

PERSPECTIVE AND DIRECTIONS FOR FUTURE STUDIES

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27.1 INTRODUCTION

Oxidative stress, an unavoidable consequence of life in an aerobic oxygen-enriched atmosphere, is a cytotoxic process that occurs in cells when antioxidant mechanisms are overwhelmed by reactive oxygen species (ROS). This imbalance causes damage to important biomolecules and cells, with potential impact on the whole organism [1]. ROS are atoms or molecules possessing one or more unpaired electrons in the outer orbit and, therefore, are prone to react chemically [2]. ROS include superoxide anions ($O_2^{\bullet-}$), hydroxyl ($\bullet OH$), alkoxyl (RO^{\bullet}), and peroxy radicals (ROO^{\bullet}), and hydrogen peroxide (H_2O_2). These radicals are common products of life in an aerobic environment, and they are responsible for oxygen toxicity. The “dark side” of oxygen is related to the fact that each oxygen atom has one unpaired electron in its outer valence shell, and molecular oxygen has two unpaired electrons. Thus atomic oxygen is a free radical, and molecular oxygen is a (free) biradical. Concerted tetravalent reduction of oxygen by the mitochondrial electron transport chain, to form water, is considered a relatively safe process; however, the univalent reduction of oxygen generates reactive intermediates. Thus the “oxygen paradox” is dangerous to all forms of life for whom it is an essential component of energy production [3]. The major sources of ROS include mitochondrial respiratory chain, xanthine/xanthine oxidase, myeloperoxidase in cytoplasm, uncontrolled

arachidonic acid (ARA) cascade, and NADPH oxidase (Fig. 27.1). Over 90% of ROS production occurs “accidentally” in mitochondria during metabolism of oxygen when some of electrons passing “down” the electron transport chain leak away from the main path and go directly to reduce oxygen molecules to the superoxide anion [4]. NADPH oxidase generates superoxide radical by the one-electron reduction of oxygen, using NADPH as the electron donor [5, 6]. The ability of NADPH oxidase inhibitors to retard ROS-mediated cytotoxicity provides strong support for the idea that ROS are generated through the activation of NADPH oxidase. In the presence of metal ions such as Fe^{2+} and Cu^{2+} , H_2O_2 is also transformed into hydroxyl radical ($\bullet OH$) through the Fenton reaction (Fig. 27.1). Hydroxyl radicals can attack polyunsaturated fatty acids in neural membrane phospholipids, forming the peroxy radical (ROO^{\bullet}), and then propagate the chain reaction of lipid peroxidation. ROS production plays an important role in cell signaling. ROS modulate the transcription factor NF- κ B through the activation of kinases that phosphorylate the inhibitory subunit of NF- κ B/I- κ B, causing its ubiquitination and release of NF- κ B from the NF- κ B complex. Free NF- κ B migrates from cytosol to the nucleus, where it binds to the κ B domain of the target gene promoter, leading to transcriptional activation of many proinflammatory enzymes, cytokines (TNF- α , IL-1 β , and IL-10), chemokines, immune receptors, and cell surface adhesion molecules (Fig. 27.1) [7], which are

