

# Prediction of Adar Editing Levels from bpRNA Structure

load the feature matrix and split into training and test sets

```
set.seed(1)

#load the feature matrix
data=read.table("rf_features.txt",header=TRUE,sep='\t')
#remove values with no NaN in editing level
data=data[is.nan(data$ave_editing_level)==FALSE,]
head(data)
```

##	ave_editing_level	edit_feat	prev_feat	next_feat	mp1	mref1	malt1	mtype1
## 1	0.58	I	S	S	44	G	A	mismatch
## 2	0.46	I	S	S	45	G	A	mismatch
## 3	0.49	B	S	S	46	G	A	mismatch
## 4	0.48	I	S	S	47	G	A	mismatch
## 5	0.59	I	S	S	48	C	A	mismatch
## 6	0.39	I	S	S	52	G	A	mismatch

##	mfeat1	mfeat1_prev	mfeat1_next	adist1	mp2	mref2	malt2	mtype2	mfeat2
## 1	S	H	I	6	None	None	None	None	None
## 2	S	H	I	5	None	None	None	None	None
## 3	S	H	B	4	None	None	None	None	None
## 4	S	H	I	3	None	None	None	None	None
## 5	S	H	I	2	None	None	None	None	None
## 6	S	I	I	-2	None	None	None	None	None

##	mfeat2_prev	mfeat2_next	adist2
## 1	None	None	None
## 2	None	None	None
## 3	None	None	None
## 4	None	None	None
## 5	None	None	None
## 6	None	None	None

The feature values are:

- edit\_feat – Single character value of structure type for the edited A at position 50. One of S,H,M,I,B,X,E
- prev\_feat – The 5' structural feature upstream of edited A
- next\_feat – The 3' structural feature downstream of edited A
- mp1 – First mutated position along the RNA sequence
- mref1 – Reference allele at first mutated position
- mtype1 – One of “mismatch”, “indel”, “wt”
- mfeat1 – Single character value of structure type for mutated base. One of S,H,M,I,B,X,E
- mfeat1\_prev – The 5' structural feature upstream of mutated base.
- mfeat1\_next – The 3' structural feature downstream of mutated base.
- adist1 – The distance (in sequence space ) of the mutated base from the edited A.

Repeat for possible second mutation ('None' if only 1 mutation present in sequence):

- mp2
- mref2
- mtype2
- mfeat2
- mfeat2\_prev
- mfeat2\_next
- adist2

```
#get training and test splits; use 80% train, 20% test split
train_indices=sample(nrow(data),0.8*nrow(data),replace=FALSE)
train_split=data[train_indices,]
test_split=data[-train_indices,]
```

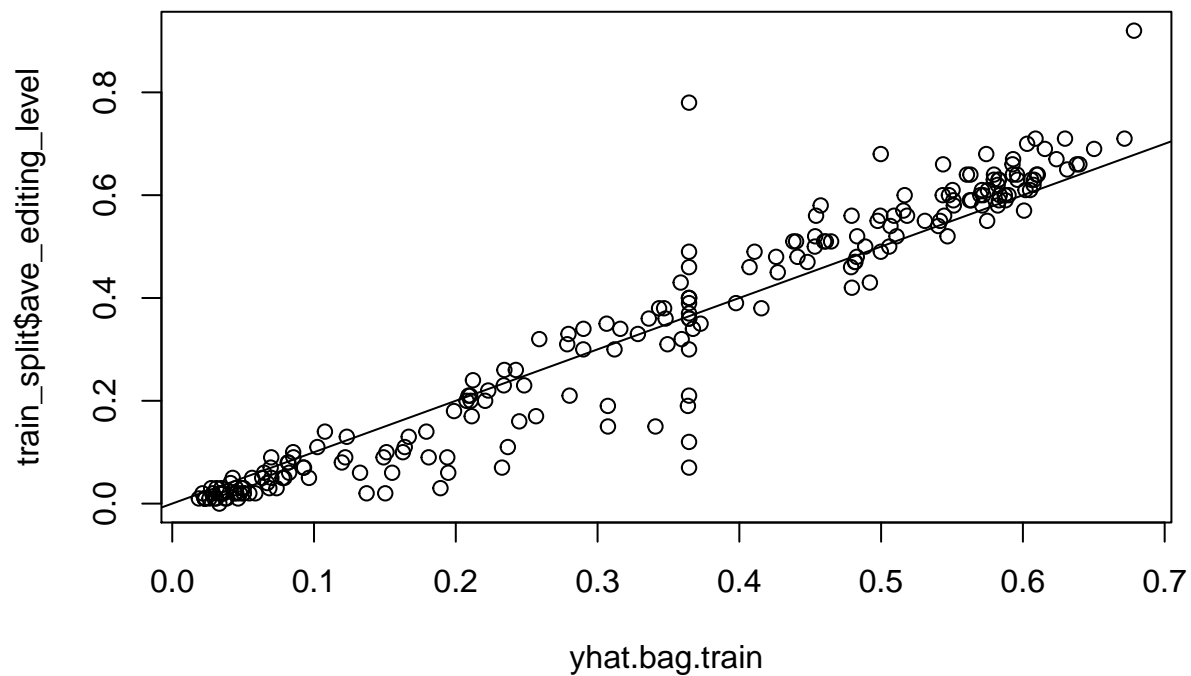
## Bagging

```
#train rf
bag.data=randomForest(ave_editing_level~.,data=data,subset=train_indices,mtry=19,importance=TRUE)
print(bag.data)

##
## Call:
##  randomForest(formula = ave_editing_level ~ ., data = data, mtry = 19,      importance = TRUE, subse
##              Type of random forest: regression
##              Number of trees: 500
## No. of variables tried at each split: 19
##
##              Mean of squared residuals: 0.02028694
##              % Var explained: 66.33
```

