

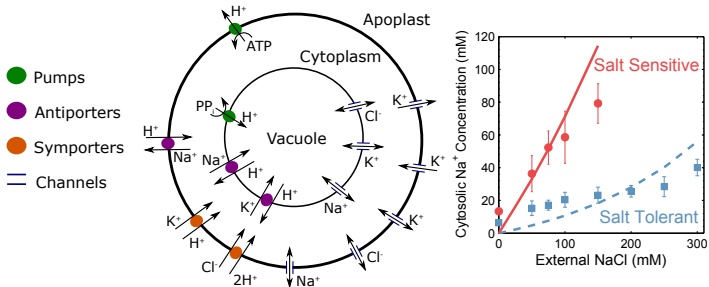
Mathematically modelling the salt stress response of individual plant cells

Kylie Foster

Supervisor: Prof. Stan Miklavcic

Phenomics and Bioinformatics Research Centre

University of South Australia



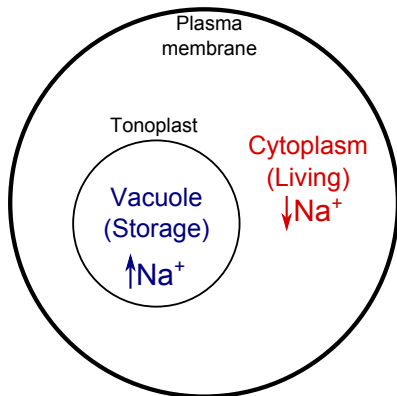
Motivation: Salinity

- Salinity is a problem globally
- In Australia high salinity affects **two-thirds** of cereal crops
- Impacts of salinity are expected to become worse
- Need to breed salt tolerant plants:
 - Requires identifying and understanding important transport processes
 - Need to identify the functions of relevant genes
 - Mathematical modelling can help!



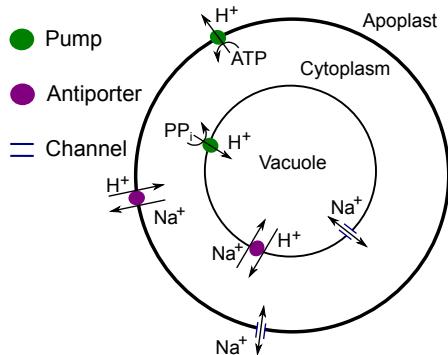
Plant cell biology: Salt stress stages

- Two main stages of salt stress:
 - **Osmotic** stress
 - Disrupts the cell's ability to take up water
 - **Ionic** stress
 - Na^+ in the living part of the cell disrupts important cell processes
- The effects of these stresses can be minimised by:
 - Storing Na^+ in the vacuole (to avoid osmotic stress)
 - Maintaining low levels of Na^+ in the cytoplasm (to avoid ionic stress)



Plant cell biology: Salinity tolerance mechanisms

- Membrane transporters control the movement of ions across cell membranes
- These include
 - H^+ pumps which power Na^+/H^+ antiporters
 - Channels that allow Na^+ to leak back into the cytoplasm

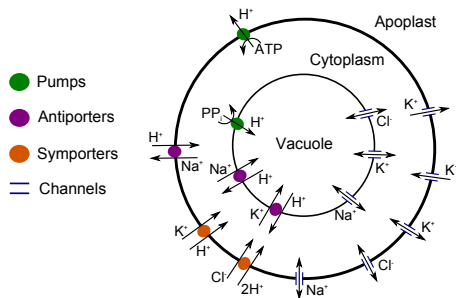


Existing cell models

- There has been extensive modelling of animal cells
 - But: Na^+ is transported by different mechanisms (Na^+/K^+ pump)
- Plant cell models:
 - Often do not include Na^+ (e.g. Hills et al., 2012)
 - Do not include transport across both membranes (Melkikh and Seleznev, 2005, 2012)

Single cell model: Active transport

- Time dependent model of the membrane transport of **water** and four mobile **ions**: Na^+ , K^+ , Cl^- and H^+
- Includes three compartments: Vacuole, Cytoplasm and Apoplast
- Constant conditions in the apoplast
- Simulates membrane transporters operating across **both** the tonoplast and plasma membrane:
 - H^+ pumps** (primary active)
 - Antiporters** (secondary active)
 - Symporters** (secondary active)
 - Channels** (passive)
- The model captures the key processes involved in salt stress response at the cell-level



Foster & Miklavcic (2015) Journal of Theoretical Biology, v385, pp 130-142

Model equations: Ion fluxes

- Conservation equations for mobile ions in the vacuole:

$$\frac{d}{dt} (V_v[H^+]_v) = J_H^{A_v} + J_H^{XNa_v} + J_H^{XK_v}$$

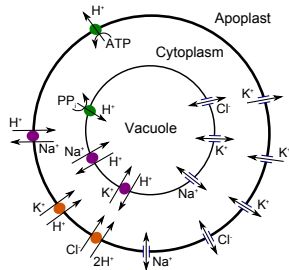
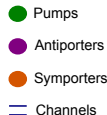
$$\frac{d}{dt} (V_v[Na^+]_v) = J_{Na}^{C_v} + J_{Na}^{XNa_v}$$

$$\frac{d}{dt} (V_v[K^+]_v) = J_K^{C_v} + J_K^{XNa_v}$$

$$\frac{d}{dt} (V_v[Cl^-]_v) = J_{Cl}^{C_v}$$

- Fluxes (J) depend on

- Concentrations in the different cell compartments (\square_a , \square_c , \square_v)
- Electric potential differences across the cell membranes
- Various transport specific parameters (channel permeabilities, antiporter reaction rates, pump rates)