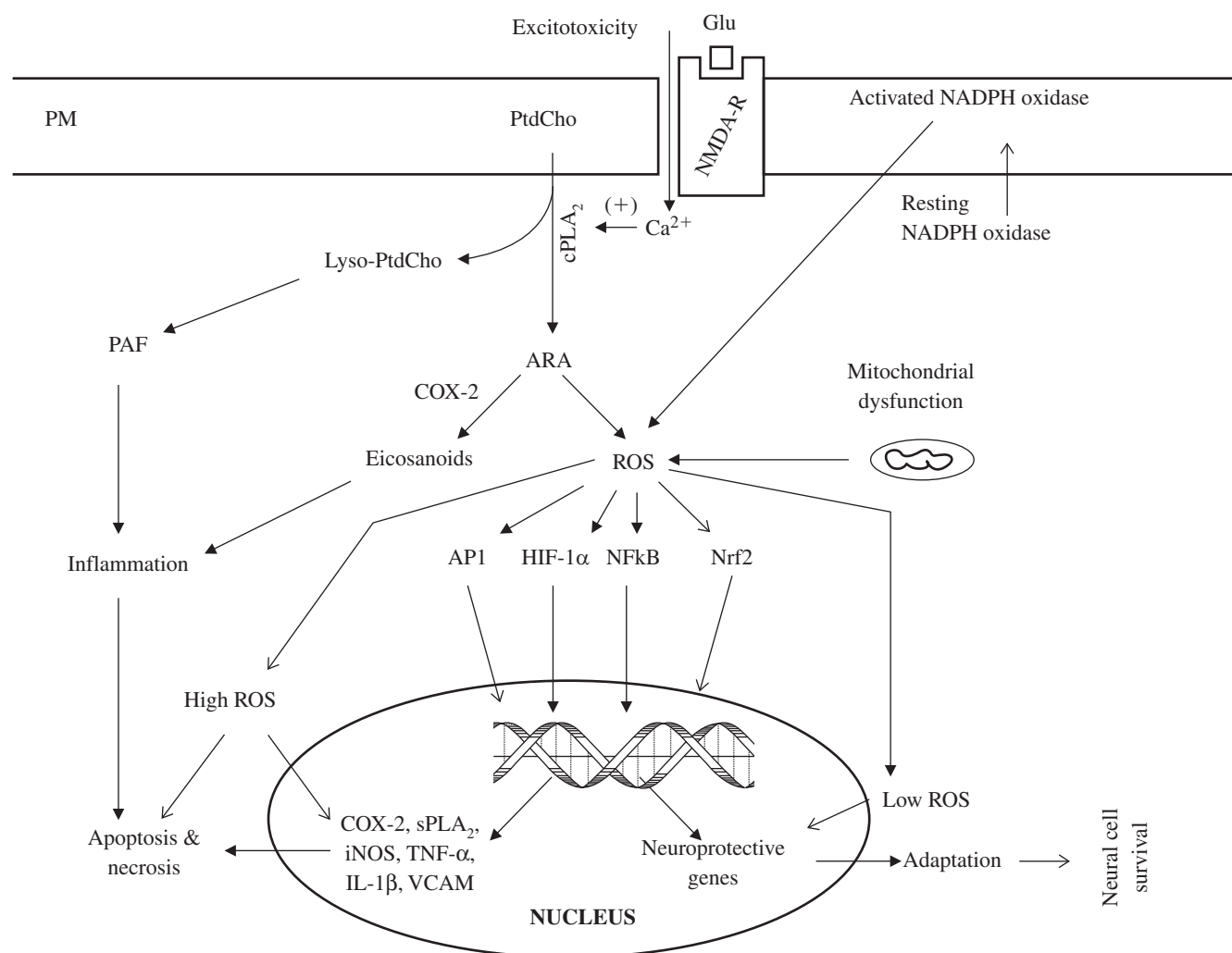


Fig. 27.1 Hypothetical model showing generation of reactive oxygen species (ROS) from various sources and effect of ROS on various transcription factors. PM, plasma membrane; NMDA-R, *N*-methyl-D-aspartate receptor; Glu, glutamate; PtdCho, phosphatidylcholine; lyso-PtdCho, lyso-phosphatidylcholine; cPLA₂, cytosolic phospholipase A₂; sPLA₂, secretory phospholipase A₂; COX-2, cyclooxygenase; ARA, arachidonic acid; NF-κB, nuclear factor κB; iNOS, inducible nitric oxide synthase; PAF, platelet-activating factor; VCAM-1, vascular cell adhesion molecule-1; AP-1, activator protein 1; HIF-1α, hypoxia-inducible transcription factor-1α; Nrf2, NF-E2-related factor 2.

involved in different cellular processes such as cell proliferation, survival, stress responses, cellular immunity, and inflammation. Dysregulation of the NF-κB pathway leads to diseases such as autoimmunity and cancer.

Although chemical and biochemical reactions for the production of ROS in vertebrates and invertebrates systems are similar, cellular heterogeneity in various tissues and organs makes the intensity of oxidative stress responses very different. The consequences of oxidative stress depend not only on cell type and nature of ROS but also on endogenous antioxidant status and cooperation among various antioxidant systems [3]. In general, the preservation of redox status is a major factor for cell survival. Brains from vertebrate and invertebrate

phyla may respond differently in terms of intensity of oxidative stress compared to visceral organs of vertebrates (liver, kidneys, and lung) and similar systems in invertebrates. The mammalian brain demands high energy and possess highly active mitochondrial metabolism with high oxygen utilization (20% of the total oxygen inspired). This high utilization of oxygen comes at a heavy biological price. As important as oxygen is for the survival of neurons and glia, it also indirectly contributes to their destruction and death over time. The reason for this is that a small percentage (an estimated 1–4%) of the oxygen that enters cells is metabolized to derivatives that gradually erode and destroy essential molecules [8]. These destructive derivatives of oxygen are often referred to

as free radicals or ROS. Brain not only possesses high levels of polyunsaturated fatty acids but also contains transition metals such as iron and copper, which are capable of generating hydroxyl radical [9]. In addition, brain contains low levels of cytosolic antioxidants compared to liver and other visceral tissues [10, 11]. Thus brain is unable to protect itself from toxic effects of high levels of ROS. Superoxide radical reacts with nitric oxide (NO^\bullet) to produce the peroxynitrite anion (ONOO^-), a nonradical product, which is as toxic as $\bullet\text{OH}$ in terms of its ability to oxidize and destroy by standard molecules. NO^\bullet and ONOO^- are often referred to as reactive nitrogen species (RNS). Like ROS, RNS oxidize lipids and protein components. Studies on ROS and RNS indicate that ROS/RNS are highly reactive and short-lived species that do not accumulate to significant levels, and it is not possible to measure them directly; rather, one must measure either the accumulation of biomolecules or the exogenously added indicators that are modified by ROS and RNS. In other words, generation of ROS and RNS leaves its footprint in the cell in the form of different oxidatively modified components.

