Quantitative modelling of the energy cost of Na<sup>+</sup> exclusion, transport and storage in plant roots under salt stress

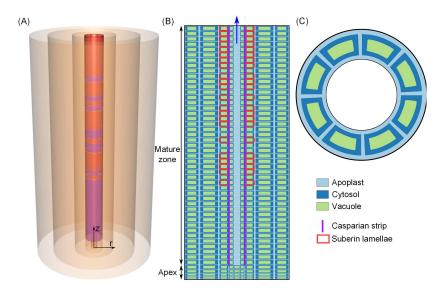
#### Kylie Foster and Stan Miklavcic

Phenomics and Bioinformatics Research Centre, School of Information Technology and Mathematical Sciences.

**University of South Australia** 

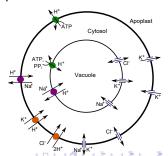


#### Schematics of our model root



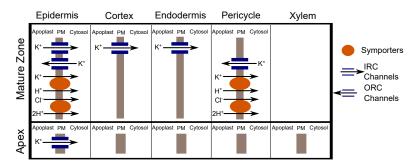
## Driving forces for ion and water transport

- Water transport is driven by hydraulic and osmotic pressure differences.
- Apoplastic and symplastic ion (Na<sup>+</sup>, K<sup>+</sup>, H<sup>+</sup> and Cl<sup>-</sup>) transport is driven by electrochemical diffusion and convection.
- Transmembrane ion transport is driven by relevant ion concentrations and transmembrane potentials:
- Channels permeable to Na<sup>+</sup>, K<sup>+</sup> (VIC, IRC and ORC) and CI<sup>-</sup>;
- K<sup>+</sup>/H<sup>+</sup> symporters and CI<sup>-</sup>/H<sup>+</sup> symporters;
- H<sup>+</sup> pumps;
- Na<sup>+</sup>/H<sup>+</sup> antiporters.



## Spatial distributions of model transport proteins

• Spatial distribution of plasma membrane transport proteins:



- PM VIC channels and H<sup>+</sup> pumps operating in all cells
- Plasma membrane Na<sup>+</sup>/H<sup>+</sup> antiporters? We have simulated a range of spatial distributions.
- Tonoplast membrane transport proteins: No active storage of Na<sup>+</sup> in the apex.



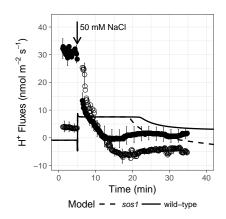
# Energy cost calculations

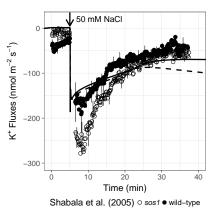
- We assume:
  - 5 ATP synthesized per O<sub>2</sub> consumed in respiration.
  - 1 H<sup>+</sup> transported across PM per ATP hydrolysed.
  - 2 H<sup>+</sup> transporter across TM per ATP hydrolysed.
- There is a single 'composite' pump operating on the model TM, so we can only calculate an upper bound for the ATP cost of storage (assuming all TM H<sup>+</sup> fluxes are through the V-ATPase).
- Energy costs calculated:
  - Cost of active efflux of Na<sup>+</sup> across plasma membranes (based on H<sup>+</sup> flux through PM antiporters).
  - Cost of actively transporting all ions across plasma membranes (based on total H<sup>+</sup> flux through PM pumps).
  - Cost of actively transport of Na<sup>+</sup> across both PM and TM.
  - Cost of actively transporting all ions across PM and TM.



#### Model validation: MIFE fluxes and electric potentials

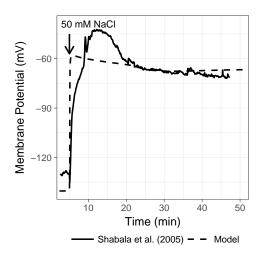
- Model parameters fitted using K<sup>+</sup> and H<sup>+</sup> flux measurements, and electric potential measurements from Shabala et. al. (2005) for wild-type *Arabidopsis* plants and *sos1* mutants.
- Mature zone fluxes:





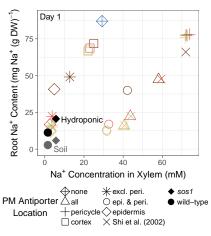
#### Model validation: MIFE fluxes and electric potentials

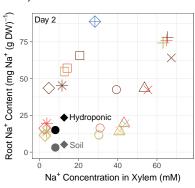
Mature zone epidermal cell electric potential results:



# Model validation: Na<sup>+</sup> content and xylem concentrations

 We also compared our model with Na<sup>+</sup> root contents and Na<sup>+</sup> concentrations in the xylem of wild-type Arabidopsis and sos1 mutants from Shi et al. (2002).





