

RESEARCH ARTICLE

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Degradation of red blood cell deformability during cold storage in blood bags

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Abstract

Red blood cells (RBCs) stored in blood bags develop a storage lesion that include structural, metabolic, and morphologic transformations resulting in a progressive loss of RBC deformability. The speed of RBC deformability loss is donor-dependent, which if properly characterized, could be used as a biomarker to select high-quality RBC units for sensitive recipients or to provide customized storage timelines depending on the donor. We used the microfluidic ratchet device to measure the deformability of red blood cells stored in blood bags every 14 days over a span of 56 days. We observed that storage in blood bags generally prevented RBC deformability loss over the current standard 42-day storage window. However, between 42 and 56 days, the deformability loss profile varied dramatically between donors. In particular, we observed accelerated RBC deformability loss for a majority of male donors, but for none of the female donors. Together, our results suggest that RBC deformability loss could be used to screen for donors who can provide stable RBCs for sensitive transfusion recipients or to identify donors capable of providing RBCs that could be stored for longer than the current 42-day expiration window.

KEYWORDS

blood banking, deformability, red blood cell, storage lesion

1 | INTRODUCTION

Red blood cells (RBCs) collected from donors for use in blood transfusions are currently stored at 4°C for up to 42 days [1,2]. During this period, RBCs can develop a storage lesion, which is characterized by a number of structural (lipid peroxidation, Band 3 aggregation, mem-

brane asymmetry), metabolic (slowed metabolism due to adenosine triphosphate and 2,3-diphosphoglycerate depletion), and morphologic transformations (discoid, echinocyte, and spherocyte) [3–6]. The storage lesion coincides with a shorter RBC circulation time arising from the rapid uptake of transfused RBCs by reticuloendothelial macrophages [7], and thus resulting in the need for more frequent

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transfusions. While the 42-day storage window is currently uniformly applied to all RBC units, the actual rate of RBC degradation is known to vary between donors [4,5,8,9]. This variability has also been observed in outcomes for chronic transfusion recipients, where some RBC units are able to maintain durable hemoglobin levels in recipients; while other units are rapidly cleared, leading to the need for repeat transfusions [10,11]. Therefore, if the rates of degradation could be established for individual donors, it may be possible to select long-lasting units for sensitive recipients, such as those requiring chronic transfusions. Similarly, it may also be possible to provide customized expiration timelines for different donors to ensure that high-quality RBC units are not prematurely outdated, while less stable RBC units are used before they cease to provide clinical benefits. A key challenge in transfusion medicine has therefore been the development of a simple biomarker to assess the quality of stored blood to optimally meet the needs of the transfusion recipient.

Independent investigation of the cellular changes associated with RBC storage lesions has so far failed to produce a reliable biomarker for storage-based degradation [12]. However, these cellular changes collectively reduce RBC deformability and thus make this parameter an attractive potential biomarker for the RBC storage lesion. Previous studies have found the deformability of cold stored RBCs to be relatively stable for the first 14 days but begins to degrade after 3 weeks of storage [5,13–16]. This change coincides with clinical evidence indicating that blood transfusion efficacy diminishes markedly after 30-day storage [17,18]. The loss of RBC deformability may directly impact on transfusion efficacy as more rigid RBC may be taken up more rapidly by the reticuloendothelial macrophages [19]. Additionally, rigid transfused RBCs are known to compromise microvascular flow by occluding blood capillaries [20]. Together, these findings from previous studies suggest that RBC deformability is a promising biomarker for the degradation of stored RBC units.

Various methods have been employed to measure deformability of stored RBCs, including bulk flow and single cell techniques. Bulk flow methods include micropore filtration [13,21,22] and ektacytometry [23–25]. These methods infer RBC deformability indirectly based on blood viscosity and only provide a populational average measurement, both of which limit the sensitivity of these methods. Single cells methods, such as micropipette aspiration [26–28] and optical tweezers [29–31], provide single-cell deformability measurements but are typically limited by sample throughput, which make them susceptible to variability and selection bias. Microfluidic techniques have been developed to overcome these limitations by enabling RBC deformability measurement with greater throughput and ease-of-use. Importantly, recent methods are beginning to provide sufficient sensitivity and repeatability to observe deformability loss in RBCs donated for blood transfusions [32–39]. These studies suggest that it may be possible to identify blood donors that can provide high-quality RBC units that could be reserved for sensitive or chronic transfusion recipients. Donor-specific RBC deformability measurement is particularly useful for its potential to explain evidence for donor-dependent storage and transfusion efficacy [1,40–43]. Therefore, measurement sensitivity and repeatability are critical properties

in efforts to assess differences between donated RBC units or between donors.

Recently, we developed a microfluidic technology to measure RBC deformability with sufficient sensitivity and repeatability for analyzing differences between healthy donors [36]. Using an accelerated aging model of RBCs stored in plastic tubes, we found that donor RBCs had degradation profiles that were highly variable between donors, but consistent for each donor. Importantly, some donors showed significant loss of RBC deformability during storage, while other donors showed little or no storage-induced loss of RBC deformability. Here, to evaluate variability between donors during cold storage in blood bags, we assessed the degradation of RBC deformability over the 42-day storage window, and for an additional 14 days thereafter, for a total of 56 days. We show that, in most cases, blood bags preserved RBC deformability during the 42-day storage window, while the degradation of RBC deformability in the subsequent 14 days were highly variable. Our results confirm that RBC deformability provides a potentially useful approach for donor-level screening to identify donors for whom the storage expiration window could potentially be lengthened.

2 | METHODS

2.1 | Blood bags

This study was approved by the University of British Columbia's Clinical Research Ethics Board (UBC REB# H19-01121) and Canadian Blood Services (CBS) Research Ethics Board (CBS REB# 2019-029). RBCs were collected and processed by CBS between January 2020 and February 2021 using the buffy coat separation method and saline-adenine-glucose-mannitol additive solution for storage in standard Fresenius blood bags [44].

2.2 | RBC storage and processing for deformability assessment

RBC units were stored according to CBS standard operating procedures, at 4°C for a period of 8 weeks, 2 weeks longer than the CBS-approved storage period of 42 days (6 weeks). Samples were analyzed on the day of RBC unit collection and processing, followed by analysis at weeks 2, 4, 6, and post-expiration at week 8.