27.2 ENDOGENOUS ANTIOXIDANT DEFENSE MECHANISMS IN VERTEBRATES AND INVERTEBRATES

Under physiological conditions, the antioxidant defense system within vertebrate and invertebrate bodies can easily neutralize the amount of ROS produced through ROS generating systems [2]. The antioxidant systems include low-molecular-weight antioxidants like glutathione and vitamin C, antioxidant enzymes such as superoxide dismutase, catalase, transferrin, and glutathione peroxidase, and the ROS defense system involving the participation of enzymes associated with DNA and membrane repair. These enzymes repair ROS-mediated oxidative damage to cellular structures [2]. Thus during normal aerobic metabolism ROS generation remains under tight control through the activities of the above-listed antioxidant defense systems. Low levels of ROS are needed for fundamental cellular functions such as growth and adaptation responses and for optimal functioning of the immune system. Oxidative stress induces a number of biochemical changes in vertebrate and invertebrate systems. The extent of these changes depends on the severity of the oxidative stress. In vertebrate and invertebrate systems, low levels of ROS promote cell proliferation, while intermediate ROS levels produce growth arrests [2, 8]. Under a sustained environmental stress, ROS are produced over a long time, and thus significant damage may occur to cell structure and functions. This damage has been implicated in a wide variety

of chronic diseases, including neurodegenerative diseases (Alzheimer disease, Parkinson disease, amyotrophic lateral sclerosis); arteriosclerosis and heart disease; strokes and ischemia/reperfusion injury; chronic inflammatory diseases (rheumatoid arthritis, lupus erythematosus, and psoriatic arthritis); and mutagenesis, cell transformation, and cancer [12, 13].

To counteract ROS-mediated damage, vertebrate and invertebrate systems have evolved several mechanisms. These mechanisms include (a) termination of the ROS using free radical scavengers and antioxidant enzymes, (b) induction of endogenous signaling systems that protect cells from toxic effects of ROS, and (c) repair of damaged cellular components. These mechanisms may also involve the activation of redox-sensitive transcription factors and increased expression of antioxidant enzymes and antiapoptotic proteins. One of the major cellular antioxidant responses in vertebrate and invertebrate systems is the induction of detoxification enzymes through the cytoplasmic oxidative stress system (Nrf2-Keap1). It is activated by ROS (Fig. 27.1). The transcription factor Nrf2 is constitutively expressed in all tissues of vertebrates and invertebrates, although amounts may vary among various cells and organs [14, 15]. The detoxification organs (kidney and liver) have the highest levels of Nrf2. Nrf2 may be further induced by cellular stressors including endogenous ROS or exogenous electrophiles [16]. Under normal conditions, Keap1 forms a complex with Nrf2 and keeps it within the cytoplasm, targeting it for ubiquitination and proteasomal degradation. This results in the maintenance of low levels of Nrf2 that mediate the constitutive expression of Nrf2 downstream genes. When vertebrate and invertebrate cells are exposed to oxidative stress, a signal involving phosphorylation and/or redox modification of critical cysteine residues in Keap1 inhibits the enzymic activity of the Keap1-Cul3-Rbx1 E3 ubiquitin ligase complex, leading to reduction in Nrf2 ubiquitination and degradation. As a result, free Nrf2 translocates from the cytosol into nucleus, where it forms a heterodimer with members of the small musculo-aponeurotic fibrosarcoma (Maf) transcription factor family [15]. These Nrf2/Maf heterodimers bind to antioxidant response elements present in the promoters of numerous antioxidant genes, including NQO-1, glutathione *S*-transferase, glutathione peroxidase (GPx), catalase, and HO-1 [14, 17, 18]. Upon recovery of cellular redox homeostasis, Keap1 travels into the nucleus to dissociate Nrf2 from the ARE. Subsequently, the Nrf2-Keap1 complex is exported out of the nucleus by the nuclear export sequence (NES) in Keap1. Once in the cytoplasm, the Nrf2-Keap1 complex associates with the Cul3-Rbx1 core ubiquitin machinery, resulting in degradation of Nrf2 and termination of the Nrf2/ARE signaling

pathway [16, 19]. The Nrf2 signaling pathway mediates multiple avenues of cytoprotection by activating the transcription of more than 200 genes that are not only crucial in metabolism of drugs and toxins, protection against oxidative stress, and inflammation but also play an integral role in stability of proteins and in the removal of damaged proteins via proteasomal degradation or autophagy [20]. Nrf2 interacts with other important cell regulators such as tumor suppressor protein 53 (p53) and nuclear factor- κ B (NF- κ B) and through their combined interactions act as the guardian of a healthy life span, protecting against many age-related diseases such as cancer and neurodegeneration. It is hypothesized that this signaling pathway plays a critical role in the determination of species longevity and that this pathway may indeed be the master regulator of the aging process [21]. Since Nrf2-mediated cellular defense response protects multiple organs or multiple tissues, activation of Nrf2 has been implicated in conferring protection against many human diseases, including cancer, neurodegenerative diseases, cardiovascular diseases, acute and chronic lung injury, autoimmune diseases, and inflammation [18].

