

# ***MSD1 regulates pedicellate spikelet fertility in sorghum through the jasmonic acid pathway***

ABE516 Project 2

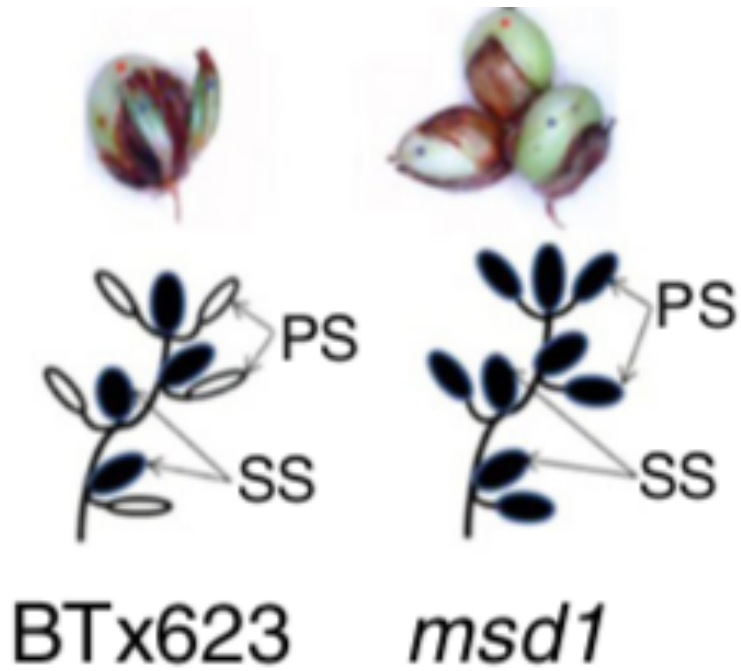
Chen Chen and Ken Youens-Clark



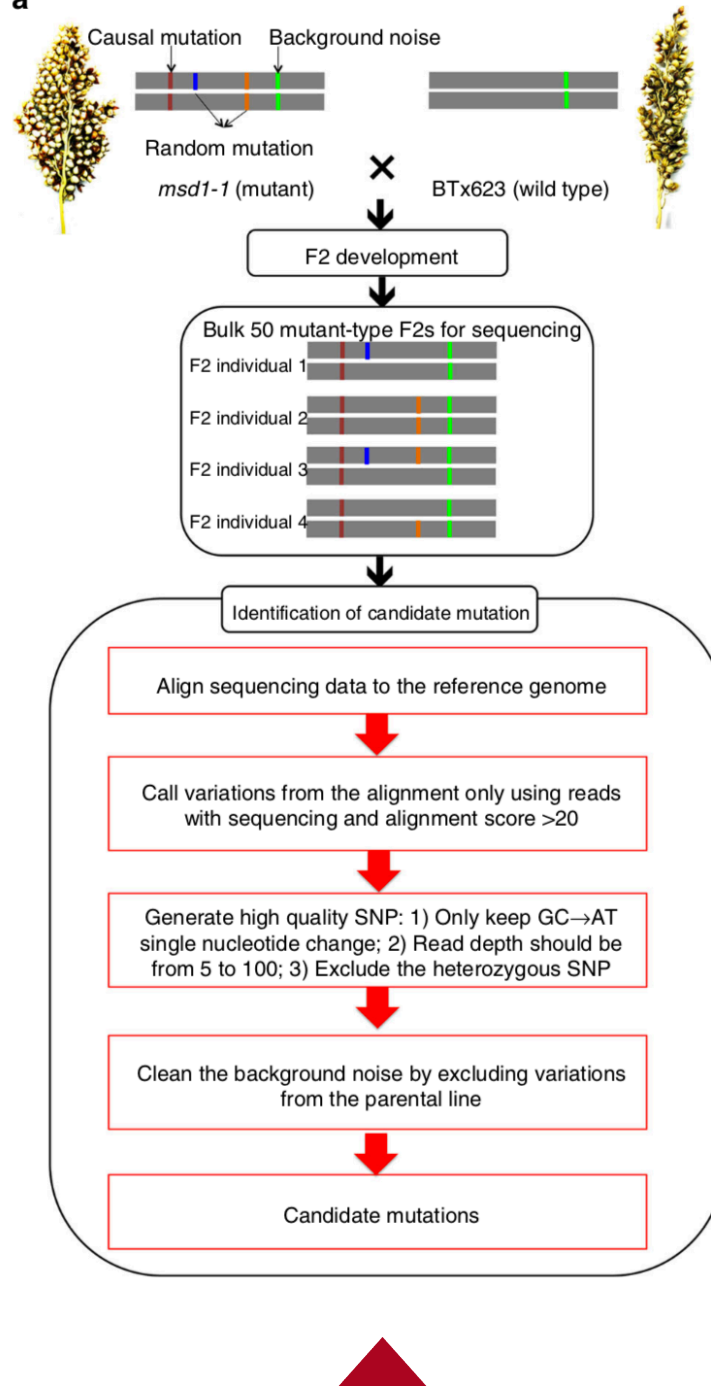
# Sorghum



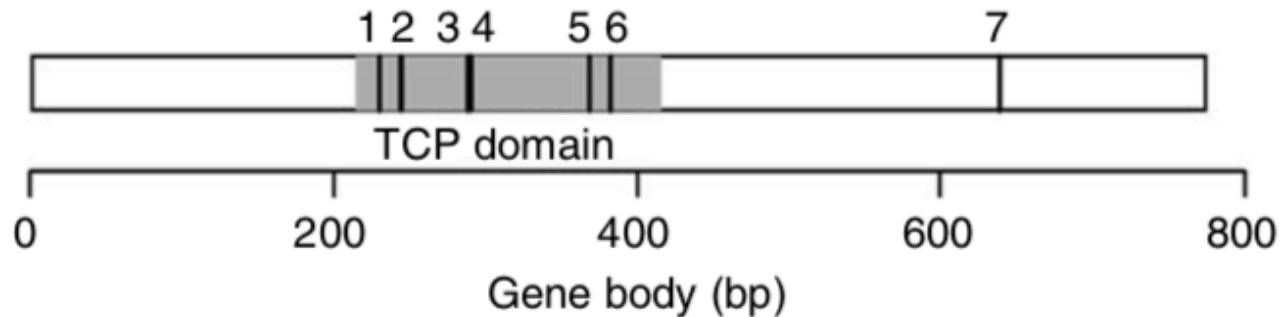
## BTx623 v *msd1*



**a**

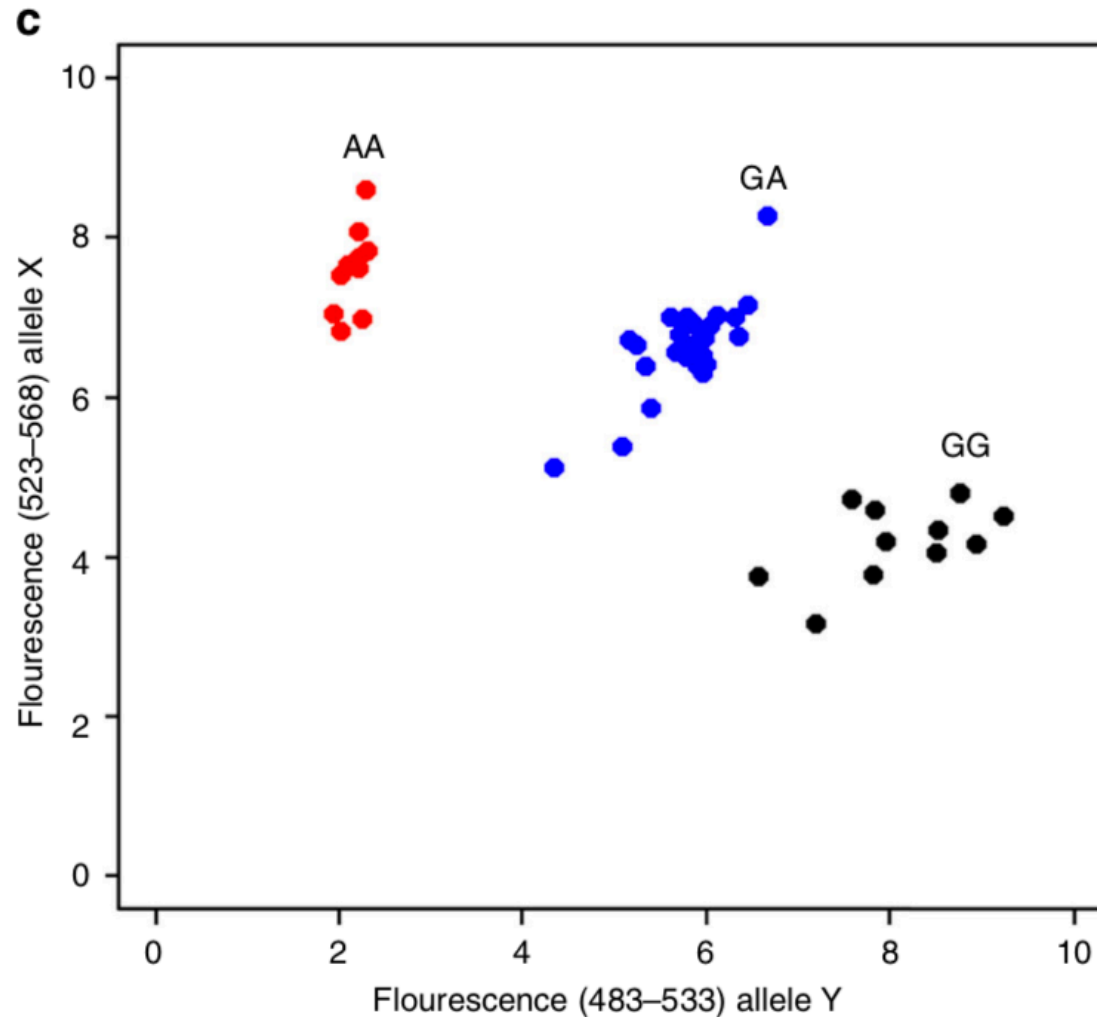


