

Perform Enrichment Analysis

Loading required libraries

```
library(clusterProfiler)
library(org.Dm.eg.db)
library(pathview)
library(biomaRt)
library(ReactomePA)
library(ggplot2)
library(RColorBrewer)
library(ggsci)
```

Read significant genes into csv file

```
sig_genes <- function(file_path){
  genes <- read.csv(file_path, row.names = 1, stringsAsFactors = FALSE)
  final_genes <- na.omit(genes)
  return(final_genes)
}
significant_genes <- sig_genes("../Enrichment analysis in R/Data/Significant genes.csv")
head(significant_genes)
```

```
##           baseMean log2FoldChange      lfcSE      stat      pvalue
## FBgn0039155   730.5958         4.619014 0.1687068   27.37895 1.294976e-76
## FBgn0003360  4343.0354         3.179672 0.1435262   22.15395 6.158877e-32
## FBgn0039827   261.9162         4.162516 0.2325888   17.89646 1.212750e-30
## FBgn0025111  1501.4105        -2.899864 0.1269205  -22.84788 1.377576e-28
## FBgn0034736   225.8764         3.511439 0.2146721   16.35722 3.632828e-21
## FBgn0035085   638.2326         2.560412 0.1372952   18.64896 5.654840e-15
##
##                padj
## FBgn0039155 1.600332e-72
## FBgn0003360 3.805570e-28
## FBgn0039827 4.995723e-27
## FBgn0025111 4.256022e-25
## FBgn0034736 8.978897e-18
## FBgn0035085 1.164708e-11
```

Convert FlyBase IDs (FBgn) to ENTREZ IDs

```
entrez_ids <- function(){
  # copy the rownames of significant_genes and store it in flybase_ids
  flybase_ids <- rownames(significant_genes)
  # convert FLYBASE ids into ENTREZID
  entrez_ids <- mapIds(org.Dm.eg.db,
```

```

        keys = flybase_ids,
        column = "ENTREZID",
        keytype = "FLYBASE",
        multiVals = "first")
# create column named ENTREZID in significant_genes that contain ENTREZID
# of sig genes
significant_genes$ENTREZID <- entrez_ids
# remove ENTREZID that contain NA
sign_genes <- significant_genes[!is.na(significant_genes$ENTREZID), ]
return(sign_genes)
}

sign_genes <- entrez_ids()

```

'select()' returned 1:1 mapping between keys and columns

```
head(sign_genes)
```

```

##           baseMean log2FoldChange      lfcSE      stat      pvalue
## FBgn0039155   730.5958         4.619014 0.1687068  27.37895 1.294976e-76
## FBgn0003360  4343.0354         3.179672 0.1435262  22.15395 6.158877e-32
## FBgn0039827   261.9162         4.162516 0.2325888  17.89646 1.212750e-30
## FBgn0025111  1501.4105        -2.899864 0.1269205 -22.84788 1.377576e-28
## FBgn0034736   225.8764         3.511439 0.2146721  16.35722 3.632828e-21
## FBgn0035085   638.2326         2.560412 0.1372952  18.64896 5.654840e-15
##           padj ENTREZID
## FBgn0039155 1.600332e-72   42865
## FBgn0003360 3.805570e-28   32007
## FBgn0039827 4.995723e-27   43689
## FBgn0025111 4.256022e-25   32008
## FBgn0034736 8.978897e-18   37572
## FBgn0035085 1.164708e-11   37991

```

Up-regulated significant genes

```

up_sig_genes <- function(){
  upregulated_genes <- sign_genes[ sign_genes$padj<0.01 & sign_genes$log2FoldChange>0 , ]
  return(upregulated_genes)
}

upregulated_sig_genes <- up_sig_genes()
upregulated_sig_genes

```

```

##           baseMean log2FoldChange      lfcSE      stat      pvalue
## FBgn0039155   730.59581         4.619014 0.16870676  27.378948 1.294976e-76
## FBgn0003360  4343.03540         3.179672 0.14352623  22.153947 6.158877e-32
## FBgn0039827   261.91624         4.162516 0.23258876  17.896463 1.212750e-30
## FBgn0034736   225.87636         3.511439 0.21467209  16.357221 3.632828e-21
## FBgn0035085   638.23261         2.560412 0.13729520  18.648955 5.654840e-15

