13771.04   13771.04   13771.04   13771.04
```

Results

subsequently, the features are aggregated by accession ID, with the value calculated using the function denoted on the `p3.config`.

```
length(exprs(qnt.filtered))
```

```
## [1] 92
```

```
result <- combineFeatures(qnt.filtered, groupBy = fData(qnt.filtered)$accession, fun=combine_
```

```
## Combined 23 features into 17 using mean
```

```
length(exprs(result))
```

```
## [1] 68
```

```
head(exprs(result))
```

```
##           iTRAQ4.114 iTRAQ4.115 iTRAQ4.116 iTRAQ4.117
## ref|NP_000843.1    43594.156   43594.156   43594.156   43594.156
## ref|NP_000916.2    27149.912   27149.912   27149.912   27149.912
## ref|NP_001073027.1  47457.660   47457.660   47457.660   47457.660
## ref|NP_001348.2    29536.990   29536.990   29536.990   29536.990
## ref|NP_001531.1    11439.398   11439.398   11439.398   11439.398
## ref|NP_001677.2     7962.812    7962.812    7962.812    7962.812
```

References

1. Domon B, Aebersold R: **Mass spectrometry and protein analysis.** *science* 2006, **312**:212–217.
2. Deutsch EW: **Mass spectrometer output file format mzML.** *Proteome Bioinformatics* 2010:319–331.
3. Gatto L, Gibb S: **MSnbase: Labelled and label-free mS2 data pre-processing, visualisation and quantification.** 2016.
4. Gatto L, Lilley K: **MSnbase - an r/Bioconductor package for isobaric tagged mass spectrometry data visualization, processing and quantitation.** *Bioinformatics* 2012, **28**:288–289.