# Identification of SNPs in MSD1 gene



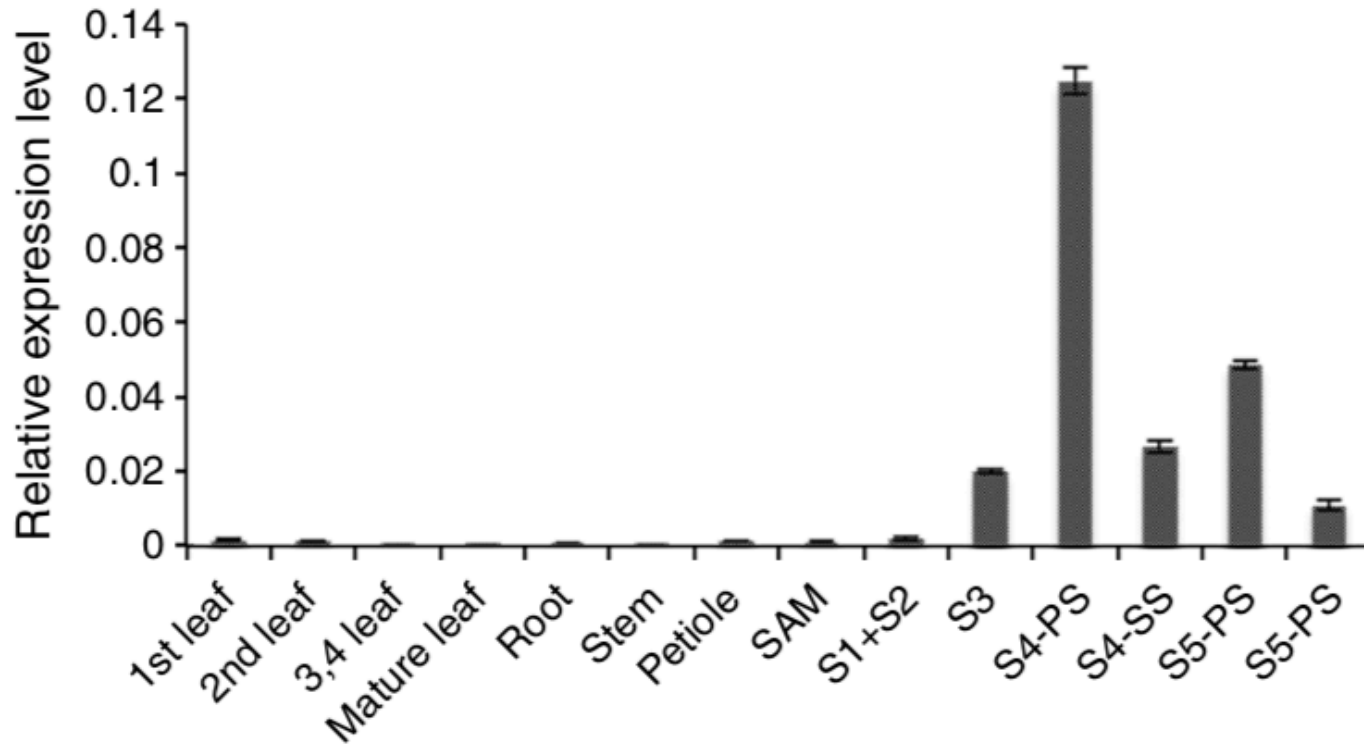
SNP ID	Mutant	Chromosome	Location	Variation	Mutation effect	Amino acid change
1	<i>msd1-7</i>	7	54912926	G/A	Missense_variant	R/Q
2	<i>msd1-5</i>	7	54912940	G/A	Missense_variant	D/N
3	<i>msd1-6</i>	7	54912985	C/T	Missense_variant	R/W