Oxidative stress also promotes both the transcriptional activity and protein stability of FoxOs, forkhead transcript factors that are expressed abundantly throughout the body of vertebrates and invertebrates [22–24]. FoxOs are major targets for insulin signaling. FoxO1 belongs to a nuclear protein subfamily that includes FoxO3a, FoxO4, and FoxO6 in mammals and its ortholog DAF-16 in *Caenorhabditis elegans* [25]. These proteins, characterized by a highly conserved central DNA binding domain and a carboxyl *trans*-activation domain, play important roles in mediating the effect of insulin or IGF on metabolism, growth, survival, differentiation of cells, oxidative stress response, DNA repair, cell cycle, and apoptosis [26]. Among the FoxO isoforms, FoxO1 preserves redox balance by promoting protein synthesis and subsequently inhibiting cell cycle arrest. This evidence indicates that FoxO1 integrates and orchestrates responses to different stress signals to maintain cellular function [22, 23, 26].

In addition, vitagenes play an important role in conferring protection against oxidative stress. These genes include genes for heat shock proteins (Hsps), thioredoxin/thioredoxin reductase system, and heme oxygenase-1 [27, 28]. Heat shock response contributes to establish a cytoprotective state in a wide variety of oxidative stress-mediated diseases, including inflammation, cancer, aging, and neurodegenerative disorders. When appropriately activated, heat shock response not only initiates and restores cellular homeostasis but rebalances redox equilibrium. Activation of this pathway is particularly important for neural cells with relatively weak endogenous antioxidant defenses [28, 29].

27.3 BIOMARKERS OF OXIDATIVE STRESS IN VERTEBRATES AND INVERTEBRATES

As stated above, direct measurement of ROS *in vivo* is difficult because of the highly reactive nature of these compounds and their minute concentrations in biological fluids. Instead, one relies on measurement of stable end products of oxidation of different molecules. ROS-mediated damage may occur to DNA, proteins, and lipids in vertebrate and invertebrate systems. Several markers of ROS-mediated damage have been described in the literature. An ideal biomarker for the detection of oxidative stress in chronic neurodegenerative and visceral diseases should be precise and reliable. It should be easy to quantify and be reproducible. It should not be subjected to wide variation in the general population and not affected by comorbid factors. To evaluate the effect of medication, the biomarker should change linearly with disease progression. Among biomarkers for oxidative stress, F₂-isoprostanes, 4-hydroxynonenal (4-HNE), and 8-hydroxy-2-deoxyguanosine have attracted considerable attention. F₂-isoprostanes and 4-HNE are compounds derived from arachidonic acid (ARA) via a free radical-catalyzed mechanism (Fig. 27.2). Several F₂-isoprostanes have been described in the vertebrate system, but it is becoming increasingly evident that 15-F_{2t}-isoprostane (15-F_{2t}-IsoP) is a good biomarker for oxidative stress [30]. Isoprostanes are generated from cell membrane-bound ARA by free radical attack. They are cleaved from the sites of their origin by phospholipase A₂ and then circulate in plasma and are excreted in urine [31, 32]. F₂-isoprostanes can be detected in biological fluids, such as urine, blood plasma, bronchoalveolar lavage, or cerebrospinal fluid (CSF), as well as in tissues. The main advantage of urinary measurements of F₂-isoprostanes is that the compounds are very stable and are not formed *ex vivo* [33]. On the other hand, in blood plasma and tissues, autooxidation may occur. Several methods are available for F₂-isoprostane detection including gas chromatography-mass spectroscopy and liquid chromatography-mass spectroscopy. In addition, a reliable ELISA technique suitable for analysis of large numbers of samples has recently been developed [32, 34, 35]. No information is available on the determination of isoprostanes in invertebrate systems. In future studies, attempts should be made to determine levels of isoprostanes in invertebrate systems by gas chromatography-mass spectroscopy and liquid chromatography-mass spectroscopy.

4-HNE is the most prevalent toxic lipid peroxidation product formed during oxidative stress. It is derived from nonenzymic oxidation of ARA. It not only modifies amino acid residues of proteins (sulfhydryl groups of cysteine, imidazole moiety of histidine, and the ϵ -amino