```

```
## FBgn0029167 3706.11653      2.197000 0.09698887 22.652087 3.326229e-13
## FBgn0085359   68.61004      4.918127 0.49598137  9.915951 2.757558e-12
## FBgn0024288   58.85110      4.585836 0.46501165  9.861766 1.611111e-11
## FBgn0029896  489.89237      2.445024 0.15194279 16.091743 2.492025e-10
## FBgn0038832  301.92236      2.541149 0.20251626 12.547876 1.365942e-07
##                padj ENTREZID
## FBgn0039155 1.600332e-72    42865
## FBgn0003360 3.805570e-28    32007
## FBgn0039827 4.995723e-27    43689
## FBgn0034736 8.978897e-18    37572
## FBgn0035085 1.164708e-11    37991
## FBgn0029167 5.138192e-10    39529
## FBgn0085359 3.786434e-09    2768869
## FBgn0024288 1.991011e-08    45039
## FBgn0029896 2.368957e-07    31612
## FBgn0038832 1.125354e-04    42468
```

ENTREZIDs of up-regulated significant genes

```
up_entrez_ids <- upregulated_sig_genes$ENTREZID
up_entrez_ids
```

```
## [1] "42865" "32007" "43689" "37572" "37991" "39529" "2768869"
## [8] "45039" "31612" "42468"
```

Down-regulated significant genes

```
down_sig_genes <- function(){
  downregulated_genes <- sign_genes[ sign_genes$padj<0.01 & sign_genes$log2FoldChange<0 , ]
  return(downregulated_genes)
}
downregulated_sig_genes <- down_sig_genes()
downregulated_sig_genes
```

```
##                baseMean log2FoldChange    lfcSE      stat      pvalue
## FBgn0025111 1501.41051      -2.899864 0.1269205 -22.847884 1.377576e-28
## FBgn0000071  342.23841      -2.679584 0.1826118 -14.673665 5.251900e-11
## FBgn0035189  215.64911      -2.974958 0.2535481 -11.733308 2.990802e-09
## FBgn0032405   79.61406      -2.672618 0.2397721 -11.146491 5.027699e-07
## FBgn0051092  153.06560      -2.327711 0.1767464 -13.169782 1.413227e-06
## FBgn0037290   67.09796      -3.010273 0.3300244  -9.121367 2.366946e-06
##                padj ENTREZID
## FBgn0025111 4.256022e-25    32008
## FBgn0000071 5.408582e-08    40831
## FBgn0035189 2.640023e-06    38124
## FBgn0032405 3.883269e-04    34627
## FBgn0051092 1.027333e-03    43105
## FBgn0037290 1.625040e-03    40613
```

ENTREZIDs of Down-regulated significant genes

```
down_entrez_ids <- downregulated_sig_genes$ENTREZID
down_entrez_ids
```

```
## [1] "32008" "40831" "38124" "34627" "43105" "40613"
```

Gene Ontology

Group up-regulated significant genes that have similar BP GO terms

```
go_up <- function(){
  go <- groupGO( gene = up_entrez_ids,
                 OrgDb = org.Dm.eg.db,
                 ont = "BP", # Biological Process
                 readable = TRUE)
  return(go)
}
go_terms_up <- go_up()
head(go_terms_up)
```

```
##           ID           Description Count GeneRatio
## G0:0000003 G0:0000003      reproduction      2      2/10
## G0:0002376 G0:0002376 immune system process      1      1/10
## G0:0008152 G0:0008152      metabolic process      3      3/10
## G0:0009987 G0:0009987      cellular process      5      5/10
## G0:0016032 G0:0016032      viral process      0      0/10
## G0:0022414 G0:0022414 reproductive process      2      2/10
##           geneID
## G0:0000003      sesB/Sox100B
## G0:0002376           Hml
## G0:0008152      sesB/CG1544/Sox100B
## G0:0009987 Kal1/sesB/CG1544/Sox100B/CG3168
## G0:0016032
## G0:0022414      sesB/Sox100B
```

Convert GO terms of up-regulated significant genes to DataFrame

```
dataframe_go_up <- function(){
  df_go_terms_up <- as.data.frame(go_terms_up)
  return(df_go_terms_up)
}
df_go_group_up <- dataframe_go_up()
```

```
df_go_group_up_top7 <- head(df_go_group_up,7)
```

Barplot of up-regulated BP GO terms

```
bar_plot_up <- function(){
p <- ggplot(df_go_group_up_top7, aes(x = reorder(Description, - Count),
y = Count, fill = Description)) +
  geom_bar(stat = "identity") +
  ggtitle("BP of up-regulated GO terms") +
  coord_flip() +
  theme_bw() +
  scale_fill_jama()+
  theme(plot.title = element_text(size = 12,
face = "bold", hjust = 0.5))+
  xlab("Description")
jpeg("../Enrichment analysis in R/outputs/BPgroup_up_barplot.jpeg")
print(p)
dev.off()
}
bar_plot_up()
```

```
## pdf
## 2
```