4	<i>msd1-1</i>	7	54912986	G/A	Missense_variant	R/Q
5	<i>msd1-2</i>	7	54913064	C/T	Missense_variant	T/M
6	<i>msd1-3</i>	7	54913078	C/T	Missense_variant	L/F
7	<i>msd1-4</i>	7	54913335	G/A	Stop gained	W/*

# Homozygous mutation (AA) mutation leads to *msd1* panicles



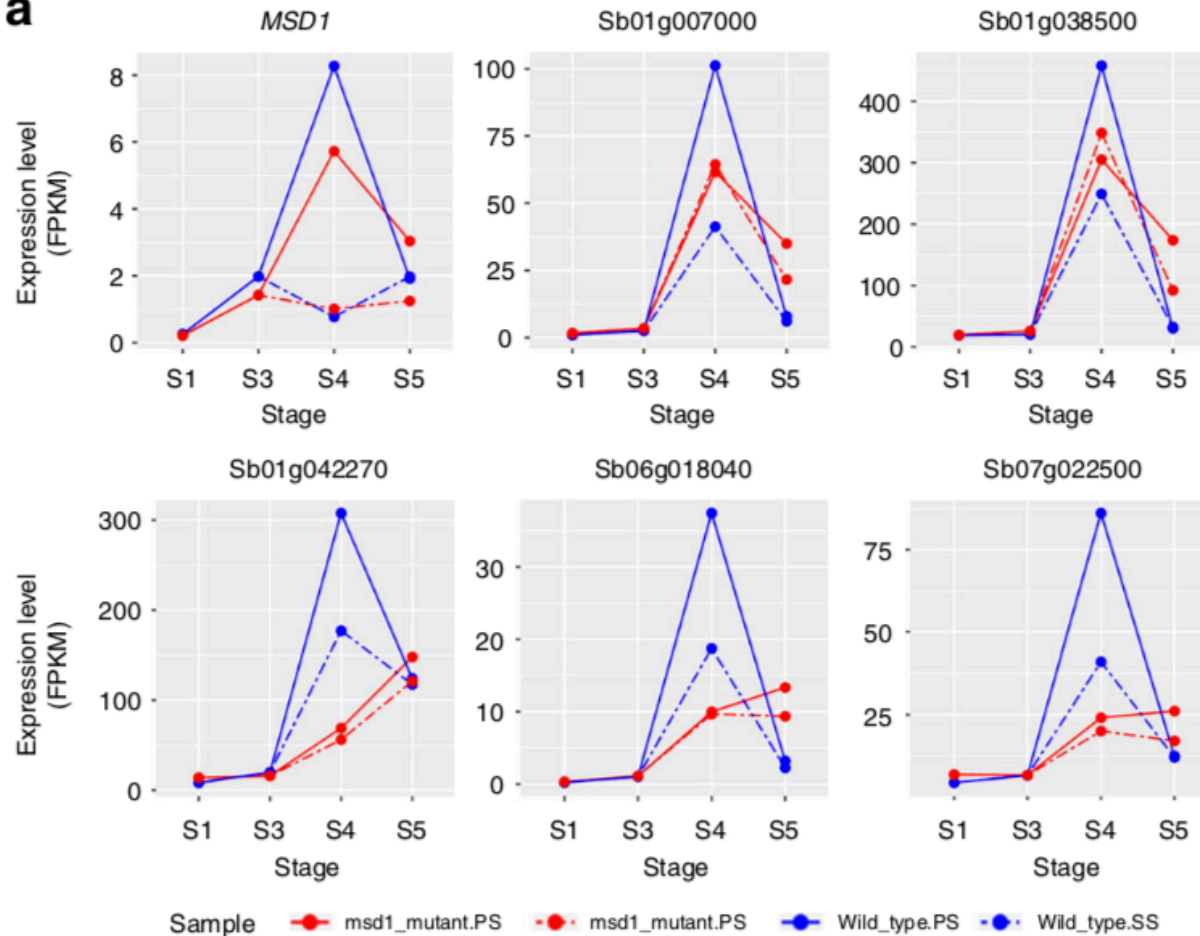
# Relative expression of MSD1 from qRT-PCR in various tissue types

