The feasibility of targeted test-trace-isolate for the control of B.1.1.7

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Abstract

The SARS-CoV-2 variant B.1.1.7 reportedly exhibits substantially higher transmission than the ancestral strain and may generate a major surge of cases before vaccines become widely available. As B.1.1.7 can be sensitively detected using the Thermo Fisher TaqPath S-gene RT-PCR test, contact tracing and isolation programs appear well-suited to slowing the spread of the new variant, which is still rare in most of the dozens of countries in which it has been identified. However, key determinants of outcomes such as data-sharing, trace success, and isolation compliance vary widely between regions, which may discourage public health agencies from explicitly redirecting existing contact tracers to contain B.1.1.7. Here we apply a branching-process model to estimate the effectiveness of implementing a B.1.1.7-focused testing, contact tracing, and isolation strategy with realistic levels of performance. Our model indicates that bidirectional contact tracing can substantially slow the spread of B.1.1.7 even in regions where a large fraction of the population refuses to cooperate with contact tracers or to abide by quarantine and isolation requests.

The frequency of the B.1.1.7 variant of SARS-CoV-2 has grown rapidly from its initial detection in October 2020 to become the dominant strain in southeastern England by 2021. Studies have estimated the new strain is between 40% and 80% more contagious^{1,2}. The rapid exponential growth of B.1.1.7, which is now found in dozens of countries, risks another and potentially higher wave of COVID-19 cases prior to widespread vaccination.

Due to the B.1.1.7 variant's characteristic Δ69-70 mutation in the spike protein, cases of the new variant can be sensitively detected and distinguished from the ancestral SARS-CoV-2 via a Thermo Fisher TaqPath RT-PCR diagnostic test for COVID-19³. Twenty million such kits are manufactured weekly⁴. As such, existing COVID-19 testing infrastructure can be used to track the transmission of the new variant. Samples testing positive by other kits can be re-screened⁵ without an emergency use authorization.

Test-trace-isolate (TTI) strategies have been widely used to mitigate the spread of SARS-CoV-2⁶. Models by the present authors⁷ and others⁸ have found that incorporating backwards tracing to identify infector individuals could dramatically increase the efficacy of tracing programs. However, testing delays, mistrust, and low compliance in some communities have undermined the confidence of health authorities in the benefits of TTI^{9,10}. Moreover, efficacy sharply decreases when caseloads are high¹¹, as is

true for SARS-CoV-2 – but not yet B.1.1. $7^{12,13}$ – in many regions.

Given the current low prevalence of B.1.1.7 in most jurisdictions and the ability to identify cases of the new variant using existing testing infrastructure, we hypothesised that TTI programs dedicated to controlling B.1.1.7 could substantially reduce the harm inflicted by the new variant prior to widespread vaccination of populations later in 2021. Such programs could be enhanced through incorporation of bidirectional tracing.

However, the effectiveness of TTI strategies varies widely from region to region due to programmatic and population-level differences in variables such as the proportion of cases who share their contact history with contact tracers; the proportion who comply with quarantine and isolation requests; and the overall rate of tracing success. Given this variation, it is unclear whether tracing programs exhibiting realistic levels of performance could feasibly dampen the spread of B.1.1.7.

To evaluate the potential benefits of applying targeted test-trace-isolate to control B.1.1.7, we applied a previously published branching-process model of COVID-19 contact tracing⁷ to estimate the change in the effective reproduction number achievable across a wide range of parameters.

Methods

In our model, each case generates a number of new cases drawn from a negative binomial distribution, according to some pre-specified incubation- and generation-time distributions (Table 1). Cases could be identified and isolated based on symptoms alone or through contact tracing. Cases were assumed to either comply with quarantine and isolation requests, or ignore them completely, according to some fixed probability of compliance; cases that comply with quarantine & isolation were assumed to generate no further cases.

Successful tracing depended on the identified case sharing their contact history with tracers, and on the contact in question taking place within the time window (measured in days pre-symptom onset for symptomatic cases, and days pre-identification for asymptomatic cases). Environmental transmission was assumed to be untraceable. Symptomatic cases required a positive test before initiating contact tracing, as in the EU¹⁴ and most US jurisdictions.