## Predictions on training split

```
#get predictions on training & test data
yhat.bag.train=predict(bag.data,newdata=train_split)
yhat.bag.test=predict(bag.data,newdata=test_split)
plot(yhat.bag.train,train_split$ave_editing_level)
abline(0,1)
```

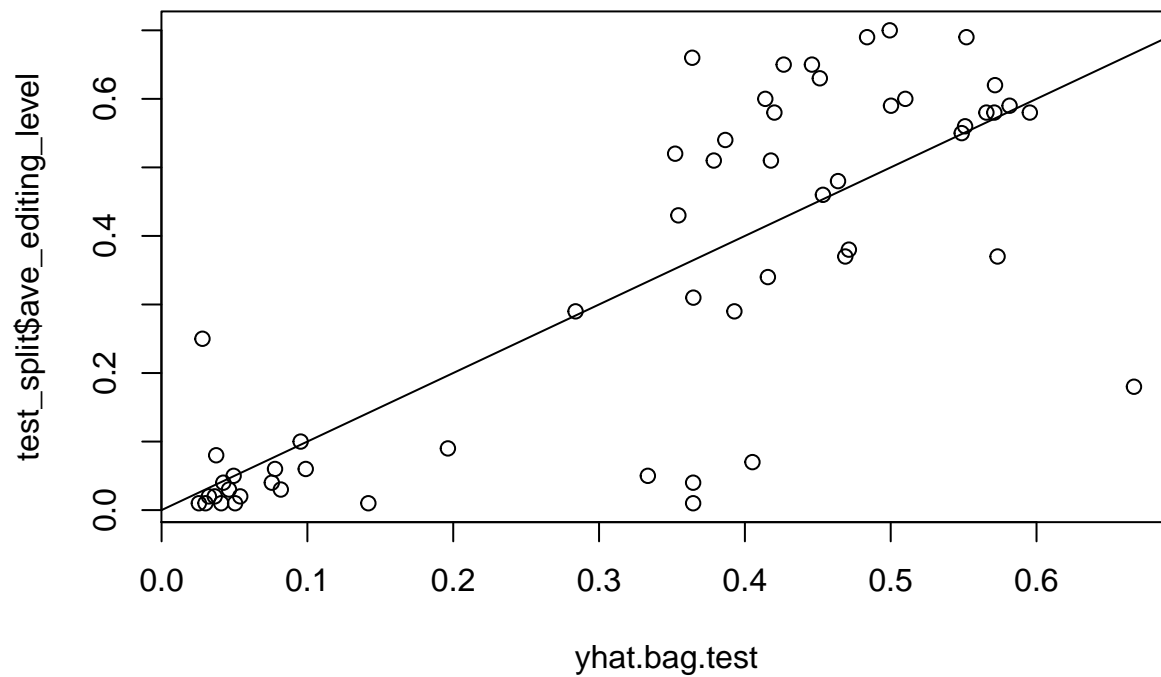


```
print(mean((yhat.bag.train-train_split$ave_editing_level)^2))
```

```
## [1] 0.00472248
```