Hiah

Medium

PM Antiporter

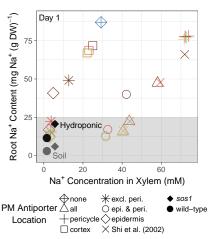
Density

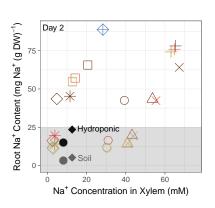


None

#### Model validation: Na<sup>+</sup> content and xylem concentrations

#### • Realistic Na<sup>+</sup> contents:

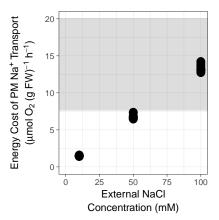


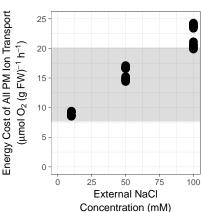




#### Magnitude of energy costs: PM ion transport

 The energy costs of transporting just Na<sup>+</sup> across just the plasma membranes are very high compared to available energy from respiration.

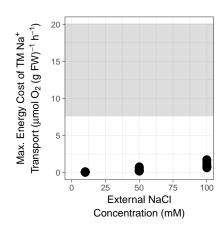


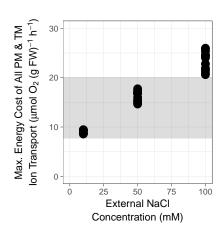




## Magnitude of energy costs: Tonoplast ion transport

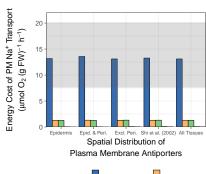
 The energy costs of transporting Na<sup>+</sup> across tonoplast membranes are relatively minor.



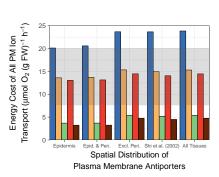


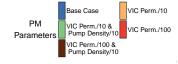
## Possible explanations for high energy costs

- The model energy costs are very high.
- Possible explanations:
  - Overestimated passive plasma membrane Na<sup>+</sup> permeability.



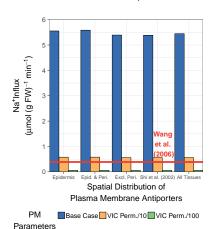


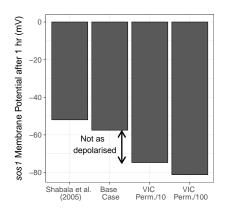




## Possible explanations for high energy costs

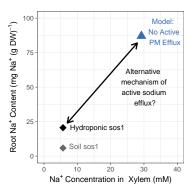
- Overestimated passive plasma membrane Na<sup>+</sup> permeability.
  - Further experimental evidence:





#### Possible explanations for high energy costs

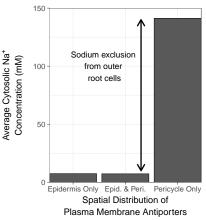
- The model energy costs are very high.
- Possible explanations:
  - There is a transport mechanism missing from the current model of Na<sup>+</sup> transport in roots, i.e. active Na<sup>+</sup> efflux through a transporter other than SOS1.
  - A much more energy efficient mechanism would be required.

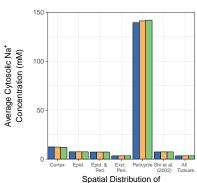


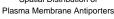


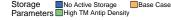
#### Control points for Na<sup>+</sup> transport: Cytosolic Na<sup>+</sup>

- What are the most important control points for maintaining low Na<sup>+</sup> levels in root cell cytosols?
  - Outer root cells.
  - NOT: Inner root cells or storage.