Model equations: Volume and electric potential

- Cell compartment volumes change over time (due to water transport):

$$\frac{dV_c}{dt} = -Q_v - Q_a$$
$$\frac{dV_v}{dt} = Q_v$$

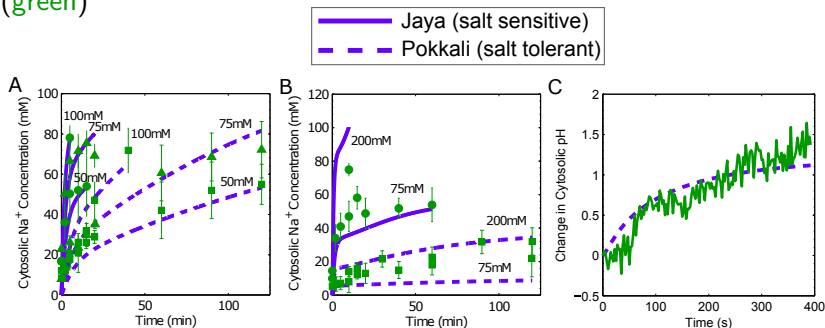
- Water flow rates across membranes (Q's) driven by osmotic pressure differences
- Transmembrane potential, $\Delta\psi$:

$$\frac{d\Delta\psi_{a/v}}{dt} = -\frac{I_{a/v}}{C_m^{a/v}}$$

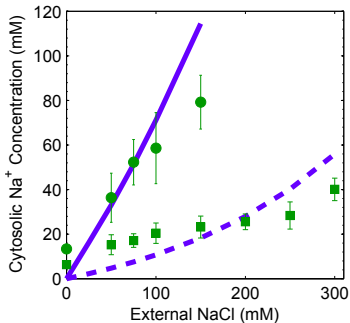
- I = the total current flowing across the membrane (determined by ion fluxes, J 's)
- C_m = membrane capacitance
- Nonlinear system of ordinary differential equations.

Comparison with experiments: Parameter fitting

- Anil et al. (2007) conducted two-photon microscopy experiments to derive cytosolic Na^+ concentrations of individual, cultured rice cells
- Kader et al. (2005) conducted experiments to estimate salt induced changes in cytosolic pH in salt tolerant rice cells
- Model simulations (purple) were performed by matching experimental conditions and fitting parameters to experimental data (green)



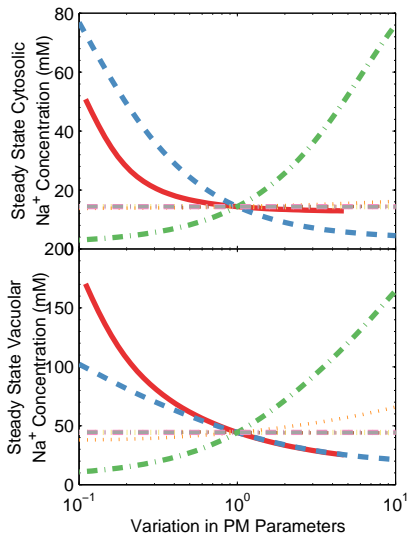
Comparison with experiments



- *No further parameter fitting*
 - Experimental cytosolic Na⁺ concentrations (**symbols**) were measured after 1 hr of exposure to different external Na⁺ concentrations
 - Model simulations (**lines**) were carried out using the parameters found from the previous experimental results
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- The model simulations show that the key differences between the cultivars are:
 - Plasma membrane Na⁺ channel permeability
 - Ability to store Na⁺ in the vacuole

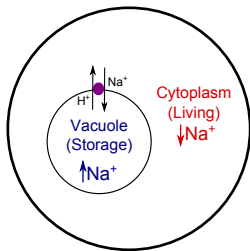
Parameter sensitivity: Plasma membrane transporters

- Effects of varying **plasma membrane** parameters:
 - H^+ pump density
 - - - Na^+/H^+ antiporter strength
 - · - · - Na^+ channel permeability
- The changes in cytosolic Na^+ concentration lead to changes in vacuolar Na^+ concentration



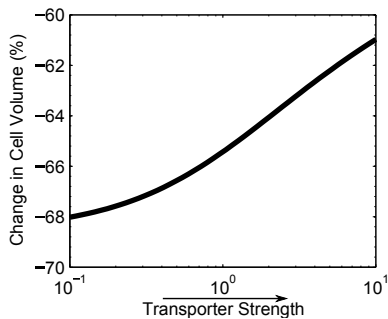
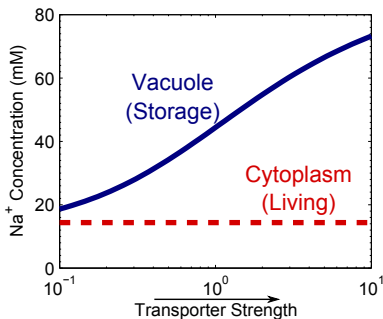
Model applications

