*Ocean acidification during rearing constrains the molecular physiology of a subarctic crustacean*

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Target Journal: Frontiers in marine science

## Abstract

Ocean acidification threatens marine crustaceans, particularly those at high latitudes where conditions are changing more rapidly. Red king crab, which supports important fisheries in Alaskan waters, were reared at pH 8.0 (ambient), pH 7.8 (moderate OA), and pH 7.5 (severe OA) for ~12 weeks from larval hatching to the first crab stage, and gene expression was examined to characterize transcriptional responses. Similar functional patterns of expression were observed in crabs reared in both the moderate and severe OA treatments, but differences were more pronounced in severe OA (8.4% of genes were differentially expressed) than in moderate OA (2.0% were differentially expressed). Expression of nearly half of all genes (44%) increased or decreased significantly with pCO2, suggesting that OA affects crab physiology in a dose-dependent manner. Variability in gene expression was low among crabs reared in OA treatments, and only a handful of biological processes were enriched in OA-induced genes (transcription regulation, nervous system), indicating that OA reduces the breadth of physiological functions in red king crab. We also found transposable element activity was high in OA-reared crab. Whether highly active transposable elements are pathological or mitigate negative effects of OA remains unknown. Downregulated functions were numerous, and included functions related to energy production, biosynthesis, metabolic and catabolic processes, and immune function. Genetic analysis of SNPs extracted from RNA-Seq data indicates that genetic composition and family representation did not differ among OA treatments, and that the transcriptional differences in OA-reared crab are attributed to phenotypic plasticity that applies broadly to the 10+ families. Our results suggest that the decreased growth and survival and longer intermolt periods previously reported in red king crab juveniles in OA treatments is likely due to a combination of metabolic limitations and vulnerability to infection or secondary stressors. Future studies should expose OA-reared red king crab to abiotic (e.g., warming) and biotic (e.g., microbial/viral) challenges to identify environmental scenarios that will most negatively impact red king crab populations, and to identify genotypes that may be more resilient to ocean change.

## 

## Introduction

Global oceans are rapidly changing as a result of increased concentrations of atmospheric greenhouse gasses. In addition to warming, deoxygenation, and sea level rise, oceans are acidifying (Cooley et al. 2022). This is occurring due to the increased partial pressure of carbon dioxide (CO2) in the ocean, which has absorbed ~30% of added atmospheric CO2 emitted since the industrial revolution [(Gruber et al., 2019)](https://paperpile.com/c/bgbcYo/Ex9Vl). As a result, the concentration of aqueous CO2, hydrogen ions, and bicarbonate ions are increasing, and ocean pH, carbonate ion concentrations, and calcium carbonate saturation states are decreasing. This shift is referred to as ocean acidification (OA) [(Feely et al., 2004)](https://paperpile.com/c/bgbcYo/DqmRT). Compared to pre-industrial pH levels, open ocean surface waters are estimated to already have decreased by pH 0.1, and will continue to decrease another 0.08 - 0.37 pH units by 2100 depending on greenhouse gas emissions (Cooley et al. 2022).

While ocean acidification is a global phenomenon, changes at high latitudes are likely to have outsized biological effects. Carbon dioxide is more soluble in those regions due to colder water, resulting in lower pH levels and carbonate saturation states [(Fabry et al., 2009; Mathis et al., 2015)](https://paperpile.com/c/bgbcYo/z3Fr9+UN5F). Increased freshwater inputs from sea ice melt and river runoff, and enhanced upwelling and respiration are likely to augment acidification in high latitude oceans [(Mathis et al., 2015)](https://paperpile.com/c/bgbcYo/z3Fr9). Conditions in those regions are therefore more likely to become undersaturated with carbonate ions and reach severely low pH levels, particularly at bottom waters [(Fabry et al., 2009; Terhaar et al., 2020)](https://paperpile.com/c/bgbcYo/UN5F+ONCt). As carbonate ions (aragonite, calcite) are critical components of calcium carbonate shells and structures, calcifying species in arctic and subarctic oceans are among the most vulnerable to the effects of acidification [(Melzner et al., 2019; Figuerola et al., 2021)](https://paperpile.com/c/bgbcYo/Sm63X+cLsPr). Effects of OA on crustaceans have significant socio-economic implications given their importance to commercial fisheries as both prey species (krill, copepods) [(Smetacek and Nicol, 2005; Mueter et al., 2021)](https://paperpile.com/c/bgbcYo/ibCbk+hAeOj) and fishery stocks (e.g. king, tanner, and snow crabs) [(Punt et al., 2014, 2015)](https://paperpile.com/c/bgbcYo/qBNZ3+fsu8s).