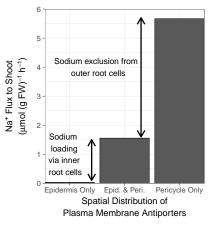


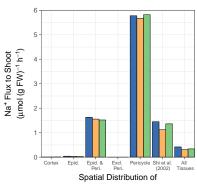
 Our Model
 Validation
 Magnitude of energy costs
 Control points
 Energy efficiency
 Summary

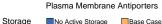
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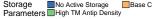
# Control points for Na<sup>+</sup> transport: Na<sup>+</sup> flux

- What are the most important control points for Na<sup>+</sup> accumulation in the shoot?
  - Outer root cells (active efflux reduces Na<sup>+</sup> flux).
  - Inner root cells (active efflux increases Na<sup>+</sup> flux).





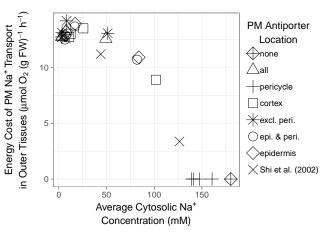






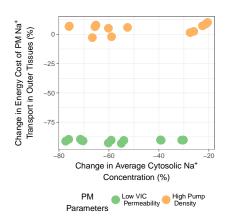
#### Na<sup>+</sup> exclusion from the outer root: Energy costs

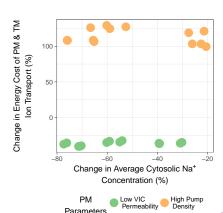
 There are significant energy costs associated with the exclusion of Na<sup>+</sup> from the outer root cells.



## Na<sup>+</sup> exclusion from the outer root: Energy efficiency

- Increasing the plasma membrane H<sup>+</sup> pump density lowers the cytosolic Na<sup>+</sup> concentration but increases the energy costs.
- Reducing the passive Na<sup>+</sup> permeability of the plasma membranes significantly reduces the energy costs.

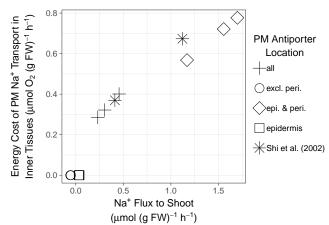






# Minimising net Na<sup>+</sup> flux into the xylem: Energy costs

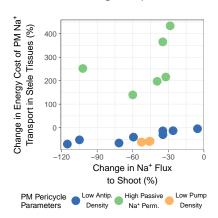
- Lower Na<sup>+</sup> flux requires *less* energy at the pericycle.
- Energy costs for Na<sup>+</sup> transport across pericycle plasma membranes are relatively small.

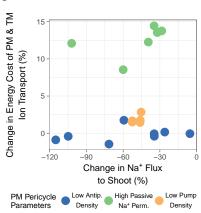




# Minimising net Na<sup>+</sup> flux into the xylem: Energy efficiency

 Reducing active loading of Na<sup>+</sup> is more energy efficient than increasing the passive unloading of Na<sup>+</sup>.

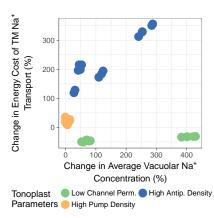


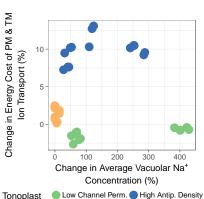


# Storage of Na<sup>+</sup> in vacuoles: Energy efficiency

- Reducing the passive Na<sup>+</sup> permeability of the tonoplast reduces the energy costs.
- Energy costs for Na<sup>+</sup> transport across tonoplast membranes are relatively small.