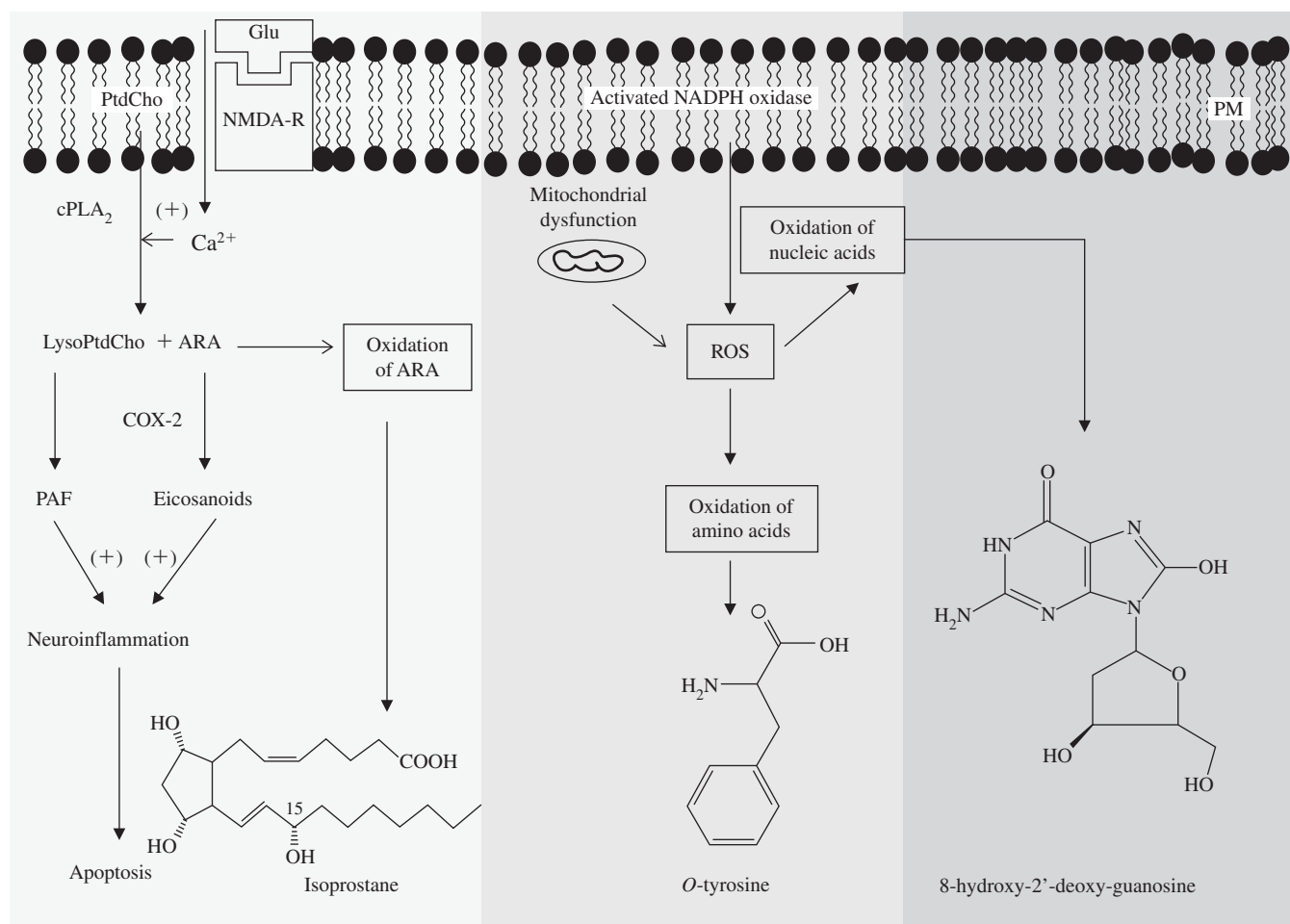


Fig. 27.2 Generation of biomarkers for oxidative stress in vertebrates. PM, plasma membrane; NMDA-R, *N*-methyl-D-aspartate receptor; Glu, glutamate; PtdCho, phosphatidylcholine; lyso-PtdCho, lyso-phosphatidylcholine; cPLA₂, cytosolic phospholipase A₂; COX-2, cyclooxygenase; ARA, arachidonic acid; PAF, platelet-activating factor; ROS, reactive oxygen species. (See color insert.)

group of lysine) but also inhibits DNA and protein synthesis and inactivates enzymes, modifies low-density lipoproteins, and modulates gene expression [36]. Mitochondrial proteins are targets of 4-HNE adduct formation following oxidative stress *in vivo* and *in vitro*. In addition, 4-HNE also inactivates the 2-oxoglutarate dehydrogenase and pyruvate dehydrogenase complexes, cytochrome *c* oxidase, and NADH-linked respiration in isolated mitochondria [37, 38]. 4-HNE produces a variety of cytotoxic effects such as the inhibition of DNA, RNA, and protein synthesis, cell cycle arrest, mitochondrial dysfunction, and apoptosis in vertebrate and invertebrate systems [39]. Thus 4-HNE is a good biomarker for oxidative stress. 4-HNE has been detected by gas chromatography-mass spectroscopy and liquid chromatography-mass spectroscopy in vertebrate systems, and studies on the use of 4-HNE as a biomarker for oxidative stress in invertebrates are beginning to appear [25].

8-Oxodeoxyguanosine (8-oxodG) is another important biomarker for oxidative stress in vertebrate and invertebrate systems [40–42]. It is synthesized *in vivo* by the attack of ROS on DNA. It is highly mutagenic, resulting in GC to TA transversions. After cleavage from DNA as a result of DNA repair, 8-oxodG is excreted in body fluids including blood, urine, and CSF of vertebrates. Another significant source of extracellular 8-oxodG may be oxidation of the nucleotide pool [43]. Urinary 8-oxodG levels are therefore considered a general biomarker of oxidative stress. Methods used for 8-oxodG detection include high-performance liquid chromatography, tandem mass spectroscopy, and a recently developed competitive ELISA [44]. These methods can be used for the determination of oxidative stress-mediated DNA damage in invertebrate systems.