- Overexpression of **tonoplast** Na^+/H^+ **antiporter** genes has been shown to increase salinity tolerance in a wide variety of plant species
- The improved salinity tolerance could be due to
 - ① Decreasing the level of Na^+ in the living cytoplasm (reduced ionic stress), and/or
 - ② Increasing storage of Na^+ in the cell vacuole (reduced osmotic stress)



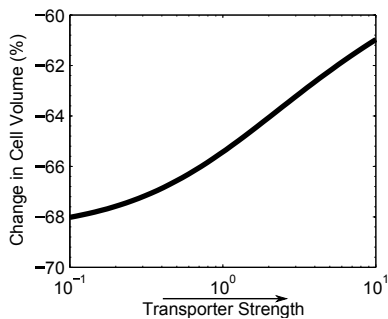
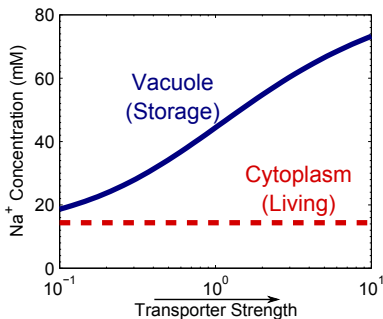
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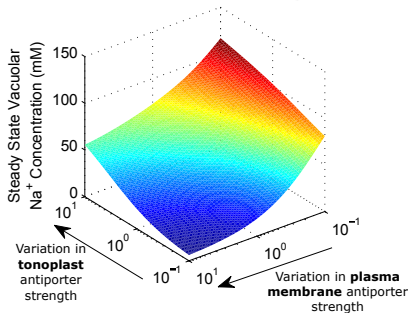
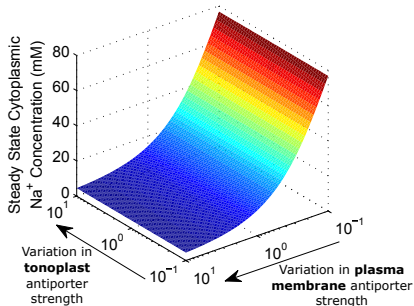
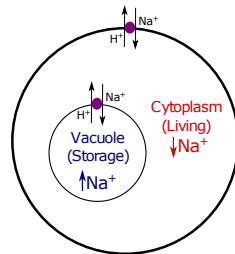
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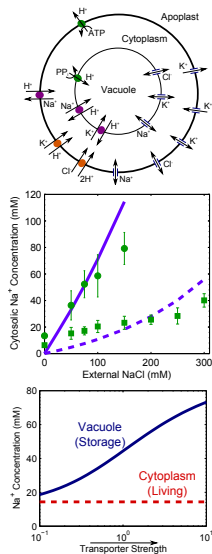
Model applications

- **Both** reduced ionic stress and reduced osmotic stress can be achieved by **simultaneously** overexpressing tonoplast and plasma membrane transporter genes
- To date this has not been achieved experimentally



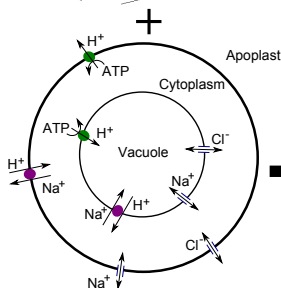
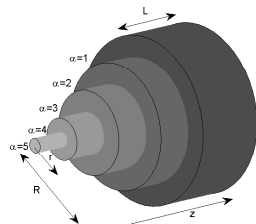
Single cell model: Summary remarks

- Our model captures the key processes responsible for salt stress response at the cell-level
- The model predictions are qualitatively and quantitatively comparable with experimental data
- The model simulations are particularly interesting in light of attempts to genetically alter Na^+ transport processes

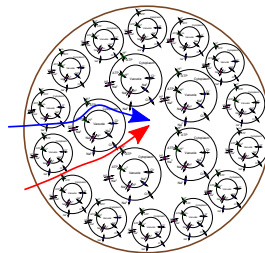


Future work: Multi-scale model

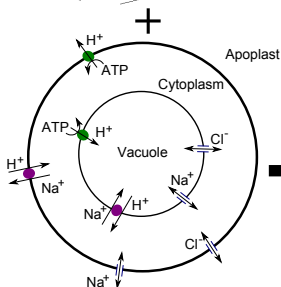
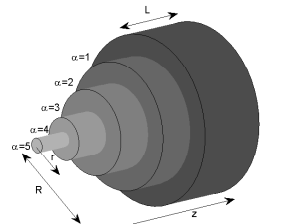
- Developing a multi-scale model of ion and water transport that combines features of:



- Organ-level model
 - Diffusion
 - Convection
- Cell-level model
 - Active extrusion of Na^+
 - Storage of Na^+ in cell vacuoles



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• Thank You!