### Predictions on test split

```
plot(yhat.bag.test,test_split$ave_editing_level)
abline(0,1)
```



```
print(mean((yhat.bag.test-test_split$ave_editing_level)^2))
```

```
## [1] 0.02331793
```

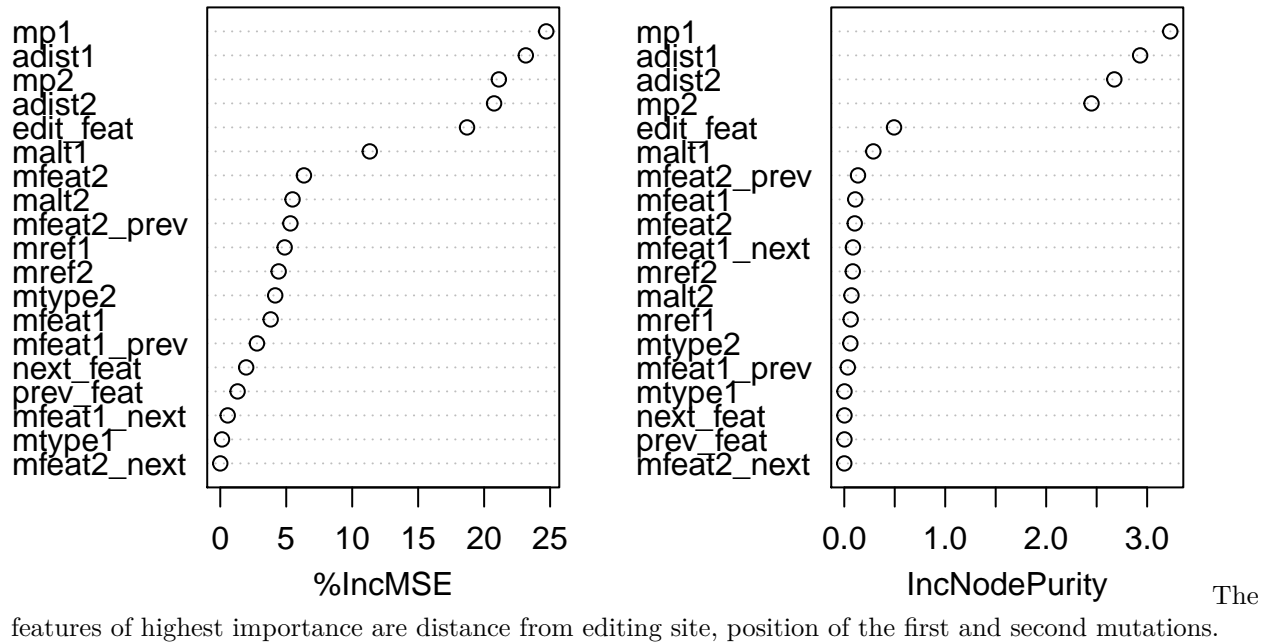
## Feature Importance

```
#get the feature importance  
importance (bag.data )
```

```
##           %IncMSE IncNodePurity  
## edit_feat  18.7039268  0.493035316  
## prev_feat   1.3142513  0.001756823  
## next_feat   1.9645418  0.002037126  
## mp1        24.7061524  3.229132949  
## mref1       4.8820341  0.063430900  
## malt1      11.3305610  0.287789138  
## mtype1      0.1320094  0.002121355  
## mfeat1       3.8201350  0.108592032  
## mfeat1_prev  2.7825562  0.034755064  
## mfeat1_next  0.5648393  0.085860348  
## adist1      23.1578734  2.930658413  
## mp2        21.1147003  2.448996284  
## mref2       4.4266836  0.084321687  
## malt2       5.4808279  0.071266847  
## mtype2      4.1661620  0.060279124  
## mfeat2       6.3435142  0.103652048  
## mfeat2_prev  5.3159174  0.135840224  
## mfeat2_next  0.0000000  0.000000000  
## adist2      20.7532765  2.675104494
```