Proteomics can be used for determining levels of not only oxidative/nitrosative protein modifications but also

protein-bound methionine oxidation [45, 46]. It is controversial whether ROS- and RNS-mediated protein modifications and oxidized methionines have a significant direct physiological and pathological impact on cellular injury. The generation of these proteins may be a secondary phenomenon. A clear delineation of the causal connections cannot be provided at present, but it is becoming increasingly evident that high levels of ROS/RNS produce distinct pathological cell consequences that greatly amplify and propagate injury, leading to irreversible cell and tissue degeneration [45, 46]. It is tempting to speculate that redox proteomics can be used to define redox molecular mechanisms associated with oxidative stress in vertebrate and invertebrate systems.

27.4 OXIDATIVE STRESS AND AGING

Although information on the effect of oxidative stress in vertebrate and invertebrate systems has been discussed by Rizvi and Pandey in this book, it is important to emphasize that production of ROS has a major impact on cell aging and tissue damage [47, 48], particularly in cardiovascular and nervous systems [49]. Levels of ROS are elevated with age, as biomarkers of lipid peroxidation (isoprostanes) are increased and antioxidant activity is decreased. However, in elderly individuals who are still healthy, the ROS level has been reported to be similar to that of young adults [50], or at least comparable to antioxidant defenses [51], supporting the view that markers of oxidative stress are not influenced by old age when good health and nutritional status are preserved. In addition, psychological stress and lifestyle factors such as smoking, lack of exercise, and status of n-3 fatty acids in the body have an impact on the level of ROS [52–54]. ROS are not only responsible for whole body accelerated aging but also for decline in cognitive functional. Thus in an elderly population (>80 years old), free radicals are involved in poorer cognitive function, loss of autonomy, loss of ability to perform daily activities, and institutionalization, as well as depressive symptoms [55].

27.5 CONCLUSION

Oxidative stress is defined as an imbalance between ROS production and their removal by antioxidant systems with increased accumulation of free radicals. Major production of ROS occurs in mitochondria during oxygen utilization. In addition, ROS may be synthesized in phagocytic cells, as well as in vascular wall and various other tissues by enzymes such as NADPH oxidase, myeloperoxidase, xanthine oxidase, cyclooxygenase, and lipoxygenase. At low concentrations, ROS are

associated with a vast array of physiological functions, such as gene expression and immune responses. At high levels, ROS react with lipids, carbohydrates, proteins, and DNA, altering their structure and function and resulting in inflammation, apoptosis, and mutagenesis.

These days, we have been empowered by lipidomics, proteomics, and genomics. These procedures can be used not only for identifying and determining levels of biomarkers for oxidative stress in various tissues of vertebrate animals and invertebrate organisms but also for detecting biomarkers levels in biological fluid samples. Identification of biomarkers for oxidative stress may lead not only to early diagnosis and follow-up of the progression of neurodegenerative and chronic visceral diseases but also to monitoring of therapeutic responses. Identification of ROS/RNS-modified proteins by proteomics in vertebrates and invertebrates can provide important information on cellular function of modified proteins. Another important challenge for future studies will be to incorporate this knowledge into a framework whereby these complex functional and regulatory alterations in the cellular proteome can be used to increase our understanding of and improve the treatment of diseases with the component of oxidative stress in vertebrate and invertebrate systems.

REFERENCES

1. Duracková Z. Some current insights into oxidative stress. *Physiol Res* 2010; 59: 459–469.
2. Halliwell B. Oxidative stress and neurodegeneration: where are we now? *J Neurochem* 2006; 97: 1634–1658.
3. Davies K.J. Oxidative stress, antioxidant defenses, and damage removal, repair, and replacement systems. *IUBMB Life* 2000; 50: 279–289.
4. Pieczenik S.R. and Neustadt J. Mitochondrial dysfunction and molecular pathways of disease. *Exp Mol Pathol* 2007; 83: 84–92.
5. Dworakowski R., Anilkumar N., Zhang M., and Shah A. M. Redox signalling involving NADPH oxidase-derived reactive oxygen species. *Biochem Soc Trans* 2006; 34: 960–964.
6. Sun G.Y., Horrocks L.A., and Farooqui A.A. The roles of NADPH oxidase and phospholipases A₂ in oxidative and inflammatory responses in neurodegenerative diseases. *J Neurochem* 2006; 103: 1–16.
7. Li X. and Stark G.R. NFkappaB-dependent signaling pathways. *Exp. Hematol* 2002; 30: 285–296.
8. Halliwell B. Biochemistry of oxidative stress. *Biochem Soc Trans* 2007; 35: 1147–1150.
9. Floyd R.A., and Hensley K. Oxidative stress in brain aging. Implications for therapeutics of neurodegenerative diseases. *Neurobiol. Aging* 2002; 23: 795–807.