**a**



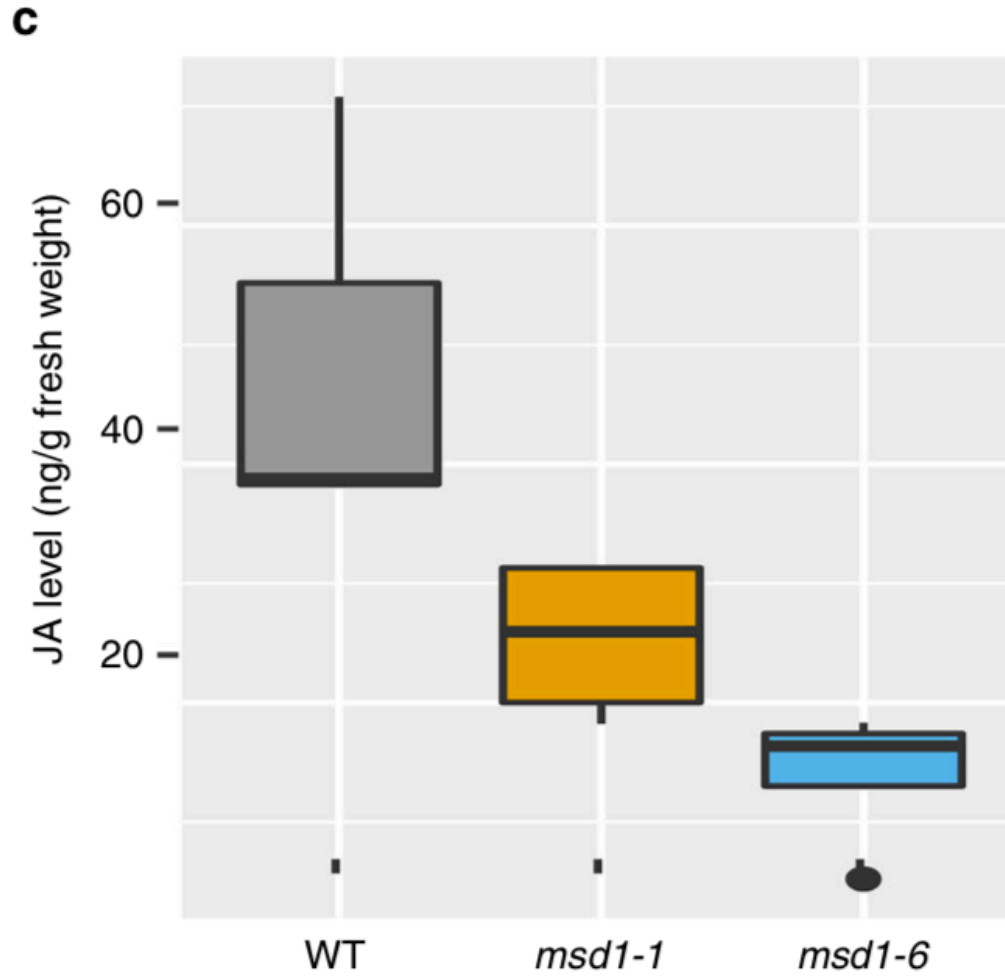
# Expression pattern of 5 target genes involved in JA synthesis