Group down-regulated significant genes that have similar MF GO terms

```
go_down <- function(){
  go <- groupGO(gene = down_entrez_ids,
    OrgDb = org.Dm.eg.db,
    ont = "MF", # Molecular function
    readable = TRUE)
  return(go)
}
go_terms_down <- go_down()
head(go_terms_down)
```

```
##           ID           Description Count GeneRatio
## GO:0003774 GO:0003774 cytoskeletal motor activity    0      0/6
## GO:0003824 GO:0003824 catalytic activity            2      2/6
## GO:0005198 GO:0005198 structural molecule activity    0      0/6
## GO:0005215 GO:0005215 transporter activity           1      1/6
## GO:0005488 GO:0005488 binding                       3      3/6
## GO:0009055 GO:0009055 electron transfer activity      0      0/6
##           geneID
## GO:0003774
## GO:0003824 CG9119/fir1
## GO:0005198
## GO:0005215 Ant2
## GO:0005488 Ama/CG9119/LpR2
## GO:0009055
```

Convert GO terms of down-regulated significant genes to DataFrame

```
dataframe_go_down <- function(){  
df_go_terms_down <- as.data.frame(go_terms_down)  
return(df_go_terms_down)  
}  
df_go_group_down <- dataframe_go_down()
```

```
df_go_group_down_top7 <- head(df_go_group_down,7)
```

Barplot of down-regulated MF GO terms

```
bar_plot_down <- function(){  
p <- ggplot(df_go_group_down_top7, aes(x = reorder(Description, - Count),  
y = Count, fill = Description)) +  
  geom_bar(stat = "identity") +  
  ggtitle("MF of down-regulated GO terms") +  
  coord_flip() +  
  theme_bw() +  
  scale_fill_jama() +  
  theme(plot.title = element_text(size = 12,  
face = "bold", hjust = 0.5)) +  
  xlab("Description")  
jpeg("../Enrichment analysis in R/outputs/MFgroup_down_barplot.jpeg")  
print(p)  
dev.off()  
}  
bar_plot_down()
```

```
## pdf  
## 2
```

Over-representation analysis

Go enrichment analysis

Enriched GO terms among up-regulated significant genes

```
enrich_go_up <- function(){  
  ego_up <- enrichGO( gene      = up_entrez_ids,  
                      OrgDb     = org.Dm.eg.db,  
                      keyType    = "ENTREZID",  
                      ont        = "CC",  
                      pvalueCutoff = 0.05,  
                      pAdjustMethod = "BH",  
                      readable    = TRUE)  
  
  return(ego_up)
```

```
}
enrichment_go_up <- enrich_go_up()
head(enrichment_go_up)
```

```
##              ID              Description GeneRatio  BgRatio
## GO:0031012 GO:0031012      extracellular matrix    2/7 271/12622
## GO:0031966 GO:0031966      mitochondrial membrane  2/7 300/12622
## GO:0030312 GO:0030312 external encapsulating structure 2/7 306/12622
## GO:0005740 GO:0005740      mitochondrial envelope    2/7 330/12622
## GO:0005918 GO:0005918      septate junction          1/7  35/12622
## GO:0070160 GO:0070160      tight junction            1/7  40/12622
##              pvalue  p.adjust  qvalue  geneID Count
## GO:0031012 0.008981758 0.04674036 0.02025898  Kal1/Hml  2
## GO:0031966 0.010926205 0.04674036 0.02025898  sesB/CG1544  2
## GO:0030312 0.011350225 0.04674036 0.02025898  Kal1/Hml  2
## GO:0005740 0.013119498 0.04674036 0.02025898  sesB/CG1544  2
## GO:0005918 0.019254363 0.04674036 0.02025898    CG3770   1
## GO:0070160 0.021978871 0.04674036 0.02025898    CG3770   1
```

Dataframe of enriched GO terms for up-regulated significant genes

```
df_go_terms_up <- function(){
  df_go_term_up <- as.data.frame(enrichment_go_up)
  return(df_go_term_up)
}
df_go_up <- df_go_terms_up()
```

```
df_go_up_top7 <- head(df_go_up, 7)
```

BarPlot of up-regulated CC enriched GO terms

```
bar_plot_enriched_up <- function(){
  jama_colors <- pal_jama("default")(7)
  p <- ggplot(df_go_up_top7, aes(x = reorder(Description, -Count),
                                y = Count, fill = p.adjust)) +
    geom_bar(stat = "identity") +
    coord_flip() +
    scale_fill_gradientn(name = "p.adjust",
                        colors = jama_colors) +
    labs(title = "Bar plot of CC of Enriched Up-regulated GO terms",
         x = "GO Term",
         y = "Gene Count") +
    theme_bw() +
    theme(plot.title = element_text(size = 10, face = "bold", hjust = 0.5))
  jpeg("../Enrichment analysis in R/outputs/CC_enriched_up_barplot.jpeg")
  print(p)
  dev.off()
}
bar_plot_enriched_up()
```

```
## pdf
## 2
```