Each outbreak was initialized with 20 index cases to minimize stochastic extinction and designated as "controlled" if it reached extinction (zero new cases) before reaching 10,000 cumulative cases. Effective reproduction numbers ($R_{\rm eff}$) were computed as the mean number of child cases produced per case.

Results

To investigate the potential for TTI to mitigate the spread of B.1.1.7, we investigated the effective reproduction number achieved across a range of data-sharing and trace-success rates (Figure 1). To account for uncertainty in the transmissibility of B.1.1.7, we explored outcomes for reproduction numbers between 1.2 and 2.0; these values assume that non-tracing interventions are already in place.

In the absence of contact tracing, identification and isolation of symptomatic cases alone reduced $R_{\rm eff}$ by 0.2 to 0.3 even when quarantine and isolation compliance was low (Figure 1, top rows). When identification and isolation left $R_{\rm eff}$ substantially greater than 1 (i.e., when base R \geq 1.4), moderate levels of tracing could have substantial effects.

When contacts were traced up to 2 days prior to symptom onset, roughly 60-70% data sharing and trace success was required to achieve an $R_{\rm eff}$ reduction of at least 0.1, relative to isolation alone. If the window was extended to 6 days pre-onset to enable more effective bidirectional tracing, roughly 45-55% data sharing and trace success was sufficient. Higher levels of data sharing and trace success could achieve substantially larger reductions: in many scenarios, 85% data sharing and trace success reduced $R_{\rm eff}$ by >0.2 in the 2-day case and >0.35 in the 6-day case.

Due to the exponential growth of uncontrolled epidemics, small reductions in $R_{\rm eff}$ can have a large impact on the total number of downstream cases arising from a given index case over a given timespan. For example, under a simple geometric series approach, reducing $R_{\rm eff}$ by 0.1 from a starting value between 1.2 and 2.0 reduces the total number of child cases after 10 generations by 37-43%; an $R_{\rm eff}$ reduction of 0.2 results in a reduction in child cases of 61-66%. Given an average generation time of 6 days, 10 generations equates to roughly 2 months – enough time, given sufficient delay in the spread of the new variant, to vaccinate a substantial fraction of the population.

Discussion

Our results suggest that regions with even moderately functional contact tracing programs focused on B.1.1.7 could substantially slow the spread of the variant. Given a 2-day window for bidirectionally tracing contacts pre-symptom onset, our model predicts that a program with 70% trace success, 70% data sharing, and 70% compliance with isolation could achieve an $R_{\rm eff}$ reduction of at least 0.1 relative to the no-tracing case. Given a 6-day window for efficient bidirectional tracing, regions with just 50% data-sharing, trace success, and isolation compliance could achieve a reduction of 0.1.

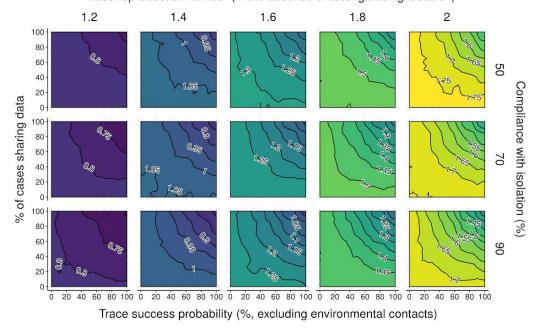
Under simple assumptions, such a reduction would reduce the number of child cases produced in two months by roughly 40%, buying time for vaccination to immunise many more people. More effective tracing programs can achieve larger reductions. Higher rates of compliance and data sharing might be achieved through home visits by contact tracers¹⁵; exoneration for anything discovered in the course of B.1.1.7 contact tracing¹⁰; and financial and other support of people in quarantine and isolation¹⁶. In principle, concentrating vaccination in communities with clusters of uncontrollable B.1.1.7 transmission could further impair viral spread and increase the sustainability of testing, tracing, and isolation for the control of COVID-19.

These results assume a high availability of suitable tests for B.1.1.7, and a rapid and consistent testing turnaround. They also take no account of any medical, demographic, geospatial or behavioural variation between cases which could influence the spread of the new variant.