To analyze the RBCs within a blood bag, a 3 ml sample was aseptically drawn from the unit through the blood administration ports, using a 27-gauge needle and syringe (BD) to limit port ripping and subsequent leakage. Blood is drawn very slowly to avoid bubbles and turbulence of flow. The ports were then covered with Parafilm to preserve the sterility of the unit throughout the storage period. The drawn sample was centrifuged at 1500 × g with no brakes for 10 min at room temperature, and supernatant was transferred to a fresh tube. The supernatant was centrifuged again at 1500 × g for 10 min to remove any remaining RBCs, transferred to a cryogenic vial and stored at –80°C for assessment of hemolysis at a later stage. Additionally,

100 μ l of the blood drawn on day 1 was also stored for hemolysis assessment. The RBC pellet was suspended in Hank's Balanced Salt Solution (HBSS, Gibco) and 0.2% Pluronic-F127 (MilliporeSigma), also known as poloxamer 407, and washed three more times, each time centrifuging for 5 min at 300 \times g with brakes on. The final RBC sample was suspended at 1% hematocrit in HBSS + 0.2% Pluronic solution and used for deformability assessment using the microfluidic ratchet device.

2.3 | Hematological parameters

At each sampling timepoint, 100 μ l of RBC sample (no washing) was used to monitor the hematological parameter changes. Mean corpuscular volume (MCV), red blood cell distribution width (RDW-CV), mean cell hemoglobin (MCH), and mean corpuscular hemoglobin concentration (MCHC) were assessed using the Sysmex system. Note that a few data points from donors 3 and 4 are missing due to equipment inaccessibility during the COVID-19 pandemic.

2.4 | Assessment of hemolysis

The hemolysis assessment was performed as previously described [45]. Briefly, frozen whole blood and supernatant samples were fully thawed. An aliquot of each supernatant sample was transferred to a fresh tube and centrifuged at 3000 \times g for 3 min. Packed, unwashed blood samples from day 1 of storage were vortexed for 30 s and diluted 1:10 with deionized (DI) water. Finally, 10 μ l of each sample was transferred to a 96-well flat bottom plate (BioVision Inc) together with 100 μ l of Drabkin's reagent (MilliporeSigma) containing Brij-35 solution (ThermoFisher). The plate was then incubated at room temperature for 15 min on a plate shaker, and absorbance was read on a microplate reader (manufacturer) at 540 nm. Hemolysis in the supernatant at each timepoint was calculated relative to whole blood at day 1, using the following formula:

$$\% \text{ Hemolysis} = \frac{\text{OD}_{\text{supernatant}} \times (1 - \text{Hct})}{\text{OD}_{\text{whole blood}}} \times 100\%$$

where Hct is the hematocrit, and $\text{OD}_{\text{supernatant}}$ and $\text{OD}_{\text{whole blood}}$ are the measured optical density from the supernatant and whole blood, respectively.

2.5 | Microfluidic device manufacturing and operation

The microfluidic ratchet device was manufactured as previously described [36,46,47]. Briefly, a mask with device features was created by photolithographic fabrication, which in turn was used to create a polydimethylsiloxane (PDMS, Sylgard-184, Ellsworth Adhesives) master. The PDMS master device was used to create secondary molds for routine device manufacture. The PDMS device is made by mixing PDMS in a 10:1 ratio of PDMS and hardener and cured in the mold in

a 65°C oven for a minimum of 2 h. Holes are manually punched in the device using Harris Uni-Core punches with 0.5 mm diameter for inlets and 2 mm for outlets. The PDMS part of the device is then bound to a thin RTV layer (RTV 615, Momentive Performance Materials LLC), followed by to a glass slide (2 \times 3 inch, Corning) for durability, using a Har-rick Plasma model PDC-001 air plasma.

Prior to sample introduction, the device is filled with PBSS + 0.2% Pluronic-F127 buffer for 15 min until fully buffered. The microfluidic ratchet device operates using an oscillatory sorting pressure and constant forward pressure system, which propagates the sample forward toward the distinct outlet based on the cell's ability to deform through the sorting matrix. The upward pressure is applied at 175 mbar for 4 s, and downward pressure is applied for 1 s at 162 mbar. The forward and sample pressures are applied at 40–45 and 50–55 mbar, respectively. The distribution of cells in each distinct outlet is counted in each outlet microchannel. Each sample is run on two separate devices, and the mean is calculated thereafter. Each device is discarded after each use.

2.6 | Statistical analysis

Statistical analysis was performed using GraphPad Prism (V8.0) software. Means and standard deviation from mean are plotted unless otherwise stated. To calculate the standard deviation for rigidity score (RS) obtained from doublet measurements, the following formula was used:

$$\sigma_{\text{RS}} = \sqrt{\frac{1}{N} \sum_{i=1}^N (M1_i - M2_i)^2}$$

where M1 and M2 are the first and second RS measurements. Correlations between data sets were calculated using Pearson *r* with 95% confidence interval.

3 | RESULTS

3.1 | Sorting RBCs based on deformability using the microfluidic ratchet device

The design of the microfluidic ratchet device to sort RBCs based on deformability has been described previously [36,46,47]. Briefly, RBCs are deformed through a series of micrometer-sized constrictions using oscillatory flow, which selectively transport cells based on their ability to squeeze through each constriction. The constrictions are arranged in a matrix, where the openings of the constrictions are varied from 7.5 μ m down to 1.5 μ m, between rows in the matrix. RBCs are sorted diagonally through the constriction matrix until reaching a limiting constriction row that prevents their transit. The RBCs then proceed horizontally along the limiting row of constrictions until they reach a specific outlet. The resolution of these constrictions is limited by our photolithographic microfabrication process and can be improved using higher-resolution equipment. Our current design enables sorting of RBCs based on their deformability into 12 fractions in different outlets. The distribution of cells after sorting could be determined by imaging