**a**





## JA levels are significantly different in WT v *msd1*



# *msd1* can be “rescued” with application of JA

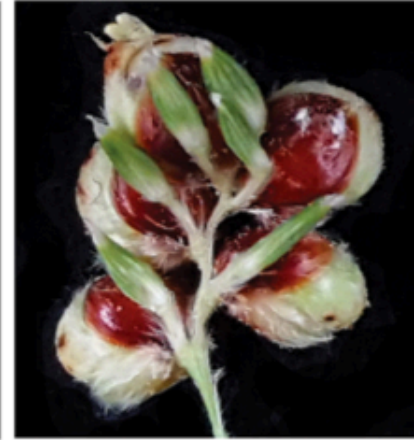
BTx623 untreated



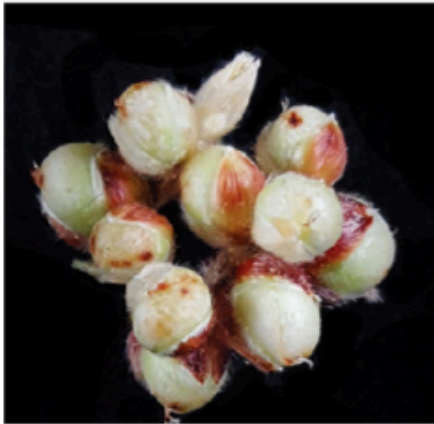
BTx623 water



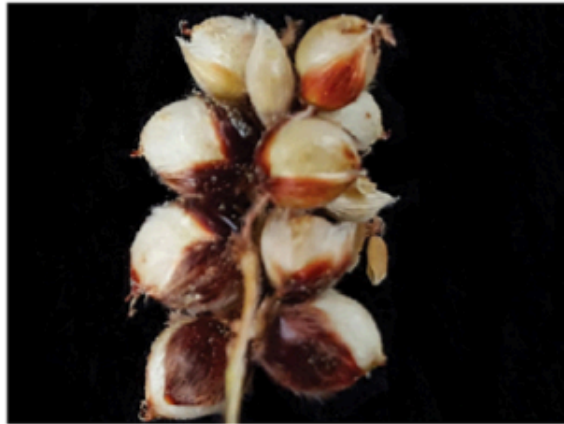
BTx623 + Me-JA



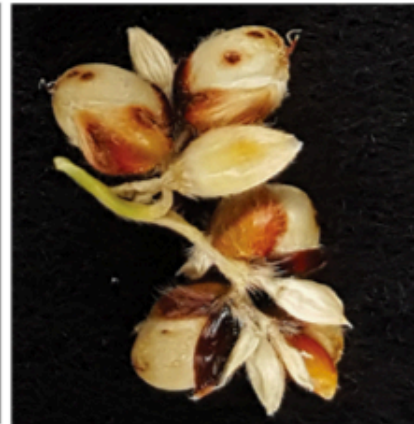
*msd1* untreated

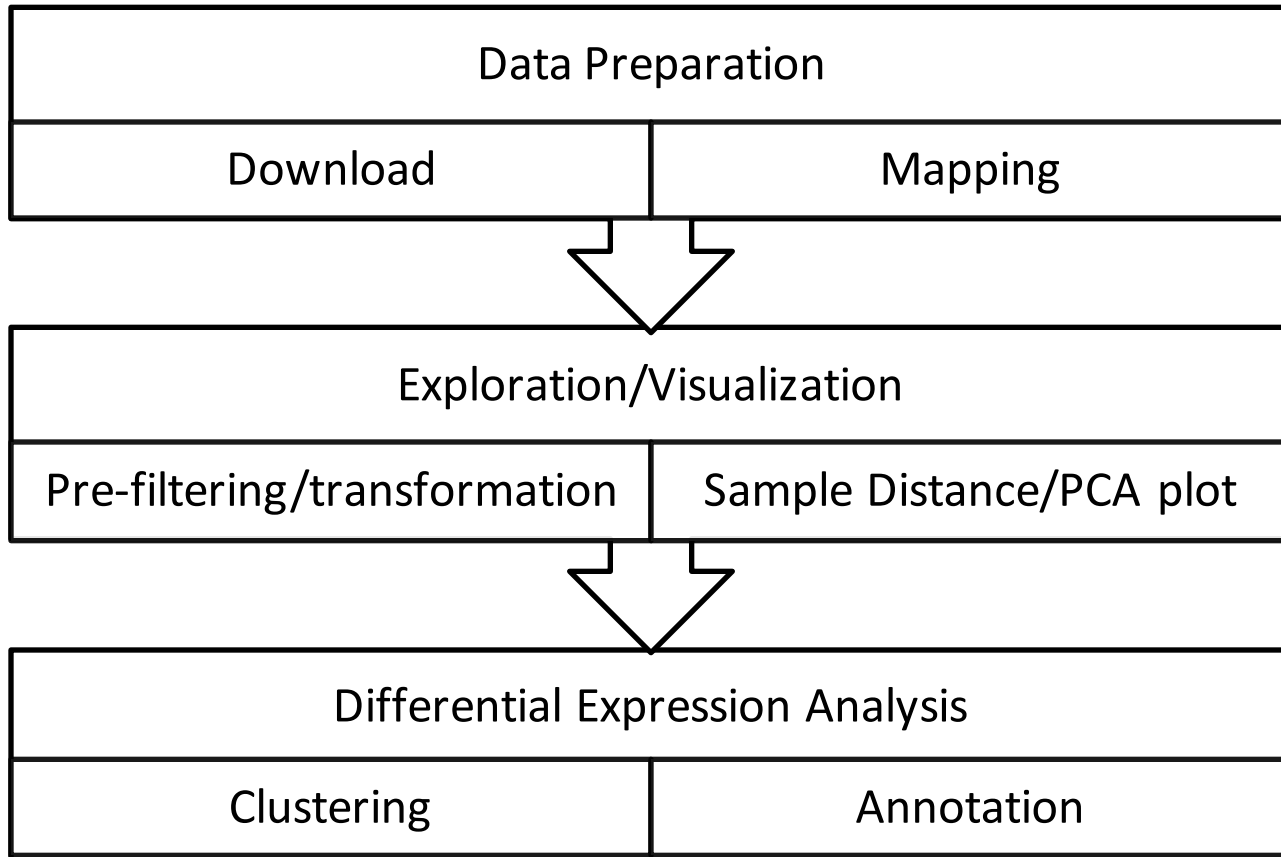


*msd1* water



*msd1* + Me-JA





Download SRA data from NCBI website and use SRAToolkit to convert them to fastq format.

Quality Check

## Basic Statistics

Measure	Value
Filename	SRR6431607.fastq
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	35654668
Sequences flagged as poor quality	0
Sequence length	250
%GC	44