Parameters

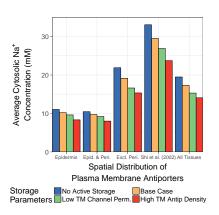




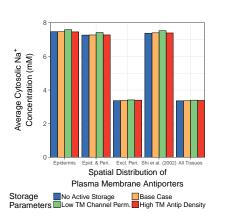
High Pump Density

# Storage of Na<sup>+</sup> in vacuoles: Benefits

- Temporary reduction in cytosolic Na<sup>+</sup> concentrations
- 8 hrs after 100 mM NaCl



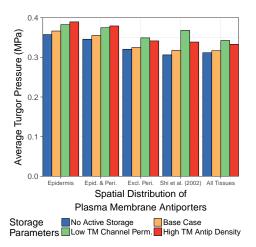
• 15 days after 100 mM NaCl





#### Storage of Na<sup>+</sup> in vacuoles: Benefits

Increase in turgor pressure





# Summary: Magnitude of energy costs

- The energy cost of ion transport under salt stress, based on the current model of Na<sup>+</sup> transport in roots, is unrealistically high.
- Possible explanations for this:
  - Na<sup>+</sup> influx is restricted by relatively low passive plasma membrane permeabilities.
  - There is an alternative transport mechanism effluxing Na<sup>+</sup>.
- Key model assumptions:
  - Ca<sup>2+</sup> transport is not included in the model.
  - Cl<sup>-</sup> is the only mobile anion.
  - H<sup>+</sup> transport across the tonoplast membrane is simulated using one 'composite pump' that represents the combined transport of H<sup>+</sup> via V-ATPase and H<sup>+</sup>-PPase.



# Summary: Energy efficiency

- Exclusion of Na<sup>+</sup> from the outer root tissues:
  - Is very energetically expensive.
  - Can be most efficiently achieved by reducing the passive uptake of Na<sup>+</sup>.
- Na<sup>+</sup> loading/unloading of the xylem:
  - It is more efficient to minimise net Na<sup>+</sup> transport into the xylem by reducing the active loading of Na<sup>+</sup> rather than by increasing the passive unloading of Na<sup>+</sup>.
- Active Na<sup>+</sup> storage in vacuoles:
  - Can be most efficiently achieved by reducing the passive leak of Na<sup>+</sup> across the tonoplast.
  - Benefits?



#### References

#### Experimental comparisons:

- Shabala, L., Cuin, T. A., Newman, I. A., Shabala, S., 2005.
   Salinity-induced ion flux patterns from the excised roots of *Arabidopsis sos* mutants. Planta 222 (6), 1041-1050.
- Shi, H., Quintero, F. J., Pardo, J. M., Zhu, J., 2002. The putative plasma membrane Na<sup>+</sup>/H<sup>+</sup> antiporter SOS1 controls long-distance Na<sup>+</sup> transport in plants. The Plant Cell Online 14 (2), 465-477.
- Wang, B., Davenport, R., Volkov, V., Amtmann, A., 2006.
   Low unidirectional sodium influx into root cells restricts net sodium accumulation in *Thellungiella halophila*, a salt-tolerant relative of *Arabidopsis thaliana*. J. Exp. Bot. 57 (5), 1161-1170.

#### References

- Spatial distribution of transport proteins:
  - Desbrosses, G., Josefsson, C., Rigas, S., Hatzopoulos, P., and Dolan, L. (2003). AKT1 and TRH1 are required during root hair elongation in Arabidopsis. J. Exp. Bot. 54, 781788.
  - Gaymard, F., Pilot, G., Lacombe, B., Bouchez, D., Bruneau, D., Boucherez, J., et al. (1998). Identification and disruption of a plant shaker-like outward channel involved in K<sup>+</sup> release into the xylem sap. Cell 94, 647655.
  - Gierth, M., Mser, P., and Schroeder, J. I. (2005). The potassium transporter AtHAK5 functions in K<sup>+</sup> deprivation-induced high-affinity K<sup>+</sup> uptake and AKT1 K<sup>+</sup> channel contribution to K<sup>+</sup> uptake kinetics in Arabidopsis roots. Plant Physiol. 137, 11051114.
  - Ivashikina, N., Becker, D., Ache, P., Meyerhoff, O., Felle, H. H., and Hedrich, R. (2001). K<sup>+</sup> channel profile and electrical properties of *Arabidopsis* root hairs. FEBS Lett. 508, 463469.
  - Lagarde, D., Basset, M., Lepetit, M., Conejero, G., Gaymard, F., Astruc, S., et al. (1996). Tissue-specific expression of *Arabidopsis* AKT1 gene is consistent with a role in K<sup>+</sup> nutrition. Plant J. 9, 195203.