Dotplot of up-regulated CC enriched GO terms

```
dot_plot_enriched_up <- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_go_up_top7, aes(x = reorder(Description, -Count),
                                y = Count, size = Count, color = p.adjust)) +
  geom_point(alpha = 0.6) +
  coord_flip() +
  scale_size_continuous(range = c(3, 8),
                        name = "Gene Count") +
  scale_color_gradientn(name = "p.adjust",
                        colors = jama_colors) +
  labs(title = "Dot plot of CC of enriched up-regulated genes",
        x = "GO Term",
        y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                   face = "bold",
                                   hjust = 0.5))

jpeg("../Enrichment analysis in R/outputs/CC_enriched_up_dotplot.jpeg")
print(p)
dev.off()
}
dot_plot_enriched_up()
```

```
## pdf
## 2
```

Network plot for enriched GO terms among up-regulated genes

```
jpeg("../Enrichment analysis in R/outputs/Network_plot_up.jpeg")
cnet_plot_up <- cnetplot(enrichment_go_up, showCategory = 3, vertex.label.cex = 1.2)
print(cnet_plot_up)
dev.off()
```

```
## pdf
## 2
```

GO graph for enriched GO terms among up-regulated significant genes

```
jpeg("../Enrichment analysis in R/outputs/GO_graph_up.jpeg")
go_graph_up <- plotGOgraph(enrichment_go_up)
```

```
##
## groupGOTerms:   GOBPterm, GOMFterm, GOCCTerm environments built.
```



```

##
## Building most specific GOs .....

## ( 33 GO terms found. )

##
## Build GO DAG topology .....

## ( 33 GO terms and 50 relations. )

##
## Attaching package: 'SparseM'

## The following object is masked from 'package:base':
##
##      backsolve

##
## Annotating nodes .....

## ( 12622 genes annotated to the GO terms. )

## Loading required package: Rgraphviz

## Loading required package: graph

## Loading required package: grid

##
## Attaching package: 'Rgraphviz'

## The following objects are masked from 'package:IRanges':
##
##      from, to

## The following objects are masked from 'package:S4Vectors':
##
##      from, to

print(go_graph_up)

## $dag
## A graphNEL graph with directed edges
## Number of Nodes = 28
## Number of Edges = 42
##
## $complete.dag
## [1] "A graph with 28 nodes."

```

```
dev.off()
```

```
## pdf
## 2
```

Enriched GO terms for down-regulated significant genes

```
enrich_go_down <- function(){
  ego_down <- enrichGO( gene      = down_entrez_ids,
                        OrgDb      = org.Dm.eg.db,
                        keyType     = "ENTREZID",
                        ont         = "MF",
                        pvalueCutoff = 0.05,
                        pAdjustMethod = "BH",
                        readable    = TRUE)

  return(ego_down)
}

enrichment_go_down <- enrich_go_down()
head(enrichment_go_down)
```

```
##              ID
## GO:0005346 GO:0005346
## GO:0015216 GO:0015216
## GO:0015215 GO:0015215
## GO:0015605 GO:0015605
## GO:1901505 GO:1901505
## GO:0015932 GO:0015932
##
##              Description
## GO:0005346      purine ribonucleotide transmembrane transporter activity
## GO:0015216      purine nucleotide transmembrane transporter activity
## GO:0015215      nucleotide transmembrane transporter activity
## GO:0015605      organophosphate ester transmembrane transporter activity
## GO:1901505      carbohydrate derivative transmembrane transporter activity
## GO:0015932      nucleobase-containing compound transmembrane transporter activity
##      GeneRatio BgRatio      pvalue    p.adjust      qvalue geneID Count
## GO:0005346      1/5 10/12435 0.004015092 0.02668318 0.01034805   Ant2      1
## GO:0015216      1/5 11/12435 0.004415890 0.02668318 0.01034805   Ant2      1
## GO:0015215      1/5 13/12435 0.005217101 0.02668318 0.01034805   Ant2      1
## GO:0015605      1/5 14/12435 0.005617513 0.02668318 0.01034805   Ant2      1
## GO:1901505      1/5 23/12435 0.009215419 0.03044605 0.01180733   Ant2      1
## GO:0015932      1/5 24/12435 0.009614543 0.03044605 0.01180733   Ant2      1
```

Dataframe of enriched GO terms among down-regulated significant genes

```
df_go_terms_down <- function(){
df_go_term_down <- as.data.frame(enrichment_go_down)
return(df_go_term_down)
```

```

}
df_go_down <- df_go_terms_down()