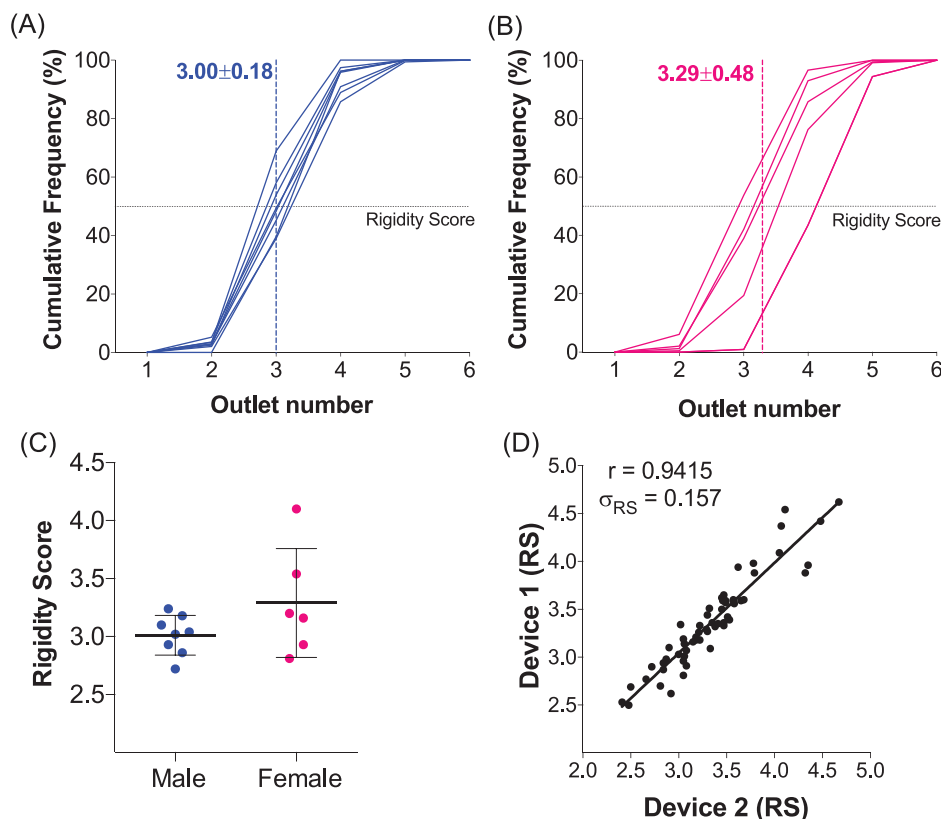


FIGURE 1 Inter-donor variability of packed red blood cell (RBC) units on the day of collection and measurement repeatability. Cumulative distribution of RBCs sorted based on deformability using the microfluidic ratchet device from male (A) and female (B) donors. A rigidity score (RS) is derived from the fractional outlet number at the 50% cross over point of the cumulative distribution. The mean RSs for male and female donors are indicated using dashed lines. (C) RS for RBCs from the day of collection for male ($n = 8$) and female ($n = 6$) donors. (D) Repeatability of the RS from doublet measurements on the same samples, which showed a Pearson's $r = 0.9415$ and a standard deviation $\sigma_{RS} = 0.157$

the flow of cells into the outlets or by counting the cells in the outlet via microscopy.

3.2 | Data analysis

After sorting each RBC sample using the microfluidic ratchet device, the distribution of RBCs in outlets 1–12 can be used to establish a cumulative distribution from the smallest outlet to the largest outlet. The cumulative distribution could then be described using the RS based on the outlet where the cumulative distribution function crosses 50% (Figure 1A,B). Fractional outlet numbers can be obtained by linear interpolation of cumulative distribution graph between outlets. The RS provides a simple metric for comparing distributions between different donor and samples [36].

3.3 | RBC deformability profiles at the time of collection

We established a baseline deformability profile of all blood bags at the time of collection. Packed RBCs in standard blood bags collected from healthy donors ($n = 14$) were obtained from CBS. Consistent with previous reports [5,36], we observed significant variability in initial RBC

deformability among donors. Specifically, donor RS ranged from 2.71 to 4.13 with a mean of 3.13 ± 0.37 . The mean RS of male donor RBCs ($n = 8$, 3.00 ± 0.18 , Figure 1A) was slightly lower than female donor RBCs ($n = 6$, 3.29 ± 0.48 , Figure 1B), but this difference was not statistically significant (Figure 1C).

3.4 | Measurement repeatability

Each RBC sample in this study was measured twice using different replicate microfluidic devices. We used this doublet data to confirm the repeatability of our measurement by plotting the RS from the first and second measurements against each other (Figure 1D). These results suggest that the RBC deformability measurements were highly repeatable with a standard deviation of 0.157 in repeated measurements.

3.5 | RBC deformability loss during cold storage

To assess RBC deformability loss during cold storage, we sampled RBCs from blood bags every 2 weeks from 0 to 8 weeks of cold storage, which is 2 weeks beyond the current 42-day storage window (Figure 2). We initially tested RBC deformability changes at days 4 and 7 ($n = 5$, data not shown), and since there were no noticeable

