## **Tutorial for siestim**

Kurnia Susvitasari

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interest:

siestim is an R package that estimates the serial interval distribution in incomplete sampling data. The input is a vector (or a list of vectors) of the serial interval data. Given the data, siestim will estimate not only the parameters (mean and standard deviation) of the serial interval distribution, but also the parameters that explain the data incompleteness, e.g. the success probability of sampling the secondary case in a transmission chain. The package is created as a complementary for paper "A Method to Estimate Serial Interval Distribution Under Partially-sampled Data", which is co-author by Kurnia Susvitasari, Paul Tupper, Jessica Stockadale, and Caroline Colijn.

To install the package, you can run the following R command

remotes::install\_github("ksusvita92/siestim")

### Once installed, you should be able to call the package using: library(siestim)

- This tutorial consists of two parts. In the first half, we will simulate a data set of observed serial intervals directly from the Compound Geometric
  - mean (mu) of the serial interval distribution, • standard deviation (sigma) of the serial interval distribution • success probability (pi) of sampling the secondary case
- proportion of non-coprimary transmission (w); see [1]. In the second half, we will simulate an SIR outbreak and then we will estimate the same parameters of interest as above. This section is to show that siestim can also be implemented in a data set that contains stochastic uncertainty.

Gamma (CGG) distribution and the Folded Gamma Difference (FGD) distribution; see [1]. Then, we will estimate the following parameters of

#### Under incomplete sampling, the infector-infectee pairs may not represent direct transmissions. We differentiate two types of transmission: coprimary and non-coprimary. Coprimary transmission is defined as a pair in which both the infector and infectee were directly infected by the

thereby, is modeled by the FGD distribution.

- Distribution-based Simulation
- same unsampled case; non-coprimary transmission includes both direct transmission and inderect transmission (a pair in which the infector and infectee is separated by at least one unsampled cases). We model the serial intervals that come from coprimary transmission using FGD distribution and those come from non-coprimary transmission using CGG distribution. The idea of siestim is to use mixture model to estimate the parameters of interest.

Folded Gamma Difference (FGD) Distribution Suppose  $X_{xi}$  and  $X_{xj}$  represent the serial intervals between transmission pair  $x \to i$  and  $x \to j$  where x is an unsampled case. Then, a random variable

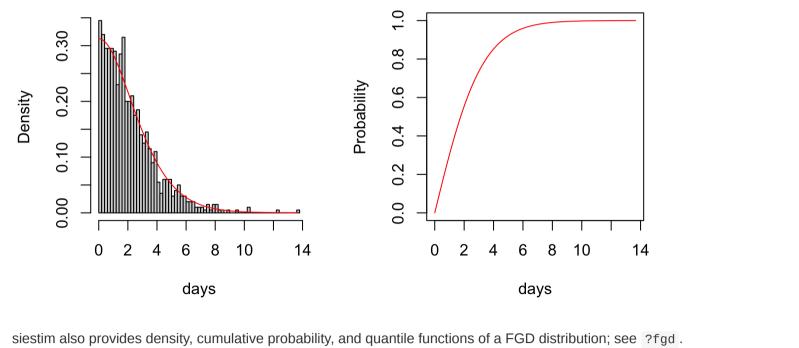
 $Y = |X_{xi} - X_{xj}|$ 

represents the symptom onset difference between i and j. In this case, the transmission pair  $i \to j$  is referred to coprimary transmission and

To generate random samples from FGD distribution of size 1000 with mean 4 days and standard deviation 2 days, you can run the following commands set.seed(12)

x < - rfgd(1000, 4, 2)# Fitting the histogram with the density

```
par(mfrow = c(1,2))
hist(x, breaks = 50, freq = F, xlab = "days", main = "FGD distribution")
lines(sort(x), dfgd(sort(x), 4, 2), col = "red")
base::plot(sort(x), pfgd(sort(x), 4, 2), xlab = "days", ylab = "Probability", main = "Cumulative Probability", tylength of the probability of th
pe = "1", col = "red")
                                                                                                                                                                                                                                                                                                                                                                         Cumulative Probability
                                                                                         FGD distribution
```



 $i o person_1 o \ldots o person_M o j$ 

#### Assuming the serial intervals in a transmission chain are identically distributed and independent, the symptom onset difference between case i and j is the convolution over all underlying serial intervals in the transmission pair $i \to j$ , that is

transmission.

can run the following commands

x < - rcgg(1000, 6, 1, .6)

set.seed(12)

