



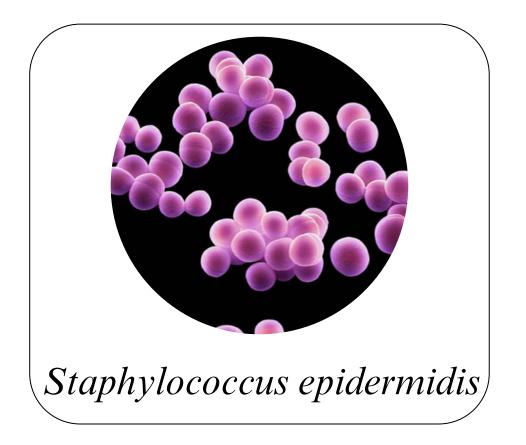
Exploring the Mechanisms behind S. epidermidis Resistance to Vancomycin using Bioinformatics Analysis

Laís Carvalho, Ângela França, Miguel Rocha

Introduction



Coagulase-negative staphylococci (CoNS), particularly *Staphylococcus epidermidis*, are significant in healthcare due to their ability to form biofilms and colonize medical devices causing infections.



This issue is exacerbated by its multidrug resistance, including resistance to vancomycin.

Premature infants, immunosuppressed patients, and those requiring long-term medical devices are especially at risk.

Aim of the work



The aim of this project was to identify the main pathways enriched in the metabolism of *S. epidermidis* with vancomycin-induced resistance.

Materials



1 Data

Gene expression data in excel spreadsheet (xlsx)

2 ShinyGO

A web-based graphical tool for enriching gene sets

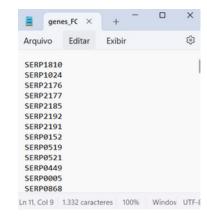
3 KEGG

Bioinformatics database that integrates genomic, chemical and functional information, the information can be accessed by python packages and modules, such as **Bio.KEGG** and **bioservices**.