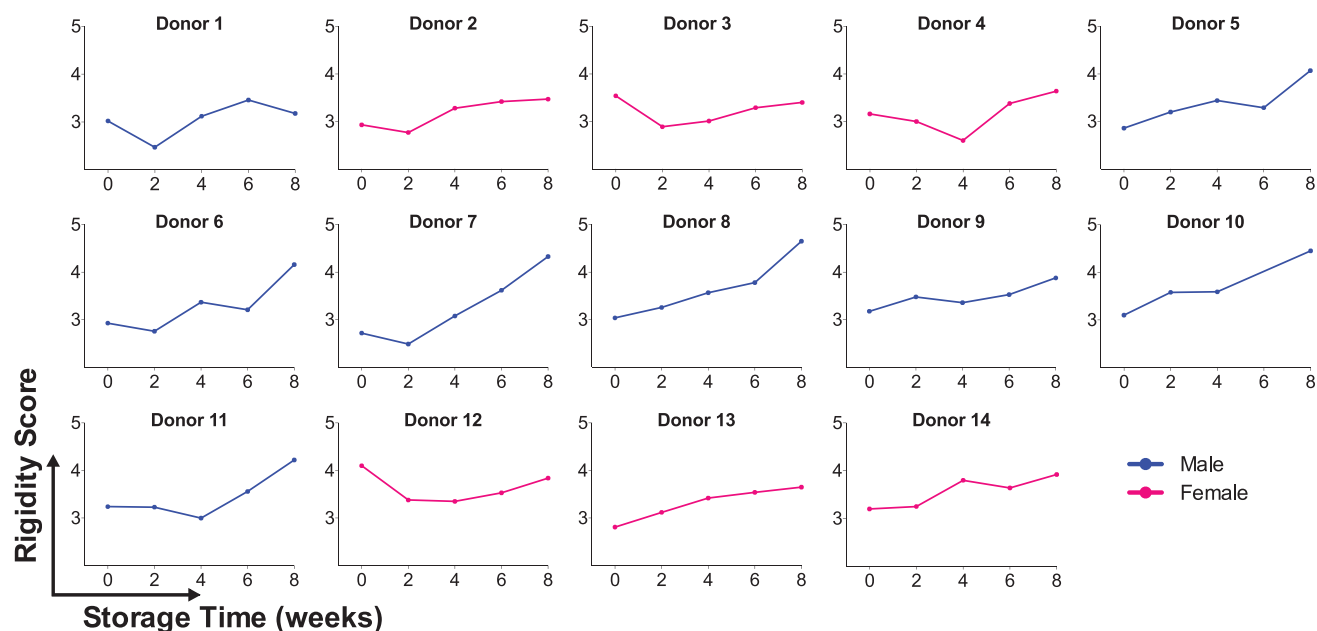


FIGURE 2 Red blood cell (RBC) deformability aging curves. Measured rigidity score (RS) for RBCs from each donor, sampled every 2 weeks over 8 weeks of cold storage. Each data point is the mean of doublet measurements

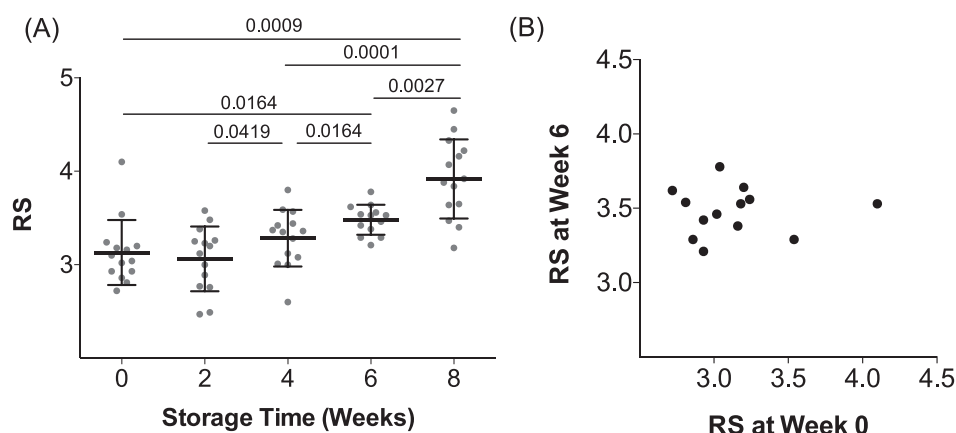


FIGURE 3 Degradation of red blood cell (RBC) deformability during cold storage. (A) Donor RBCs exhibited a progressive increase in rigidity score (RS) over 6 weeks, with an accelerated increase in RS between weeks 6 and 8. The mean rigidity scores (RSs) were 3.13 ± 0.35 (week 0), 3.06 ± 0.35 (week 2), 3.29 ± 0.30 (week 4), 3.48 ± 0.16 (week 6), and 3.91 ± 0.42 (week 8). (B) Correlation between RS on the day of processing and the day of expiration (week 6), $r = 0.0086$

changes, we continued to sample every 2 weeks of storage. From 0 to 4 weeks, the stored RBC units showed no detectable deformability loss. At the expiry date of 6 weeks, the stored RBC units showed detectable loss in deformability, as reflected by an increased RS of 0.35 ($p < 0.05$; Figure 3A). From weeks 6 to 8, the stored RBC units exhibited a dramatic loss of deformability. In fact, the average RBC deformability loss was greater from weeks 6 to 8 ($\Delta RS = 0.42$) than for the loss from weeks 0 to 6. While it should be noted that RBC deformability alone does not necessarily predict transfusion efficacy, this punctuated loss of RBC deformability after 6 weeks strongly supports the current 42-day storage window. Interestingly,

some donated RBC units showed an initial increase in deformability from weeks 0 to 2, which was followed by a progressive deformability loss thereafter. These results are confirmed by doublet measurements and are also consistent with earlier studies that showed RBC units can often recover some of their deformability upon initial storage [5,48,49].

We further evaluated whether the deformability of fresh RBC is predictive of the rate of RBC deformability loss during storage by relating the RS of RBCs at the time of collection to the RS at the end of the 42-day storage window. We found no correlation between the two (Figure 3B). In fact, regardless of initial RS, all RBC units converged to RS of

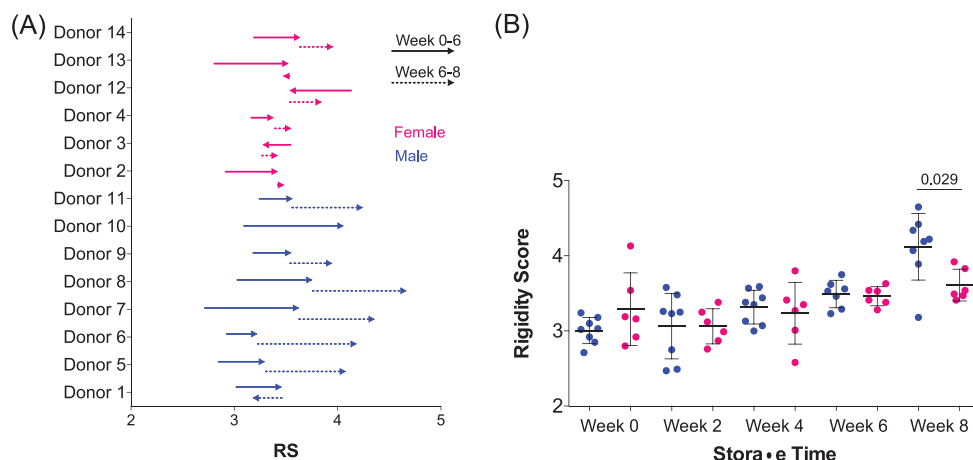


FIGURE 4 Comparison of red blood cell (RBC) deformability loss profiles between male and female donors. (A) Changes in rigidity scores (RSs) from weeks 0 to 6 and weeks 6 to 8 of cold storage. Arrows indicate direction of change. (B) RS for male and female donors every 2 weeks of storage for 8 weeks