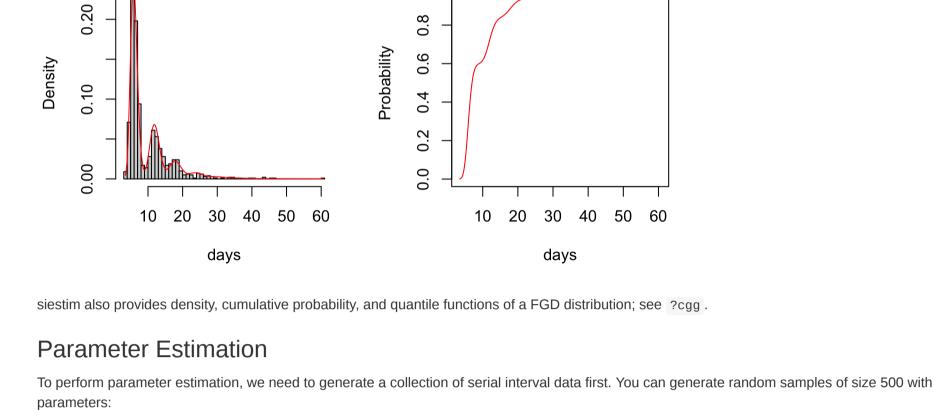
Compound Geometric Gamma (CGG) Distribution

 $Y = X_{iperson_1} + \ldots + X_{person_M i}$ In this case, Y is modeled by CGG distribution having parameters  $(\mu, \sigma, \pi)$ ;  $\pi$  is the success probability of sampling the secondary case.

Suppose that the transmission pair  $i \to j$  is separated by  $m=1,\ldots,M$  unsampled cases; if m=0, it means that  $i \to j$  is a direct

```
# Fitting the histogram with the density
par(mfrow = c(1,2))
hist(x, breaks = 50, freq = F, xlab = "days", main = "FGD distribution")
lines(sort(x), dcgg(sort(x), 6, 1, .6), col = "red")
base::plot(sort(x), pcgg(sort(x), 6, 1, .6), xlab = "days", ylab = "Probability", main = "Cumulative Probabilit
y", type = "1", col = "red")
            FGD distribution
                                                   Cumulative Probability
                                              1.0
```

To generate random samples from CGG distribution of size 1000 with mean 6 days, standard deviation 1 days, and success probability 0.6, you



# true parameters mu <- 4; sigma <- 2; pi <- .75; w <- .6

#> 4.156 2.055 0.789 0.547

*#*> 0.475 0.375 0.064 0.100

#> [1] "Successful convergence"

#> Convergence message:

• population size: 500

• initial infected case: 1

par(oma=c(0, 0, 0, 5))

• epidemic duration: 100 days

• reproduction number,  $R_0$ : 2

#> Standard error: #> mu sigma pi

To estimate the parameters of interest, you can run

 $iv \leftarrow c(5,3,.5,.5)$  # initial values of the estimates

 $1b \leftarrow rep(0, 4) \# lower bound of the estimates$ 

by running these following commands

•  $\mu=4$  days •  $\sigma=2$  days •  $\pi = .7$ • w = .6

set.seed(12) n < - rbinom(1, 500, w)x <- c(rcgg(n, mu, sigma, pi), rfgd(500-n, mu, sigma))

```
ub <- c(10, 5, 1, 1) # upper bound of the estimates
myest <- siestim(x, iv, lb, ub)</pre>
myest
#> ---- Serial Interval Estimation ----
#> Parameter estimations:
#> mu sigma pi w
```

As well, you can estimate the 95% confidence intervals for each estimate by running the following command getci(myest, .95) #> lower upper #> mu 3.2243807 5.0876834 #> sigma 1.3202107 2.7890577 #> pi 0.6638437 0.9133689 0.3519951 0.7427887 **SIR Simulation** You can generate an influenza-like outbreak using siestim using a function called simOutbreak. The function generates an SIR outbreak together with, for every infected case, the DNA sequences and epidemiological data (who infected whom, time of infection and recovery).

#### • generation interval: $\Gamma(4,2)$ • importation rate: 0.01 • transversion rate: $5 imes 10^{-5}$

Suppose you want to generate an outbreak with these parameters:

lines(0:100, myoutbreak\$dynam\$ninf, col = "red") lines(0:100, myoutbreak\$dynam\$nrec, col = "blue")

• transition rate:  $10^{-4}$ Then, you can run set.seed(12)

```
myoutbreak <- simOutbreak(500, c(4,2))</pre>
#> Attempt 1 : Generate outbreak with ncases = 388
#> Attaching package: 'dplyr'
#> The following objects are masked from 'package:stats':
       filter, lag
#> The following objects are masked from 'package:base':
#>
#>
      intersect, setdiff, setequal, union
```

base::plot(0:100, myoutbreak\$dynam\$nsus, type = "1", col = "darkgreen", xlab = "day", ylab = "Count")

```
legend(par('usr')[2], par('usr')[4], bty = 'n', xpd = NA, legend = c("susceptible", "infected", "recovery"), fill
= c("darkgreen", "red", "blue"))
     500
                                                           susceptible
                                                             infected
    400
                                                           recovery
Count
    300
    200
     100
          0
                   20
                           40
                                    60
                                             80
                                                     100
                               day
```

