

CZECH TECHNICAL UNIVERSITY IN PRAGUE Faculty of Nuclear Sciences and Physical Engineering



Estimating patient's life expectancy after a successful kidney transplant using machine learning methods

Odhad délky života pacienta po úspěšné transplantaci ledviny pomocí metod strojového učení

Bachelor's Degree Project

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Academic year: 2022/2023

Katedra: matematiky Akademický rok: 2022/2023

ZADÁNÍ BAKALÁŘSKÉ PRÁCE

Student:

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Studijní program:

Aplikovaná informatika

Název práce (česky):

Odhad délky života pacienta po úspěšné transplantaci ledviny pomocí

metod strojového učení

Název práce (anglicky):

Estimating patient's life expectancy after a successful kidney

transplant using machine learning methods

Pokyny pro vypracování:

- 1) Prozkoumejte současný přístup k transplantacím ledvin, jeho problémy a výzvy. / Investigate the current approach to kidney transplantation, its problems and challenges.
- 2) Prozkoumejte příslušné metody strojového učení a metody pro hodnocení přesnosti modelu. /
 - Explore applicable machine learning methods and model accuracy evaluation methods.
- 3) Vyčistěte, předzpracujte a rozšiřte stávající datovou sadu. / Clean, preprocess and extend the existing dataset.
- 4) Vytvořte prediktivní model strojového učení pro odhad délky života pacienta a ohodnoť
te jeho přesnost. /
 - Create a predictive machine learning model estimating a patient's life expectancy and evaluate its accuracy.
- 5) Navrhněte úpravy skórovacího algoritmu pro transplantace ledvin na základě výsledků prediktivního modelu. /
 - Design an updated kidney matching compatibility scoring algorithm based on the prediction model.
- 6) Prozkoumejte možnost integrace dosažených výsledků do nástroje pro správu transplantací TX Matching.
 - Evaluate the possibility of integrating achieved results into kidney transplantation management tool TX Matching.

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Datum zadání bakalářské práce:

31.10.2022

Datum odevzdání bakalářské práce: 2.8.2023

Doba platnosti zadání je dva roky od data zadání.

V Praze dne 31.10.2022

garant oboru

vedoucí katedry

děkan

Acknowledgment: I am grateful to Ing. Tomáš Kouřim for his expert guidance and to Dr. Pavel Strache able support and insightful feedback throughout this project. I would also like to exappreciation to PaedDr Eliška Rafajová for her language assistance.	
Author's declaration: I declare that this Bachelor's Degree Project is entirely my own work and I have listed all in the bibliography.	ll the used sources
Prague, August 2, 2023	Kyrylo Stadniuk

Název práce:

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Abstrakt: Abstrakt max. na 10 řádků. Abstrakt max. na 10 řádků.

Klíčová slova: klíčová slova (nebo výrazy) seřazená podle abecedy a oddělená čárkou

Title:

Estimating patient's life expectancy after a successful kidney transplant using machine learning methods

Author: Kyrylo Stadniuk

Abstract: Max. 10 lines of English abstract text. Max. 10 lines of English abstract text.

Key words: keywords in alphabetical order separated by commas

Contents

1	Intr	oductio	n	8
2	Med	lical Ba	ckground	9
	2.1		story of kidney transplantation	9
		2.1.1	Early Animal Experiments	9
		2.1.2	Early Human Transplantation	9
		2.1.3	First Successes	10
		2.1.4	Attempts in Immunosuppression	11
		2.1.5	Gloom Then Revolution	12
		2.1.6	Plateau	12
		2.1.7	Tissue Typing	12
		2.1.8	Antilymphocyte Serum	13
		2.1.9	Conclusion and challenges of the field	13
	2.2	Immun	nology	13
	2.3		nology of kidney transplant	14
		2.3.1	Immune system activation Peritransplant	14
		2.3.2	Stimulation of Adaptive Alloimmunity	15
		2.3.3	T Cell-mediated rejection	16
		2.3.4	B Cell-mediated rejection	16
		2.3.5	Transplant Tolerance	16
		2.3.6	Factors Influencing Rejection Beyond the Graft - Microbiome	17
3	Mac	hine Le	earning Background	18
	3.1		vised Learning	18
		3.1.1	Linear Regression	19
		3.1.2	Logistic Regression	20
		3.1.3	Support Vector Machines	21
	3.2	Unsup	ervised Learning	24
		3.2.1	Principal Component Analysis (PCA)	25
		3.2.2	Gaussian Mixtures	25
	3.3	Data P	reparation	25
		3.3.1	Handling Categorical Features	26
		3.3.2	Feature Scaling	26
		3.3.3	Handling Missing Feature Values	27
	3.4	Model	Training and Hyperparameter Tuning	27
	3.5	Surviva	al Analysis	28
		3.5.1	Performance Metrics	33

		3.5.2 Cox Proportional Hazards Method	34
		3.5.3 Penalized Cox Models	35
		3.5.4 Random Survival Forests	35
	3.6	Overview of Machine Learning Libraries and Tools	35
		3.6.1 Comparison	35
	3.7	Conclusion	35
	D 4		26
4		a Preparation and Analysis	36
	4.1	Data Loading	36
	4.2	Data preprocessing pipeline	37
	4.3	Exploratory Data Analysis	38
		4.3.1 Survival Data	38
		4.3.2 Age	40
		4.3.3 Donor Type	40
		4.3.4 Gender	42
		4.3.5 The Use of Dialysis	42
		4.3.6 Race	44
	4.4	Dataset building, Exclusion criteria and noice reduction	44
5	Mac	hine Learning Model	45
	5.1	Problem Formulation	45
	5.2	Model selection	45
	5.3	Results	46
		5.3.1 Coxnet	46
		5.3.2 Random Survival Forest	48
		5.3.3 Comparison	50
	5.4	Scoring algorithm	50
	5.5	Limitations	50
	5.6	Further work	51
	5.0	Tuttici work	31
6	App	lications	53
	6.1	Existing Solutions	53
		6.1.1 Txmatching	53
	6.2	KidneyLife	53
	6.2	KidneyLife	53 53
	6.2	KidneyLife	
	6.2	KidneyLife	53

Chapter 1

Introduction

The goal of this paper is to explore fields of kidney transplantation and machine learning, create and apply machine learning model in real-world application.

Chapter 2

Medical Background

2.1 The history of kidney transplantation.

2.1.1 Early Animal Experiments

Advancements in surgical methods and techniques at the beginning of the 20th century eventually led to experiments with organ transplantation. On March 1st, 1902, Emerich Ullman, a physician at the Vienna Medical School, performed the first recorded organ transplantation. He performed an autograft, meaning the transplantation where the donor and the recipient are the same individual. Ullmann utilized the method of vascular suturing developed by Ervin Payr, to connect the dog's kidney to the vessels of its neck. The transplant was successful - the kidney produced urine. The dog was presented the same day to Vienna medical society eliciting significant interest and discussion.

The same year other similar transplantations were made. Another physician, Alfred von Decastello, performed a dog-to-dog kidney allograft at the Institute of Experimental Pathology in Vienna. The kidney produced urine for a while but then stopped working. Later Ullman performed a dog-to-goat kidney xenograft (cross-species transplant), and to his surprise kidney produced some urine, but later stopped.

In Lyon in the department headed by Mathieu Jabourday, his assistants Carrel, Briau and Villard were working on new methods of vascular suturing. In 1902, Alex Carrel published the method of vessel anastomosis now referred to as Carrel's seam. This technique represented a significant improvement over existing methods and effectively addressed the common issues of thrombosis, hemorrhage, stricture, and embolism[13].

Later Carrel moved to the United States where he continued his research on vessel suturing and organ transplantations at The Rockefeller Institute for Medical Research. There he perfected his method and while performing autografts and allografts documented what later would be recognized as "rejection". For his works in 1912, he got the Nobel Prize in Medicine. By this time, his method of suturing had been widely adopted in human surgeries[19].

2.1.2 Early Human Transplantation

The first recorded human renal xenograft was performed by Mathieu Jaboulay in 1906. He chose a pig and a goat as donor animals and performed two xenografts. One kidney was transported to the arm and the second to the thigh. Each kidney functioned for one hour[1, 4].

The second and third human transplants performed by Ernst Unger were far more known. On December 10, 1909, he performed a kidney transplant from a stillborn baby to a baboon. Even though the kidney produced no urine, the postmortem showed that vascular anastomosis (connection of vessels) was

performed successfully. This inspired Unger to perform another transplantation that same month, but this time monkey-to-human xenograft. The kidney was transplanted from an ape to dying from renal failure young woman. The kidney never worked.

These early experiments demonstrated that technically kidney transplantation was possible, but the mechanism of rejection was not yet fully understood. Carrel in his famous lecture about the future of transplantation (1914) to the International Surgical Society mentioned that the works of his colleague at the Rockefeller center J.B. Murphy might seriously impact the development of the field. Murphy found that irradiation and benzol treatment increased the graft survival of cancer in mice. This observation inspired Carrel to conduct his own experiments, wherein he irradiated recipients and found prolonged graft survival, but these experiments were never formally published[11].

The period of the 1930s and 1940s was rather stagnant compared to the beginning of the century. European surgical centers that studied transplantology before were in decline. Moyo Clinic in the US was conducting some cautious experiments without considering Carrol's works and attempts at immunosuppression.[13] However, there was a notable event during this period - the first human-tohuman transplantation. It was performed by Yurii Voronyi (in literature for some reason he is referred to as Voronoy) on March 3, 1933, in Kherson, Ukraine. The recipient was a 26-year-old woman admitted to the hospital on March 3, 1933, with mercury chloride poisoning induced by a suicide attempt the previous day that resulted in acute renal failure. Transplantation seemed the only viable option. It was known from previous experiments by other scientists that no xenograft ever was successful so humanto-human transplantation was the only feasible choice. The option of injuring a living person by organ removal was not even considered. It was known from the physiology that kidneys save their function a couple of hours after the reperfusion with ringer-solution and that organs keep some sterility a couple of hours after the host's death. So temporary cadaver transplantation until the woman's own kidneys would regenerate seemed to be a reasonable option. The transplantation was performed to the thigh's artery and vein using Carrel's seam with some modifications. After some time the kidney started to produce urine for a while but then eventually the allograft failed and 48 hours after the surgery the patient dies. The reason for graft failure was blood group mismatch and too long warm ischemia time - 6 hours, so the kidney began to degrade, resulting in an immune reaction to dead kidney cells and kidney blood cells. Voronyi performed another 5 such transplantations, which he considered as a bridge therapy until the recipient's own kidneys would recover. Kidneys produced urine for different durations from 1 to 7 days with 2 patients eventually recovering and living normally thereafter[12].

2.1.3 First Successes

In 1946, at the Peter Bent Brigham Hospital in Boston, a group of surgeons: Hufnagel, Hume, and Landsteinerhuman performed kidney transplantation under local anesthetic on the arm vessels. The short period of kidney functioning may have helped the patient to recover from acute renal failure. It ignited the hospital's interest in renal transplantation.

Simonsen in Denmark, Dempster in London, and Küss in Paris concluded that it is preferable to place the kidney in the pelvis. Further, both Simonsen and Dempster deduced that the immune response was responsible for graft failure and both hypothesized that the humoral mechanism of rejection was probable.

In the early 1950s, two groups of surgeons based in Paris and Chicago performed pelvic kidney transplants without immunosuppression. In Paris, Jean Hamburger reported the first live-related kidney transplant between a mother and her child. he transplanted kidney began to function immediately. It functioned for 22 days until it was rejected.

A series of nine transplantations with the thigh position of the allograft was closely studied in Boston and the first usage of hemodialysis for the preparation was recorded in Boston by David Hume in 1953. In

some of these cases, mild successes were achieved using the adrenocorticotropic hormone (more known as cortisone). It was hypothesized that the endogenous immunosuppression of uremia was responsible for the results rather than the drug regimen. Hume's findings were substantial as he concluded that prior blood transfusions, blood group matching between the donor and recipient, and host bilateral nephrectomy could be beneficial for the success of the transplant. These conclusions were later confirmed by subsequent studies.

These attempts in the early 1950s taught technical aspects of kidney transplantation and with increased confidence on December 23, 1954 in Boston Joseph Murray performed kidney allograft from one identical twin to another, bypassing the rejection barrier. From that time many similar surgeries were performed in Boston. This caused a lot of talks and predictions but all of them were negated when one of such recipients got pregnant and gave birth to a completely normal infant. However, in retrospective, it didn't bring anything new scientifically, because the technical possibilty of kidney transplantation was evident and the cases of successful skin allografts between identical twins were known for decades, but nonetheless it was an important milestone that aroused the interest in further experiments [1, 4].

2.1.4 Attempts in Immunosuppression

In 1948 at Mayo Clinic patients handicapped by rheumatoid arthrytis were given already mentioned cortisone, adrenal cortical hormone with mild immunosuppressive properties, that relieved their condition. This popularized the research on adrenal cortical hormones, but later it was concluded that the steroid effect was clinically insignifficant for transplantation. After that, the experiments with irradiation, abandoned by Carrel and Murphy, were revitalized. Joan Main and Richmond Prehn showed that weakening of the immune system of adult mice by radiation and consequent skin and bone marrow transplantation from the same donor resulted in skin transplant acceptance. This encouraged teams in Boston and Paris to pursue the similar approach in humans.