```

BarPlot of down-regulated MF enriched GO terms

```

bar_plot_enriched_down<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_go_down, aes(x = reorder(Description, - Count) ,
                             y = Count, fill = p.adjust)) +
  geom_bar(stat = "identity") +
  coord_flip() +
  scale_fill_gradientn(name = "p.adjust",
                       colors = jama_colors) +
  labs(title = "Bar plot of MF of Enriched down-regulated Genes",
        x = "GO Term",
        y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                   face = "bold",
                                   hjust = 0.5))
jpeg("../Enrichment analysis in R/outputs/MF_enriched_down_barplot.jpeg",
width = 700, height = 800)
print(p)
dev.off()
}
bar_plot_enriched_down()

```

```

## pdf
## 2

```

Dotplot of down-regulated MF enriched GO terms

```

dot_plot_enriched_down<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_go_down, aes(x = reorder(Description, -Count),
                             y = Count, size = Count,
                             color = p.adjust)) +
  geom_point(alpha = 0.6) +
  coord_flip() +
  scale_size_continuous(range = c(3, 8),
                        name = "Gene Count") +
  scale_color_gradientn(name = "p.adjust",
                        colors = jama_colors) +
  labs(title = "Dot plot of MF of enriched down-regulated genes",
        x = "GO Term",
        y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                   face = "bold",

```

```

                                                    hjust = 0.5))
jpeg("../Enrichment analysis in R/outputs/MF_enriched_down_dotplot.jpeg",
width = 700, height = 800)
print(p)
dev.off()
}
dot_plot_enriched_down()

```

```

## pdf
## 2

```

Network plot for enriched GO terms among down-regulated genes

```

jpeg("../Enrichment analysis in R/outputs/Network_plot_down.jpeg")
cnet_plot_down <- cnetplot(enrichment_go_down, showCategory = 7, vertex.label.cex = 1.2)
print(cnet_plot_down)
dev.off()

```

```

## pdf
## 2

```

GO graph for enriched GO terms among down-regulated significant genes

```

jpeg("../Enrichment analysis in R/outputs/GO_graph_down.jpeg")
go_graph_down <- plotGOgraph(enrichment_go_down)

```

```

##
## groupGOTerms:   GOBPterm, GOMFterm, GOCCterm environments built.

##
## Building most specific GOs .....

## ( 41 GO terms found. )

##
## Build GO DAG topology .....

## ( 41 GO terms and 50 relations. )

##
## Annotating nodes .....

## ( 12435 genes annotated to the GO terms. )

```

```
print(go_graph_down)
```

```
## $dag
## A graphNEL graph with directed edges
## Number of Nodes = 12
## Number of Edges = 13
##
## $complete.dag
## [1] "A graph with 12 nodes."
```

```
dev.off()
```

```
## pdf
## 2
```

Pathway enrichment analysis

Extract sig gene names from ensembl database

```
ensembl_gene_names <- function() {
  # Connect to the Ensembl database
  ensembl <- useMart("ensembl", dataset = "dmelanogaster_gene_ensembl")
  # list of Drosophila melanogaster FlyBase gene IDs
  flybase_gene_ids <- rownames(sign_genes)
  # Convert FlyBase gene IDs to external gene identifiers
  gene_info <- getBM(attributes = c("ensembl_gene_id", "external_gene_name"), filters = "flybase_gene_id", values = flybase_gene_ids, mart = ensembl)
  return(gene_info)
}
gene_info_result <- ensembl_gene_names()
head(gene_info_result)
```

```
##   ensembl_gene_id external_gene_name
## 1   FBgn0000003      7SLRNA:CR32864
## 2   FBgn0000008                      a
## 3   FBgn0000014          abd-A
## 4   FBgn0000015          Abd-B
## 5   FBgn0000017          Abl
## 6   FBgn0000018          abo
```

Up-regulated Dmel_XXXX ids

```
dmel_up <- function(){
  filtered_up_sig_genes <- gene_info_result[gene_info_result$ensembl_gene_id %in% rownames(upregulated_genes), ]
  dmel_up_identifiers <- paste("Dmel", filtered_up_sig_genes$external_gene_name, sep = "_")
  return(dmel_up_identifiers)
}
dmel_up_genes <- dmel_up()
dmel_up_genes
```

```
## [1] "Dmel_sesB"      "Dmel_Sox100B" "Dmel_Hml"      "Dmel_CG3168"  "Dmel_gas"
## [6] "Dmel_CG3770"    "Dmel_CG15695" "Dmel_Kal1"     "Dmel_CG1544"  "Dmel_CG34330"
```