Assuming that we sample only p proportion of infected cases and we do not know the true transmission pairs. In this case, we need to reconstruct all plausible transmission pairs from the genomic data. We say that case i and j are linked as an infector-infectee pair if their genomic distance is

You can sample p=.6 of the infected cases to reconstruct the transmission cloud with the genomic distance cutoff  $\varepsilon=18$  SNPs using

## Select by group

Reconstruct a Transmission Cloud

mytc <- createTC(myoutbreak, .6, 18, seed = 12)</pre>

col\_pal = c(unsampled = "black", sampled = "gold"),

# true transmission tree adegenet::plot(mytc\$tt, thin = F,

node value = 1,

 $arrow_size = .5$ ,  $node_size = 5$ ,  $edge_width = .5,$  $node\_width = .5,$ height = 800,width = 600, label = F)

# plot transmission cloud

tc\_nodes <- mytc\$tt\$linelist %>% filter(group == "sampled") %>%

library(dplyr)

select(id)

library(visNetwork)

x\_axis = "inf.times", node\_color = "group",

less than a non-negative threshold  $\varepsilon$ . We refer all plausible transmission pairs as *transmission cloud*.

```
tc_edges <- data.frame(from = mytc$tc$inf.source, to = mytc$tc$inf.ID, arrows = "to")</pre>
 visNetwork(tc_nodes, tc_edges, width = "100%", height = "500px") %>%
   visOptions(highlightNearest = list(enabled = TRUE, algorithm = "hierarchical")) %>%
   visNodes(size = 25) %>%
   visEdges(width = 5, color = "black") %>%
   visLegend() %>%
   visLayout(randomSeed = 12)
Parameter Estimation
```

# Sometimes we may be uncertain about which cases are truly linked. For a given case, there may be multiple other cases that may have infected them (see the transmission cloud above). To handle this extra source of uncertainty, we sample a collection of transmission trees consistent with it,

and then estimate the parameters of interest over all sampled trees.

mytt <- lapply(1:100, function(i){</pre>

in parallel; see ?parallel::detectCores() for detail.

group\_by(inf.ID) %>% slice\_sample(n = 1) %>%

#> Parameter estimations: mu sigma pi #> 3.955 2.104 0.814 0.898

mu sigma pi w

#> Standard error:

 $0.05^{-1}$ 

0.00

0

mytc\$tc %>%

pull(si)

})

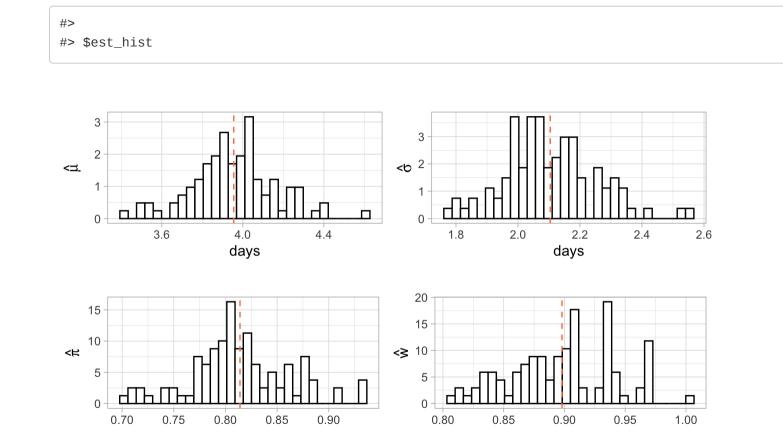
To sample, say, 100 transmission trees from given transmission cloud, you can run

myest #> ---- Serial Interval Estimation -----

myest <- siestim(mytt, iv, lb, ub, list(ncore = parallel::detectCores()))</pre>

```
#> 0.536 0.433 0.108 0.072
To plot the siestim output, you can use
 siestim::plot(myest)
 #> $density
        Gamma-distributed Serial Interval
                                                       Mixture Density
    0.25
    0.20
 density
0.10
                                                   0.10
```

We estimate the serial interval distribution using the collection of serial interval data sets above. The following command allows you to run siestim



0.05

0.00

15

10

— est. per data — final est.

You can also estimate the 95% confidence interval using

lower upper

Commun **14**, 4830 (2023).←

getci(myest, .95)

#> mu 2.9040124 5.006162 #> sigma 1.2556631 2.952586 #> pi 0.6022847 1.000000 0.7563758 1.000000

10

· - · cop. trans. - - non-cop. trans.

15

```
The function siestim allows you to include prior distributions on each parameter \mu, \sigma, \pi, w. For more detail, see ?siestim.
   1. Stockdale, J.E., Susvitasari, K., Tupper, P. et al. Genomic epidemiology offers high resolution estimates of serial intervals for COVID-19. Nat
```