In 1958, Murray's team transplantation on humans utilizing the Main-Prehn method conducted lethal total body irradiation (TBI) on two patients with additional bone marrow transplant. Ten more recipients were irradiated with sub-lethal TBI, but without donor bone marrow transplant. As a result 11 patients passed away within a month, the only survivor had sub-lethal TBI without transplanted bone marrow and he got kidney from his non-identical twin brother. This was rather revolutionary - for the first time kidney was not rejected from non-identical twin. The kidney functioned for 20 years. Jean Hamburger and his team performed another fraternal twin transplant utilizing the same irradiation technique. The transplant functioned for 26 years finishing with the recipient's death for rejection-unrelated reason.

Between 1960 and 1962 Kuss and Hamburger performed four successful transplantations between non-twin patients with following TBI. This gave promise that the transplantations could be done in non-twins and potentially between anybody. The research continued.

It was obvious that TBI is not the best choice and that it is necessary to find a substitution. In 1959, Schwarz and Damesehek from Tufts University published paper that described how an anticancer drug 6-mercaptopurine (6-MP) lowered immune response to foreign proteins in rabbits. Roy Calne, a training surgeon at Royal Free Hospital, London, dissapointed with TBI in prolongation of kidney allograft survival in dogs, noticed Schwarz and Dameshek's paper and performed his own experiment in dogs and found that it signifficantly prolonged dog's survival. Charles Zukoski and David Hume found the same outcomes.

6-MP was used in three tranplantations at Royal Free Hospital, but without success, However Kuss and assosiates reported one prolonged graft survival from a nonrelated donor. The TBI was main agent and intermittent usage of azathioprine and prednisone was used as an additional therapy.

Gertrude Elion and George Hitchings provided Roy Calne with the 6-MP derivative - azathioprine. Calne showed even longer graft survival with azathioprine. Both Elion and Hitchings were awarded with

the Nobel Prize for the development of 6-MP and azathioprine. In 1961 Azathioprine became available for human use.

2.1.5 Gloom Then Revolution

In 1963 National Research Council organized a small conference constisting of 25 transplant clinitians and scientists to review the status of kidney transplantation at the moment. 'the discussion was quite depresive. Clinitians presented their results, that were rather discouraging: less that 10% of hundreds performed transplantations survived for more than three months, from patients with TBI only six got to the one year mark. Murray reported that from his first ten patients on 6-MP one survived for a year, others passed away within 6 months, so it was concluded that drugs were not more effective than radiation.

The gloom continued untill Tom Starzl, until then unknown, did his presentation where he described his protocol that allowed graft survival for more than one year in 70% of cases. He was not believed at first, but then he showed medical records of his patients and he was eventually believed. The only thing that differed from other protocols with 6-MP was that addition of prednison. This was a sensation. In the first year after the presentation, 50 new tranplantation programs were founded in US alone. And his protocol became medical world standard for the next 20 years.

2.1.6 Plateau

During the period from 1964 to 1980 nothing groundbreaking had happened, although the steady development was seen. Dialysis become available and thanks to the acumulated exprerience the dosages became more precise. The brain death was accepted and the body was supported for a while to save organs for transplantation.

Hemodialysis for renal failure was created by Willem Kolff from Holland during WWII. But it couldn't be used for chronic renal failure untill 1960 when was invented Teflon arteriovenous conduits for long-term vascular access.

Acceptance of brain death as a real death. Before the mid 60s the kadaver transplantation was limited by the ischemic damage. Now the additional organs were available from "heartbeating kadavers".

Cold for organ preservation. This was suggested in 1905 by Carrel's colleague Charles Guthrerie. Initially, Starzl used total body hypothermia to protect donor organs, but by 1960 switched to infusing cold solution into the portal vein to protect donor livers. In 1963 the infusion of cold solution intravenous in the transplanted kidney has become a standard.

As the organ preservation for more than 6 hours was achieved in mid 60s the exchange of organs between centers has became practical. Initially sharing was local and informal, that roused the worry that the organs could be distributed unequally and that they could be transported outside of the US. This led to Congress passing the National Transplant Act in 1984. The Southeastern Organ Procurement Foundation (SEOPF), founded in 1969 and eventually composed of 12 hospitals in several cities, served as the template for the United Network of Organ Sharing (UNOS) that controls organ allocation and placement, monitors performance of transplant centers and organ procurement organizations, collects data, and controls quality. They kindly provided us with data for this paper.

2.1.7 Tissue Typing

Although tissue typing was suggested by Alexis Carrel in the beginning of 20th century it could not be proven and used until 1958 when Jean Dausset discovered the first human leukocyte antigen (HLA). Testing for antibodies was not reliable until 1964 when Paul Terasaki invented a microxytotoxicity assay. Test included mixing donor's lymphocytes and recipient's serum and quickly has became the standard

and was name crossmatch. For a couple of years Terasaki performed typing for most of U.S. transplant centers and found a couple of observations: 1) Positive cross-match test predicts hyperacute rejection. 2) matching can reliably identify optimal donor within a family. It was assumed that the same would work for non-related recipients.

However, when in 1970 Terasaki reviewed his large database of cadaver renal allografts he found no correlation with the typing. This raised a lot of agitation in tissue typing community and his grant even was temporally suspended until others didn't report the same. Now it is conclded that the

2.1.8 Antilymphocyte Serum

Next mark was cyclosporine, a fungal derivative with immunosuppressive properties discovered in 1976 by Jean-François Borel. It revolutionized the renal and extrarenal transplants, proving to be much better than the previous drug azathioprine. However it also had to be combined with prednisone to gain those results. It was used until 1989 when even more potent drug was discovered - Tacrolimus. It helps even when the cyclosporine with prednisone has no effect.

Tom Starzl discovered that donor leukocyte chimerism was present in patients who had maintained successful kidney or liver grafts for up to three decades.

chimerism is an important cause (not the consequence) of successful transplantation,

successful engraftment is the result of the responses of coexisting donor and recipient cells each to the other causing reciprocal clonal exhaustion followed by peripheral clonal deletion

2.1.9 Conclusion and challenges of the field

The ultimate goal is immunosuppression without drugs because drugs are often toxic and the proper dosing might be tricky.

2.2 Immunology

The immune system is a sophisticated defense mechanism that evolved to protect multicellular organisms from pathogens such as bacteria, fungi, viruses, and parasites. It consists of many cells and tissues that compose a complex system that detects, evaluates, and responds to the invader. The immune system is divided into humoral and cell-mediated immunity. Humoral is mediated by soluble immunoglobulin proteins referred to as antibodies, while cell-mediated involves pathogen-specific T Lymphocytes that either destroy the invader or assist other cells in doing so. Both are essential for a complete immune response.

Lymphocyte is a type of white blood cell that is responsible for both humoral and cell-mediated immune responses. There are two types of lymphocytes: T lymphocytes (T cells) and B lymphocytes (B cells). B cells mediate humoral response by producing antigen-specific antibodies. An antigen is any molecular structure that binds to an antibody or specific surface T cell receptor, triggering an immune response. Once B-cell encountered an antigen it starts to produce antibodies specific to it, antibodies then bind to it, marking the invader for destruction. T cells when encountering an antigen start to proliferate forming an army of T cells that will eliminate the invader and will form long-term memory about the pathogen.

Physical barriers: epithelia and mucous membranes constitute the first line of defense. To activate the immune system the pathogen must first breach physical barriers. The immune system categorizes pathogens by common characteristics and designs its response accordingly. Pathogen detection and categorization rely on the interaction between pathogen and T-cell receptors, as well as soluble antibodies.

Binders for T cell receptors and antibodies can be the whole pathogen's body, its part, or molecules excreted by it.

Pathogens are recognized and categorized by molecular patterns that are associated with a particular pathogen and are referred to as pathogen-associated molecular patterns (PAMPs). Pathogen recognition receptors (PRRs), which are excreted by white blood cells, bind to PAMPs initiating the cascade of events that will mark a pathogen for destruction.

Pathogen-host interaction is a continuous arms race, as pathogens usually have a short life cycle and can modify their DNA to elude the host's recognition systems. The generation of diversity in developing cells is designed to combat this. When lymphocytes are developing in bone marrow random PRRs are generated, then cells are tested on non-reactivity to host cells. If the test is passed the cell is released into circulation. The principle of recognizing self vs. non-self is called tolerance.

There are two interconnected systems of response: innate and adaptive. Innate includes primitive built-in cellular and molecular mechanisms aimed at preventing infections and quickly demolishing common pathogens. It consists of physical and molecular barriers as well as PRRs that are encoded in DNA and therefore are inherited. Innate immunity provides a fast and effective response which however is not very specific and cannot differentiate small differences. Adaptive immunity is constituted by both humoral, where antibodies neutralize and eradicate extracellular microbes and toxins, and cell-mediated immunity, where T lymphocytes exterminate intracellular invaders.

Adaptive immunity is much slower but more able to recognize small differences. It typically starts to act within 5 to 6 days after initial exposure. Because it takes time to create an army of cells with specific receptors. After pathogen extermination, some of the lymphocytes with the specific receptor become memory cells, making it easier to fight this type of pathogen.[13]

In conclusion, the immune system is a complex network of molecules, cells, tissues, and organs that cooperate in protecting the organism from pathogens. The system can be divided into two main branches: innate and adaptive, which cooperate in protecting the host from infections while developing long-term immunity to specific pathogens. Understanding the mechanisms of the immune system is essential to understanding the domain of kidney transplantation.

2.3 Immunology of kidney transplant

The process of transplantation inevitably includes termination of blood flow, and, as a result, oxygenation. Therefore cell is unable to generate sufficient amount of energy to maintain homeostasis, leading to damage or death. Damage or death is associated with DAMP release that might be detected by both innate and adaptive immunity.

2.3.1 Immune system activation Peritransplant

The process of transplantation inevitably includes termination of blood flow, and, as a result, oxygenation. Therefore cell is unable to generate sufficient amount of energy to maintain homeostasis, leading to damage or death. Damage or death is associated with DAMP release that might be detected by both innate and adaptive immunity. Mostly it is the ancient innate immunity that is activated with its soluble arm - complement system.

Damage Signals Many DAMPS are recognized by the same PRRs that mediate response to PAMPs. These DAMPS include molecules that are normally hidden from the immune system and are produced during ischemia, such as extrecellular ATP, heat shock proteins (HSPs), uric acid, etc. Likewise, oxidative stress and decline in intracellular potassium may act as intracellular damage signals.

Complement Complement system is comprised of series of proteine kinases that are sequentially activated resulting in membrane attack complex (MAC) formation. MAC include complement components C5 to C9, which are inserted into pathogen cell membrane resulting in compromising cell integrity leading to cell death.

There are three pathways of complement system activation: the classical pathway, the alternative pathway, and the mannose-binding lectin (MBL) pathway. The classical pathway is activated by IgM and IgG antibodies and participates in antibody-mediated rejection, that will be discussed further. Alternative complement is always active and therefore must be controlled by a regulatory proteins, to prevent inadequate responses. The MBL pathway is activated by damaged endothelium, a cell tissue that covers organs and vessels, and carbohydrates present on pathogens. Either pathway results in C3 convertase that cleaves C3. This cleavage leads to a cascade of reactions that culminate in MAC formation.

Long ischemia time results in endothelial cell damage that is acossiated with ischemia-reperfusion injury (IRI). IRI activates MBL and alternative complement pathways.

Gene silencing using small interfering RNA (siRNA) might be a promising instrument in organ transplantation, because it can be applied to an allograft during cold reperfusion and it has been shown to mitigate IRI in animal models. Other strategies of supressing local complement activation would also be useful.

2.3.2 Stimulation of Adaptive Alloimmunity

Immune response to a graft occurs in two main stages: afferent and efferent arms. In afferent stage, recipient lymphocytes are stimulated by donor antigens and start to proliterate and send signals to other cells. In efferent arm, leukocytes migrate to the transplanted organ and donor specific antibodies are produced.

For the immune system to be activated graft must express antigens that will be considered by the host's immune system as foreign. These include ABO antigens, human leukocyte antigens (HLA), and polymorphic non-HLA "auto-antigens".

ABO Blood Group Antigens

ABO system is used to group blood into groups, based on presence or absense of antigens on a blood cell surface. There are four major blood groups: A, B, O and AB.[20]

When allocating an organ to transplant the first thing that is considered is ABO blood group antigens compatibility. ABO antigens are expressed almost by any cell in the allograft, and if the transplantation to be carried out in ABO-incompatible donor and recipient it would result in a hyperacute antibodymediated rejection.