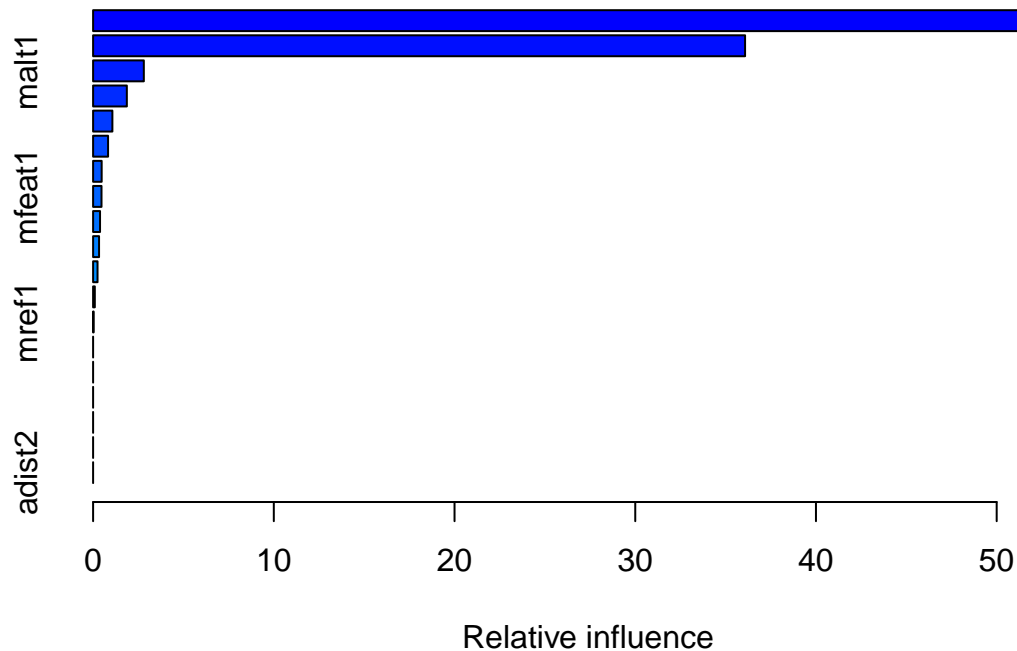
```
varImpPlot(bag.data)
```

bag.data



## Boosting

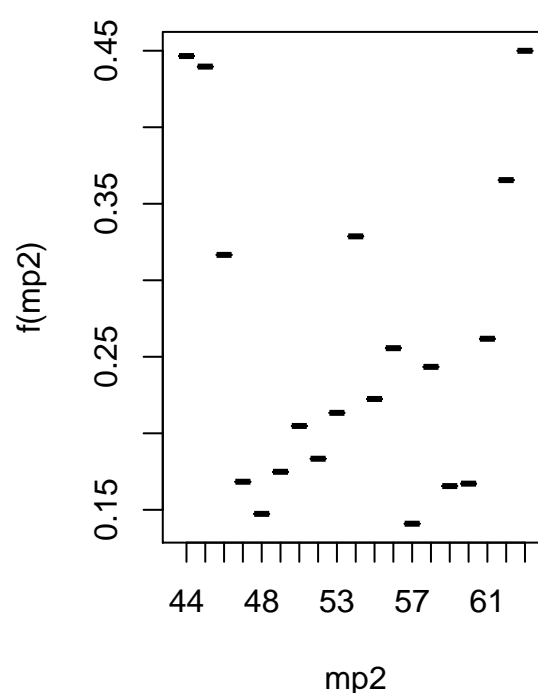
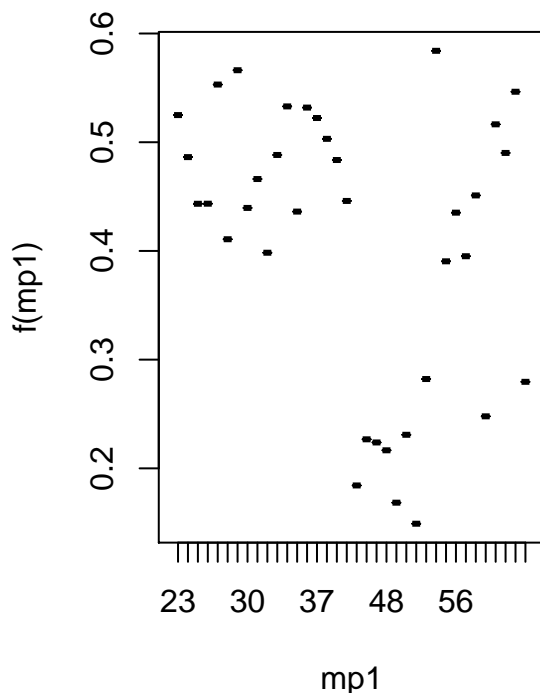
```
boost.data=gbm(ave_editing_level~.,data=train_split,distribution="gaussian",n.trees=5000,interaction.depth=4)
summary(boost.data)
```



```
##          var      rel.inf
## mp1      mp1 55.33374191
## mp2      mp2 36.07809806
## malt1    malt1 2.80729944
## edit_feat edit_feat 1.86652344
## mfeat2_prev mfeat2_prev 1.06718314
## mfeat1_next mfeat1_next 0.82839213
## mfeat2     mfeat2 0.47408074
## mfeat1     mfeat1 0.46410669
## mtype2     mtype2 0.38417724
## mref2      mref2 0.32889475
## malt2      malt2 0.24439359
## mfeat1_prev mfeat1_prev 0.09503127
## mref1      mref1 0.02807759
## prev_feat  prev_feat 0.00000000
## next_feat  next_feat 0.00000000
## mtype1     mtype1 0.00000000
## adist1     adist1 0.00000000
## mfeat2_next mfeat2_next 0.00000000
## adist2     adist2 0.00000000
```

