

Biophysics 210: Biological Light Microscopy
Kurt Thorn
Syllabus

Discussion section meets Tuesdays from 1-2:30pm in MH2100

Labs meet Thursday or Friday from 2-5pm (location varies)

Week 8: Measuring Cellular Processes; Fluorescent Biosensors

Goals: Understand methods for measuring dynamical processes in cells, both for tracking motion of biomolecules and for following chemical changes in cells using biosensors.

Discussion Section: May 19th

Labs: May 21st and 22nd

Lectures (watch before discussion section):

- [Measuring Dynamics: Photobleaching and Photoactivation](#)
- [Measuring Dynamics: Fluorescent Speckle Microscopy](#)
- [Förster Resonance Energy Transfer \(FRET\) Microscopy](#)
- [Fluorescent Protein Indicators](#)

Required Additional Reading:

- [To, T. L., Piggott, B. J., Makhijani, K., Yu, D., Jan, Y. N., & Shu, X. \(2015\). Rationally designed fluorogenic protease reporter visualizes spatiotemporal dynamics of apoptosis in vivo. *Proceedings of the National Academy of Sciences*, 112\(11\), 3338-3343.](#)

Additional Reading (optional):

- [Lippincott-Schwartz, J., Altan-Bonnet, N., & Patterson, G. H. \(2003\). Photobleaching and photoactivation: following protein dynamics in living cells. *Nature Cell Biology*, S7-14.](#)
- [McKinney, S. A., Murphy, C. S., Hazelwood, K. L., Davidson, M. W., & Looger, L. L. \(2009\). A bright and photostable photoconvertible fluorescent protein. *Nature methods*, 6\(2\), 131-133.](#)
- [Salmon, E. D., & Waterman, C. M. \(2011\). How we discovered fluorescent speckle microscopy. *Molecular biology of the cell*, 22\(21\), 3940-3942.](#)

- [Danuser, G., & Waterman-Storer, C. M. \(2006\). Quantitative fluorescent speckle microscopy of cytoskeleton dynamics. *Annu. Rev. Biophys. Biomol. Struct.*, 35, 361-387.](#)
- [Berney, C., & Danuser, G. \(2003\). FRET or no FRET: a quantitative comparison. *Biophysical journal*, 84\(6\), 3992-4010.](#)
- [Chen, T. W., Wardill, T. J., Sun, Y., Pulver, S. R., Renninger, S. L., Baohan, A., ... & Kim, D. S. \(2013\). Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature*, 499\(7458\), 295-300.](#)

Discussion Section Topic: We will discuss FRAP / FLIP / photoactivation and other techniques for tracking movement as well as different strategies for biosensor design and their strengths and weaknesses.

Lab: We will demonstrate Ca^{++} imaging using GCaMP and photoactivation and tracking of mEos2.