Donors with blood group O are so called "universal donors". Organs from them can be safely transplanted to recipients with any ABO blood group. Whereas, recipient with AB group can safely receive organ from recipient with any ABO blood group and is called a "universal recipient".[9]

HLA

Histocompatibility antigens are genetically encoded antigens that cover cell surfaces. They differ between individuals of the same species and therefore trigger an immune response in case of allograft. In all vertebraes histocompatibility antigens are divided into single major histocompatibility complex (MHC) and numerous minor histocompatibility (miH) systems. In case of either MHC or miH incompatibility the result is an immune response to the graft, more severe in case of MHC than miH. Rejection

1able 2.1: MH	C class division
MHC class I	MHC class II

MHC class I	MHC class II
HLA-A	HLA-DR
HLA-B	HLA-DP
HLA-C	HLA-DQ

in MHC-compatible donor-recipient pair is usually delayed, in some cases forever. Although, sometimes miH mismatch might be so severe that it would be comparable to full MHC mismatch.

MHC antigens are proteins that cover cell surfaces to help the immune system to recognize self vs. non-self. Major histocompatibility comples is divided into MHC class I and MHC class II. MHC class I cover surfaces of most cells and are liable for activation of cytotoxic CD8 cells, that help to find and destroy infected cells. MHC class II are found on certain immune cells and play crucial role in immune response coordination. In humans MHC class 1 are divided into three subgroups each, as can be seen on table

In clinical practice, clinicians asses and try to match donors and recipient according to the number of HLA-A, -B, and -DR mismatches, ranging from zero mismatches (0-0-0) to a maximum of 6 mismatches (2-2-2). Generally more emphasis is placed on DR loci due to capability of CD4 T cell activation, which might trigger both humoral and cellular adaptive immune responses.

Minor histocompatibility proteins can act as antigens, although weaker than MHC. However if prior sensitisation exists it could result in severe immune response that might result in graft loss.

2.3.3 T Cell-mediated rejection

T cell-mediated rejection or TCMR is the most common type of allograft rejection, as it still happens in 20% of transplantations mostly within first 6 months posttrasplant. Immune system cells migrate through vessels to the graft, become activated and start to attack the organ. Complement may also play role in it.

2.3.4 B Cell-mediated rejection

B cells are immune system cells that produce antibodies. Alloantibodies are antibodies that react to donor-specific HLA antigens and might cause hyperacute rejection, acute antibody-mediated rejection (ABMR), and chronic ABMR. About 30% of patients have sensitivities and have certain HLA antibodies. It might cease transplantation or require antibody suppression strategy. Even low amount of antibodies below crossmatch cutoff doubles the risk of ABMR and increases the risk of graft failure by 76%. Additionally, donor specific antibodies might develop posttransplant and cause an actue ABMR.

Acute AMBR is rarely seen in patients withour prior sensitization and is highly difficult to treat. AMBR is characterized by decline in allograft function, presense of DSA and signs of acute vascular injury. A progressive reduction in graft function over time is observed almost universally.

2.3.5 Transplant Tolerance

Taking into account the detrimental effect of long-term immunosuppression one of the primary objectives in transplantation is the induction of immunologic non-responsiveness (tolerance) to an allograft. The are a couple a pathways of immune non-responsiveness generation described in literature, however it haven't gone further animal models yet.

2.3.6 Factors Influencing Rejection Beyond the Graft - Microbiome

Human body is very complex system where every subsystem influences other subsystems and the whole system in general. It is clear that gut microbiome has a profound influence on immune system. It is possible microflora on the allograft might cause rejection. Immunosupression, profilactory antibbiotics, diet changes and other restrictions assossiated with organ transplantation result in decrease in gut microbiome diversity that result in systematic inflammation, that might contribute to alloimunnity, as well as autoimunnity.

Chapter 3

Machine Learning Background

Machine learning is a subfield of computer science that consists of building algorithms capable of processing large amounts of data, finding patterns, and performing actions such as predictions or generating new data. It is an intersection of many fields of science, such as statistics, theory of probability, linear algebra, calculus, and certainly, computer science.

Machine learning excels in problems that are either overly complex or have no known algorithm.[9] It can help us generate knowledge. We can extract previously unknown correlations from the data and build knowledge. It might make fewer errors in decision-making than humans.

Based on the problem and, therefore, on our approach to building a dataset and the model, machine learning can be divided into four subfields: *supervised*, *semi-supervised*, *unsupervised*, and *reinforce-ment learning*. *Supervised learning* means that data is labeled, and we want to predict labels for the unlabeled data. The term labeled data is explained in the following section dedicated to supervised learning. Unsupervised learning deals with unlabeled data.

Semi-supervised learning deals with partially labeled data, and we need to label it fully either manually, or using techniques such as *clustering*.

In *reinforcement learning*, we create an environment, set up rewards for performing certain actions and punishment for others, and let the machine (actor) perform actions that produce the highest reward.

Every field is affected by human errors, and medicine is no exception. Machine learning also makes mistakes, but if we manage to get at least 1% fewer errors than humans make, this will be a substantial achievement. The human body is a complex system, where it is very difficult to comprehend all processes and and how they relate to each other. In addition, machine learning can help us gain insight into them through accumulated data and discover new relations between them.

In this chapter, we will cover all theoretical backgrounds that might prove useful for solving our problem, including classical machine learning, statistical survival analysis, basic steps that are required to **create machine learning systems**, and data preprocessing. We will begin by exploring supervised learning.

3.1 Supervised Learning

Supervised learning is the process of training a model on data where the outcome is known, to make predictions for data where the outcome is not known[12]. *Classification* and *regression* are common supervised learning tasks. In this section we will define these problems and the necessary terminology, and describe commonly used algorithms that are used to solve these types of problems.

In supervised learning the *dataset* is the collection of labeled examples $\{(\overline{x}_i, y_i)\}_{i=1}^N$, where each individual \overline{x}_i is called a *feature vector*. A feature vector is a vector that in each its dimension j = 1, ..., D

contains a value that describes an example in some way. This value is called a *feature* and is denoted as $x^{(j)}$. The *label* y^i might be either a finite set of classes $\{1, 2, ..., C\}$, in case of a classification task, or a real number, a vector, a matrix or graph, in case of a regression. The goal of supervised learning algorithm is to create a model using the dataset that will take the feature vector as an input and produce a label or a more complex structure as an output.

Classification is a problem of assigning a label to an unlabeled example. This problem is solved by a classification learning algorithm that takes a labeled set of examples as input and produces a model that takes an unlabeled example as input and outputs a label. If the set of labels has only two classes we talk about *binary classification*. Consequently, if the set of labels has three or more classes, it is a *multiclass classification*. Some algorithms are binary classifiers by definition while others are multiclass classifiers. It is possible to create an *ensemble* out of binary classifiers that will be able to perform multiclass classification. An ensemble is a combination of algorithms that are connected to perform one task.

Regression is a problem of predicting a *target value* given an unlabeled example. The problem is solved by a regression learning algorithm that takes a set of labeled examples as input, and produces a model that takes an unlabeled example as input and outputs a target value.

Classification and regression tasks are similar in many ways and often for each classifier there is an equivalent regressor, and vice versa. In the following subsections we are going to explore some techniques for supervised learning.

3.1.1 Linear Regression

Linear regression is a popular regression learning algorithm. The model produced is a linear combination of all features.

The problem formulation we are trying to solve is as follows: Given a collection of labeled examples $\{(\overline{x}_i, y_i)\}_{i=1}^N$, create a model

$$f_{\overline{w},b}(\overline{x}) = \overline{wx} + b,\tag{3.1}$$

where N is the size of the collection, \overline{x}_i is a *feature vector* of D dimensions of example $i=1,\ldots,N$, every feature $x_i^{(J)} \in \mathbb{R}$, $y_i \in \mathbb{R}$ is the target value. \overline{w} is a D-dimensional vector of parameters and $b \in \mathbb{R}$. Notation $f_{\overline{w},b}(\overline{x})$ means that f is parametrized by \overline{w} and b.

To train the linear regression means to find optimal values (\overline{w}^*, b^*) of parameters \overline{w} and b so that the model makes as accurate predictions as possible. In graphical terms, it means finding such a hyperplane that fits data points from the training set as well as possible, as shown in image 3.1.

To find optimal parameters we need to minimize the following expression:

$$\frac{1}{N} \sum_{i=1}^{N} (f_{\bar{w},b}(\bar{x_i}) - y_i)^2.$$
 (3.2)

It is called *mean squared error (MSE)*, the *loss function* that comprises of *squared error loss* $(f_{\bar{w},b}(\bar{x_i}) - y_i)^2$, another loss function that evaluates individual predictions. The loss function measures the model's overall performance (MSE) or evaluates each prediction (square error loss).

There is a *closed-form solution* for finding optimal values (\overline{w}^*, b^*) . A closed-form solution is a simple algebraic expression that gives the result directly. In case of linear regression, it is the *normal equation*, and it looks like the following:

$$\overline{\mathbf{w}}^* = (\mathbf{x}^{\mathrm{T}}\mathbf{x})\mathbf{x}^{\mathrm{T}}\mathbf{y}. \tag{3.3}$$

Where x^T means transposed feature matrix x.

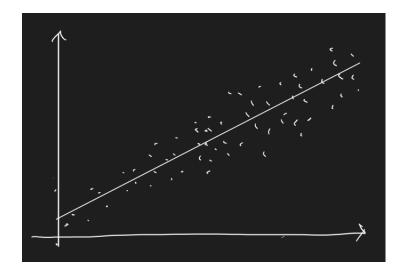


Figure 3.1: Linear regression for two-dimensional data

We could select another loss function, but according to Andriy Burkov, it would be a different algorithm. For example, we could take the absolute difference between $f(x_i)$ and y_i but that would create problems as the derivative of absolute value is not continuous. Therefore the function is not smooth, which might create unnecessary complications during the optimization process.

Linear models are usually resilient to overfitting because they are simple. The model overfits when it learns the intricacies of the training dataset so well that it remembers actual values instead of learning the underlying pattern. Such model is unable to make accurate predictions when confronted with unseen data. More on overfitting in section 3.6.

3.1.2 Logistic Regression

Logistic regression is a binary classifier that estimates the probability of an example belonging to a particular class. If the predicted probability of the instance belonging to a class is greater than 50%, then the model concludes that it belongs to the class (referred to as positive class and labeled as 1). Otherwise, it predicts that the example does not belong to that class (but belongs to the negative class, labeled 0). Logistic regression comes from statistics where its mathematical formulation is similar to a regression, hence the name. Multiclass classification is available in softmax regression, a multiclass variant of logistic regression.

As with linear regression, in logistic regression, we want to model y as a linear combination of \overline{x} , but in this case, it is not that straightforward.

The logistic regression model looks like the following:

$$f_{\bar{w},b}(\bar{x}) \stackrel{def}{=} \frac{1}{1 + e^{-(wx+b)}}.$$
 (3.4)

Similar to linear regression, our task is to find optimal values (\overline{w}^*, b^*) for parameters \overline{w} and b.

Once we found (\overline{w}^*, b^*) for the 3.4, in other words, we trained the model, we can apply the model 3.4 on features x_i from an example (x_i, y_i) . The output value lies in the range $0 . If <math>y_i$ is the positive class, the likelihood of y_i being a positive class is given by p. Consequently, if y_i is the negative class, the likelihood for it being the negative class is given by 1 - p.

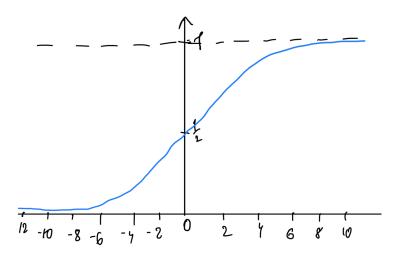


Figure 3.2: Logistic function

In the figure we can see that if the y has a value lower than $\frac{1}{2}$, it has negative x values and will be marked as a negative class. If y is greater than $\frac{1}{2}$, it is positive. Although, depending on the context, the threshold may be different.

In logistic regression, instead of *minimizing* MSE we are trying to *maximize* the *likelihood function*. In statistics, the likelihood function tells how likely the example is according to our model. The objective function in logistic regression is called *maximum likelihood*. It looks like the following:

$$L_{\bar{w},b} \stackrel{def}{=} \prod_{i=1...N} f_{\bar{w},b}(\bar{x}_i)^{y_i} (1 - f_{\bar{w},b}(\bar{x}_i))^{(1-y_i)}. \tag{3.5}$$

On the other hand, due to the exponential function in equation 3.5, it is better to use the *log-likelihood* instead, to make calculations easier. As *Log* is a strictly increasing function, maximizing it is the same as maximizing its argument. The solution to this optimization problem is the same as the solution to the original problem. The log-likelihood function looks like the following:

$$LogL_{\bar{w},b} \stackrel{def}{=} ln(L_{\bar{w},b}(\bar{x})) \sum_{i=1}^{N} y_i ln f_{\bar{w},b}(\bar{x}) + (1 - y_i) ln (1 - f_{\bar{w},b}(\bar{x})). \tag{3.6}$$

Unfortunately, there is no closed-form solution for this optimization problem. Nonetheless, the function is convex, hence gradient descent (or any other optimization algorithm) pretty much guarantees the finding of the global minimum, provided that the learning rate is not too large and enough time is given.