3.48 ± 0.16 . These results show that the RBC deformability loss profile during storage cannot be predicted by initial RBC deformability. Therefore, determining this profile for each donor will require multiple samples over storage time.

3.6 | Differences in RBC stability during storage between male and female donors

We investigated the differences in RBC deformability loss profiles between male donors ($n = 8$) and female donors ($n = 6$). In the first 6 weeks of storage, we observed no donor-specific variation in RBC deformability loss. From weeks 6 to 8, we observed a dramatic loss of RBC deformability for the male donors, but not female donors ($p < 0.05$; Figure 4). In fact, an accelerated RBC deformability loss in the final 2 weeks of storage was observed for the majority of the male donors ($\Delta RS = 0.485$) and for none of the female donors ($\Delta RS = 0.172$). These results suggest that certain donors are able to provide more stable RBCs and that these donors are more likely to be female than male.

3.7 | Hematological parameters over 6 weeks of storage

We monitored standard hematological parameters (Figure 5) of cold stored RBC units including MCV, RDW, MCH, and MCHC. Overall, the hematological parameters stayed within the normal range (Figure 5, grey shaded area), with the notable exception of MCHC levels, which dropped slightly below accepted values of 315–355 g/L (Medical Council of Canada reference values [50]) at the 6-week expiration date. We related the general hematological data for each sample to the deformability of the matching RBCs at week 0 and at week 6 of storage. We found a slight positive correlation between increase in MCV and increase in the RS over time for male donors ($r = 0.7504$), but not for female donors. Male donors also showed a slight negative correla-

tion between RS and MCHC ($r = -0.6373$). There were no correlations between any other parameters and changes in deformability.

3.8 | Hemolysis levels in blood bags

We also measured hemolysis at all-time points for stored RBC units (Figure 6). In Canada, the maximum allowable hemolysis is 0.8% at the time of expiry (6 weeks). We found that the majority of RBC samples did not show hemolysis above the standard safety level of 0.8% until week 8. The exceptions were donor 9, which had 1.15% hemolysis at week 0 and donor 13, which had 1.11% hemolysis at week 6. By week 8, half of the donor bags ($n = 3$ male, $n = 2$ female) were above the acceptable hemolysis threshold of 0.8%.

4 | DISCUSSION

In this study, we investigated the progressive loss of RBCs deformability under standard cold storage conditions. Using the microfluidic ratchet device, we sorted RBCs into 12 fractions based on deformability and derived an RS based on the distribution of RBCs within these fractions. RSs were obtained in freshly donated blood bags and over 8 weeks of storage, which is 2 weeks longer than the standard storage window. We observed consistent loss of RBCs deformability during storage but the rate and magnitude of this loss was donor-specific and was not predicted based on the deformability of freshly donated RBCs. After 6 weeks of storage, RBCs from both male and female donors converged to a similar deformability. However, from weeks 6 to 8, the RBC deformability loss accelerated dramatically for male donors but not for female donors. Together, these results demonstrate how RBC deformability could be used as a potential biomarker for the quality of donated RBC units, as well as how different storage window may be appropriate for certain donors. Investigating these questions further will require larger studies with greater numbers of RBC units.

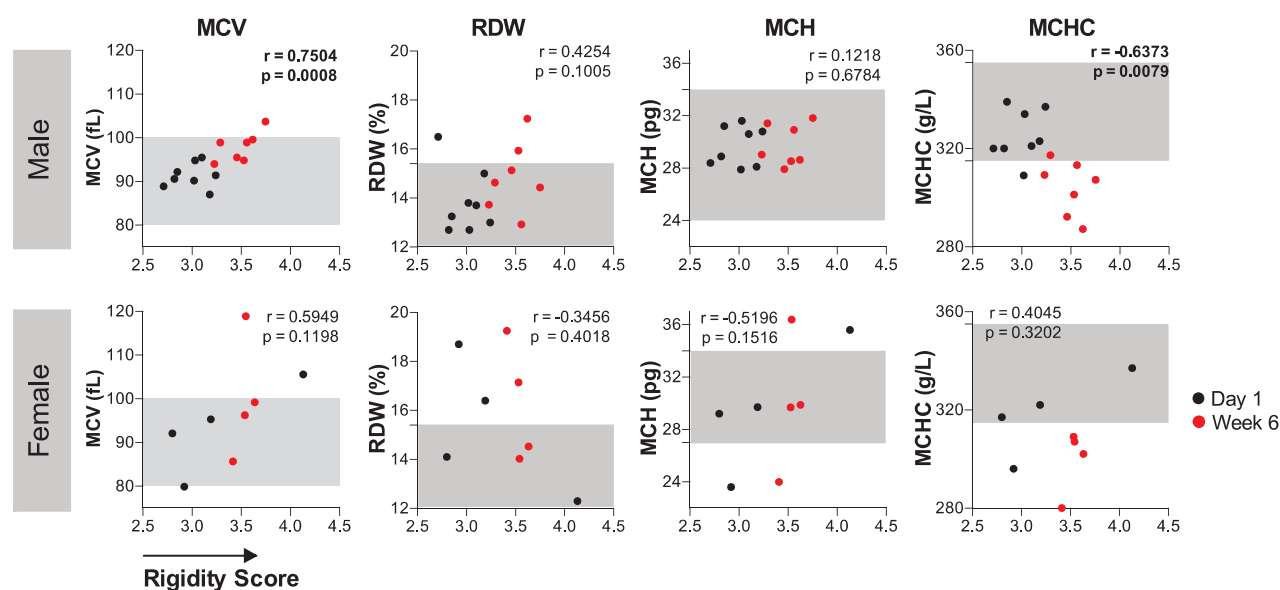


FIGURE 5 Correlation between hematological parameters and rigidity score (RS). Male donors showed minor correlation between rigidity score (RS) and mean corpuscular volume (MCV) ($r = 0.7504$; $p = 0.0008$), as well as between RS and mean corpuscular hemoglobin concentration (MCHC) ($r = -0.6373$; $p = 0.0079$). Female donors showed no correlations between RS and hematological parameter. Greyed areas indicate Medical Council of Canada reference values