Ocean acidification has broad but variable physiological effects on crustaceans (Bednaršek et al. 2021). Many are capable of maintaining hemolymph pH homeostasis by acid-base regulation [(Pane and Barry; Meseck et al., 2016)](https://paperpile.com/c/bgbcYo/hXQnc+fyGG3), and have exoskeletons composed of both calcium carbonate and chitin, which may protect against direct dissolution [(Pörtner, 2008; Ries et al., 2009)](https://paperpile.com/c/bgbcYo/2I0yK+SnPuX), although see Dickenson et al (2021). Still, tissue and shell growth [(Long et al., 2017)](https://paperpile.com/c/bgbcYo/uWfS2), molt cycle [(Whiteley, 2011; Long et al., 2021)](https://paperpile.com/c/bgbcYo/tnE2j+8MiSJ), exoskeleton properties [(Siegel et al., 2022)](https://paperpile.com/c/bgbcYo/MobLl), metabolic activity [(Dissanayake and Ishimatsu, 2011; Thor et al., 2018)](https://paperpile.com/c/bgbcYo/OHMpC+ujtzk), reproduction [(Swiney et al., 2015; Meseck et al., 2016)](https://paperpile.com/c/bgbcYo/lMd85+fyGG3), behavior [(Dodd et al., 2015; Clements and Comeau, 2019)](https://paperpile.com/c/bgbcYo/sVT5N+FAhhg), and immune function [(Meseck et al., 2016; Shields, 2019)](https://paperpile.com/c/bgbcYo/fyGG3+mAevT) can all be altered by acidified conditions. Effects are most acutely observed in early life stages (larvae, juveniles), and vary greatly by species, severity of OA, and duration of exposure [(Bednaršek et al., 2021; Siegel et al., 2022)](https://paperpile.com/c/bgbcYo/MobLl+wQz00). The diversity of responses in crustaceans, and all marine invertebrates for that matter, has highlighted the need for a more mechanistic understanding of how organisms function in acidified conditions.

In this study, we examine the molecular physiology of OA-exposed juvenile red king crab, *Paralithodes camtschaticus*, one of several crab species that are highly valued fisheries in the Alaskan waters. Laboratory studies on red king crab from the Bering Sea indicate that OA affects their physiology such that development is altered, and growth and survival typically decreases [( Long et al., 2013; Long et al., 2013, 2019)](https://paperpile.com/c/bgbcYo/k5lvR+4gdpO+1pb0O). OA also interacts with warming by increasing intermolt duration and decreasing survival ([Swiney et al. 2017](https://doi.org/10.1093/icesjms/fsw251)). Little is known about the mechanisms underlying these impacts to red king crab. Metabolic changes are likely, as indicated by higher oxygen consumption when juvenile red king crab are first exposed to OA, but which does not last for the duration of a 3-week exposure [(Long et al., 2019)](https://paperpile.com/c/bgbcYo/k5lvR). A recent study examined broad-scale gene expression patterns in multiple life stages of red king crab ([Stillman et al. 2020](https://doi.org/10.1017/S002531541900119X)), and found that cuticular processes were affected in juveniles, while there were no pronounced changes to larval gene expression.

Here, we provide a high-resolution description of the physiological response of red king crab to ocean acidification. We leverage RNA-Seq to examine genome-wide expression patterns, and construct libraries from at least 13 individuals per treatment, rather than pools of individuals which can obscure genotype-dependent variation. We leverage a recently published draft *P. camtschaticus* genome which is large (7.29 Gbp) and highly repetitive [(Veldsman et al., 2021b)](https://paperpile.com/c/bgbcYo/DkoIn). Crab were reared from hatching to the first crab stage (C1) over 3 months in three carbonate chemistrytreatments, thus capturing transcriptional differences among crab that are reared in historically ambient conditions (pH 8.0), and those acclimated to a moderately (pH 7.8) and severely (pH 7.5) acidified environments.

## Methods

#### Animal collection

Ovigerous females for this study were collected in Bristol Bay in October 2016 in crab pots during the commercial fishery. Animals were transported to Kodiak in the live hold of a fishing vessel. Females were held communally in tanks at the Kodiak Fisheries Research Center supplied with flow through sand-filtered seawater pumped from Trident Basin, Kodiak, at local ambient temperature and salinity. Crabs were fed to excess on chopped frozen fish and squid. Embryos were monitored and, as they approached hatching, 48 females were moved to individual 48L tubs with flow-through seawater. Tanks were monitored for larvae. Larvae for this experiment were collected during peak hatch from as many females as possible over a period of 3 days: April 10-12, 2017. Larvae from the same 21 females on the first 2 days of larval stocking and from 20 of those 21 on the third (one female completed hatching after the second day of stocking) were pooled and used to stock larval rearing tanks.