3.1.3 Support Vector Machines

Support vector machine (SVM) is a widely-used and powerful machine learning algorithm that can perform a wide range of tasks, including linear and nonlinear classification, regression, and outlier detection on small- to medium-sized datasets.

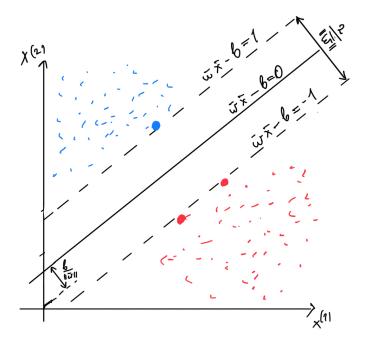


Figure 3.3: SVM demonstration for two-dimensional dataset

Linear SVM

In its classical formulation, the support vector machine is a binary classifier. Classes are called positive and negative and are labeled +1 and -1, respectively.

The model is described by the equation

$$f(x) = sign(\overline{wx} - b).$$

The function sign returns +1 if the input is positive, and -1 if it is negative. To train the SVM means to find optimal values (\overline{w}^*, b^*) of parameters \overline{w} and b so that the model makes as accurate predictions as possible. The process of finding (\overline{w}^*, b^*) is called training.

The concept behind support vector machines is demonstrated in Figure 3.3. The image consists of two classes represented by red and blue dots, divided by a solid line termed the *decision boundary* $\overline{wx} - b = 0$, with two dashed lines by its sides known as *support vectors* $\overline{wx} - b = 1$ and $\overline{wx} - b = -1$. Support vectors are defined by the closest instances of a class to the decision boundary. These instances are emphasized in the figure.

The distance between the closest instances of two classes is called *margin* and is equal to $\frac{2}{\|\overline{w}\|}$, where $\|\overline{w}\|$ is the Euclidean norm and \overline{w} is a parameter vector of the same dimensionality as the feature vector. Thus, the smaller the norm, the larger the margin. The larger the margin, the better the model's generalization. The primary objective of the model is to find the largest possible margin $\frac{2}{\|\overline{w}\|}$, so, to do that we need to *minimize* the Euclidian norm defined by the expression

$$||\overline{w}|| = \sqrt{\sum_{j=1}^{D} (w^{(j)})^2}.$$

The fundamental assumption of support vector machines is that classes are linearly separable, implying their instances can be separated by a hyperplane (decision boundary) with no examples of one class

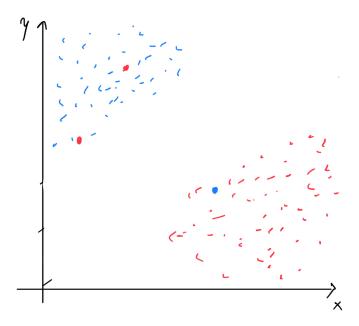


Figure 3.4: Linearly non separable dataset

lying among the ones of the opposite class. It is illustrated in the figure 3.4. In this case, the algorithm won't be able to find an optimal solution with no instances lying between the support vectors and the decision boundary. Consequently, the model is highly sensitive to outliers.

Every optimization problem requires constraints, and for the support vector machine, they are the following:

- 1. $\overline{wx_i} b \geqslant +1$ if $y_i = +1$
- 2. $\overline{wx_i} b \le -1$ if $y_i = -1$.

These two equations can be reduced to one $y_i(\overline{wx} - b) \ge 1$.

The optimization problem we want to solve is the following: Minimize $\|\overline{w}\|$ subject to constraint $y_i(\overline{wx_i} - b) \ge 1$ for i = 1, ..., N, where N is the number of features. This problem can be modified so that the quadratic programming techniques could be used in the optimization process. The modified formula is $\frac{1}{2} \|\overline{w}\|^2$, and minimization of it would also mean minimization of $\|\overline{w}\|$. The updated optimization problem looks like this:

$$min\frac{1}{2}||\overline{w}||^2 \text{ such that } y_i(\overline{wx_i} - b) \ge 1, i = 1, ..., N$$
(3.7)

Handling Noise

To introduce the ability of SVM to handle nonlinearly separable data (but not to the extreme), we define the hinge loss function: $max(0, 1 - y_i(\overline{wx_i} - b))$. It is zero if the constraints 1 and 2 are satisfied. If it is not, the data point does not lie on the right side of the decision boundary. The function value is proportional to the distance from the decision boundary. The resulting cost function looks like the following:

$$C\|\overline{w}\|^2 + \frac{1}{N} \sum_{j=1}^{N} \max(0, 1 - y_i(\overline{wx_i} - b)),$$
 (3.8)

where C is the hyperparameter that determines the trade-off between increasing the size of the decision boundary and ensuring that each x_i lies on the correct side of the decision boundary. Its value is chosen experimentally. C handles the trade-off between classifying the training data well and classifying future examples well (generalization). For higher values of C, the misclassification error will be almost negligible, so the algorithm will try to find the highest margin without considering it. For lower values of C, the algorithm will try to make fewer mistakes by sacrificing the margin size. (A larger margin is better for the generalization.) Lower values lead to wider streets and more margin violations, higher values lead to narrower streets and fewer margin violations.

SVM with the hinge loss function is called *soft-margin SVM* while the original formulation that optimizes the Euclidian norm is referred to as *hard-margin SVM*. *Soft margin classification* tries to mitigate the downsides of the *hard margin classification* by trying to find a balance between keeping the margin as large as possible and mitigating the margin outliers (instances that lie on the margin or on the opposite side).

Handling Non-linearity

We can adapt SVM to work with nonlinearly separable datasets by applying the kernel trick. The kernel trick means transforming the original space to a higher dimensional one during the cost function optimization with the hope that, in higher dimensional space, it will become linearly separable. In mathematical language: the kernel trick is mapping $\varphi: \overline{x} \to \varphi(\overline{x})$, where $\varphi(\overline{x})$ is a vector of higher dimensionality than \overline{x} . The kernel trick allows us to save a lot of non-necessary computations.

There are multiple kernel functions. The most widely used are linear, polynomial, radial basis function (RBF),

3.2 Unsupervised Learning

Unsupervised learning deals with a dataset that does not have labels. There are three main branches of unsupervised learning: clustering, dimensionality reduction and anomaly detection. Clustering is a method that identifies similar instances and groups them into sets. It has applications in data analysis, namely, exploratory data analysis (EDA), customer segmentation, dimensionality reduction, and anomaly detection. Clustering might be either soft, where an instance has a score of belonging to a particular cluster, or hard, where an instance belongs to only one class. The score might be the distance from the cluster centroid or an affinity (similarity score).

Dimensionality reduction is useful for visualization and for the acceleration of learning. Datasets often have a lot of redundant data or the task requires a lot of features. Many algorithms, such as linear models, SVMs, decision trees, might have their performances compromised due to high-dimensional data. So called *curse of dimensionality* states that high dimensional data can cause slow learning and prevent us from getting an optimal model. Consequently, the reduction of the data dimensionality might be a good idea. However, it is worth noting that a dimensionality reduction algorithm might lose some useful information. A lot of modern algorithms, such as neural networks or ensemble algorithms, handle high dimensional data very well, and dimensionality reduction techniques are used less than in the past. However, they are still used for data visualization and cases when we need to build an interpretable model while we are limited in theb number of algorithms we can use.

Anomaly (outlier) detection involves the detection of instances strongly deviating from the norm. These instances are called *outliers* or anomalies while regular ones are referred to as *inliers*. Anomaly detection has many applications. For example, it can be used as a data preprocessing step to remove outliers from the dataset, which might improve the performance of the resulting model. In addition, it is used in the *fraud detection* task and the detection of faulty products in manufacturing facilities.

Novelty detection is closely related to anomaly detection. The only difference is that novelty detection assumes that the training dataset was not contaminated by outliers while anomaly detection does not make this assumption.

3.2.1 Principal Component Analysis (PCA)

Principal components are vectors that define a new coordinate system. The first vector goes in the direction of the highest variance. The second vector is orthogonal to the first one and goes in the direction of the second highest variance, and so on. If we were to reduce dimensionality to $D_{new} < D$, we would pick D_{new} largest principal components and project instances onto them.

It is not advised to choose the number of dimensions arbitrarily. It is recommended to choose a number of dimensions that preserves a large amount of variance (e.g. 95%), or in case of visualization to reduce the number of dimensions down to two or three. There are different versions of PCA; kernel PCA, Incremental PCA (online or batch PCA), and Randomized PCA.

3.2.2 Gaussian Mixtures

Gaussian mixtures is a common algorithm that can be used for anomaly detection. Gaussian mixtures assume that the dataset is generated by several Gaussian distributions. Any instance lying in a region of low density is an anomaly. The density threshold has to be specified. If one gets too many false positives (good products labeled as faulty) they need to decrease the threshold. Consequently, if we get too many false negatives (faulty products labeled as good) the threshold has to be increased. Gaussian mixtures belong to soft clustering. Gaussian mixtures require the number of clusters to be specified. It needs to be run a couple of times to avoid suboptimal solutions.

3.3 Data Preparation

Due to factors such as curse of dimensionality and inherent noise, we cannot load raw data to an algorithm and expect good performance. Most often, the raw data has too many features and most of them have very little predictive power. We need to build a dataset first. *Feature engineering* is responsible for transforming raw data into a dataset. It is a labor-demanding process that requires creativity and, most importantly, domain knowledge.

The objective of this stage is to create *informative* features or features with *high predictive power*. For example, in our task of predicting survival time, donor-recipient blood group compatibility or recipient's age is likely to have much higher predictive power than the donor's or recipient's citizenship.

Moreover, it is possible to create new features with higher predictive power out of those with low predictive power. For example, the calculation of *estimated Glomerular Filtration Rate (eGFR)*, the metric of kidney function estimated on a patient's age, gender, and serum creatinine level, could potentially give more information to the learning algorithm than all features separately.

In the following subsections, we will cover some popular feature engineering techniques.

3.3.1 Handling Categorical Features

The majority of machine learning algorithms primarily operate with numerical features. To handle categorical features (the ones with only a few possible values), such as the age group or a blood group, we can use *one-hot encoding* to convert them to several binary ones. For instance, let's consider a blood group feature comprised of four primary blood groups: A, B, AB, and O. We can convert each blood group into a vector of four numerical values:

$$A = [1, 0, 0, 0]$$

$$B = [0, 1, 0, 0]$$

$$AB = [0, 0, 1, 0]$$

$$O = [0, 0, 0, 1]$$

This technique will increase the dimensionality of the dataset but this is a trade-off we have to make because if we were to assign a number to each group (1 to A, 2 to B, etc.), that would imply gradation or ranking among these categories, while there is none.

However, if the categorical feature does suggest some gradation, for example, university marks as "fail", "average", "good", or "excellent", an enumeration of each value will be appropriate. This practice of assigning a number to categories that have ranking is called *ordinal encoding*.

Binning (or *bucketing*) is the technique used for converting numerical values into multiple binary features called *bins* or *buckets*. For example, a patient's age can be transformed into age-range bins: 0 to 18 years old, 18 to 25 y.o., 25 to 40 years old, and so on. This technique might help an a laearning algorithm learn better, particularly with smaller datasets.

3.3.2 Feature Scaling

Different ranges of feature values might pose a problem to some machine learning algorithms as they do not handle them very well. It might result in a slower training time or a poorer performance. This problem is solved by *normalization* and *standardization* scaling techniques.

Normalization (also known as *min-max scaling*) is a technique of converting an actual range of numerical feature values into a standard range of values: [-1, 1] or [0, 1] without losing any information. The normalization formula for value $x^{(j)}$ for feature j, looks like the following:

$$\bar{x}^{(j)} = \frac{x^{(j)} - min(j)}{max(j) - min(j)},$$

where min(j) and max(j) are minimal and maximal values of feature j.

Standardization is a scaling technique that scales numerical data in such a way that after scaling, it has properties of the *standard normal distribution* with the mean μ =0 (average value) and the standard deviation from the mean σ = 1. The standardization formula for value $x^{(j)}$ for feature j, looks like the following:

$$\hat{x}^{(j)} = \frac{x^{(j)} - \mu^{(j)}}{\sigma^{(j)}}.$$

Typically, standardization is used for supervised learning, in case feature values are formed by standard distribution (bell curve) or a feature has outliers. In other cases, the normalization is preferred.

3.3.3 Handling Missing Feature Values

Datasets frequently have missing values and to handle them, we have one of the following options:

- 1. **Removal of rows with missing values**. The most direct and straightforward approach to managing missing data. If missing values are sparse or the dataset is large enough, the usage of this technique would be appropriate.
- 2. **Feature removal**. If the dataset has a feature with an excessive amount of missing values relative to its size, it is better to remove the feature.
- 3. **Regression imputation**. This technique implies the filling in a missing feature value with predictions of a machine learning regression algorithm.
- 4. **Mean/median imputation**. This method involves the filling of missing feature values with their mean or median value.
- 5. **Constant value imputation**. This technique entails the filling the missing values with clearly too high or too low values. The motivation is for the algorithm to discern the value as an outlier while considering other features. This method is not recommended as it can introduce bias.