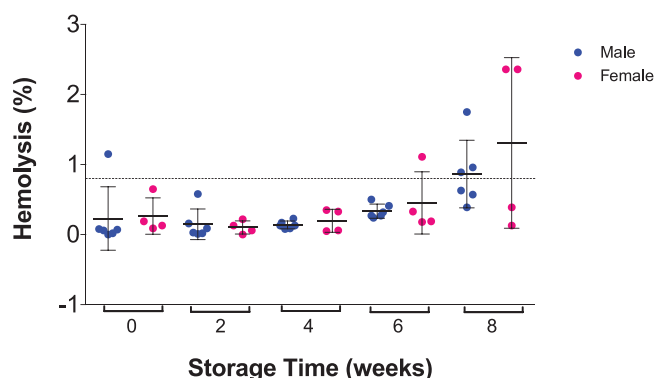


FIGURE 6 Hemolysis during storage in bags for male and female donors. The majority of donor blood bags did not show hemolysis above the 0.8% threshold, except for one male donor at week 0 and female donor at week 6. Half of all the blood bags ($n = 3$ male and $n = 2$ female) showed hemolysis above the 0.8% threshold at 8 week

The observed differences between male and female donors in their RBC deformability loss profiles are consistent with other differences between male and female blood. For example, RBCs from males have been shown to be smaller in size, as well as greater in hematocrit, MCV, hemoglobin concentration, viscosity, and RBC fragility compared to RBCs from females [42]. Some of these differences have been attributed to the female sex hormone estrogen, which has been shown to protect RBCs from deformability loss [43] but also has a major impact on the regulation of erythropoiesis [44]. Furthermore, difference in distribution and function of estrogen receptors on RBCs [45], as well as differences in serum estradiol concentration may [46] affect

intracellular signaling and better protect against oxidative stress in female RBCs. These differences could collectively explain the accelerated deformability loss observed for RBCs from male donors upon storage past the 42-day storage window.

Profiling the loss of RBC deformability for individual donors could serve to guide the selection of blood units prior to transfusion. It is well-established that clinical efficacy of RBC units in blood transfusions declines with the age of the blood bag [33–35,37,38] and that the loss of RBC deformability corresponds with this decline in clinical efficacy [47]. This study demonstrates that loss of RBC deformability can be profiled over the course of storage. We observed that RBC deformability was generally preserved during the 42-day storage expiration window. However, beyond the 42-day storage expiration window, RBC deformability loss was accelerated and varied significantly between donors. Donor-specific variability in deformability loss of stored RBCs is consistent with previous reports [5,36,39] and may strongly impact post-transfusion outcomes [32,51,52]. Additionally, there is evolving evidence that the quality of stored RBCs may be defined by sub-population of cells with low deformability. Our deformability-based cell sorting approach is well-positioned to investigate these hypotheses by more carefully examining the less deformable sub-populations. Consequently, deformability-based sorting of RBCs could be a valuable tool for investigating challenges associated with RBC product quality.

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CONFLICT OF INTEREST

Hongshen Ma is listed as inventors on a patent related to this work.

AUTHOR CONTRIBUTIONS

Hongshen Ma supervised the study. Hongshen Ma, Emel Islamzada, and Kerry Matthews conceived the idea. Emel Islamzada, Kerry Matthews, and Emel Islamzada performed the experimental work. All authors wrote the manuscript.

