# **Overlap Consensus Assembly Project**

Ken Reese, Kevin Boehme, Shaun Miller

# **Methods**

# **Error Handling**

The overlap consensus approach to contig assembly has an innate and flexible method for overcoming sequencing error. Our overlap consensus assembler performs a pairwise comparison between all reads, calculating a maximum overlap score between the two and generates a NxN matrix where N is the number of reads to assemble. This overlap scoring approach inherently accounts for errors in the reads by allowing for high overlap scores even in the presence of a few rare errors.

#### **Branching Nodes**

Our algorithm was an implementation of a greedy travelling salesman algorithm. We arbitrarily chose the first contig in the matrix and chose its highest scoring overlap (that wasn't itself). After choosing a best scoring overlapping contig we remove all other branches from the original contig. Then, we select the newly chosen contig and perform the same operation on it, namely choosing the highest scoring overlapping contig (that is still available) and then removing all other branches. We broke contigs when the overlap score was sufficiently low.

This approach maximizes each consecutive overlap score starting at any arbitrary contig until reaching each contig. It does not, however, guarantee that the absolute maximum path will be found.

# **Assembly Quality**

In order to assess the quality of our assembly we focused on the following informative metrics: number of contigs, largest contig size, N50, and mean contig size. The results for the following datasets are found below, with a discussion on the quality in the next section.

#### small.easy.fasta

Number of contigs: 1

N50: 903

Mean contig length: 903.00 Minimum contig length: 903 Maximum contig length: 903

# synthetic.small.noerror.fasta

Number of contigs: 2

N50: 557

Mean contig length: 401.00 Minimum contig length: 245

Maximum contig length: 557

# synthetic.large.noerrors.fasta

Number of contigs: 22

N50: 1125

Mean contig length: 212.86

Minimum contig length: 50

Maximum contig length: 1560

# real.small.error.fasta

Number of contigs: 1

N50: 1238

Mean contig length: 1238.00
Minimum contig length: 1238
Maximum contig length: 1238

# real.large.error.fasta

Number of contigs: 268

N50: 279

Mean contig length: 212.03

Minimum contig length: 100

Maximum contig length: 6700

# **Assembly Comparison**

Metrics for the comparative analysis were produced by running each assembler on dataset 6 (a large, but not crazy large dataset - 19.fastq).

#### Our OLC assembler

Number of contigs: 3

N50: 3222

Mean contig length: 2876.33 Maximum contig length: 3659

# Cap3

Number of contigs: 6836

N50: 526

Mean contig length: 369.82 Maximum contig length: 5228

# Our de Bruijn

Average (mean) contig size: 79.42

N-50:100

Total contigs generated: 40,286

Largest contig size: 1,974

#### Discussion

We compared our results with that of CAP3. We used the default settings and found CAP3 ran very quickly (12 minutes). We also thought it would be enlightening to compare our OLC results and those of Cap3 with our previously implemented de Bruijn Graph assembler.

Comparing our assembler to Cap3 we see that ours wasn't able to produce as quality of an assembly, although our N-50 and mean contig sizes were larger than Cap3 was able to produce. A major consideration is the time and memory efficiency of the assembler and in both respects Cap3 is superior to our assembler.

Comparing just our overlap graph with our de Bruijn graph shows that the overlap approach produces much fewer, and larger contigs. We believe this is because of how dynamic the overlap scoring algorithm is and how well it handles sequencing errors. It is worth mentioning that while our de Bruijn graph produced far worse assemblies it ran much quicker (20 seconds). The de Bruijn graph approach to genome assembly has many limitations especially when it comes to long stretches of homopolymeric regions. However, we believe that running a preliminary assembly using a de Bruijn graph coupled with a more sophisticated and computationally expensive overlap consensus could be a potentially powerful approach to read assembly.

#### **BLAST Results**

#### **CAP3 Results**

The longest contig of 1 analysis, 5315 base pairs long, mapped extremely well to Chromosome 19, Chromosome 15, and Chromosome 1. In Chr19, we see if mapping to the OR4G3P gene, an olfactory receptor gene, part of the largest gene family in the human genome. The match in Chr15 maps to OR4G6P, a very similar olfactory receptor gene. Based on the level of similarity (both have over 99% identity (2 mismatches for Chr19, 23 for Chr15, and that out of 5228 potential matches), and 100% Query coverage), we may assume that these olfactory receptor proteins detect very similar substances (then again, small differences in olfactory sensors do tend to produce large differences in sensation).

For entertainment, we may perhaps unjustifiably assert that these could be receptors helping humans tell the difference between their own flatulence and the flatulence of others-similar, yet profoundly different propositions! The Chr1 position is similar, but with a lower quality (significantly lower Max Score vs. the other matches)- it aligns to gene OR4G4P- another olfactory receptor family gene.

In other words, CAP3's largest contig aligned excellently to a well-preserved sequence corresponding to a olfactory reception genes.

#### **Improvements**

#### **Assembly**

Our assembler could benefit from a number of improvements. First, the speed and efficiency of our code and algorithm have a lot of room for improvement. Attempting our algorithm on a full human chromosome proved to be an impossible task. For this assembler to be at all viable we would need to rework many slower portions of the code as well as bring fundamental improvements to the overlap consensus algorithm mainly in the form of avoiding costly pairwise comparisons between irrelevent reads.

Another major improvement would be to implement a trimming algorithm before performing the alignment. For example, the Fastq format provides important quality metrics that should be taken into account by any serious assembler, however ours does not make use of this data. A quick preprocessing of reads based on their quality metrics would allow our assembler to prune/trim low quality reads and thus improve our downstream assembly.

Finally, our assembler could benefit from improvements in usability. We have many chunks of the assembly broken down into separate scripts but don't have a coherent pipeline to connect them all.

# **Overcoming Memory Limitations**

For the smaller datasets, we were able to fit the entire matrix in memory. However, for the larger dataset with 100,000 reads, storing the matrix in memory proved to be more difficult. To overcome this, we tried two different approaches, outlined in brief below.

The first approach involved associating each read with a number (corresponding to its line number in the reads file) in order to compress the keys. We implemented this method using Python and used a large amount of memory on the supercomputer. For a reason unknown at the time of this writing, this method repeatedly failed after several hours on the supercomputer.

Our second approach was written in Java, and used an on disk key-value store to store the matrix. This enabled "relatively" fast access to individual entries in the matrix. The biggest limitation with this method is the amount of time it took to build the key-value store from the matrix file. Because we began this solution after several other attempts failed, we were not able

to run this method for an extended period, and were able to produce only a limited number of contigs.

Conclusion: Although calculating the entire 100,000 by 100,000 matrix proved to be tractable (the Secali program computed this matrix on a desktop computer in around 20 hours), using the output matrix to produce contigs proved to be rather more difficult, in terms of space and time complexity.

# **Project Suggestions**

- 1. Perhaps a preliminary assignment where we detail our approach and talk it through with a TA. The TA can then tell us of potential things to consider when implementing our approach.
- 2. Perhaps consider removing the synthetic datasets and include only the very small dataset (for testing purposes) and then the large human reads with errors. In our experience, we made sure our program worked on the very small dataset then went straight to work on the bigger more realistic datasets. The medium size synthetic datasets were somewhat of an afterthought and didn't really contribute to our assembly report (except that we ran them because it's required).