The “mp1” and “mp2” features are the most important variables (these are the positions along the sequence of mutation 1 and 2). We produce partial dependence plots for these two variables. These plots illustrate the marginal effect of the mp1 and mp2 variables after integrating out the other variables.

```
par(mfrow=c(1,2))
plot(boost.data,i="mp1")
plot(boost.data,i="mp2")
```



We use

the boosted model to predict on the test data:

```
yhat.boost=predict(boost.data,newdata=test_split,n.trees=5000)
mean((yhat.boost-test_split$ave_editing_level)^2)
```

```
## [1] 0.02125189
```

Experiment with the boosting shrinkage parameter (increase to 0.2 from default of 0.001)

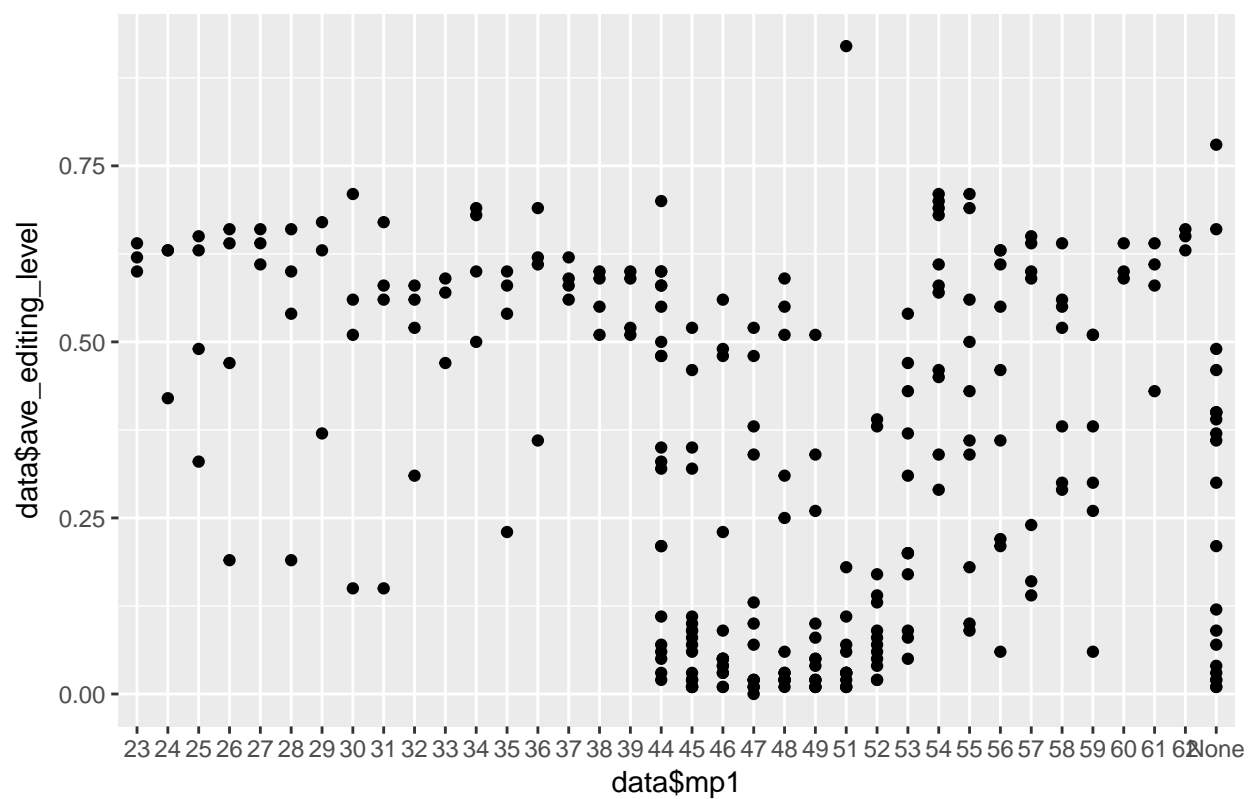
```
for(shrinkage_val in c(0.005, 0.01, 0.05, 0.1, 0.2))
{
  print(shrinkage_val)
  boost.data=gbm(ave_editing_level~.,
                 data=train_split,