KEGG pathway enrichment analysis among up-regulated significant genes

```
kegg_enrichment_up <- function(){
kegg_up <- enrichKEGG(gene = dmel_up_genes,
                      organism = "dme", # Drosophila melanogaster
                      pvalueCutoff = 0.05)
  return(kegg_up)
}
kegg_enrich_up <- kegg_enrichment_up()
```

```
## Reading KEGG annotation online: "https://rest.kegg.jp/link/dme/pathway"...
```

```
## Reading KEGG annotation online: "https://rest.kegg.jp/list/pathway/dme"...
```

```
head(kegg_enrich_up)
```

```
##                                category
## dme04512 Environmental Information Processing
## dme00785                                Metabolism
## dme00380                                Metabolism
## dme00310                                Metabolism
## dme01210                                Metabolism
##                                subcategory      ID
## dme04512 Signaling molecules and interaction dme04512
## dme00785 Metabolism of cofactors and vitamins dme00785
## dme00380      Amino acid metabolism dme00380
## dme00310      Amino acid metabolism dme00310
## dme01210      Global and overview maps dme01210
##                                Description
## dme04512      ECM-receptor interaction - Drosophila melanogaster (fruit fly)
## dme00785      Lipoic acid metabolism - Drosophila melanogaster (fruit fly)
## dme00380      Tryptophan metabolism - Drosophila melanogaster (fruit fly)
## dme00310      Lysine degradation - Drosophila melanogaster (fruit fly)
## dme01210      2-Oxocarboxylic acid metabolism - Drosophila melanogaster (fruit fly)
##      GeneRatio BgRatio      pvalue  p.adjust qvalue      geneID Count
## dme04512      1/2 11/3587 0.006124707 0.01946050      NA Dmel_CG3168      1
## dme00785      1/2 18/3587 0.010012453 0.01946050      NA Dmel_CG1544      1
## dme00380      1/2 21/3587 0.011676297 0.01946050      NA Dmel_CG1544      1
## dme00310      1/2 35/3587 0.019422401 0.01997453      NA Dmel_CG1544      1
## dme01210      1/2 36/3587 0.019974528 0.01997453      NA Dmel_CG1544      1
```

DataFrame of KEGG enriched among up-regulated significant genes

```
dataframe_kegg_up <- function(){
  df_kegg_up_genes <- as.data.frame(kegg_enrich_up)
```

```

    return(df_kegg_up_genes)
}
df_kegg_up <- dataframe_kegg_up()
head(df_kegg_up)

```

```

##                                category
## dme04512 Environmental Information Processing
## dme00785                                Metabolism
## dme00380                                Metabolism
## dme00310                                Metabolism
## dme01210                                Metabolism
##                                subcategory      ID
## dme04512 Signaling molecules and interaction dme04512
## dme00785 Metabolism of cofactors and vitamins dme00785
## dme00380      Amino acid metabolism dme00380
## dme00310      Amino acid metabolism dme00310
## dme01210      Global and overview maps dme01210
##
##                                Description
## dme04512      ECM-receptor interaction - Drosophila melanogaster (fruit fly)
## dme00785      Lipoic acid metabolism - Drosophila melanogaster (fruit fly)
## dme00380      Tryptophan metabolism - Drosophila melanogaster (fruit fly)
## dme00310      Lysine degradation - Drosophila melanogaster (fruit fly)
## dme01210      2-Oxocarboxylic acid metabolism - Drosophila melanogaster (fruit fly)
##      GeneRatio BgRatio      pvalue  p.adjust qvalue      geneID Count
## dme04512      1/2 11/3587 0.006124707 0.01946050      NA Dmel_CG3168      1
## dme00785      1/2 18/3587 0.010012453 0.01946050      NA Dmel_CG1544      1
## dme00380      1/2 21/3587 0.011676297 0.01946050      NA Dmel_CG1544      1
## dme00310      1/2 35/3587 0.019422401 0.01997453      NA Dmel_CG1544      1
## dme01210      1/2 36/3587 0.019974528 0.01997453      NA Dmel_CG1544      1

```

Barplot for KEGG enriched up-regulated significant gene

```

bar_plot_kegg_enriched_up<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_kegg_up, aes(x = reorder(Description, - Count),
                             y = Count, fill = p.adjust)) +
  geom_bar(stat = "identity", width = 0.8) +
  coord_flip() +
  scale_fill_gradientn(name = "p.adjust",
                       colors = jama_colors) +
  labs(title = "Bar plot of Enriched KEGG pathway among up-regulated Genes",
        x = "Enriched pathway",
        y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                   face = "bold",
                                   hjust = 0.5))
jpeg("../Enrichment analysis in R/outputs/kegg_enriched_up_barplot.jpeg",
width = 700, height = 800)
print(p)
dev.off()