Methods



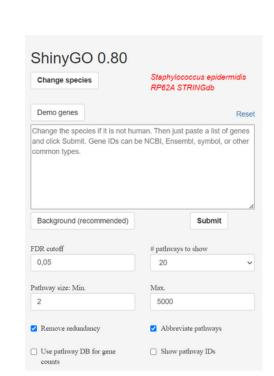


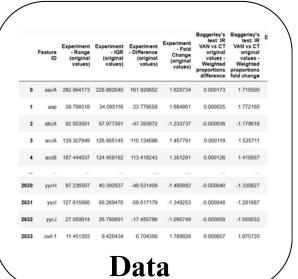


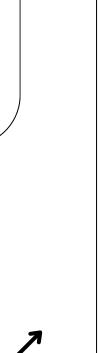
Applying the

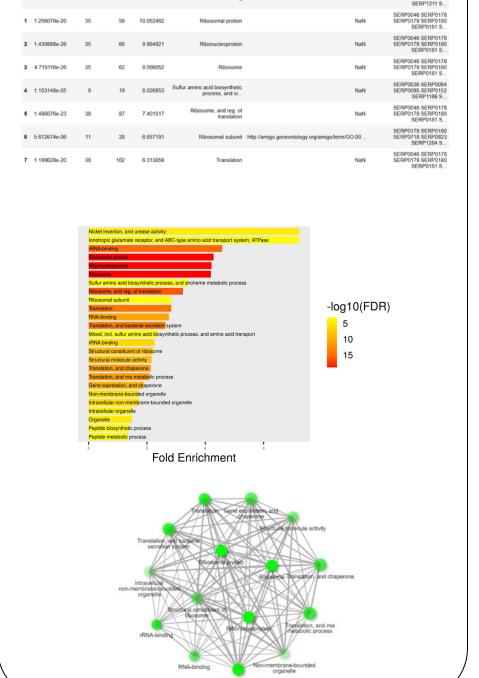
gene list in

ShinyGO

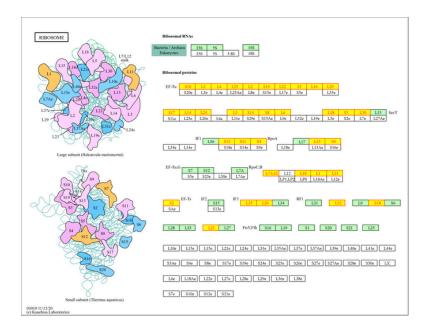




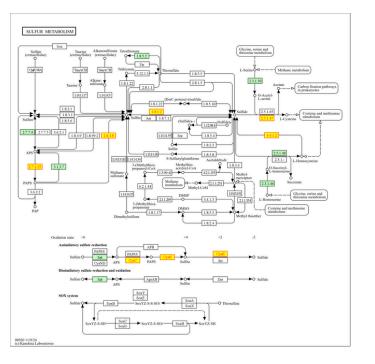




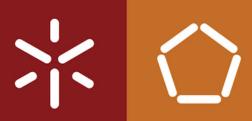
Results



Get the metabolic pathways



Data processing



Feature ID	Experiment - Range (original values)	Experiment - IQR (original values)	Experiment - Difference (original values)	Experiment - Fold Change (original values)	Baggerley's test: IR VAN vs CT original values - Weighted proportions difference	Baggerley's test: IR VAN vs CT original values - Weighted proportions fold change	Baggerley's test: IR VAN vs CT original values - Test statistic	Baggerley's test: IR VAN vs CT original values - P- value
aacA	292.994173	228.882040	161.929852	1.628734	0.000173	1.715500	1.487221	0.136957
aap	39.799318	34.085116	23.779558	1.684861	0.000025	1.772160	1.648674	0.099215
abcA	92.503501	57.977391	-47.393972	-1.233737	-0.000036	-1.178618	-1.005391	0.314709
accA	139.307849	126.905145	110.134086	1.457791	0.000119	1.525711	3.404836	0.000662
accB	187.444037	124.459192	113.418243	1.351291	0.000126	1.415937	1.959460	0.050059
•••	•••							
уусН	97.226507	40.392937	-49.531409	-1.400982	-0.000040	-1.330827	-1.087926	0.276628
yycl	127.615565	58.269470	-59.517179	-1.349253	-0.000048	-1.281687	-0.906014	0.364929
yycJ	27.050614	26.780691	-17.455786	-1.095749	-0.000009	-1.050033	-0.674502	0.499992
zwf-1	11.451203	9.420434	6.704356	1.788828	0.000007	1.875720	1.591939	0.111399
zwf-2	232.208325	230.292300	-147.181758	-1.402003	-0.000123	-1.342866	-1.575461	0.115150

Data

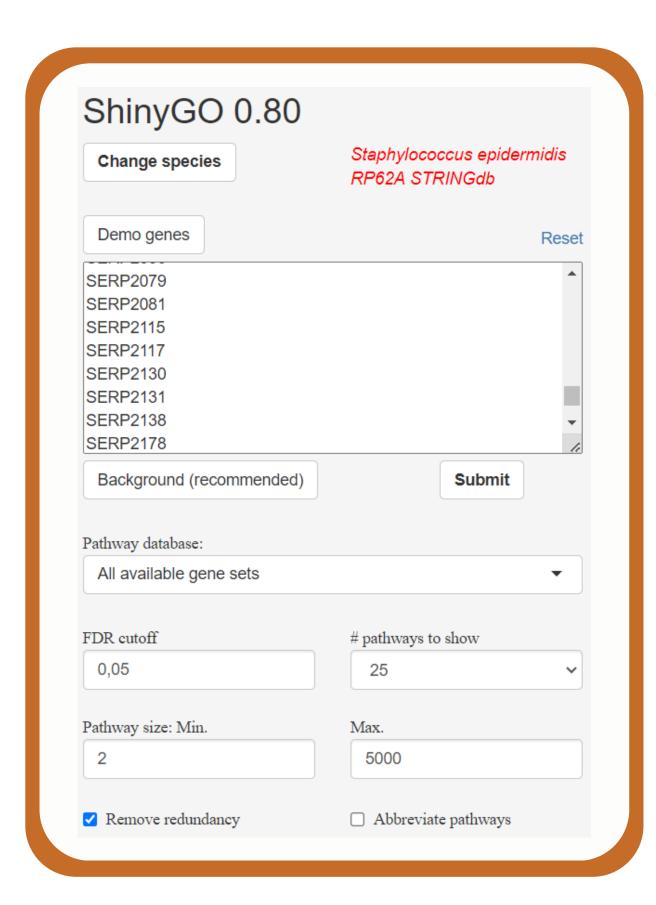
The data was filtered to keep only the genes with FDR p-values < 0.05 and a Fold Change > 2.