#### Experimental Design

Experiments were performed in 180L conical bottomed tanks. Each tank was randomly assigned to one of three pH treatments: 1) Local ambient (pH ~ 8.1), 2) pH 7.8, and 3) pH 7.5, with five replicate tanks per treatment. Tanks were stocked with larvae in a random order. Flowthrough water for this experiment was passed through a 5 μm filter and UV sterilized and flow into each tank was 2 L/min. . The temperature and salinity of all tanks were allowed to vary with seasonal change of the intake water. The pH in each tank was adjusted via direct bubbling of CO2 controlled by feedback from Honeywell controllers connected to an in-tank Durafet III pH probe. The temperature and pH in each tank was measured daily using a Durafet III pH probe calibrated with TRIS buffer (Millero, 1986). Water samples were taken once a week from each tank, poisoned with HgCl2 and analyzed for dissolved inorganic carbon (DIC) and total alkalinity (TA). DIC and TA were measured using a VINDTA 3C (Marianda, Kiel, Germany) and a 5012 Coulometer (UIC Inc.) according to DOE (1994) using Certified Reference Material from the Dickson Laboratory (Scripps Institute, San Diego, CA, USA; (Dickson et al. 2007)). The seacarb package (Lavigne and Gattuse 2012) in R (V3.6.1, Vienna, Austria) was used to calculate the other parameters of the carbonate system.

Water temperatures increased from about 5°C at the beginning of the experiment to about 10°C by the end and averaged about 7.2°C throughout. pHs were well controlled to within 0.01 pH units in experimental treatments (Table 1).

**Table 1** Water parameters during the experiment. Temperature and pH were measured daily; salinity, dissolved inorganic carbon (DIC) and alkalinity were measured weekly; all other parameters were calculated. Values are mean ± standard deviation.

|  |  |  |  |
| --- | --- | --- | --- |
| **Treatment** | **Ambient** | **Moderate OA** | **Severe OA** |
| Temperature (C) | 7.24 ± 1.40 | 7.25 ± 1.40 | 7.23 ± 1.44 |
| Salinity | 31.267 ± 0.142 | 31.277 ± 0.149 | 31.288 ± 0.163 |
| pHT | 8.05 ± 0.03 | 7.79 ± 0.05 | 7.50 ± 0.06 |
| pCO2  (µatm) | 370.74 ± 26.92 | 703.89 ± 90.45 | 1414.71 ± 287.82 |
| HCO3- (mmol/kg) | 1.89 ± 0.08 | 1.96 ± 0.05 | 2.00 ± 0.04 |
| CO3-2 (mmol/kg) | 0.11 ± 0.01 | 0.06 ± 0.01 | 0.03 ± 0.01 |
| DIC (mmol/kg) | 2.01 ± 0.08 | 2.06 ± 0.05 | 2.10 ± 0.05 |
| Alkalinity (mmol/kg) | 2.16 ± 0.08 | 2.12 ± 0.05 | 2.09 ± 0.06 |
| ΩAragonite | 1.66 ± 0.09 | 0.96 ± 0.13 | 0.52 ± 0.19 |
| ΩCalcite | 2.65 ± 0.15 | 1.53 ± 0.21 | 0.83 ± 0.31 |

#### Larval rearing

Larvae were stocked at 50 larvae/L, or approximately 9,000 larvae per tank, and reared according to Swingle et al. (2013), except that in this experiment we used ambient incoming seawater rather than elevated to avoid potential interactive effects between pH and temperature. In brief, larvae were fed daily to excess on a diet of *Artemia* *sp.* enriched with DC DHA Selco (Inve Aquaculture) except during the non-feeding glaucothoe stage. Daily, five larvae were removed from each tank and the stage of each was determined. During the intermolt period between each successive stage, all the larvae were gently removed from tanks and the tanks were thoroughly cleaned to remove any microbial buildup. Once the larvae molted to the glaucothoe stage, artificial seaweed was provided as a settling substrate. Glaucothoe continued to be monitored until they molted to the first crab stage.