It is often impossible to tell which data imputation method would work the best and therefore, it should be checked experimentally.

3.4 Model Training and Hyperparameter Tuning

It is a common practice to divide a dataset into three parts

- Training set (70% of the dataset)
- Validation set (15% of the dataset)
- Test set (15% of the dataset)

The training set, being the largest of them, is employed to train the machine learning model. Validation and test sets, which are of identical sizes and often called hold-out sets, are used in subsequent stages of model evaluation.

The rationale behind the use of separate training and validation sets is to prevent overfitting - a situation when the model performs well on the training data but poorly on the unseen data. Overfitting can occur if the model is tested and evaluated on the same dataset. As a result, the model may memorize the training examples and fail to make accurate predictions on the unseen data. To alleviate this, we use the validation set to fine-tune the model, and the test set to assess its performance before deploying it to production.

A typical workflow involves training the model on the training set, validation on the validation set using the selected metric, then adjusting the model's parameters to improve its performance. This process is repeated until no substantial improvement is observed. Finally, the model's performance is assessed on the test set. This iterative process is referred to as hyperparameter tuning.

An alternative to the three-set technique is *k-fold cross-validation*. This technique involves splitting the dataset into k subsets, or folds, of equal size. One fold is used as a validation set, while the other k-1 folds constitute a training set. The model is trained exactly k times, with each fold serving as a validation

set only once. The only drawback is that it is highly computationally demanding, particularly with a high k value and larger datasets, as the model will be trained k times.

A hyperparameter is a parameter specified before model training, in contrast to regular parameters that are calculated during training. Each model possesses a different set of hyperparameters and they profoundly influence the model's performance. The number of trees in Random Forest and the C hyperparameter in Support Vector Machines are examples of hyperparameters. The task of finding the optimal combination of hyperparameters is called hyperparameter tuning. One strategy might be to select hyperparameters manually and observe their impact on the performance. However, to utilize the grid search is a better way.

Grid search is a standard way of performing hyperparameter fine-tuning. It includes defining hyperparameters to experiment with, providing values for each hyperparameter to be tested, and training a model for each possible combination of hyperparameters. The performance of each individual model is assessed using k-fold cross-validation and the best combination of hyperparameters is selected. This approach is used in sci-kit-learn's implementation - GridSearchCV.

Grid search proves to be effective when dealing with relatively few hyperparameter combinations. However, with larger number of hyperparameter combinations, it is advisable to use RandomizedSearch (RandomizedSearchCV in sci-kit-learn). This method is very similar to grid search but instead of trying every possible combination of provided values, it tests only a specified number of randomly selected hyperparameter combinations. The primary advantage of this method over grid search lies in more control over computational power and the time dedicated to hyperparameter tuning.

3.5 Survival Analysis

Survival analysis, also known as time-to-event analysis, is a statistical method used to analyze the time until an event of interest occurs. Its name originates from clinical and biological research, where these methods are used to analyze survival time, hence the name. These methods, however, found their uses in areas far beyond clinical settings: in business to predict the time until the customer "churns" from a subscription, and in engineering, to estimate the product longevity or their parts. In social sciences, estimate the longevity of a marriage or a student dropout rate in an academic setting.

Censoring

The most distinct feature of these methods is the ability to handle censored data. Censoring refers to a circumstance when the information about survival time is only partially known. For example, in the dataset used in this work, there are 370 000 patients that were reportedly alive at the time of the last date of observation. And we do not know what happened to them after that date — they are censored.

Look at the figure 3.5. On the y-axis, we can see individual patients, while the x-axis corresponds to the study timeline (the right side is the end of the study). Cross (X) denotes an occurrence of the event, and circle (O) corresponds to the subject's exit from the study.

There are two types of censoring: left and right censoring. Right censoring, the most common one, occurs when we know that event did not happen up to a certain point — the patient didn't attend follow-ups after a certain point in time or the study ended when he was still alive. As shown in Figure 3.5, patients E and F are right-censored, as the event didn't happen during the study, and B is also right-censored, as the subject dropped out of the study. Left censoring, being much less common, describes the situation when the subject's survival time is unknown, but we know it is less than a specific time. For example, in epidemiologic studies, we may not know when the patient was infected, but we know when the first symptoms occurred.

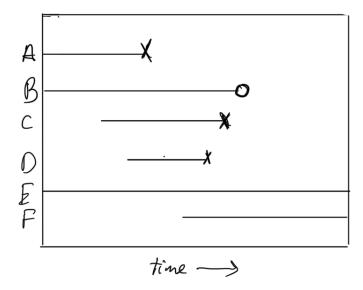


Figure 3.5: caption

Censoring Assumptions

There are three types of censoring assumptions: random, independent and non-informative. They are have similarities, but there are differences that we are going cover in the following paragraphs. We need censoring assumptions to be able to handle censored subjects.

In case of *random censoring*, subjects censored at time *t* are asumed to have the same failure rate as remaining subjects provided the same survival experience. Censoring is *independent* if it is random within any subgroup of interest. If we consider only one subgroup, then we wouldn't see the difference between random and independent. For example, let's take a three year disease occurrence study with 100 subjects at risk. Individuals are followed for three years and by the end of the study 20 of them contract the disease. We calculate the three-year disease risk as 20% and three-year survival as 80%. Now, suppose we want to continue the study for an additional two years on the remaining 80 individuals. However 40 of them refuse to continue in the study, and therefore are lost to follow-up (censored). Out of 40 remaining subjects, 5 contract the disease. How would we estimate the five-year survival?

And these assumptions come to a rescue. Under an assumption of random and independent censoring, we would assume that those who remained in the study are no different from those who left. Therefore we would estimate that out of 40 censored individuals, 5 contracted the disease - the same amount, as with those who remained. Consequently, we would calculate the five-year risk as 20 individuals in the first three-year period, plus 5 out of 40 observed, plus 5 estimated out 40 censored and we would get 30% five-year risk of disease contraction and 70% five-year survival under random and independent censoring assumptions. In this case random and idependent censoring are the same, as no predictor variables were considered.

Let's expand our example to illustrate the difference between random and independent censoring. Let's introduce another group to the study: group B (the first is group A) with 100 individuals. In the first three years, 40 contracted the disease and 10 left the study. So, the calculated three-year risk for group B is 40%. Out of 50 remaining, between 3 and 5 years 10 contracted the disease. The risk is 20% for years between 3 and 5. Under independent assuption, we estimate that out of 10 censored, 2

contracted the disease. Let's calculate the five-year risk for the group B: 40 got the disease in the first three years, plus 10 out of 50 observed in the 3-5 year period, plus 2 estimated out of 10 censored, and we would get 52% five-year risk and 48% five-year survival for group B under independent censoring assumptions.

As we can see, the five-year risk in two groups differs signifficantly (30% against 52%) and the censoring proportion is also very different (50% against 17%) hence, the overall *censoring is not random*. However, it is random within groups A and B, therefore, the *censoring is independent*. Because independent censoring is random censoring conditional on each level of covariates.

If instead, in group B 30 subjects out of 60 were censored at the three year mark, the censoring proportion would be the same in both groups and the overall censoring would be *random*, as those censored would be the representatives of those remained at risk.

The opposite of independent censoring is *non-independent* censoring. Let's illustrate it with an example. Consider a drug study, where some subjects are censored due to occurance of side-effects. Most likely those censored due to side effects are not representative of those who are still in the study. If they indeed are more vulnerable to a health outcome, we would likely overestimate their survival under an assumption of independent censoring, introducing bias. Henceforth the independent/non-independent censoring affects the accuracy the most. Many analytical techniques, such as Kaplan-Meier survival estimation, the log rank test, the cox model operate under an assumption of independent censoring in presence of right-censored data.

Non-informative censoring distribution of time-to-event T provides no information about the distribution of time-to-censorship C, otherwise the censoring is informative. To best illustrate what is non-informative censoring, let's illustrate what is an informative censoring. Let's take a group of subjects under random and independent censoring assumptions. Every time the subject A gets an event, subject B selected randomly leaves the study (e.g. B is A's relative). If the censored subjects are representative of subjects at risk it would be random and independet censoring. Here the censoring mechanism is directly related to event occurance, so the censoring is informative.

Survival function

Because we are predicting a numerical value(rather values), from the machine learning point of view, survival analysis might be considered a regression, but instead of one numerical value we predict a continuous value - survival function or hazard function, and, obviously, the dataset is censored. Survival function (also survivor function) S(t) shows us the probability of patient surviving (event doesn't happen) at a given time t and can be denoted as

$$S(t) = P(T > t). \tag{3.9}$$

Where t is any specific *time* of interest, T is random variable for subject's survival time. For instance, if we want to know if a patient is going to live for more than 5 years after kidney transplant, t is equal to 5 and we ask whether T is greater than t (probability question). The function is declining in the range from 0 to infinity. As it is a probability, the function value ranges only from 0 to 1. Theoretically, the graph of the survival function must be smooth, but in reality it is represented by a step function.

Hazard function

Hazard function h(t) tells us the probability of given event *happening* at a given point of time t, provided the event did not happen before time t, and is denoted as

$$h(t) = \lim_{\Delta t \to 0} \frac{P(t \le T < t + \Delta t | T \ge t)}{\Delta t}.$$
 (3.10)

Subject's survival time T lies between t and $t + \Delta t$ provided that survival time T is greater or equal than t. Sometimes, the hazard function is called a *conditional failure rate*. It is a rate because it is a conditional probability per unit of time Δt . As it is not a probability, but a rate, the scale for this ratio is from 0 to infinity — depends on the measure of time in days, weeks or years. When we consider the limit of the expression as the time interval approaches zero we basically get the instantenious potential of failing at time t per unit time, given survival up to time t.

Cumulative hazard function is basically area under the hazard function that allows to say which group has a greater risk.

The relationsip between the two

There is a clear relationship between the survival function S(t) and the hazard function h(t) – if we know one, we can determine the other. The relationships are the following:

$$S(t) = exp\left[-\int_{0}^{t} h(u)du\right]$$
(3.11)

Equation 3.11 tells that the survival function S(t) is equal to the exponential of the negative integral of the hazard function from zero to t.

$$h(t) = -\left[\frac{dS(t)/dt}{S(t)}\right]$$
(3.12)

Equation 3.12 tells us that the hazard function is equal to the negative derivative of the survival function S(t) with respect to t divided by S(t).

Considering the facts that survival function describes the probability of patient surviving to a given point of time *t* and hazard function shows us the probability of person dying at any given point of time *t*, we can say that they provide complementary information about survival and risk over time. Of the two discussed functions the survival function is used much more often as it is more appealing in the context of survival analysis and in the practical part of this paper(bachelor project/thesis) we are going to estimate exactly the survival function.

Take a look at figure 3.6. *a)* shows us a graph of the estimated survival function and b) shows us a graph of the estimated hazard function for a random patient from the dataset used. As we can see the survival function is declining over time, while the hazard function increases.

Kaplan-Meier Survival Curves

Kaplan-Meier is a non-parametric method of survival function creation. It is *non-parametric* because it does not take into account any covariates, or parameters, and requires only the survival time and the censoring indicator. It works under an independent censoring assumption.

The general Kaplan-Meier formula for plotting the survival function is the following:

$$\hat{S}(t_{(f)}) = \hat{S}(t_{(f-1)}) \times \hat{P}(T > t_{(f)}|T \ge f_{(f)}),$$

That can be read as the survival function \hat{S} of time $t_{(f)}$ is equal to the probability of surving past the previous time point $t_{(f-1)}$ times the conditional probability of surviving past the time $t_{(f)}$ [13].

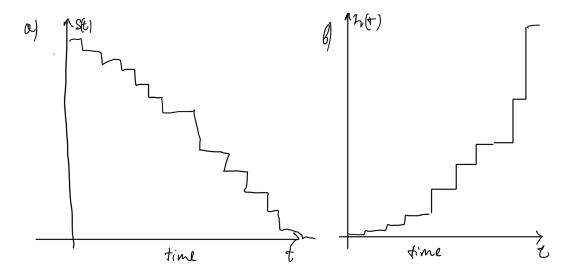


Figure 3.6: (Don't forget to provide example images both for the survival and hazard functions for some random patient from the dataset)

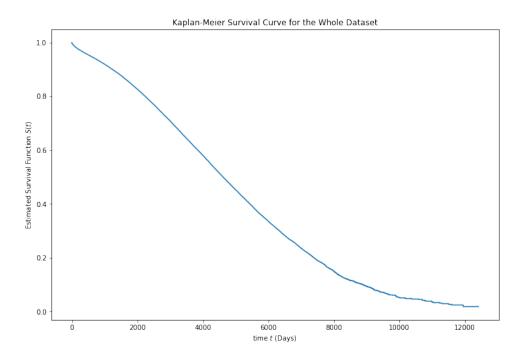


Figure 3.7: Kaplan-Meir survival curve for the whole dataset

These survival curves are often compared using the log-rank Test. *Log-rank test* is a way to compare two survival functions, that is often used in studies, where there is a target group and a placebo (control) group to assess the efficacy of the studied thing by comparing the survival curves of the two groups [13].