Our results suggest that TTI programs could help slow the spread of B.1.1.7 in regions where it is currently rare, providing vital time for vaccination to end the COVID-19 pandemic. As the efficacy of TTI programs is limited at high caseloads¹¹, these findings indicate that tracing programs should immediately prioritise controlling B.1.1.7 over less transmissible – but currently more widespread – strains.

a 2-day manual tracing window

Base reproduction number (in the absence of testing/tracing/isolation)



b 6-day manual tracing window

Base reproduction number (in the absence of testing/tracing/isolation)

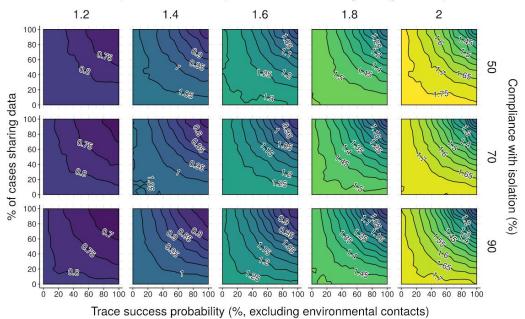


Figure 1. Evaluating the efficacy of bidirectional contact tracing for controlling B.1.1.7. Neighbour-averaged contour plots, showing R_{eff} achieved by bidirectional manual contact tracing with a tracing window of (a) 2 or (b) 6 days pre-symptom onset, under different combinations of trace success probability (x-axis), rate of data sharing with manual contact tracers (y-axis), rate of compliance with isolation and quarantine (row) and base reproduction number (columns). Other disease parameters are specified in Table 1. Isolation of symptomatic cases is sufficient to reduce R even when no traces succeed and/or no cases share their data with contact tracers. "Trace success probability" refers to trace attempts that are not

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Table 1: Parameters of the branching-process model.

Parameter	Value	Sources and Notes
% asymptomatic carriers	40%	17–21
Relative infectiousness of asymptomatic carriers	45%	Informed by viral loads and tracing results described in 17,21-25
% environmental transmission	5%	26,27
Proportion of pre-symptomatic transmission	38%	Informed by ^{21,22,24,25,28–33}
Generation time skew parameter (α)	0.397	Corresponds to pre-symptomatic transmission rate specified above.
% of symptomatic cases identified without tracing	50%	34
% of cases who comply with isolation	50%, 70%, 90%	Assumed
Test sensitivity	70%	35.36
$R_{ m base}$ (before test/trace/isolate)	1.0 to 2.0	Assumes a pre-B.1.1.7 <i>R</i> of ~1.0 ^{1,2} .
Overdispersion	0.11	37
Number of initial cases	20	Assumed
Incubation period	6.0 ± 2.1 days (lognormal distribution)	1,38,39
Delay from onset to isolation	3.8 ± 2.4 days (Weibull distribution)	40
Delay for testing	1 ± 0.3 days (gamma distribution)	Assumed
Delay for manual tracing	1.5 ± 4.8 days (lognormal distribution); median 0.5 days	Previous reports suggest most contacts can be traced within one day, but some take much longer 41

Supplementary Methods

Structure of the model - Infection dynamics

A new case is infected at some **exposure time**, equal to zero if the case is an index case and otherwise drawn from the **generation time distribution** of its parent case (see below). If not asymptomatic, the case develops symptoms at some **onset time** drawn from an **incubation time distribution**. Asymptomatic cases do not develop symptoms, but are still assigned an onset time for the purpose of determining their generation-time distribution (see below).

The number of child cases infected by the case is drawn from a negative binomial distribution, with mean equal to the appropriate reproduction number (see below) and heterogeneity determined by the overdispersion parameter k. The exposure times of these child cases are drawn from a skewed-normal **generation time distribution** centered on the symptom onset of their parent ⁴⁰, with an SD parameter (ω) of 2 and a skew parameter (α) chosen to give a pre-specified probability of pre-symptomatic transmission (for a symptomatic parent) (Table S1 & Fig. S38). The generation time distribution for an asymptomatic parent is centered on its "effective" onset time (see above). The shape of the generation-time distribution is the same for all cases.

The expected number of children produced by a case depends on its symptomatic status, and is determined by the overall R_0 value, the proportion of asymptomatic carriers p_{asym} , and the relative infectiousness x_{asym} of asymptomatic carriers (expressed as a fraction of R_0). Given a reproduction number for asymptomatics of $R_{asym} = R_0 \cdot x_{asym}$, the reproduction number of symptomatic cases that produces the desired overall R_0 is given by $R_{sym} = R_{asym} \cdot \frac{p_{asym}}{1-p_{asym}}$.