The list of genes generated to be applied in ShinyGO had to have the names of the genes changed to match the identifiers used in the STRING.

All genes (2 genes) corresponding to tRNA or pseudogenes were eliminated.

Applying the gene list in ShinyGO





The input to this tool is the list of genes of interest, and it is also necessary to select the species of the study organisms and the number of pathways you want to observe.

The other parameters have been left by default (FDR cutoff, maximum and minimum pathway size and search databases).

Get the metabolic pathways





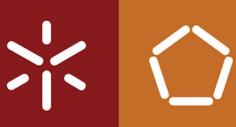
Obtain the graphic representation of the enriched pathways

The ShinyGO tool allows graphical visualization of the enriched pathways. However, limitations such as the difficulty in integrating data from different databases, the lack of annotations on the organism studied or the outdated nature of the online tool can interfere with the results and make it impossible to visualize the pathways graphically.

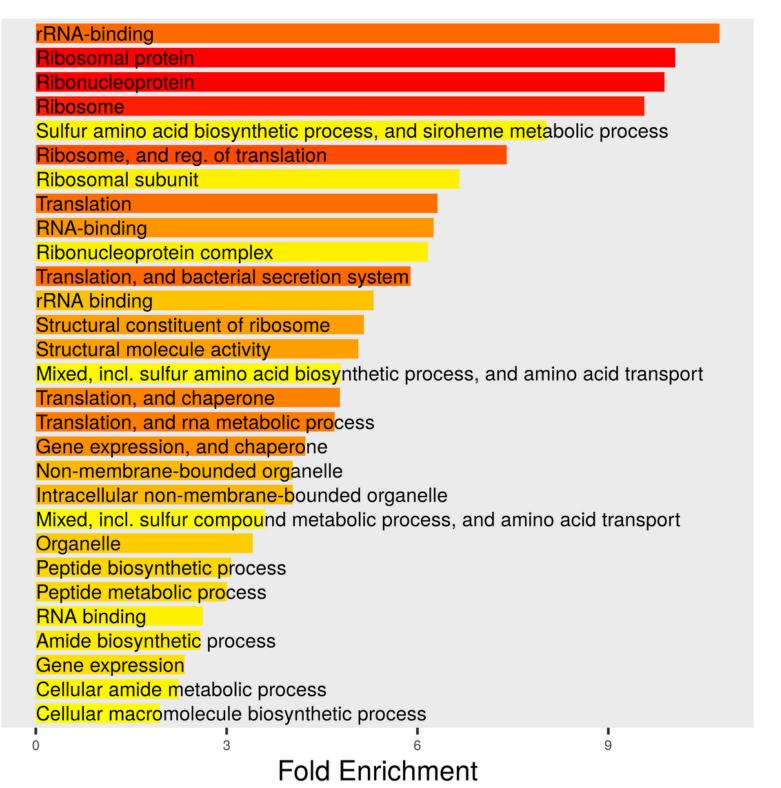
To obtain the graphs of the enriched paths, the following functions were developed:

```
In [1]: # Useful python packages
        import pandas as pd
        from Bio.KEGG import REST
        import re
        from bioservices import KEGG
In [2]: def gene_name_format(gene_list, term):
             """Function that receives a list of genes and adds the specified term (usually the identification term of the study organism
             gene list form = []
             for gene in gene list:
                gene list form.append(term + gene)
            return gene list form
        def get pathways shinyGO(file path):
            """It uploads the table (result) obtained to ShinyGO and formulates a dictionary that stores the name of the enriched pathway
            df ShinyGO = pd.read csv(file path)
            pathway dict = {}
            for index, row in df ShinyGO.iterrows():
                pathway = row['Pathway']
                gene = row['Genes']
                if pathway not in pathway dict:
                    pathway dict[pathway] = []
```