3.5.1 Performance Metrics

Survival prediction models play an important role in healthcare. They are often used to estimate the risk of developing a particular decease and have an important role in guiding the clinical management of patients. It is, therefore, crucial to assess their performance properly. Similar to machine learning, this process of model evaluation is referred to as *model validation*. There are three aspects we can asses our model on:

- 1. Overall performance, which is the distance between the predicted and observed survival time
- 2. **Discrimination**, or the model's ability to distinguish between high- and low-risk patients.
- 3. Calibration, which is the agreement between the observed and predicted survival times [21].

The absence of bias in a situation when the validation set contains censored instances is a sign of good performance measure. Otherwise, in the presence of the high levels of censoring, the evaluation would be unreasonably optimistic[21].

In this section we are going to cover three measures of discrimination and one measure that assesses both the discrimination and calibration (overall performance), that were used in this work.

3.5.1.1 Harrel's and Uno's Concordance Indices

One way to measure discimination is *concordance*. *Concordance measures* quantify the rank correlation between the predicted risk and the observed survival times. Their values usually range between 0.5 and 1, where 0.5 means that the there is no discrimination whatsoever, and 1 corresponds to the ideal discimination.[21]

Concordance probability is a probability that from the two randomly selected patients (i, j), one with shorter survival time has the higher predicted risk. Mathematically:

$$C = P(\eta_i > \eta_i | T_i < T_i),$$

where η_i and η_j are the predicted risk scores, and T_i and T_j are the survival times[21].

Harrel's concordance index C_H takes into account all suitable subjects where the shorter survival time corresponds to an event. C_H is estimated as the proportion of these pairs, where the patient with the shorter survival time has the higher predicted risk. Exists a modified version of this estimator $C_H(\tau)$, that only considers patients with $T_i < \tau$ and may provide more stable estimates[21].

According to the Scikit-survival's documentation and Rahman et.al., Harrel's concorance index is biased in the presence of censoring, and the higher the censoring is, the more biased it is going to get. Uno et. al. proposed a modified concordance index $C_U(\tau)$ that uses weights based on the probability of being censored. They found that their estimator is robust to the choice of τ , but made a remark that the error of the estimate might be quite large if there's too little instances beyond this time-point [21, 22].

3.5.1.2 Time-dependent Area under the ROC

The area under the receiver operating characteristics curve (ROC AUC) is a popular performance measure for binary classification tasks. In survival analysis, it is used to determine how well estimated risk scores can distinguish diseased patients from healthy ones[22].

$$ext{BS}^c(t) = rac{1}{n} \sum_{i=1}^n I(y_i \leq t \wedge \delta_i = 1) rac{(0 - \hat{\pi}(t|\mathbf{x}_i))^2}{\hat{G}(y_i)} + I(y_i > t) rac{(1 - \hat{\pi}(t|\mathbf{x}_i))^2}{\hat{G}(t)},$$

Figure 3.8: change to a real equation!

In binary classification, the receiver operating characteristic (ROC) is a curve that plots the true positive rate (TPR or sensitivity) against false positive rate (FPR). The latter is the ratio of negative instances that are falsely classified as positive. It is equal to 1– the true negative rate (the ratio of negative instances that are correctly classified, often referred to as specifity). The former is the ration of positive instances classified as positive [9].

In survival analysis, we extend the ROC to continuous outcomes, where a patient is alive at the start of the observation, but might experience an event at some point later. Specificity and sensitivity, therefore, become time-dependent measures. Here we consider *cumulative cases* and *dynamic controls* at any given point of time t. Cumulative cases are all subjects who experienced an event prior to or at time t. While *dynamic controls* are those who are yet to experience the event after time t. By calculating the ROC AUC for any given time point t, we can tell how well the model can distinguish patients who fail by a given time $t_i < t$ from subjects who fail after this time $t_i > t$. It is useful only if we want to predict an event happening in a period up to time t, rather than at a specific time-point t [22].

3.5.1.3 Time-dependent Brier Score

Time-dependent ROC AUC and concordance index are great to assess the overall discrimination among all time points (mean AUC and c-index) or the discrimination at any individual time point (the ROC graph), but they tells us nothing about the accuracy of individual predictions [22]. We would want something similar to regression performance measures used in machine learning. Fortunately, such metric exists. Time-dependent Brier score is a modification of mean squared error (MSE) that handles right censored data.

While concordance index and time-dependent ROC AUC measure only discrimination, the time-dependent Brier score measures both discrimination and calibration, making it a metric of "overall performance". It is defined by the following equation:

where $\pi(\hat{t}|\mathbf{x})$ is a model's predicted probability of remaining event-free up to time point for feature vector \mathbf{x} , and $\frac{1}{\hat{G}(t)}$ is the inverse probability of censoring weight [22].

3.5.2 Cox Proportional Hazards Method

Intro:

The Cox proportional hazards model is defined in terms of a hazard at time t for a subject with given vector of explanatory variables \mathbf{X} :

$$h(t, \mathbf{X}) = h_0(t) \times exp\left[\sum_{i=1}^{p} \beta_i X_i\right]$$
(3.13)

where $h_o(t)$ is a baseline hazard function. Due to the fact that the exponential function has no t, \mathbf{X} is called *time-independent*. The exponential function ensures that the function is non-negative, satisfying the definition of the hazard function.

Despite the fact that the equation 3.13 contains the baseline hazard function, it is not specified. Fortunately, we can calculate the hazard ratio, a measure of effect, without having to estimate it. Similarly, the hazard function $h(t, \mathbf{X})$ and the survival function $S(t, \mathbf{X})$ can be estimated without the estimation of the baseline function. So with minimal assumptions we can estimate everything we need (h,S and HR).

Why cox ph is popular: Being a semi-parametric, this model is a safe choice due to the fact that it consistently delivers a sufficiently reliable result. The risk of choosing the wrong model, as it often happens with parametric models, is practically non-existent. However, if one is sure that a parametric suits the problem, they should use the parametric model.

Optimisation: Similar to logistic regression, the CoxPH uses the *maximum likelihood* function 3.5 to calculate its parameters $(\hat{\beta}_i)$. However, due to the fact that the maximal likelihood considers only a part of patients, namely those who experienced an event, the formula is called *partial likelihood*.

Hazard ratio: Hazard ratio is a measure of influence of an intervention on the outcome. A hazard ratio is defined as the hazard for one individual divided by the hazard for the other, and is calculated with the following formula:

$$\hat{HR} = \frac{h(t, X^*)}{h(t, X)} = exp\left[\sum_{i=1}^{p} \hat{\beta}_i(X^* - X)\right]$$
(3.14)

PH assumption: As can be seen, the equation does not contain t and the basic hazard function, making it a *proportional hazard assumption*.

3.5.3 Penalized Cox Models

Ridge m

Lasso m

Elastic Net m

3.5.4 Random Survival Forests

useless with large amounts of data

3.6 Overview of Machine Learning Libraries and Tools

for the survival analysis the python library scikit-survival was used

3.6.1 Comparison

3.7 Conclusion

Chapter 4

Data Preparation and Analysis

In this chapter we age going to look into the UNOS dataset. Make sense of the dataset. Explore important features and their relationship with each other. Look into survival time for

The dataset provided by the IKEM (Institute of Clinical and Experimental Medicine in Prague) that I had from the beginning was not suitable for any meaningful analysis. That is why it was decided to look for the dataset elsewhere.

The data reported here have been supplied by the United Network for Organ Sharing as the contractor for the Organ Procurement and Transplantation Network. The interpretation and reporting of these data are the responsibility of the author(s) and in no way should be seen as an official policy of or interpretation by the OPTN or the U.S. Government.

The dataset is not for the open use. If you are interested in testing the results achieved in this paper, you need to acquire the data first. The requirements for the data acquirement are written here

The dataset consists of 993 806 of records for both transplanted patients and ones from the waiting list, and 450 features comprised of waiting list data and already transplanted patients for kidney and pancreas transplant from October 1, 1987 to the present. Kidney transplants have 490 172 records.

The following features will be considered.

4.1 Data Loading

The data were provided in a form of a MongoDB database dump. We cannot work with the database dump. So it was necessary to run the database first and import the database dump there. I set up the

Feature description	Type	Abbreviation
Donor Age		
Recipient Age		
Donor Type		
Donor gender		
Recipient gender		
Donor blood group		
Recipient blood group		
Recipient on dyalisis		
Recipient creatinine at the time of tx		

Table 4.1: Features

database in a Docker container (Docker is container management system + **explain what is container**) locally on my PC, as the university cluster unfortunatelly does not have Docker. The data from the database and table kidpan were then exported to CSV, compressed into zip and uploaded to the cluster.

The pandas DataFrame method read_csv() loaded data for too long to work comfortably (5 minutes), as the CSV file had the size of 80GB, so it was decided to use parquet file instead. It was done by dumping the pandas DataFrame into Parquet database file using DataFrame.to_parquet() method. Parquet is used for efficient cloud computing. Parquet provides more efficient way of loading data, as it works on the principles of databases, so the loading time of the whole dataset was decreased to 38 seconds. Additionally, it allows for specifying what columns to load, reducing the data loading time to 21 seconds. Thus using this technology has signifficantly improved the workflow.

4.2 Data preprocessing pipeline

In this section I will describe the data pipeline that I use to create the dataset out of the raw data. The pipeline can be found in github repository of this paper: survival_pipeline.py.

The work with the pipeline is pretty straightforward: we initialize the class and call the *load()* method. As is shown in the following block of python code:

Two main constants of the class are *categorical_values* and *numerical_values*. Categorical and numerical features must be specified there. It is important for following preprocessing steps.

The main method of the class *ScikitSurvivalDataLoader* is *load()*. This method loads the data into the pandas DataFrame, applies exclusion criteria (more on that later), handles NaN values and returns X and y, X being numerical (categorical values were handled with OneHot encoding and numerical values were scaled) and y having format of (PSTATUS, PTIME), first one is the boolean censoring indicator (True event happened, False - otherwise), PTIME is the number of days survived. This format is required by the Scikit-survival Library to build survival estimators.

The first step is to load the data into pandas DataFrame from the parquet file. Fortunately, pandas has support for this kind of files. It is performed by the Pandas method *read_parquet(path, engine, columns)*. In *path* we need to specify the path to the parquet file, *engine* specifies what parquet library should be used, I use 'auto', it tries *pyarrow* if it doesn't work it uses *fastparquet*. In *columns* we need to specify the columns we want to load. (explain more what is pyarrow and parquet)

Description of feature engineering step:

The next step is to divide the dataset into training, validation and test sets, the reasons behind that, were explained in the datapreprocessing section of the previous chapter. These sets are then assigned as class variables to the class and are sent to preprocessing method _handle_nan(), where the NaN values are filled with median, specific value, or examples with such values are deleted with the pandas DataFrame method drop_na(), depending on the fill_na_with_median boolean parameter.

After the NaN handling step, the training set is send to the method _get_X_y() where the numerical values are standartized and categorical are encoded with the OneHot encoding with the Scikit-Survival methods standardize() and encode_categorical(). Numerical and categorical values then comprise the X set, directly used in the training. The target value set is constructed with Surv.from_arrays() utility that

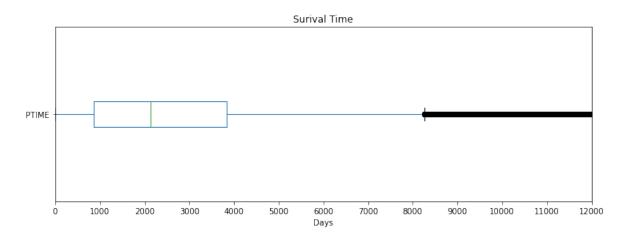


Figure 4.1: Box plot for the survival time

accepts event and survival time and builds the y value acceptable to scikit-survival algorithms. The class variable df is then set to None with the goal of memory optimisation. X and y are then returned.

When we need the validation and the test sets, we just call methods $get_validate_X_y()$ and $get_test_X_y()$ which will provide us with the sets for the hyperparameter tuning step with the validation set and the final evaluation step with the test set.

4.3 Exploratory Data Analysis

In this section we are going to cover all the most signifficant features. The data were not adjusted to limit the influence of other factors, so the correlations I am trying to make here might not be fully correct, however some are confirmed in literature.

4.3.1 Survival Data

In this subsection we are going to explore the *y* axis that is going to be used for the training of the survival estimators. The *y* value consists of censoring status, which is a boolean value, and time to event, which is a numerical value representing the survival time or the time at which it was censored. The *y* value is called the *survival data*. The column PSTATUS is a censoring status, while the PTIME column represents the time-to-event variable.