                 distribution="gaussian",
                 n.trees=5000,
                 interaction.depth=4,
                 shrinkage=shrinkage_val,verbose=F)
  yhat.boost=predict(boost.data,newdata=test_split,n.trees=5000)
  print(mean((yhat.boost-test_split$ave_editing_level)^2))
}
```

```
## [1] 0.005
## [1] 0.02435091
## [1] 0.01
## [1] 0.02757491
## [1] 0.05
## [1] 0.0335663
## [1] 0.1
## [1] 0.03733136
## [1] 0.2
## [1] 0.04807122
```

## Feature values vs editing levels

```
p1=ggplot(data=data,aes(x=data$mp1,y=data$ave_editing_level))+
  geom_point()+
  ggtitle("Position of first mutation along sequence")
p1
```

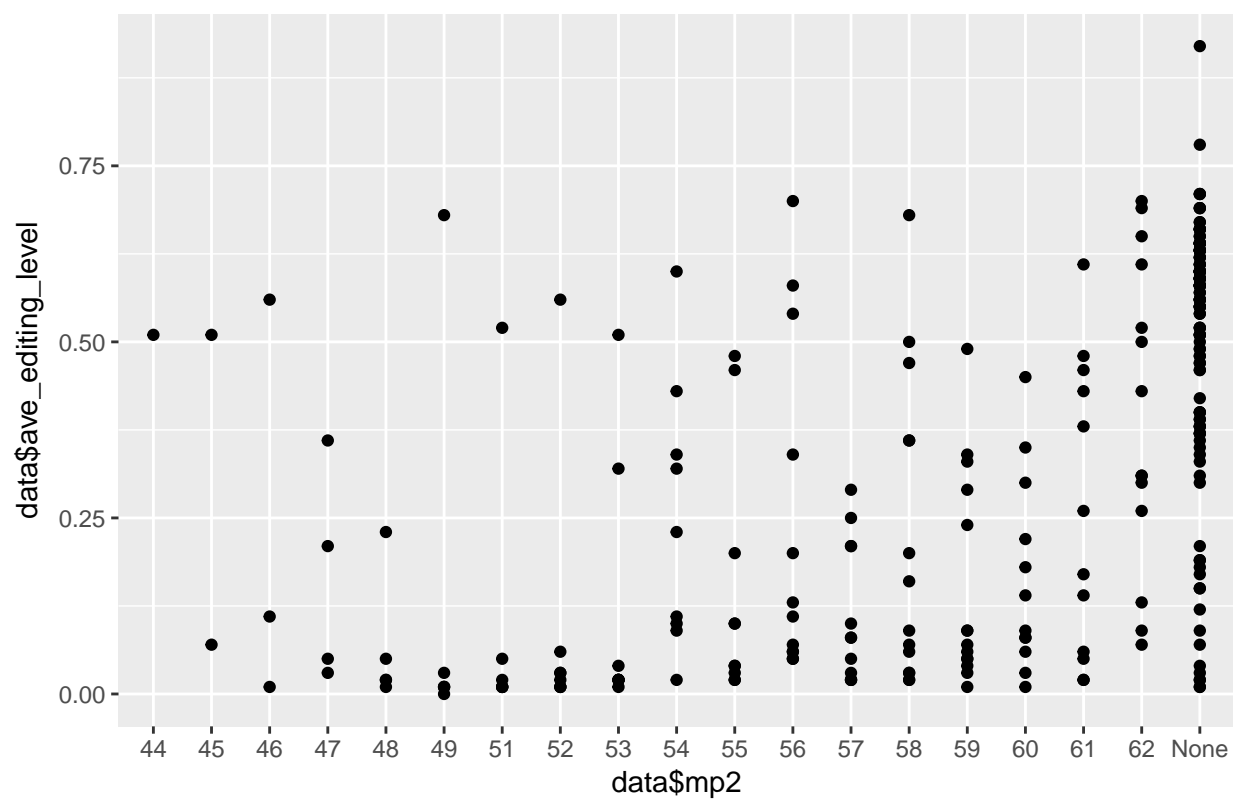
Position of first mutation along sequence



```
p2=ggplot(data=data,aes(x=data$mp2,y=data$ave_editing_level))+
  geom_point()+
  ggtitle("Position of second mutation along sequence")
p2
```