Structure of the model - Infection control

Once symptoms develop, a case is **identified** by public health authorities with probability p_{isol} , with the delay from onset to identification drawn from a **delay distribution**. Identified cases are instructed to isolate, and each case complies with that order with probability p_{comply} . Cases that comply with isolation generate no further child cases after their time of identification; cases that do not are unaffected. Asymptomatic cases cannot be identified from symptoms, but may be identified via contact tracing from other cases (see below); once identified, they are instructed to isolate as above. Tracing can also cause symptomatic cases to be isolated earlier than they would be from symptoms alone.

An identified case is **tested**, which takes time drawn from a test time distribution and returns a positive result with probability equal to the sensitivity of the test (since the model does not consider uninfected individuals, the specificity of the test is also not considered). For asymptomatic cases, or symptomatic cases identified prior to symptom onset, a positive test result is required to initiate contact tracing; symptomatic cases that have already developed symptoms can either be traced immediately upon identification, or require a positive test result prior to tracing, depending on model settings.

Whether before or after a test result is obtained, the contacts of an identified case can also be **traced**. Tracing can only proceed outward from a case if they share their contact history with a manual contact tracer (see below). Tracing can identify the children of the traced case (forward tracing) or its parent (reverse/backward tracing). The speed and success probability of tracing depends on several factors:

- If the contact between the trace originator and the trace occurred environmentally (determined with probability p_{env}), tracing cannot take place.
- If transmission was not environmental, the contact can be traced manually if:
 - The trace originator shares their contact history with a manual contact tracer (determined independently for each individual case with probability $p_{share\ manual}$);
 - The time between contact (as above) and the identification time or symptom onset of the trace initiator (whichever came first) is less than the **contact-tracing window** of the manual tracing system;
 - The trace is successfully traced by the contact tracer (determined independently for each individual case with probability $p_{trace\ manual}$).
- Otherwise, then the trace fails and the tracee is not traced.

Cases that are successfully traced are identified at a time equal to the **trace initiation time** of the trace originator plus a delay time drawn from the appropriate **trace delay distribution**. Identified contacts are quarantined, with an effect identical to isolation and governed by the same compliance variable (i.e. a case either complies with both quarantine and isolation, or neither). Quarantined cases identified through tracing can then be isolated, tested, and traced as described above. If a case is isolated through tracing earlier than they would have been otherwise, child cases whose exposure time would be later than their parent's new isolation time are eliminated, as are their descendents.

Run initiation and termination

A simulation of an outbreak under the branching-process model is initialised with a given number of index cases (by default 20, in order to reduce the probability of stochastic elimination) and proceeds generation by generation until either no further child cases are generated (extinction) or the run exceeds one of:

- 1. A **cumulative case limit** of 10,000 cases, reached if the total number of cases ever exceeded that number, or
- 2. A **time limit** of 52 weeks, reached if the latest exposure time across all cases ever exceeded that number.

In practice, virtually all runs either went extinct or reached the cumulative case limit; across all scenarios tested for all datasets used in Figures 2-4, the overall percentage of runs that terminated as a result of exceeding the time limit was less than 0.02%, and the highest percentage observed for any single scenario was 1.3%. The cumulative case limit, meanwhile, was selected to minimise the chance of a run that would otherwise go extinct being terminated prematurely while preserving computational tractability; in

test runs with a cumulative case limit of 100,000 cases, fewer than 2% of extinct runs in any scenario had a cumulative case count of over 10,000.

A terminated run was deemed "controlled" if it reached extinction, and uncontrolled otherwise. The control rate for a scenario was computed as the proportion of runs for that scenario that were controlled. 95% credible intervals on the control rate were computed by beta-binomial conjugacy under a Beta(1, 1) uniform prior, as the 2.5th and 97.5th percentiles of the beta distribution Beta(1+k, 1+n-k), where n is the total number of runs for that scenario and k is the number of controlled runs. Effective reproduction numbers were computed as the mean number of child cases produced across all cases in a run, averaged across all runs in the scenario. For main figures, 1000 runs were performed per scenario; for figures, either 500 or 1000 runs were performed, as specified in the figure captions.