#### RNA Extraction & Sequencing

For each OA treatment 13-15 juvenile crabs at the first juvenile instar (C1) stage were sampled for RNA-seq (2-4 crab from each replicate tank, Table 2), placing whole crabs in RNAlater per manufacturer’s instructions (Thermofisher Scientific, Waltham, MA). To standardize among tanks/treatments and ensure that all crabs were at the same point in their molt cycle, sampling date varied by tank to target the C1-stage, and occurred five days after 100% of crabs in a tank reached the C1-stage. RNA isolation, library construction, and sequencing were performed by the University of Oregon. Briefly, RNAlater-preserved crabs were homogenized with silica beads using a Spex Geno/Grinder®, then RNA was isolated following TRIzol™ Reagent protocol for total RNA (Invitrogen Inc., Carlsbad, CA). The purity and quality of RNA were assessed with a NanoDrop™ Spectrophotometer (ThermoFisher Scientific) and a *Fragment Analyzer*™ (Agilent Technologies, Boulder, CO). Stranded mRNA-Seq libraries were constructed with the NuGen Universal Plus mRNA kit (Tecan Genomics, Inc, Männedorf, Switzerland). Libraries were prepared for sequencing using TruSeq RNA sample prep kits (Illumina). Paired-end sequencing was conducted on two lanes of a Hi-Seq 4000 with 100-bp read length.

**Table 2** RNA-Seq sampling scheme.

|  |  |  |  |
| --- | --- | --- | --- |
| **Treatment** | **Individuals / Libraries** | **Replicate Tanks** | **Replicates per Tank** |
| Ambient (pH 8.0) | 14 | 5 | 2 or 3 |
| Moderate OA (pH 7.8) | 13 | 4 | 3 or 4 |
| Severe OA (pH 7.5) | 15 | 5 | 3 |

#### Bioinformatics

Raw sequence data from each of the two lanes was demultiplexed, concatenated by library, then trimmed using Cutadapt v3.5 [(Martin, 2011)](https://paperpile.com/c/bgbcYo/FEfqt) to remove Illumina adapters, poly-A tails, flanking N bases, reads less than 50bp, and low-quality ends from reads using minimum quality scores of 20 and 15 for the 5’ and 3’ ends, respectively. Raw and trimmed data were inspected using FastQC [(Andrews, 2010)](https://paperpile.com/c/bgbcYo/E1C6m) and MultiQC [(Ewels et al., 2016)](https://paperpile.com/c/bgbcYo/VbFa9). Reads were aligned to the draft Red king crab (*Paralithodes camtschaticus*) genome (Genbank accession GCA\_018397895.1) [(Veldsman et al., 2021a, 2021b)](https://paperpile.com/c/bgbcYo/DkoIn+kuGmz) using Bowtie2 v2.4.2 with the preset option --sensitive [(Langmead and Salzberg, 2012; Tong et al., 2020)](https://paperpile.com/c/bgbcYo/jfoCv+7FwWg). The number of fragments aligning to gene coding regions of the *P. camtschaticus* genome was quantified using featureCounts v2.0.3 [(Liao et al., 2014)](https://paperpile.com/c/bgbcYo/sRekq) with settings -p --countReadPairs to count paired-end fragments and -C and -B to exclude chimeras and singletons, respectively. Gene functions were identified by querying coding sequences of the *P. camtschaticus* genome, derived from gene annotations published along with the *P. camtschaticus* genome [(Veldsman et al., 2021a)](https://paperpile.com/c/bgbcYo/kuGmz), against the Uniprot/Swissprot database [(UniProt Consortium, 2021)](https://paperpile.com/c/bgbcYo/rZePT) using blastx from blast v2.11.0 (e-value < 1–10) [(Camacho et al., 2009)](https://paperpile.com/c/bgbcYo/4xVS9).

#### Gene expression analysis

Analyses were performed in R v4.1.2 using RStudio interface v2021.09.1 (R Core Team, 2021; RStudio Team, 2020). Unless otherwise specified, significance thresholds were alpha = 0.05 and representations of spread in data are 1 standard deviation.

##### RNA-Seq pre-processing for gene expression analysis

Gene counts were filtered to remove outlier samples and low-frequency genes. Outlier samples were identified using principal component analysis (PCA), which was performed on variance-stabilizing transformed counts of the top 500 genes using vsd and plotPCA from *DESeq2* v1.34.0 (Love et al., 2014). Genes with mean count <10 across all samples or those with counts <30 across at minimum 10% of the samples were discarded, and differences in the number of remaining fragments per sample among treatments was tested using ANOVA. This filtered gene count dataset was used in comparative gene expression analyses as raw counts, or transformed counts via variance-stabilization transformation in *DESeq2*.