10. Droge W. Oxidative stress and aging. *Adv Exp Med Biol* 2003; 543: 191–200.
11. Reiter R.J. Oxidative processes and antioxidative defense mechanisms in the aging brain. *FASEB J* 1995; 9: 526–533.
12. Emerit J, Edeas M, Bricaire F. Neurodegenerative diseases and oxidative stress. *Biomed Pharmacother* 2004; 58: 39–46.
13. Reuter S., Gupta S.C., Chaturvedi M.M., and Aggarwal B.B. Oxidative stress, inflammation, and cancer: how are they linked? *Free Radic Biol Med* 2010; 49: 1603–1616.
14. Motohashi H., and Yamamoto M. Nrf2-Keap1 defines a physiologically important stress response mechanism. *Trends Mol Med* 2004; 10: 549–557.
15. Wakabayashi N., Slocum S.L., Skoko J.J., Shin S., and Kensler T.W. When NRF2 talks, who's listening? *Antioxid Redox Signal* 2010; 13: 1649–1663.
16. Nguyen T., Yang C.S., and Pickett C.B. The pathways and molecular mechanisms regulating Nrf2 activation in response to chemical stress. *Free Radic Biol Med* 2004; 37: 433–441.
17. Kobayashi M., and Yamamoto M. Nrf2-Keap1 regulation of cellular defense mechanisms against electrophiles and reactive oxygen species. *Adv Enzyme Regul* 2006; 46: 113–140.
18. Zhang D.D. Mechanistic studies of the Nrf2-Keap1 signaling pathway. *Drug Metab Rev* 2006; 38: 769–781.
19. Giudice A., Arra C., and Turco M.C. Review of molecular mechanisms involved in the activation of the Nrf2-ARE signaling pathway by chemopreventive agents. *Methods Mol Biol* 2010; 647: 37–74.
20. Hirotsu Y., Katsuoka F., Itoh K., and Yamamoto M. Nrf2 degon-fused reporter system: a new tool for specific evaluation of Nrf2 inducers. *Genes Cells* 2011; 16: 406–415.
21. Lewis K.N., Mele J., Hayes J.D., and Buffenstein R. Nrf2, a Guardian of Healthspan and Gatekeeper of Species Longevity. *Integr Comp Biol* 2010; 50: 829–843.
22. Kaestner K.H., Knochel W., and Martinez D.E. Unified nomenclature for the winged helix/forkhead transcription factors. *Gene Dev* 2000; 14: 142–146.
23. Nakae J., Barr V., and Accili D. Differential regulation of gene expression by insulin and IGF-1 receptors correlates with phosphorylation of a single amino acid residue in the forkhead transcription factor FKHR. *EMBO J* 2000; 19: 989–996.
24. Zheng J., Mutcherson R. 2nd, and Helfand S.L. Calorie restriction delays lipid oxidative damage in *Drosophila melanogaster*. *Aging Cell* 2005; 4: 209–216.
25. Lin K., Dorman J.B., Rodan A., and Kenyon C. Daf-16: an HNF-3/Forkhead family member that can function to double the life-span of *Caenorhabditis elegans*. *Science* 1997; 278: 1319.
26. Accili D., and Arden K.C. FoxOs at the crossroads of cellular metabolism, differentiation, and transformation. *Cell* 2004; 117: 421–426.
27. Calabrese V., Mancuso C., Sapienza M., Puleo E., Calafato S., Cornelius C., Finocchiaro M., Mangiameli A., Di Mauro M., Giuffrida A., and Castellino P. Oxidative stress, and cellular stress response in diabetic nephropathy. *Cell Stress Chaperons* 2007; 12: 299–306.
28. Calabrese V., Signorile A., Cornelius C., Mancuso C., Scapagnini G., Ventimiglia B., Ragusa N., and Dinkova-Kostova A. (). Practical approaches to investigate redox regulation of heat shock protein expression and intracellular glutathione redox state. *Methods Enzymol* 2008; 441: 83–110.
29. Calabrese V., Cornelius C., Dinkova-Kostova A.T., Calabrese E.J., and Mattson M.P. Cellular stress responses, the hormesis paradigm and vitagenes: novel targets for therapeutic intervention in neurodegenerative disorders. *Antioxid Redox Signal* 2010; 610: 285–308.
30. Cracowski J.L., Durand T., and Bessard G. Isoprostanes as a biomarker of lipid peroxidation in humans: physiology, pharmacology and clinical applications. *Trends Pharmacol Sci* 2002; 23: 360–366.
31. Morrow J.D., Awad J.A., Boss H.J., Blair I.A., and Roberts L. J. II. Non-cyclooxygenase-derived prostanoids (F₂-isoprostanes) are formed *in situ* on phospholipids. *Proc Natl Acad Sci USA* 1992; 89: 10721–10725.
32. Morrow J.D. The isoprostanes: their quantification as an index of oxidant stress status in vivo. *Drug Metab Rev* 2000; 32: 377–385.
33. Pratico D., Barry O.P., Lawson J.A., Adiyaman M., Hwang S.W., Khanapure S.P., Iuliano L., Rokach J., and FitzGerald G.A. IPF2 α -I: an index of lipid peroxidation in humans. *Proc Natl Acad Sci USA* 1998; 95: 3449–3454.
34. Klawitter J., Haschke M., Shokati T., Klawitter J., and Christians U. Quantification of 15-F₂t-isoprostane in human plasma and urine: results from enzyme-linked immunoassay and liquid chromatography/tandem mass spectrometry cannot be compared. *Rapid Commun Mass Spectrom* 2011; 25: 463–468.
35. Proudfoot J., Barden A., Mori T.A., Burke V., Croft K.D., Beilin L.J., and Puddey I.B. Measurement of urinary F₂-isoprostanes as markers of in vivo lipid peroxidation—a comparison of enzyme immunoassay with gas chromatography/mass spectrometry. *Anal Biochem* 1999; 272: 209–215.
36. Esterbauer H., Schaur R.J., and Zollner H. () Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 1991; 11: 81–128.
37. Humphries K.M. and Szveda L.I. Selective inactivation of alpha-ketoglutarate dehydrogenase and pyruvate dehydrogenase: reaction of lipoic acid with 4-hydroxy-2-nonenal. *Biochemistry* 1998; 37: 15835–15841.
38. Musatov A., Carroll C.A., Liu Y.C., Henderson G.I., Weintraub S.T., and Robinson N.C. Identification of bovine heart cytochrome c oxidase subunits modified by the lipid peroxidation product 4-hydroxy-2-nonenal. *Biochemistry* 2002; 41: 8212–8220.
39. Yang Y., Sharma R., Sharma A., Awasthi S., and Awasthi Y.C. Lipid peroxidation and cell cycle signaling:

- 4-hydroxynonenal, a key molecule in stress mediated signaling. *Acta Biochim Pol* 2003; 50: 319–336.
40. Machella N., Regoli F., Cambria A., and Santella R.M. Oxidative damage to DNA: an immunohistochemical approach for detection of 7,8-dihydro-8-oxodeoxyguanosine in marine organisms. *Mar Environ Res* 2004; 58: 725–729.
 41. Machella N., Regoli F., Cambria A., and Santella R.M. Application of an immunoperoxidase staining method for detection of 7,8-dihydro-8-oxodeoxyguanosine as a biomarker of chemical-induced oxidative stress in marine organisms. *Aquat Toxicol* 2004; 67: 23–32.
 42. Machella N., Regoli F., and Santella R.M. Immunofluorescent detection of 8-oxo-dG and PAH bulky adducts in fish liver and mussel digestive gland. *Aquat Toxicol* 2005; 71: 335–343.
 43. Haghdoust S., Czene S., Naslund I., Skog S., and Harms-Ringdahi M. Extracellular 8-oxo-dG as a sensitive parameter for oxidative stress in vivo and in vitro. *Free Radic Res* 2005; 39: 153–162.
 44. Chiou C.C., Chang P.Y., Chan E.C., Wu T.L., Tsao K.C., and Wu J.T. Urinary 8-hydroxydeoxyguanosine and its analogs as DNA marker of oxidative stress: development of an ELISA and measurement in both bladder and prostate cancers. *Clin Chim Acta* 2003; 334: 87–94.
 45. Lindahl M., Mata-Cabana A., and Kieselbach T. The disulfide proteome and other reactive cysteine proteomes: analysis and functional significance. *Antioxid Redox Signal* 2011; 14: 2581–2642.
 46. Ghesquiere B., Jonckheere V., Colaert N., Van Durme J., Timmerman E., Goethals M., Schymkowitz J., Rousseau F., Vandekerckhove J., and Gevaert K. Redox proteomics of protein-bound methionine oxidation *Mol Cell Proteomics* 2011; 14: 2581–2642.
 47. Beckman K.B., and Ames B.N. The free radical theory of aging matures. *Physiol Rev* 1998; 78: 547–581.
 48. Finkel T., and Holbrook N.J. Oxidants, oxidative stress and the biology of ageing. *Nature* 2000; 408: 239–247.
 49. McEwen B.S. Sex, stress and the hippocampus: allostasis, allostatic load and the aging process. *Neurobiol Aging* 2002; 23: 921–939.
 50. Cals M.J., Succari M., Meneguzzo E., Pontezieri C., Bories P.N., Devanlay M., Desveaux N., Gatey M., Luciani L., Blonde-Cynober F., and Coudray-Lucas C. Markers of oxidative stress in fit, health-conscious elderly people living in the Paris area. The Research Group on Ageing (GERBAP). *Nutrition* 1997; 13: 319–326.
 51. Kostka T., Draai J., Berthouze S.E., Lacour J.R., and Bonnefoy M. Physical activity, aerobic capacity and selected markers of oxidative stress and the anti-oxidant defence system in healthy active elderly men. *Clin Physiol* 2000; 20: 185–190.
 52. Gidron Y., Russ K., Tissarchondou H., and Warner J. The relation between psychological factors and DNA-damage: a critical review. *Biol Psychol* 2006; 72: 291–304.
 53. Moller P., Wallin H., and Knudsen L.E. Oxidative stress associated with exercise, psychological stress and life-style factors. *Chem Biol Interact* 1996; 102: 17–36.
 54. Farooqui A.A. (2009). Beneficial effects of fish oil on human brain. Springer, New York.
 55. Maugeri D., Santangelo A., Bonanno M.R., Testai M., Abbate S., Lo Giudice F., Mamazza C., Puglisi N., and Panebianco P. Oxidative stress and aging: studies on an East-Sicilian, ultraoctagenarian population living in institutes or at home. *Arch Gerontol Geriatr Supp* 2004; 9: 271–277.