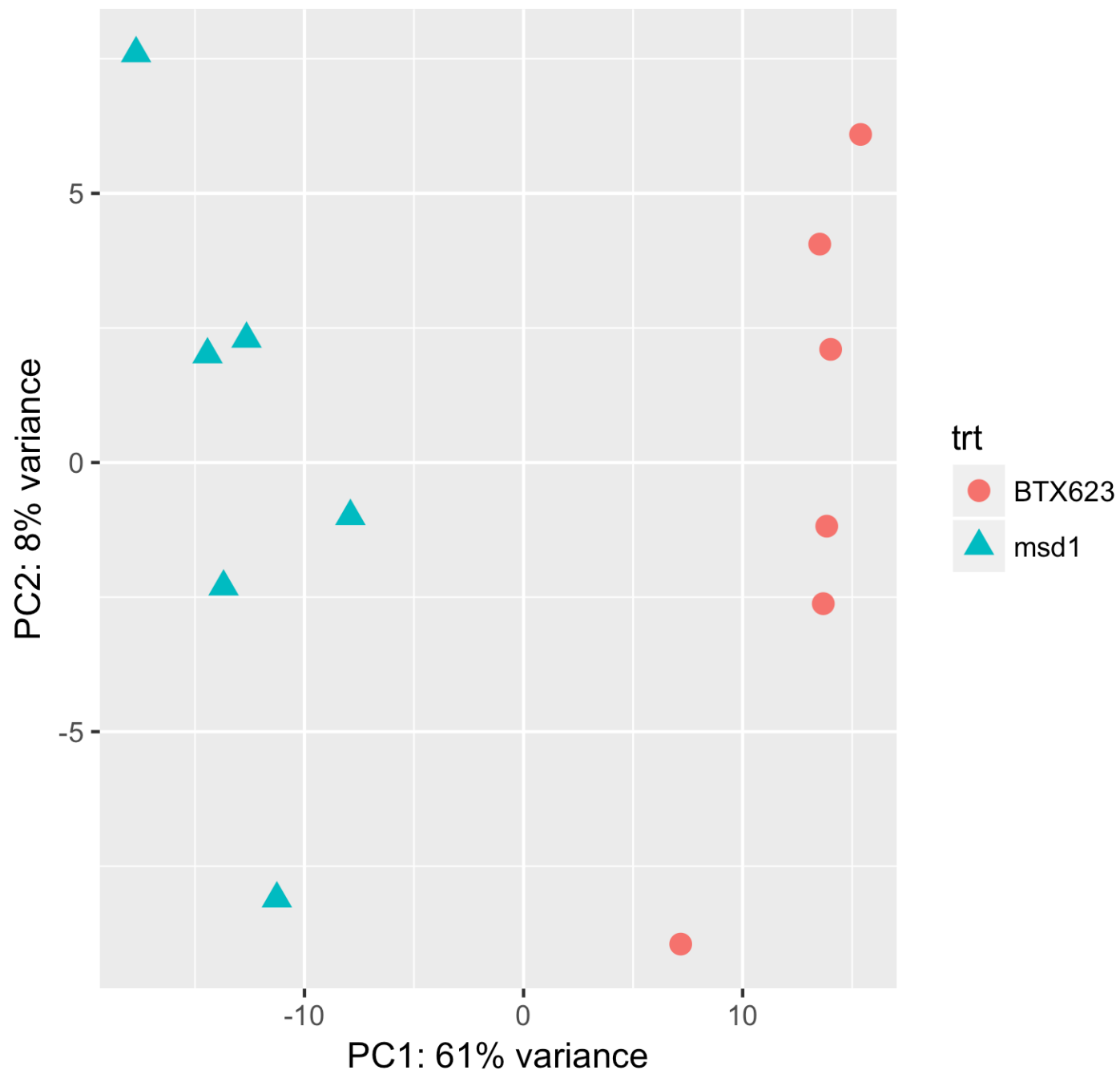
We used the STAR read aligner (Dobin et al. 2013) to align the reads for our current experiment to the Sorghum Bicolor reference genome.

<b>BTX623 (wild type)</b>	<b>Msd1 (mutant)</b>
SRR6431605.bam	SRR6431615.bam
SRR6431606.bam	SRR6431616.bam
SRR6431607.bam	SRR6431621.bam
SRR6431608.bam	SRR6431622.bam
SRR6431609.bam	SRR6431627.bam
SRR6431610.bam	SRR6431628.bam

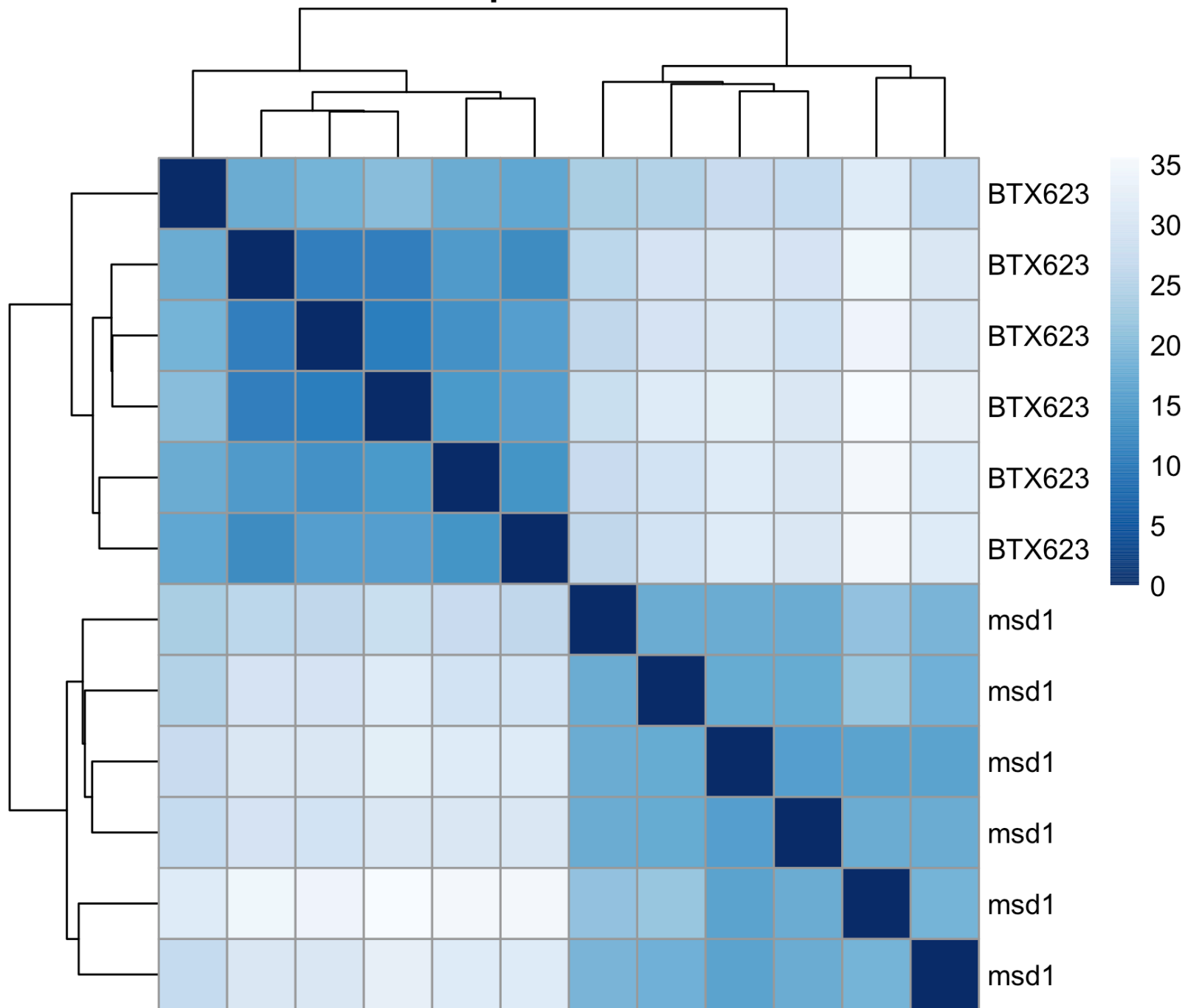
For this analysis, we used 2M sequences for each of the 12 samples.