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REFERENCES

- Dumont LJ, Aubuchon JP. Evaluation of proposed FDA criteria for the evaluation of radiolabeled red cell recovery trials. *Transfusion* 2008; 48(6):1053–60. <https://doi.org/10.1111/j.1537-2995.2008.01642.x>
- Bratosin D, Estaquier J, Ameisen JC, Montreuil J. Molecular and cellular mechanisms of erythrocyte programmed cell death: impact on blood transfusion. *Vox Sanguinis*. 2002;83:307.
- Bennett-Guerrero E, Veldman TH, Doctor A, Telen MJ, Ortel TL, Reid TS, et al. Evolution of adverse changes in stored RBCs. *Proc Natl Acad Sci U S A*. 2007;104(43):17063–8. <https://doi.org/10.1073/pnas.0708160104>
- D'Alessandro A, Liumbruno G, Grazzini G, Zolla L. Red blood cell storage: the story so far. *Blood Transfus*. 2010;8(2):82–8. <https://doi.org/10.2450/2009.0122-09>
- Matthews K, Myrand-Lapierre M-E, Ang RR, Duffy SP, Scott MD, Ma H. Microfluidic deformability analysis of the red cell storage lesion. *J Biomech*. 2015;48(15):4065–72. <https://doi.org/10.1016/j.jbiomech.2015.10.002>
- Tinmouth A, Fergusson D, Yee IC, Hébert PC. Clinical consequences of red cell storage in the critically ill. *Transfusion* 2006;46(11):2014–27.
- Yoshida T, Prudent M, D'Alessandro A. Red blood cell storage lesion: causes and potential clinical consequences. *Blood Transfus*. 2019;17(1):27–52. <https://doi.org/10.2450/2019.0217-18>
- Höglman CF, Meryman HT. Red blood cells intended for transfusion: quality criteria revisited. *Transfusion* 2006;46(1):137–42. <https://doi.org/10.1111/j.1537-2995.2006.00681.x>
- Pavenski K, Saldenberg E, Lavoie M, Tokessy M, Branch DR. Red blood cell storage lesions and related transfusion issues: a Canadian blood services research and development symposium. *Transfus Med Rev*. 2012;26(1):68–84. <https://doi.org/10.1016/j.tmr.2011.07.003>
- Franco RS. Measurement of red cell lifespan and aging. *Transfus Med Hemother*. 2012;39(5):302–7. <https://doi.org/10.1159/000342232>
- Cohen RM, Franco RS, Khera PK, Smith EP, Lindsell CJ, Ciralo PJ, et al. Red cell life span heterogeneity in hematologically normal people is sufficient to alter HbA1c. *Blood* 2008;112(10):4284–91. <https://doi.org/10.1182/blood-2008-04-154112>
- Remy KE, Spinella PC. Red blood cell storage age - what we know from clinical trials. *Expert Rev Hematol*. 2016;9(11):1011–3. <https://doi.org/10.1080/17474086.2016.1243051>
- Berezina TL, Zaets SB, Morgan C, Spillert CR, Kamiyama M, Spolarics Z, et al. Influence of storage on red blood cell rheological properties. *J Surg Res*. 2002;102(1):6–12. <https://doi.org/10.1006/jsre.2001.6306>
- Card RT, Mohandas N, Mollison PL. Relationship of post-transfusion viability to deformability of stored red cells. *Br J Haematol*. 1983; 53(2):237–40. <https://doi.org/10.1111/j.1365-2141.1983.tb02016.x>
- Huruta RR, Barjas-Castro ML, Saad STO, Costa FF, Fontes A, Barbosa LC, et al. Mechanical properties of stored red blood cells using optical tweezers. *Blood* 1998;92(8):2975–7.
- Nagaprasad V, Singh M. Sequential analysis of the influence of blood storage on aggregation, deformability and shape parameters of erythrocytes. *Clin Hemorheol Microcirc*. 1998;18(4):273–84.
- Ng MSY, David M, Middelburg RA, Ng ASY, Suen JY, Tung J-P, et al. Transfusion of packed red blood cells at the end of shelf life is associated with increased risk of mortality - a pooled patient data analysis of 16 observational trials. *Haematologica* 2018;103(9):1542–8. <https://doi.org/10.3324/haematol.2018.191932>
- Goel R, Johnson DJ, Scott AV, Tobian AAR, Ness PM, Nagababu E, et al. Red blood cells stored 35 days or more are associated with adverse outcomes in high-risk patients. *Transfusion* 2016;56(7):1690–8. <https://doi.org/10.1111/trf.13559>
- Bratosin D, Mazurier J, Tissier JP, Estaquier J, Huart JJ, Ameisen JC, et al. Cellular and molecular mechanisms of senescent erythrocyte phagocytosis by macrophages. A review. *Biochimie* 1998;80(2):173–95. [https://doi.org/10.1016/s0300-9084\(98\)80024-2](https://doi.org/10.1016/s0300-9084(98)80024-2)
- Chin-Yee I, Stachuk L, Milkovich S, Xenocostas A, Ellis C. Transfusion of red blood cells under shock conditions in the rat microvasculature. *Blood* 2004;104:2713.
- Reid HL, Barnes AJ, Lock PJ, Dormandy JA, Dormandy TL. A simple method for measuring erythrocyte deformability. *J Clin Pathol*. 1976;29(9):855–8.
- Selvan R, Parthasarathi P, Iyengar SS, Ananthamurthy S, Bhat-tacharya S. Estimation of membrane bending modulus of stiffness tuned human red blood cells from micropore filtration studies. *PLoS One*. 2019;14(12):e0226640. <https://doi.org/10.1371/journal.pone.0226640>
- Wen Z, Gao T, Yan Z, Song L, Dou H, Sun D, et al. Biophysical meanings of orientation and deformation of RBCs in shear flow field of low viscosity with new ektacytometry. *Sci China*. 1998;41(2):195–202. <https://doi.org/10.1007/BF02882727>
- Baskurt OK, Hardeman MR, Uyuklu M, Ulker P, Cengiz M, Nemeth N, et al. Comparison of three commercially available ektacytometers with different shearing geometries. *Biorheology* 2009;46(3):251–64. <https://doi.org/10.3233/BIR-2009-0536>
- Dondorp AM, Kager PA, Vreeken J, White NJ. Abnormal blood flow and red blood cell deformability in severe malaria. *Parasitol Today*. 2000;16(6):228–32.
- Evans E, Mohandas N, Leung A. Static and dynamic rigidities of normal and sickle erythrocytes. Major influence of cell hemoglobin concentration. *J Clin Invest*. 1984;73(2):477–88. <https://doi.org/10.1172/JCI111234>
- Nash GB, O'Brien E, Gordon-Smith EC, Dormandy JA. Abnormalities in the mechanical properties of red blood cells caused by Plasmodium falciparum. *Blood* 1989;74(2):855–61.
- Hebbel RP, Leung A, Mohandas N. Oxidation-induced changes in microrheologic properties of the red blood cell membrane. *Blood* 1990;76(5):1015–20.
- Mills JP, Qie L, Dao M, Lim CT, Suresh S. Nonlinear elastic and viscoelastic deformation of the human red blood cell with optical tweezers. *Mech Chem biosyst*. 2004;1(3):169–80.