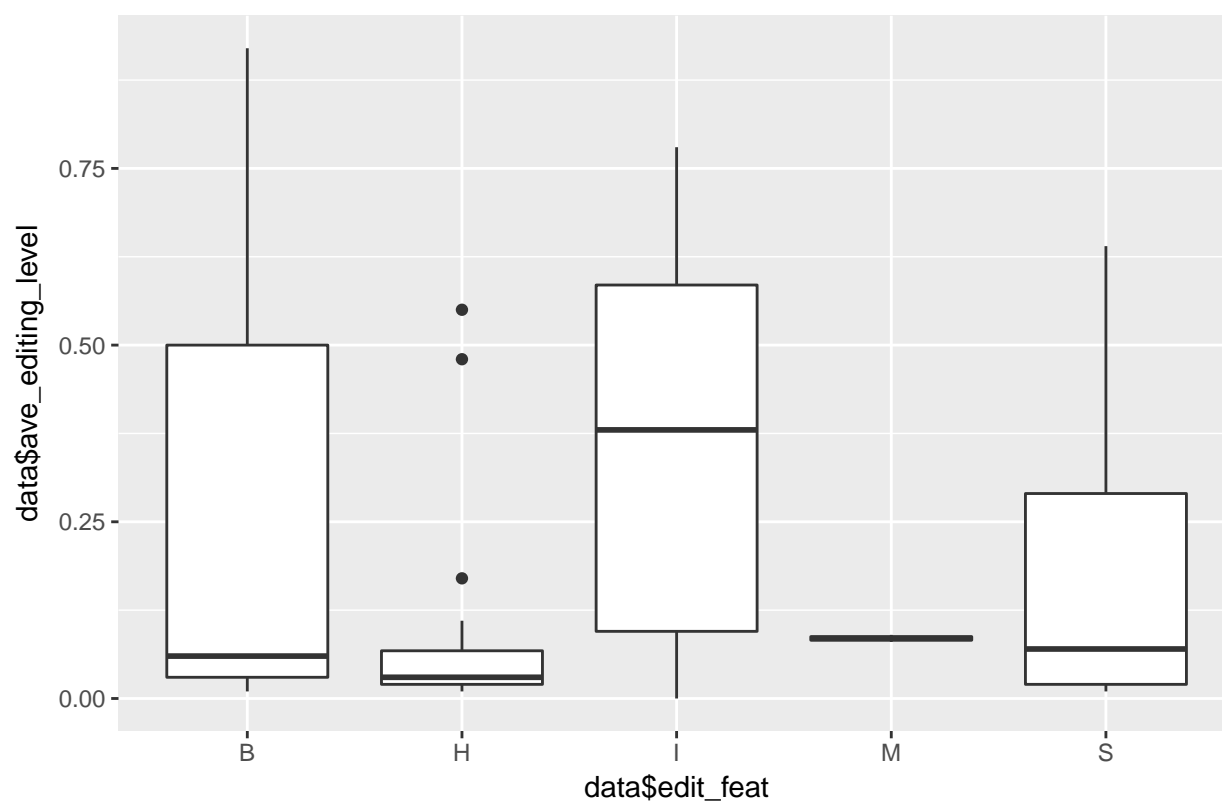


Position of second mutation along sequence



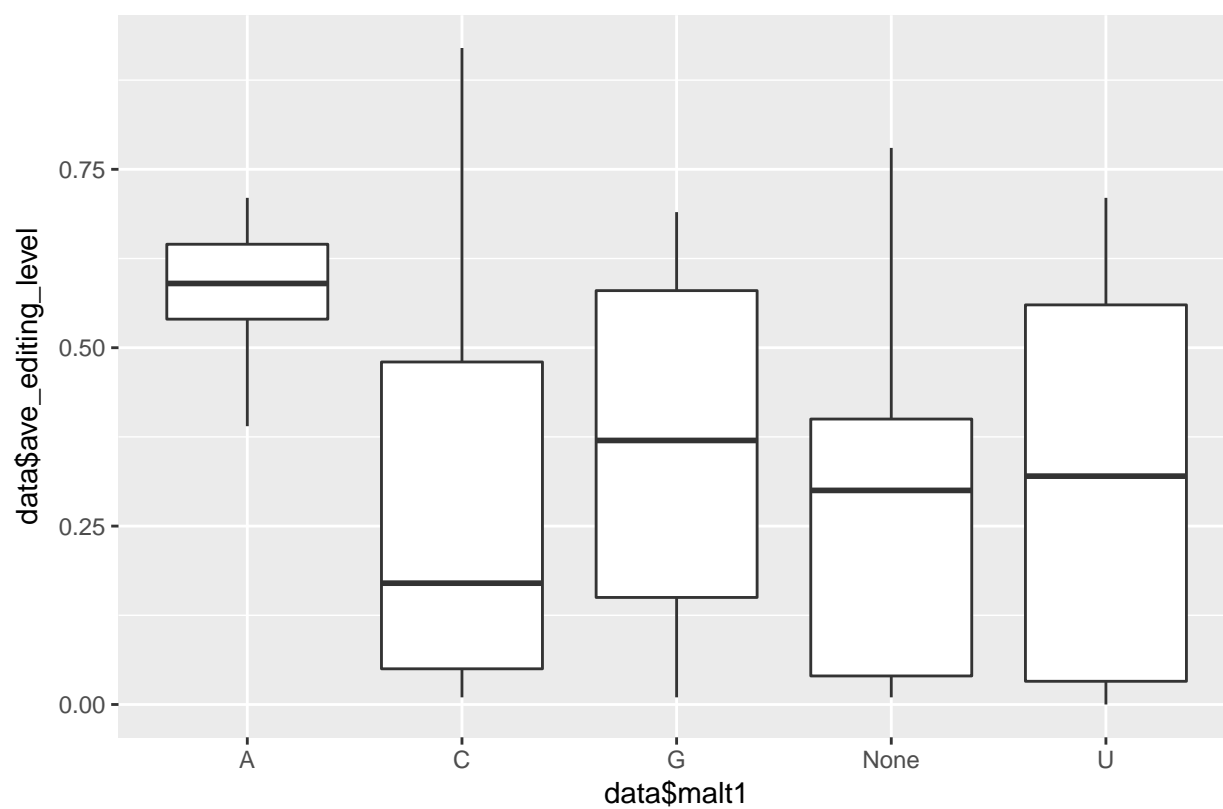
```
p3=ggplot(data=data,aes(x=data$edit_feat,y=data$ave_editing_level))+
  geom_boxplot()+
  ggtitle("Structural feature of the editing site")
p3
```

Structural feature of the editing site



```
p4=ggplot(data=data,aes(x=data$malt1,y=data$ave_editing_level))+  
  geom_boxplot() +  
  ggtitle("First mutation -- new base ")  
p4
```

First mutation -- new base



```
p5=ggplot(data=data,aes(x=data$malt2,y=data$ave_editing_level))+
  geom_boxplot() +
  ggtitle("Second mutation -- new base")
p5
```