# Stage4



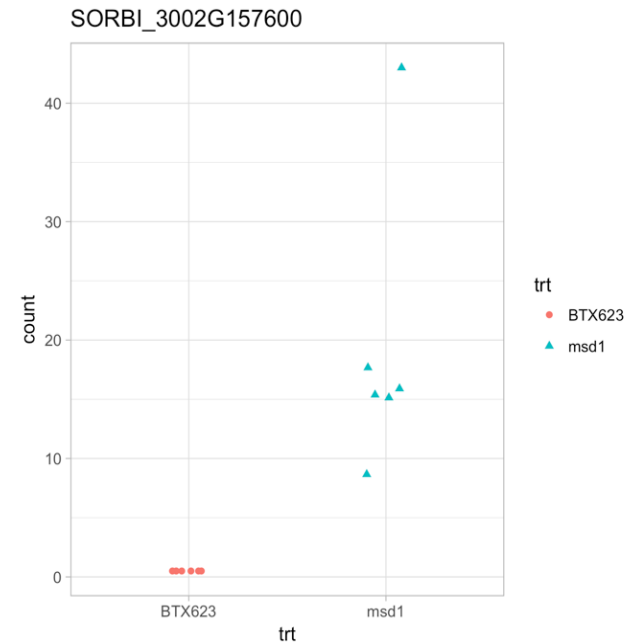
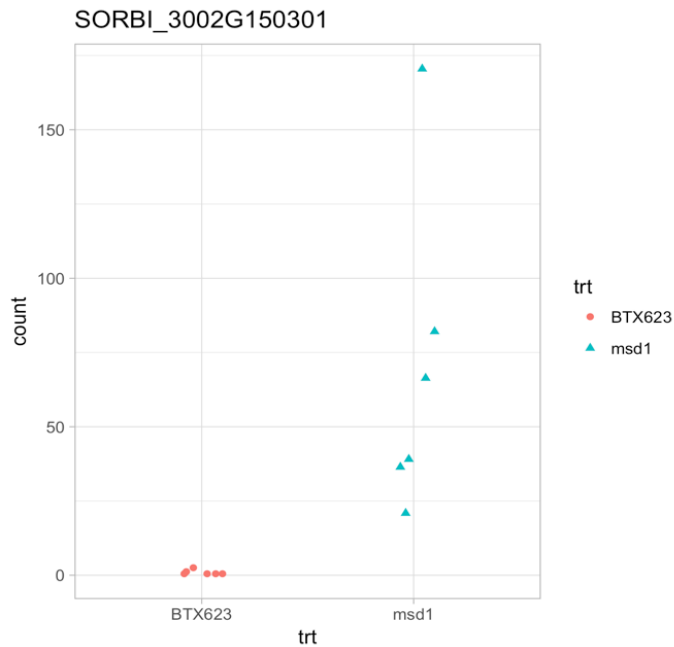
Sample Distances



We used DESeq2 to do differential expression analysis with FDR=0.05.