30. Dao M, Lim CT, Suresh S. Mechanics of the human red blood cell deformed by optical tweezers. *J Mech Phys Solids*. 2003;51(11–12):2259–80. <https://doi.org/10.1016/j.jmps.2003.09.019>
31. Hénon S, Lenormand G, Richert A, Gallet F. A new determination of the shear modulus of the human erythrocyte membrane using optical tweezers. *Biophys J*. 1999;76(2):1145–51. [https://doi.org/10.1016/S0006-3495\(99\)77279-6](https://doi.org/10.1016/S0006-3495(99)77279-6)
32. Barshtein G, Rasmusen TL, Zelig O, Arbell D, Yedgar S. Inter-donor variability in deformability of red blood cells in blood units. *Transfus Med*. 2020;30(6):492–6. <https://doi.org/10.1111/tme.12725>
33. Dhabangi A, Ainomugisha B, Cserti-Gazdewich C, Ddungu H, Kyeyune D, Musisi E, et al. Effect of transfusion of red blood cells with longer vs shorter storage duration on elevated blood lactate levels in children with severe anemia: the TOTAL randomized clinical trial. *JAMA*. 2015;314(23):2514. <https://doi.org/10.1001/jama.2015.13977>
34. Fergusson DA, Hébert P, Hogan DL, Lebel L, Rouvinez-Bouali N, Smyth JA, et al. Effect of fresh red blood cell transfusions on clinical outcomes in premature, very low-birth-weight infants: the ARIPI randomized trial. *JAMA*. 2012;308(14):1443–51. <https://doi.org/10.1001/2012.jama.11953>
35. Heddle NM, Cook RJ, Arnold DM, Liu Y, Barty R, Crowther MA, et al. Effect of short-term vs. long-term blood storage on mortality after transfusion. *N Engl J Med*. 2016;375(20):1937–45. <https://doi.org/10.1056/NEJMoa1609014>
36. Islamzada E, Matthews K, Guo Q, Santoso AT, Duffy SP, Scott MD, et al. Deformability based sorting of stored red blood cells reveals donor-dependent aging curves. *Lab Chip*. 2020;20(2):226–35. <https://doi.org/10.1039/C9LC01058K>
37. Lacroix J, Hébert PC, Fergusson DA, Tinmouth A, Cook DJ, Marshall JC, et al. Age of transfused blood in critically ill adults. *N Engl J Med*. 2015;372(15):1410–8. <https://doi.org/10.1056/NEJMoa1500704>
38. Steiner ME, Ness PM, Assmann SF, Triulzi DJ, Sloan SR, Delaney M, et al. Effects of red-cell storage duration on patients undergoing cardiac surgery. *N Engl J Med*. 2015;372(15):1419–29. <https://doi.org/10.1056/NEJMoa1414219>
39. Tzounakas VL, Georgatzakou HT, Kriebardis AG, Voulgaridou AI, Stamoulis KE, Foudoulaki-Papazizos LE, et al. Donor variation effect on red blood cell storage lesion: a multivariable, yet consistent, story. *Transfusion* 2016;56(6):1274–86. <https://doi.org/10.1111/trf.13582>
40. Dern RJ, Gwinn RP, Wiorkowski JJ. Studies on the preservation of human blood. I. Variability in erythrocyte storage characteristics among healthy donors. *J Lab Clin Med*. 1966;67(6):955–65.
41. Hess JR, Sparrow RL, Van Der Meer PF, Acker JP, Cardigan RA, Devine DV. Red blood cell hemolysis during blood bank storage: using national quality management data to answer basic scientific questions. *Transfusion* 2009;49(12):2599–603. <https://doi.org/10.1111/j.1537-2995.2009.02275.x>
42. Jordan A, Chen D, Yi Q-L, Kaniyas T, Gladwin MT, Acker JP. Assessing the influence of component processing and donor characteristics on quality of red cell concentrates using quality control data. *Vox Sang*. 2016;111(1):8–15. <https://doi.org/10.1111/vox.12378>
43. Mcateer MJ, Dumont LJ, Cancelas J, Rugg N, Vassallo R, Whitley P, et al. Multi-institutional randomized control study of haemolysis in stored red cell units prepared manually or by an automated system. *Vox Sang*. 2010;99(1):34–43. <https://doi.org/10.1111/j.1423-0410.2010.01313.x>
44. Acker JP, Hansen AL, Kurach JDR, Turner TR, Croteau I, Jenkins C. A quality monitoring program for red blood cell components: in vitro quality indicators before and after implementation of semiautomated processing. *Transfusion* 2014;54(10):2534–43. <https://doi.org/10.1111/trf.12679>
45. Han V, Serrano K, Devine DV. A comparative study of common techniques used to measure haemolysis in stored red cell concentrates. *Vox Sanguinis*. 2010;98(2):116–23. <https://doi.org/10.1111/j.1423-0410.2009.01249.x>
46. Guo Q, Duffy SP, Matthews K, Deng X, Santoso AT, Islamzada E, et al. Deformability based sorting of red blood cells improves diagnostic sensitivity for malaria caused by *Plasmodium falciparum*. *Lab Chip*. 2016;16(4):645–54. <https://doi.org/10.1039/C5LC01248A>
47. Guo Q, Duffy SP, Matthews K, Islamzada E, Ma H. Deformability based cell sorting using microfluidic ratchets enabling phenotypic separation of leukocytes directly from whole blood. *Sci Rep*. 2017;7(1):6627. <https://doi.org/10.1038/s41598-017-06865-x>
48. Henkelman S, Dijkstra-Tiekstra MJ, De Wildt-Eggen J, Graaff R, Rakhorst G, Van Oeveren W. Is red blood cell rheology preserved during routine blood bank storage? *Transfusion* 2010;50(4):941–8. <https://doi.org/10.1111/j.1537-2995.2009.02521.x>
49. Gifford SC, Derganc J, Shevkopyas SS, Yoshida T, Bitensky MW. A detailed study of time-dependent changes in human red blood cells: from reticulocyte maturation to erythrocyte senescence. *Brit J Haematol*. 2006;135(3):395–404. <https://doi.org/10.1111/j.1365-2141.2006.06279.x>
50. Medical Council of Canada. Normal values Available from: <https://mcc.ca/objectives/normal-values/>. Accessed 21 June 2021.
51. Chassé M, McIntyre L, English SW, Tinmouth A, Knoll G, Wolfe D, et al. Effect of blood donor characteristics on transfusion outcomes: a systematic review and meta-analysis. *Transfus Med Rev*. 2016;30(2):69–80. <https://doi.org/10.1016/j.tmr.2016.01.002>
52. Heddle NM, Cook RJ, Liu Y, Zeller M, Barty R, Acker JP, et al. The association between blood donor sex and age and transfusion recipient mortality: an exploratory analysis. *Transfusion* 2019;59(2):482–91. <https://doi.org/10.1111/trf.15011>

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