To best visualize the time-to-event variable we are going to use a box plot. A box plot is a simple, yet powerful statistical graph based on quartiles, that allows to quickly make sense of the data distribution. (odkaz na statistics for data scientists) It is based on three quartiles: the first(Q_1), the second(Q_2) and the third (Q_3). First quartile corresponds to 25 percentile and means that 25% of the datapoints are below it. The second quartile corresponds to 50% percentile, or median, and it means that below and above that point lies an equal amount of data points. The third quartile corresponds to 75 percentile and it means that below it lies 75% of data points. The quartiles form the box: the first quartile forms the left edge(or bottom edge, in case of horizontal box plot), the third quartile forms the right edge (or top edge) of the box and the median is drawed inside of the box. The box itself represents interquartile range (IQR), that is calculated as $IQR = Q_3 - Q_1$. The lines that lie beyond the box are called whiskers and indicate a range for "a bulk of the data". The whiskers extend to the furthest points outside of the box, except they cannot be longer than 1,5 times the IQR. The values lying outside of the whiskers are considered outliers.

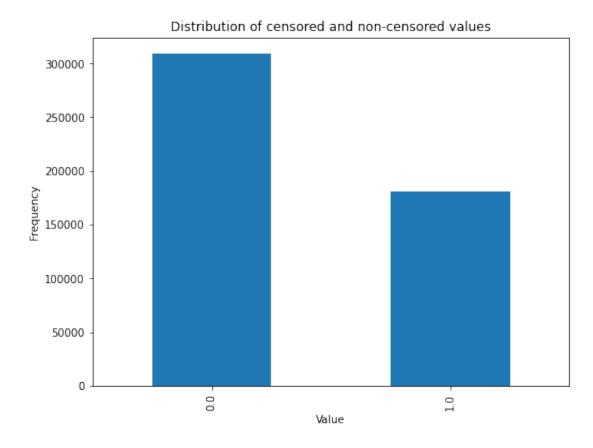


Figure 4.2: Bar chart of PSTATUS colum that provides information on censoring. 0 corresponds to censored instance, 1 corresponds to event happening. There are 308823 censored instances, and 181349 times event happened.

Look at the Figure 4.1, there you can see the box plot of patient survival time (PTIME column). The Q_1 is equal to 867 days (2.4 years), the median is 2136 days (5.85 years) and the third quantile Q_3 is equal to 3828 days (10.5 years). The interquartile range (IQR) is equal to 2961 days. This makes up the box. The left whisker extends from 0 day up to the first quartile of 867 days. The right whisker is much longer, and extends from the third quartile up to the $Q_3 + 1.5 * IQR$, which in our case is equal to 8269,5 days. The values above 8269,5 can be considered outliers. As can be seen from the figure (almost straight black line, consisting of individual dots) there are a lot of them. There are less than 10 000 (get the actual value) outliers, which is not that much compared to the 490 000 of total kidney transplantations. It is not the wisest choice to simply remove the outliers, as they still might have useful information to the model. However it has to be estimated experimentally. The handling of outliers will be described in the section dedicated to the dataset building.

Look at the figure 4.2, where there is plotted the distribution of the PSTATUS column values. 0 corresponds to censored instances, 1 corresponds to event happening. As can be seen, the distribution is quite uneven, and there is much larger amount of censored instances than ones with the event happening, as there are 308 823 censored instances and only 181 349 non-censored ones. The percentage of censoring is 63%, which is quite high.

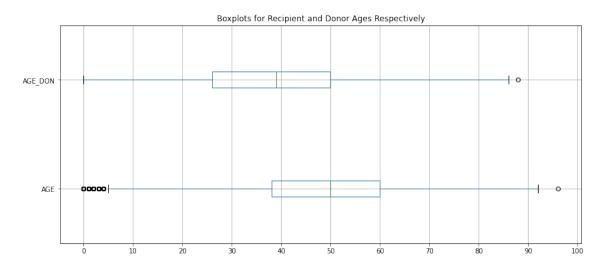


Figure 4.3: Box plot for the recipient age versus donor age.

4.3.2 Age

In this subsection we are going to explore both the donor and recipint ages in the dataset, and will see what the literature tells about their importance to long-term survival.

As can be seen on the box plot in the Figure 4.3, in this dataset donors are usually younger than the recipients by median 11 years. Median recipient age in the dataset is 50, median donor age is 39. The vast majority, 75% of the recipients aged lie between 38 and 60 years old, making IQR of 22, while 75% of the donor age lie between 26 and 50 years old, making IQR of 24. Interestingly, there are not many outliers, as the whiskers cover ages from about 5 to 93, covering the most of human life range.

In the Figure 4.4 you can see the hexagonal binning for the recipient age vs. the survival time. The hexagonal binning is a substitute for a scatter plot for large datasets, as scatter plots do not handle large data sets very well. The hexagonal binning plot consists of colored hexagons, and the darker the color is, the more instances lie in it.

The figure 4.4 more or less corresponds to box plots 4.1 and 4.3, as the majority of colored hexons lie in box ranges for age and the survival time.

4.3.3 Donor Type

In this subsection we are going to explore the influence of donor type (living or diseased) on the survival. Recipients with kidneys from living donors live longer, this is a well established fact[16] [17].

Let's look at the Figure 4.5, where are plotted two Kaplan-Meier survival curves for all patients from the dataset, without taking into account other covatiates. On the graph we can see that the survival probability of living donor transplant is indeed significantly higher than the survival probability of deceased.

This is the case because often there is no time to make full HLA screening, that may allow for HLA mismatches. Additionally, deceased transplants may suffer from mild kidney damage due to the delay in transplantation. While living donor transplants are often performed between siblings that have similar HLA, that creates better compativility.

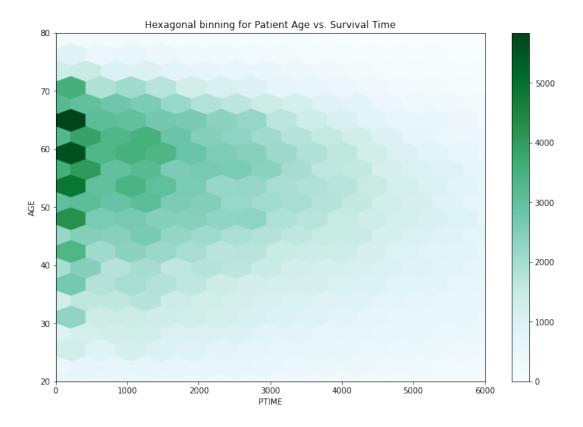


Figure 4.4: Hexagonal binning for the recipient age versus survival time. The brighter the color of the hexagon is, the more instances lie in it.

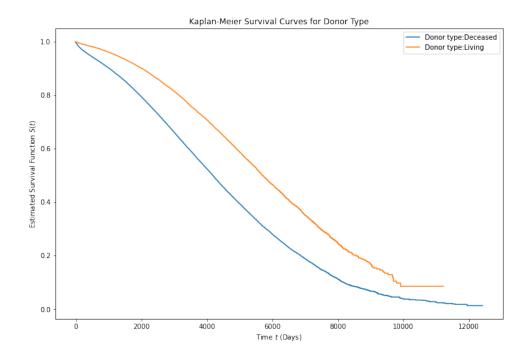


Figure 4.5: Kaplan-Meir survival curve donor types

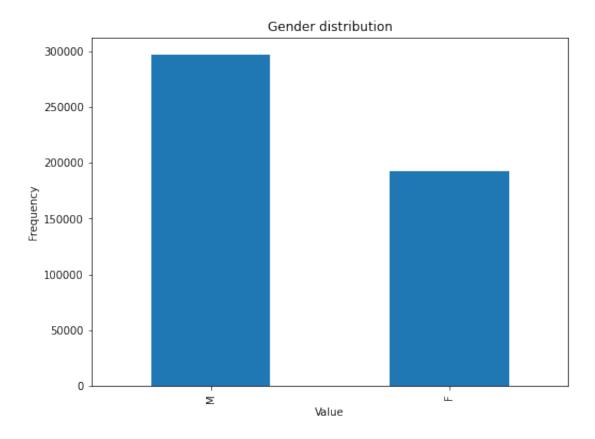


Figure 4.6: Bar chart for gender

4.3.4 Gender

In this subsection we are going to explore the gender distribution in our dataset, and its influence on survival.

Let's explore the distribution of gender in our dataset. Take a look at the figure 4.6. There are 297279 men and 192882 women - the 35% difference. Despite the fact that the chronic kidney disease is more common in women, the end stage kidney failure and therefore kidney transplantation is more common in men[19].

Let's take a look at gender's influence on survival. In the Figure 4.7 we can see the Kaplan-Meier survival curves for men and women on the whole dataset. As can be seen from the graph, females generally have less risk than their male counterparts. Women usually live longer [14]. Quite signifficant factor is the difference between male and female immune responses - males usually have greater risk to get an infection, than females, and the intesity of the infection is higher[15]. Furthermore, the influence of immunosuppresants make the problem of infection even worse.

4.3.5 The Use of Dialysis

In this subsection we are going to explore the influence of dialysis on survival. In the figure 4.8 we can see two survival curves for the patients who were on dialysis, and for those who were not. The whole dataset was used. As can be seen in the Figure, the patients who were on dialysis before the transplantation have a greater risk, while those who were not. This agrees with [20].

Why exactly this is the case, unfortunately, I was not able to find. The literatures just states it as fact.

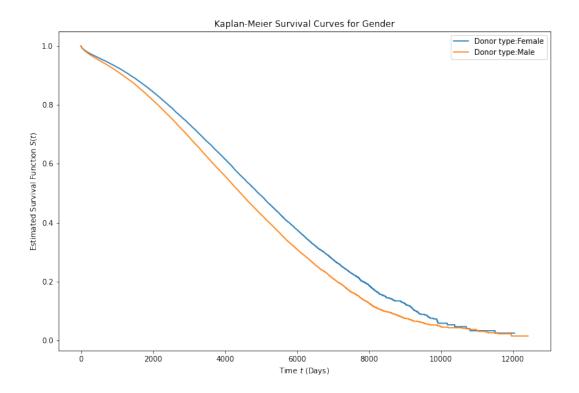


Figure 4.7: Kaplan-Meir survival curve for genders

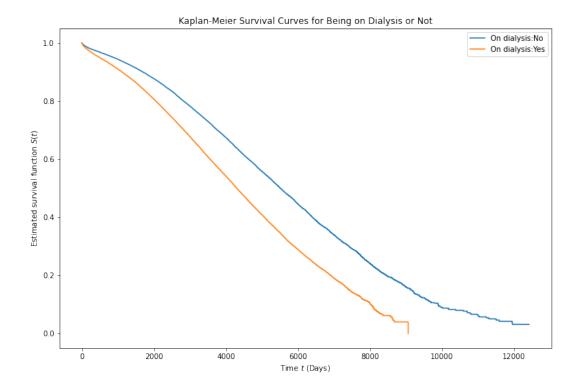


Figure 4.8: Kaplan-Meir survival curve for using dialysis or not

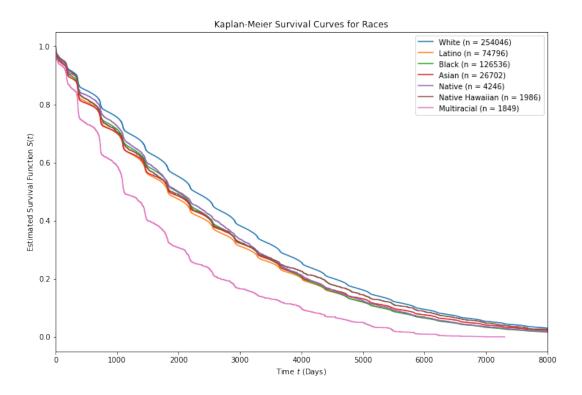


Figure 4.9: Kaplan-Meir survival curve for ethnicities

4.3.6 Race

In this subsection we are going to explore the survival curves for different ethnic groups. In the Figure [18] are plotted 7 survival curves for different ethnicities. As can be seen on the image, 5 ethnicities share about the same survival probability during the cource of 8000 days, while two of them differ substantially from the other. White americans had higher survival probability than other ethnical groups, later converging to the others. This corresponds to ([18]) examining graft survival.

The least survival probability had multiracial group. Their number, however, was the lowest - only 1849 instances, so it is not enough to make any conclusions. They might be later removed, as the definition is too vague and there are not many instances.

4.4 Dataset building, Exclusion criteria and noice reduction

Chapter 5

Machine Learning Model

5.1 Problem Formulation

Predicting the survival time after a successful kidney transplant can be approached in three ways: as a regression problem, classification problem, or through survival analysis.