```

```
}
bar_plot_kegg_enriched_up()
```

```
## pdf
## 2
```

Dotplot for Enriched KEGG pathway among up-regulated significant gene

```
dot_plot_kegg_enriched_up<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_kegg_up, aes(x = reorder(Description, -Count),
                             y = Count, size = Count,
                             color = p.adjust)) +
  geom_point(alpha = 0.6) +
  coord_flip() +
  scale_size_continuous(range = c(3, 8),
                        name = "Gene Count") +
  scale_color_gradientn(name = "p.adjust",
                        colors = jama_colors) +
  labs(title = "Dot plot of Enriched KEGG pathway among up-regulated genes",
        x = "GO Term",
        y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                   face = "bold",
                                   hjust = 0.5))
jpeg("../Enrichment analysis in R/outputs/kegg_enriched_up_dotplot.jpeg",
width = 800, height = 800)
print(p)
dev.off()
}
dot_plot_kegg_enriched_up()
```

```
## pdf
## 2
```

Visualize a specific pathway

visualize the top enriched KEGG pathway among up-regulated sig genes

```
visualize_top_path_up <- function(){
  pathview(gene.data = up_entrez_ids,
            pathway.id = "dme04512",
            species = "dme",
            kegg.dir = "../Enrichment analysis in R/outputs/")
}
visualize_top_path_up()
```



```
## Info: Getting gene ID data from KEGG...

## Info: Done with data retrieval!

## Info: Working in directory C:/Users/HP/Desktop/Mentor M. Souady/Bulk RNAseq analysis/Enrichment analysis

## Info: Writing image file dme04512.pathview.png
```

Browse the top enriched pathway among up-regulated sig genes

```
browseKEGG(kegg_enrich_up, 'dme04512')
```

Downregulated Dmel_XXXX ids

```
dmel_down <- function(){
  filtered_down_sig_genes <- gene_info_result[gene_info_result$ensembl_gene_id %in% rownames(downregulated_genes)]
  dmel_down_identifiers <- paste("Dmel", filtered_down_sig_genes$external_gene_name, sep = "_")
  return(dmel_down_identifiers)
}
dmel_down_genes <- dmel_down()
dmel_down_genes
```

```
## [1] "Dmel_Ama"      "Dmel_Ant2"     "Dmel_fir1"     "Dmel_CG9119"   "Dmel_CG1124"
## [6] "Dmel_LpR2"
```

KEGG pathway enrichment analysis among down-regulated significant genes

```
kegg_enrichment_down <- function(){
  kegg_down <- enrichKEGG(gene = dmel_down_genes,
                          organism = "dme", # Drosophila melanogaster
                          pvalueCutoff = 0.05,
                          pAdjustMethod = "BH")
  if (is.null(kegg_down) || nrow(kegg_down) == 0) {
    print("No enriched pathways found")
  }
  return(kegg_down)
}
kegg_enrich_down <- kegg_enrichment_down()
```

```
## [1] "No enriched pathways found"
```

Reactome pathway enrichment analysis among down-regulated significant genes

```

reactome_down_genes <- function(){
  reactome_enrichment <- enrichPathway(gene = down_entrez_ids,
                                         organism = "fly", # Drosophila melanogaster
                                         pvalueCutoff = 0.05)

  return(reactome_enrichment)
}
reactome_enriched_path_down <- reactome_down_genes()
head(reactome_enriched_path_down)

```

```

##                                ID
## R-DME-8964038 R-DME-8964038
## R-DME-8964043 R-DME-8964043
## R-DME-1268020 R-DME-1268020
## R-DME-5365859 R-DME-5365859
## R-DME-425397  R-DME-425397
## R-DME-5362517 R-DME-5362517
##
##                                Description
## R-DME-8964038                      LDL clearance
## R-DME-8964043                      Plasma lipoprotein clearance
## R-DME-1268020                      Mitochondrial protein import
## R-DME-5365859                      RA biosynthesis pathway
## R-DME-425397  Transport of vitamins, nucleosides, and related molecules
## R-DME-5362517                      Signaling by Retinoic Acid
##
##      GeneRatio BgRatio      pvalue    p.adjust      qvalue geneID Count
## R-DME-8964038      1/3 12/4698 0.007644902 0.03851761 0.009356504 43105      1
## R-DME-8964043      1/3 18/4698 0.011452699 0.03851761 0.009356504 43105      1
## R-DME-1268020      1/3 19/4698 0.012086383 0.03851761 0.009356504 32008      1
## R-DME-5365859      1/3 19/4698 0.012086383 0.03851761 0.009356504 34627      1
## R-DME-425397      1/3 26/4698 0.016514591 0.03851761 0.009356504 32008      1
## R-DME-5362517      1/3 28/4698 0.017777358 0.03851761 0.009356504 34627      1