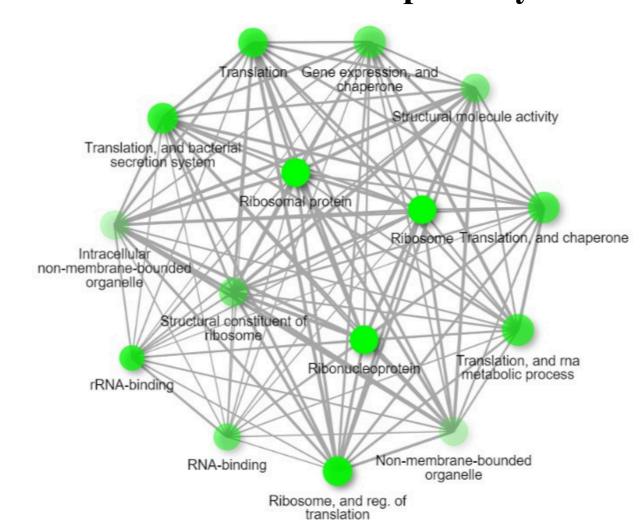
Results ShinyGO



Barplot of enriched metabolic pathways



Network of enriched pathways



-log10(FDR)

5

10

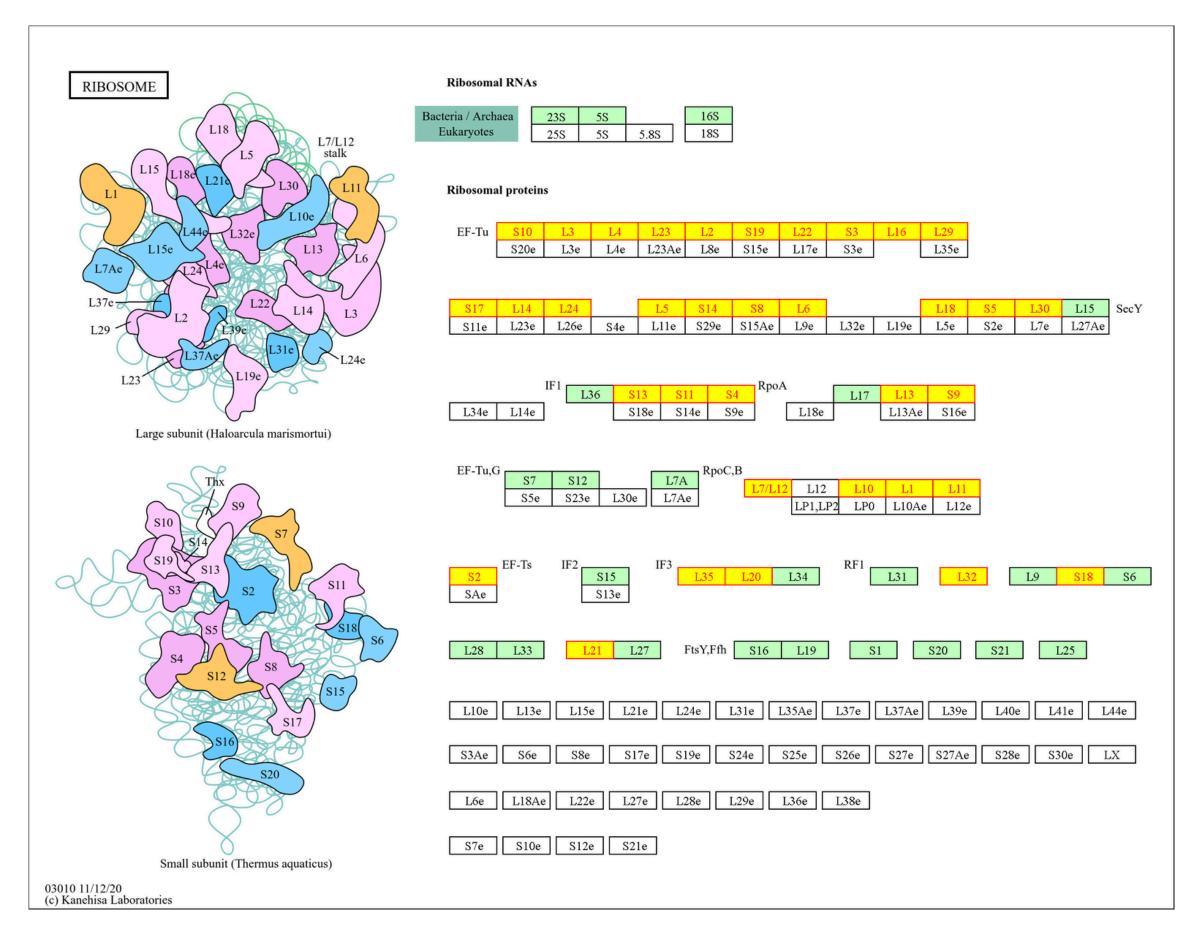
15

20

25

Results



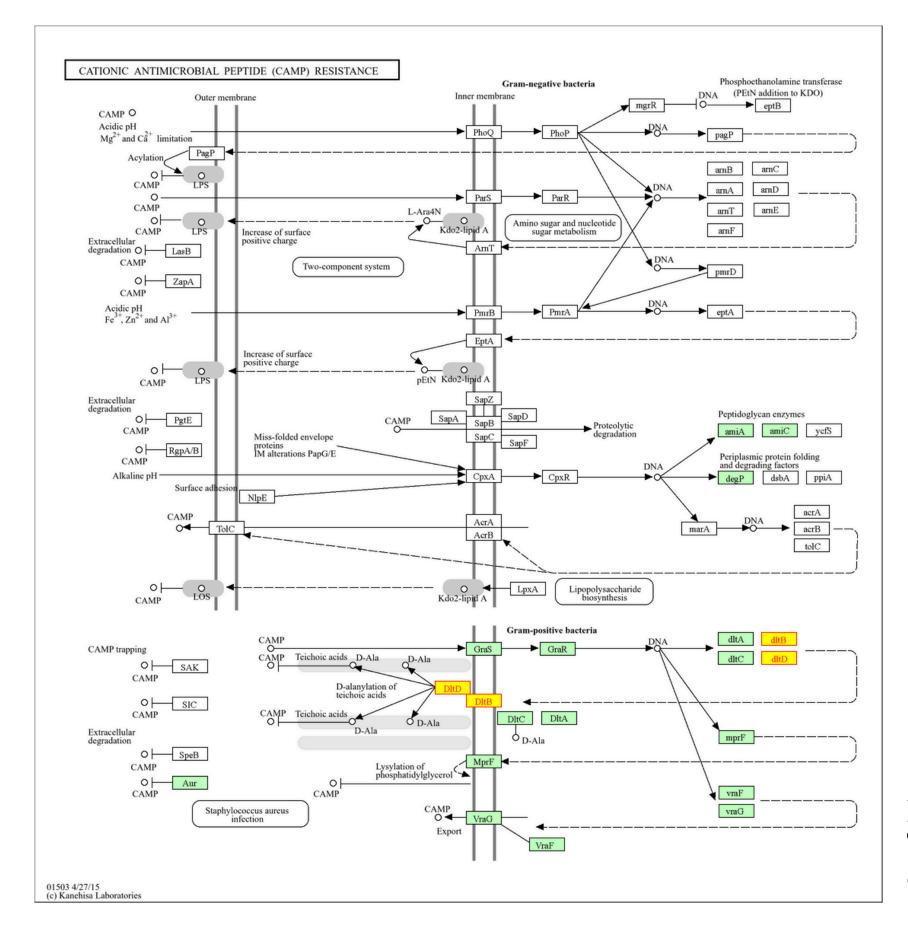


Ribosome

The main enriched metabolic pathway. The genes highlighted in yellow are those present in the set of genes studied (fold change > 2).

Results





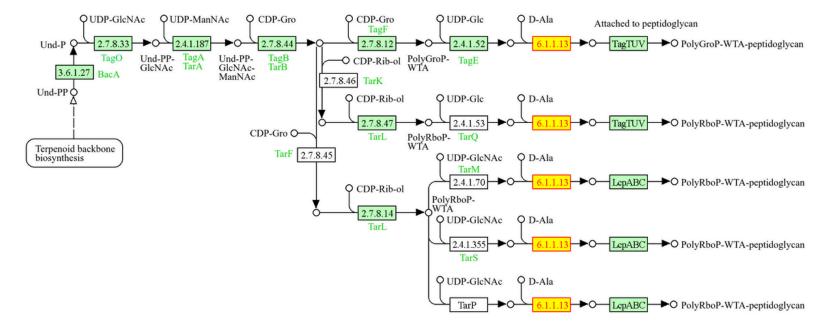
This pathway is related to mechanisms to overcome the antimicrobial action of cationic proteins of the host's innate immune system, and may include modification of the bacteria's surface charge what can influence the interacttion of vancomycin with the cell wall and, thus, vancomycin efficacy.