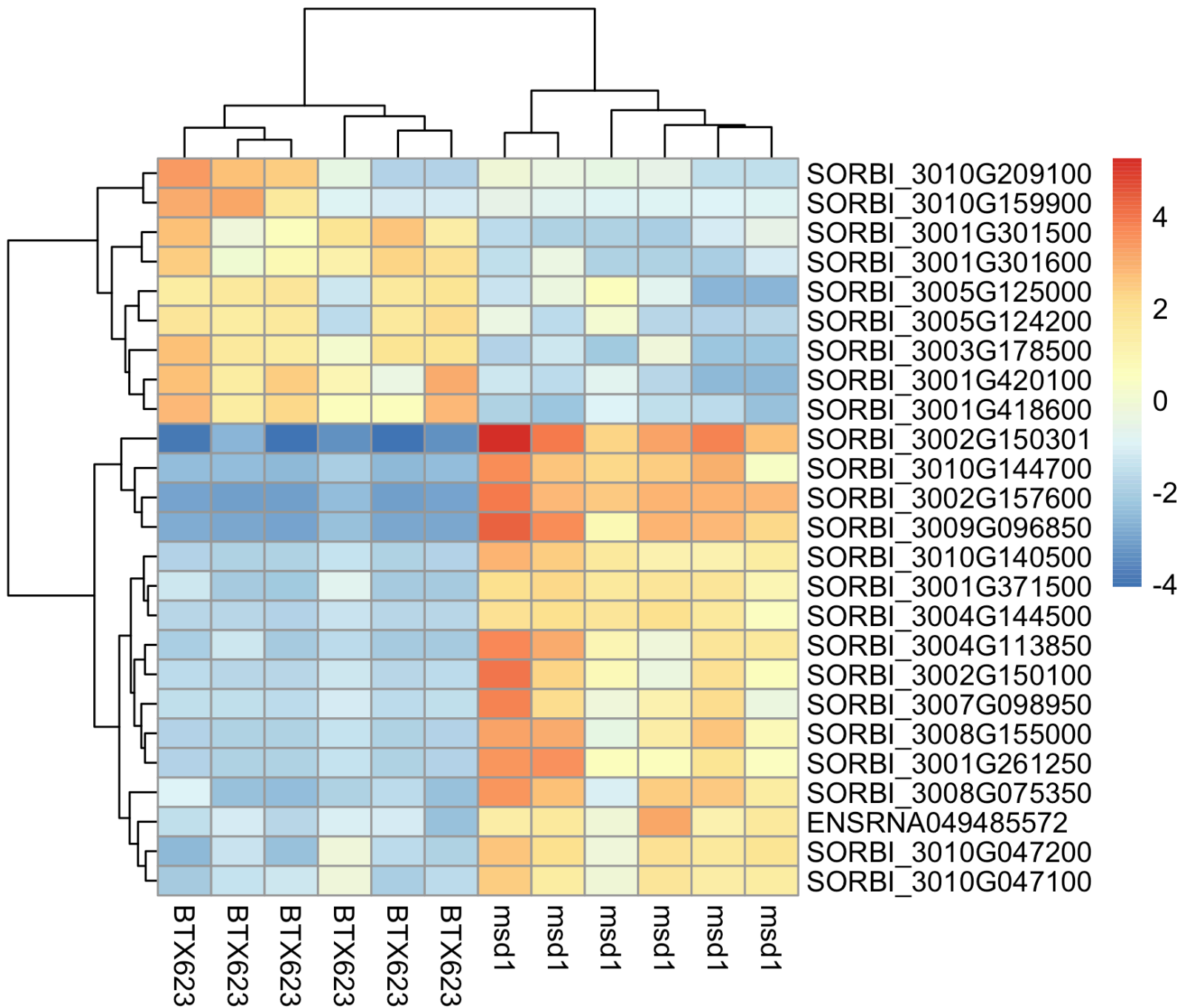
## With 556 non-zero read counts

LFC>0 (up-regulated)	73 (13%)
LFC<0 (down-regulated)	60 (11%)

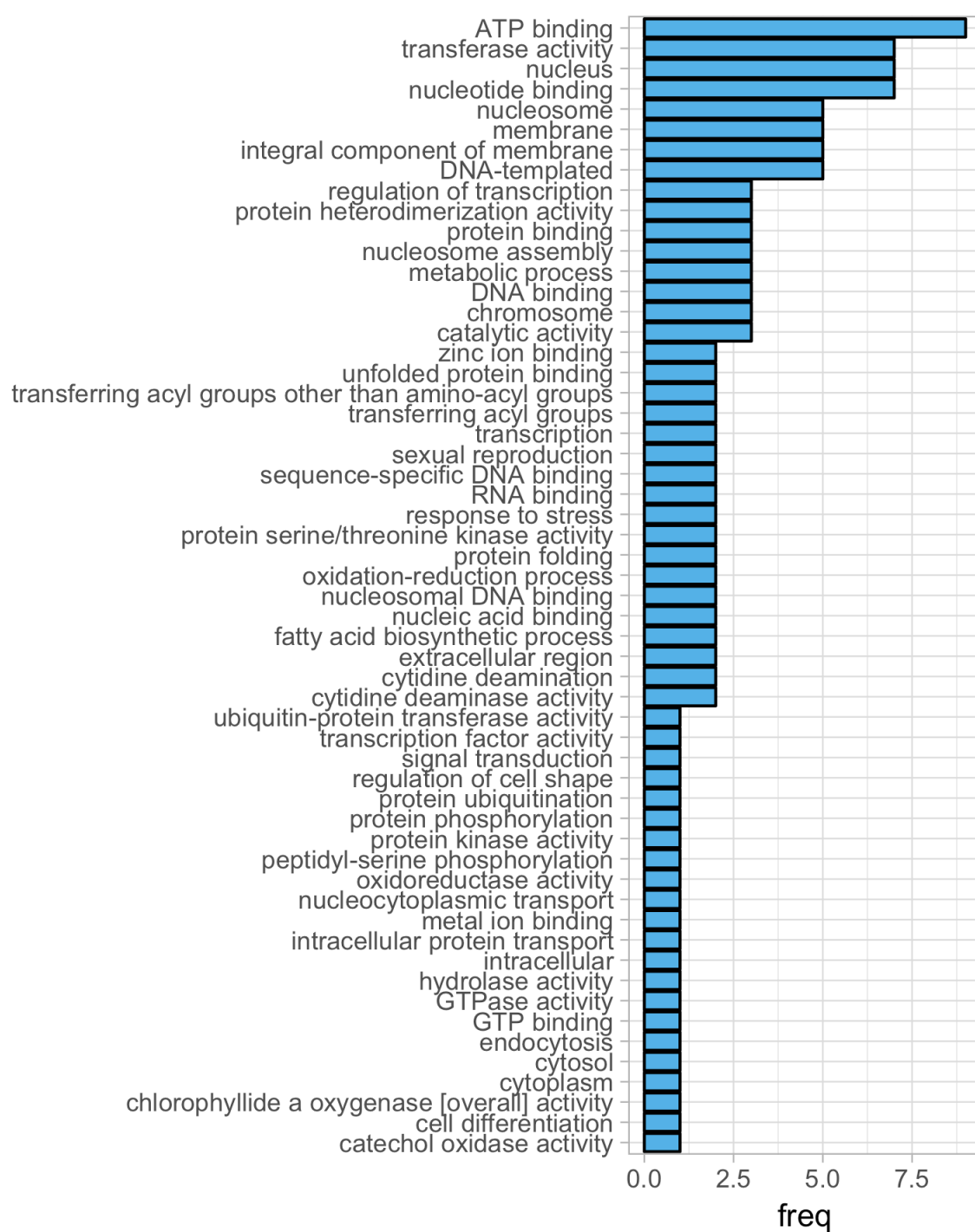




# Gene Clusters



func



## Conclusion:

Using the first 10 million sequences out of 36 million sequences, we detected 133 differentially expressed genes between stage-4 BTX623 and msd1 samples.

Among them, 73 genes are upregulated and 60 are downregulated.

With clustering method and gene ontology we found some interesting patterns.

Clustering of samples is very consistent with the background information.

Further analysis like pathway analysis and network analysis is needed.