A *regression* model may seem an intuitive choice, as we want to predict a numerical value – the survival time. But it is not the best option for the following reasons:

- 1. **The censored dataset**. The dataset has a high level of censoring 76%. The dataset contains the number of days survived, along with the survival status. Including both living and deceased patients would introduce too much noise to the model, making it highly inaccurate. It is impossible to predict the number of days survived with regression methods based on a dataset comprised of both living and deceased patients.
- 2. Censoring removal would produce bias and signifficantly reduce the dataset. We could remove all censored instances, but that would reduce the dataset from 500 000 to roughly 120 000 examples. It would also introduce significant bias, as the dataset would contain only deceased patients, and most of them passed away before the introduction of modern techniques for treating the rejection. As a result, the model created from such a dataset would be highly inaccurate.
- 3. **Regression predicts only one single number.** It poses a problem, especially over extended time frames, as there are too many factors that we can't account for, leading to incorrect predictions.

Another way of formulating the problem is *classification*. We can theoretically divide the dataset into groups: "less than one year", "one to five years", "five and more", or even more groups and train a classifier based on them, as it was done by et al.. And again, we would face problems of censoring and bias mentioned above. So the classification is also not the best option.

A more appropriate way of problem formulation is in terms of *survival analysis*. Survival analysis methods handle censoring and provide a better form of prediction: survival function or hazard function, which represents survival probability or the failure rate at each moment in time, respectively.

5.2 Model selection

The algorithms provided by the scikit-survival do not handle large datasets very well (never ending training process and worse results probably due to the noise) that is why I chose to train different models for different demographics, as one specific model for one specific demographic will perform better that one model trained for all demographics. In addition, the living donor transplantation differs a bit from the diseased transplantation, that might introduce some noice into the model.

Dataset	Uno c-index	IBS	Mean AUC
Living	0.705	0.135	0.704
Deceased	0.681	0.165	0.714
Kaplan-Meier (for IBS reference)	-	0.247	-

Table 5.1: Coxnet performance on the test set

Model	Uno c-index	IBS	Mean AUC
Coxnet (standard hyperparams, living)	0.668		
Coxnet (the best hyperparams, living)	0.705	0.135	0.704
Kaplan-Meier (for IBS reference)	-	0.247	-

Table 5.2: Cox before and after fine tuning.

The way I approach the model selection model automation with the class SurvivalEstimators defined in estimator_automation.py.

Run the following class and short list the most promising models. In this case it is survival gradient boosting and random survival forests.

5.3 Results

In this section we are going to discuss two models, Coxnet and Random Survival Forest. These models were chosen because they both are able to generate survival functions in the scikit-survival library, unlike others that only estimate the risk score. Unfortunately, the older version of scikit-survival was used - 0.14.0, due to the limitation in python version on cluster where these models were trained. The survival function will later be used in the application KidneyLife to visually illustrate the probability of survival in each moment in time.

5.3.1 Coxnet

Coxnet, or an elastic net, is a linear model, so it is fast even with large datasets, a bit worse results, compared to the Random survival forest. It makes prediction both in a form of the risk score or a survival function.

As can be seen in the table 5.1, the Coxnet performed the best for the living group, the worst for the deceased, and somewhere in between for the both living and deceased.

Discussion of the AUC figure:

As was covered in 3.4, the hyperparameter tuning is usually performed with either GridSearch or RandomizedSearch. Unfortunately, the older Python version on the cluster did not allow to install the newest version of scikit-survival, where the GridSearch was implemented. So, the hyperparamer tuning was performed with a custom script that is designed to imitate the GridSearchCV but without the k-fold cross-validation. The script optimizes for the Integrated Brier Score (IBS) that directly tells the accuracy of predictions. The script can be found: here (add link).

Discussion of the Brier Figure:

The coxnet has only two hyperparameters: L1 ratio and alpha, the latter is calculated by fitting the model and we then need to choose the best of them. L1 ratio defines the relative weight of the l_1 and l_2 penalty.

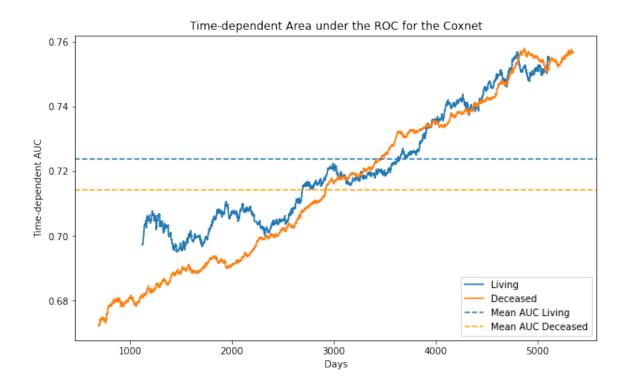


Figure 5.1: AUC curve for coxnet

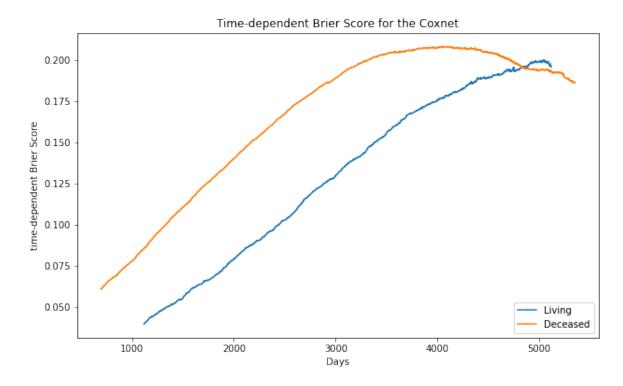


Figure 5.2: Time-dependent Brier score for the Coxnet

Feature description	Importance	Abbreviation
Donor Age		
Recipient Age		
Donor Type		
Donor gender		
Recipient gender		
Donor blood group		
Recipient blood group		
Recipient on dyalisis		
Recipient creatinine at the time of tx		

Table 5.3: Coxnet Feature Importance

Dataset	Uno c-index	IBS	Mean AUC
Living	0.727	0.129	0.743
Deceased	0.678		
Kaplan-Meier (for IBS reference)	-	0.247	-

Table 5.4: Random Survival Forest performance on the test set

On the table 5.3 we can see the importances of features for the prediction. Features with zero influence were omitted.

5.3.2 Random Survival Forest

Random survival forest is a powerful ensemble machine learning algorithm, that is comprised of multiple submodels, and therefore it takes a lot of time to train, a lot of time to make a prediction, depending on the selection of hyperparameters, making it a bit difficult to work with, especially during the hyperparameter tuning. Extremely memory hungry. The prediction is a survival function or a hazard score. It was covered in detail in (**RSF subsection**). The living and deceased subsets had 34951 and **70 000** instances respectively.

As can be seen from the table 5.4, the survival forest performed the best for the living group, the worst for the deceased, and somewhere in between for the both living and deceased, just as it was in case with the Coxnet.

discussion of AUC image:

discussion of the Brier image: On the Figure 5.4 is plotted the time dependent Brier score for the Random Survival Forest. As we can see, it increases over time, meaning that predictions get worse over time. This totally makes sense, as the more time passes after the transplant, the less pretransplant information, that was used for training, has influence on the survival.

The fine-tuning was performed with a custom script, described in the previous subsection dedicated to the coxnet. The hyperparameters that were fine-tuned are the following: n_estimators, max_depth, min_sample_split and max_features.

On the table 5.5 we can see the performance of regular RSF with the standard hyperparameters compared to the performance of RSF with fine-tuned hyperparams.

Feature importance can be seen on the table 5.6.

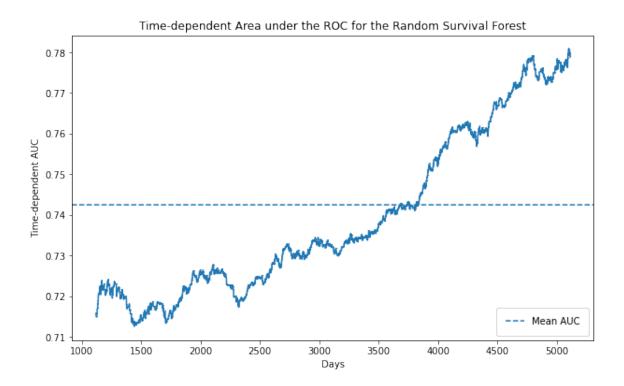


Figure 5.3: AUC for RSF

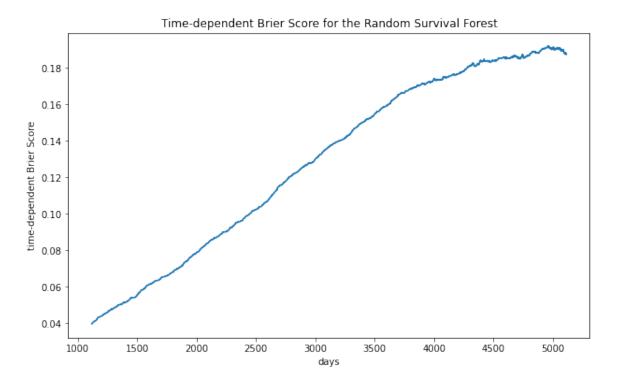


Figure 5.4: Time-dependent Brier score for the Random Survival Forest

Model	Uno c-index	IBS	Mean AUC
Random Survival Forest (standard hyperparams, living)	0.708	0.160	
Random Survival Forest (the best hyperparams, living)	0.723	0.129	0.743
Kaplan-Meier (for IBS reference)	-	0.247	-

Table 5.5: RSF before and after fine tuning.

Feature description	Importance	Abbreviation
Donor Age		
Recipient Age		
Donor Type		
Donor gender		
Recipient gender		
Donor blood group		
Recipient blood group		
Recipient on dyalisis		
Recipient creatinine at the time of tx		

Table 5.6: Feature Importance for RSF

5.3.3 Comparison

image with the two BS and AUC graphs for each model

As can be seen on the table 5.7 and on the image ..., the Random survival forest performed better than the elastic net. However, there are some differences in timeframes, where at certain points in time the coxnet performed better than the RSF.

AUC RSF vs. Coxnet:

Brier Coxnet vs. RSF:

5.4 Scoring algorithm

the cumulative hazard suits the place of transplantation score very well

5.5 Limitations

these models probably aren't suitable for KEP (check results), be they're slow, but are good for prediction estimated survival and when it is best to intervene.

the RSF deceased could be better fine-tuned

Model	Uno c-index	IBS	Mean AUC
Random Survival Forest (living)	0.723	0.129	0.742
Coxnet (living)	0.705	0.135	0.704
Random Survival Forest (deceased)			
Coxnet (deceased)	0.681	0.165	0.714

Table 5.7: Model comparison on the test set

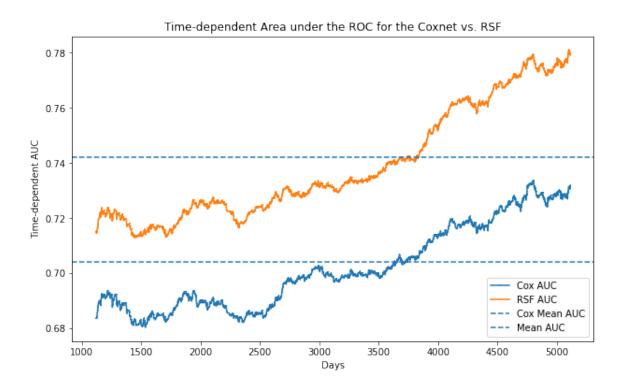


Figure 5.5: AUC: Coxnet vs. RSF, Living

5.6 Further work

more thorough hyperparameter tuning, especially with the RSF on the deceased dataset. another model for follow up data deep survival neural network

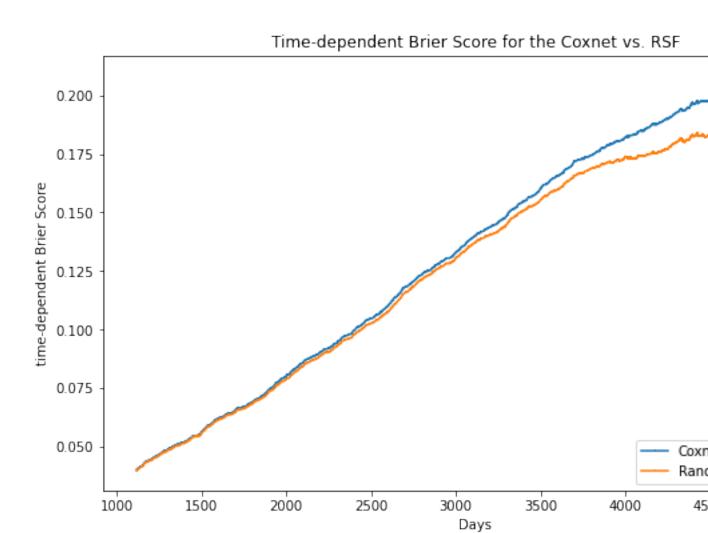


Figure 5.6: Brier: Coxnet vs. RSF, Living

Chapter 6

Applications

txmatching is something totally different, so it was decided to create separate application for accessing the model.

6.1 Existing Solutions

6.1.1 Txmatching

Txmatching is

6.2 KidneyLife

6.2.1 Frontend

6.2.2 Backend

For backend was used flask. Flask is popular python web framework. The model was saved as a binary in pickle format and the back end provides an api to access the model from the web page.

Conclusion

Text of the conclusion...

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[23]