Cationic antimicrobial peptide (CAMP) resistance

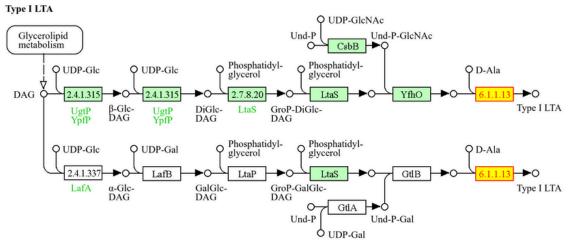
Image of the metabolic pathway obtained from KEGG. The genes highlighted in yellow are those present in the set of genes studied (fold change > 2).

TEICHOIC ACID BIOSYNTHESIS

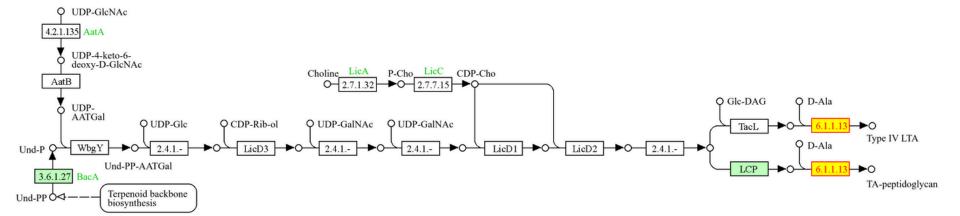
Wall teichoic acid (WTA)



Lipoteichoic acid (LTA)



Type IV LTA and TA-peptidoglycan



Results

The synthesis of teichoic acid is an essential process in the formation of the cell wall in *S. epidermidis* and other Gram-positive bacteria.

Teichoic acid biosynthesis

Image of the metabolic pathway obtained from KEGG. The genes highlighted in yellow are those present in the set of genes studied (fold change > 2).

Final considerations and next steps



Final considerations

Highly expressed genes are present in pathways that may explain the resistance of *S. epidermidis* to vancomycin;

Directing laboratory research to resistance mechanisms.

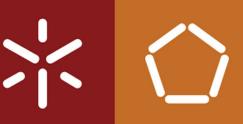
Next steps

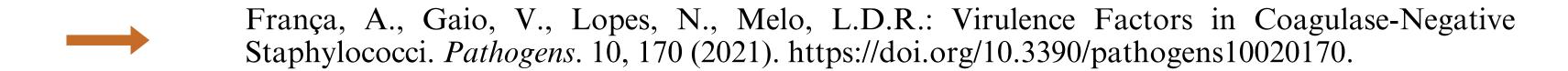
Further explore the additional pathways acquired;

Rerun the analysis for another strain;

Rerun the analysis for underexpressed genes.

References





Rogers, K.L., Fey, P.D., Rupp, M.E.: Coagulase-Negative Staphylococcal Infections. *Infect. Dis. Clin. North Am.* 23, 73–98 (2009). https://doi.org/10.1016/j.idc.2008.10.001.

Kanehisa, M., Goto, S., Kawashima, S., Okuno, Y., Hattori, M.: The KEGG resource for deciphering the genome. *Nucleic Acids Res.* 32, D277–D280 (2004). https://doi.org/10.1093/nar/gkh063.

Ge, S.X., Jung, D., Yao, R.: ShinyGO: a graphical gene-set enrichment tool for animals and plants. *Bioinformatics*. 36, 2628–2629 (2020). https://doi.org/10.1093/bioinformatics/btz931.

Loureiro, Rui João, Roque, Fátima, Rodrigues, António Teixeira, Herdeiro, Maria Teresa, & Ramalheira, Elmano. (2016). O uso de antibióticos e as resistências bacterianas: breves notas sobre a sua evolução. *Revista Portuguesa de Saúde Pública*, 34(1), 77-84. https://doi.org/10.1016/j.rpsp.2015.11.003



Exploring the Mechanisms behind S. epidermidis Resistance to Vancomycin using Bioinformatics Analysis

Laís Carvalho, Ângela França, Miguel Rocha