```

DataFrame of Reactome enriched pathways among down-regulated significant genes

```

dataframe_reactome_down <- function(){
  df_reactome_down_genes <- as.data.frame(reactome_enriched_path_down)
  return(df_reactome_down_genes)
}
df_reactome_down <- dataframe_reactome_down()
head(df_reactome_down)

```

```

##                                ID
## R-DME-8964038 R-DME-8964038
## R-DME-8964043 R-DME-8964043
## R-DME-1268020 R-DME-1268020
## R-DME-5365859 R-DME-5365859
## R-DME-425397  R-DME-425397
## R-DME-5362517 R-DME-5362517
##
##                                Description

```

## R-DME-8964038							LDL clearance	
## R-DME-8964043							Plasma lipoprotein clearance	
## R-DME-1268020							Mitochondrial protein import	
## R-DME-5365859							RA biosynthesis pathway	
## R-DME-425397							Transport of vitamins, nucleosides, and related molecules	
## R-DME-5362517							Signaling by Retinoic Acid	
##	GeneRatio	BgRatio	pvalue	p.adjust	qvalue	geneID	Count	
## R-DME-8964038	1/3	12/4698	0.007644902	0.03851761	0.009356504	43105	1	
## R-DME-8964043	1/3	18/4698	0.011452699	0.03851761	0.009356504	43105	1	
## R-DME-1268020	1/3	19/4698	0.012086383	0.03851761	0.009356504	32008	1	
## R-DME-5365859	1/3	19/4698	0.012086383	0.03851761	0.009356504	34627	1	
## R-DME-425397	1/3	26/4698	0.016514591	0.03851761	0.009356504	32008	1	
## R-DME-5362517	1/3	28/4698	0.017777358	0.03851761	0.009356504	34627	1	

Visualize Reactome Pathway Enrichment Results

Barplot for Reactome enriched pathways among down-regulated significant gene

```
bar_plot_reactome_enriched_down<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_reactome_down, aes(x = reorder(Description, - Count) ,
                                y = Count, fill = p.adjust)) +
  geom_bar(stat = "identity", width = 0.8) +
  coord_flip() +
  scale_fill_gradientn(name = "p.adjust",
                      colors = jama_colors) + labs(title = "Bar plot of
                                Enriched pathway",
                                y = "Gene Count") +
  theme_bw() +
  theme(plot.title = element_text(size = 10,
                                face = "bold",
                                color = "red",
                                align = "center"))
jpeg("../Enrichment analysis in R/outputs/reactome_enriched_down_barplot.jpeg",
width = 700, height = 800)
print(p)
dev.off()
}
bar_plot_reactome_enriched_down()
```

```
## pdf
## 2
```

Dotplot for Reactome enriched pathways among downregulated significant gene

```
dot_plot_reactome_enriched_down<- function(){
jama_colors <- pal_jama("default")(7)
p <- ggplot(df_reactome_down, aes(x = reorder(Description, -Count),
                                y = Count, size = Count,
                                color = p.adjust)) +
  geom_point(alpha = 0.6) +
```

```

coord_flip() +
scale_size_continuous(range = c(3, 8),
                        name = "Gene Count") +
scale_color_gradientn(name = "p.adjust",
                       colors = jama_colors) +
labs(title = "Dot plot of enriched Reactome pathways among down-regulated genes",
     x = "GO Term",
     y = "Gene Count") +
theme_bw() +
theme(plot.title = element_text(size = 10,
                                face = "bold",
                                color = "red"),
      legend.title = element_text(size = 10,
                                   face = "bold",
                                   color = "red"),
      legend.text = element_text(size = 10,
                                  face = "bold",
                                  color = "red"),
      axis.title = element_text(size = 10,
                                 face = "bold",
                                 color = "red"),
      axis.text = element_text(size = 10,
                                face = "bold",
                                color = "red"),
      panel.grid = element_line(linetype = "dotted",
                                color = "red",
                                size = 1))
jpeg("../Enrichment analysis in R/outputs/reactome_enriched_down_dotplot.jpeg",
width = 700, height = 800)
print(p)
dev.off()
}
dot_plot_reactome_enriched_down()

```

```

## pdf
## 2

```

visualize the top enriched Reactome pathway for down-regulated sig genes

Take the generated URL and browse it

```

visualize_reactome_path_up <- function(){
  # Convert the Reactome ID to a URL for visualization
  reactome_url <- paste0("https://reactome.org/PathwayBrowser/#/", "R-DME-8964038")
  # Print the URL for manual review
  print(reactome_url)
}
visualize_reactome_path_up()

```

```
## [1] "https://reactome.org/PathwayBrowser/#/R-DME-8964038"
```