##### Global patterns

Global gene expression differences among OA treatments were explored with PCA, using prcomp from the R package *vegan* v.2.5-7 on all gene counts that were transformed via variance-stabilization. Principal components that explained a significant amount of variance were identified using the scree test [(Cattell, 1966)](https://paperpile.com/c/bgbcYo/LaUVr). Global differences among treatments was assessed by permutational pairwise permANOVA with pairwise.adonis from the *pairwiseAdonis* package, which is a wrapper for adonis from the *vegan* package.

##### Variation in gene expression

Global differences in the variation of gene expression among OA treatments was assessed with a test of multivariate homogeneity of group dispersions. To do so, we used betadisper from the *vegan* package to calculate per-sample distances to group medians in multivariate space, then analysis of variance to compare per-sample distances among treatments for all genes, and for differentially expressed genes. Pairwise comparisons and associated permuted *p*-values identified which treatments differed. We then examined gene-wise variation in expression by treatment for genes that were upregulated, downregulated, or not differentially expressed for each treatment. The within-treatment coefficient of variation (CV=SD/mean) was calculated for each gene, providing a method of comparing variation in gene expression relative to the mean for each treatment [(Reed George F. et al., 2003)](https://paperpile.com/c/bgbcYo/Zx3jZ). CV was summarized by calculating the mean CV of genes that were upregulated, downregulated, and were not differentially expressed for each treatment.

##### Differential gene expression analysis

Differentially expressed genes among pH treatments were identified using *DESeq2* with default settings [(Love et al., 2014; Costa-Silva et al., 2017)](https://paperpile.com/c/bgbcYo/8Kv4y+ScTlp). *DESeq2* uses raw count data to generate generalized linear models and internally corrects for library size, therefore counts were not transformed prior to differential expression analysis. In addition to examining the transcriptional responses of crabs to OA treatments, differentially expressed genes were used to characterize the gene modules with expression that correlated with pCO2.

##### Gene co-expression network analysis

We identified groups of co-expressed genes (i.e. gene modules) with expression profiles that correlated with OA treatment using weighted gene co-expression network analysis (WGCNA v1.70-3, [(Langfelder and Horvath, 2008)](https://paperpile.com/c/bgbcYo/2j2Kz)). Briefly, a weighted gene network was constructed from transformed gene counts with a signed adjacency matrix using the soft thresholding power 15, and minimum module size 75. Modules were merged if their eigengene expression correlated at R > 0.75, and those with eigengenes that correlated with pCO2 concentration at alpha=0.05 were determined to be associated with OA treatment. Modules with positive and negative correlations were designated as those with upregulated and downregulated expression profiles, respectively.

##### Functional Analyses

Gene sets of interest were characterized by Gene Ontology (GO) enrichment analyses. Two enrichment analyses were performed for each of the three pairwise treatment contrasts (Figure 2) to determine the functions of differentially expressed genes that were upregulated (L2FC > 0.5) and downregulated (L2FC < 0.5) in response to the more severe OA treatment. To identify processes that are likely critical to the function of OA-reared crab, enrichment analysis was performed on a subset of differentially expressed genes that were upregulated in OA compared to ambient: those that had very consistent expression levels (genes with within-treatment CV < 3%). To characterize functions that respond to pCO2 in dose-dependent manner, enrichment analysis was performed on co-expressed gene modules (from WGCNA analysis) for which eigengenes correlated with pCO2 concentration, filtered to retain genes that either correlated individually with pCO2 (Gene Significance *p*-value < 0.05) or were differentially expressed. For all gene sets, genes were filtered for those that map to the Uniprot/Swissprot database [(UniProt Consortium, 2021)](https://paperpile.com/c/bgbcYo/rZePT), and enriched GO terms were identified by entering UniprotID’s into the Gene-Enrichment and Functional Annotation Tool from DAVID v2021 [(Sherman et al., 2022)](https://paperpile.com/c/bgbcYo/PFN7c) to identify enriched biological processes, which were defined as those with modified Fisher Exact *p*-values (EASE Scores) <0.05.

Given that not all transposable elements are assigned to transposition-related GO terms and may be overlooked by traditional enrichment analyses, gene sets were also interrogated for transposable element composition. The proportion of genes that were transposable elements was estimated for each gene set described above by searching within the protein names of annotated genes for the words “transposon”, “transposable”, “LINE” (representing long interspersed nuclear elements), “retrotransposable element”, “transposable element”, “mobile element jockey” (a LINE), and “pol polyprotein”. Pol polyprotein was included as it is core to the replication of retrotransposons, but may also be associated with retroviral activity [(Havecker et al., 2004)](https://paperpile.com/c/bgbcYo/MWApy). These terms were determined to represent the majority of TEs by manual review of annotated genes.

#### Genetic analysis

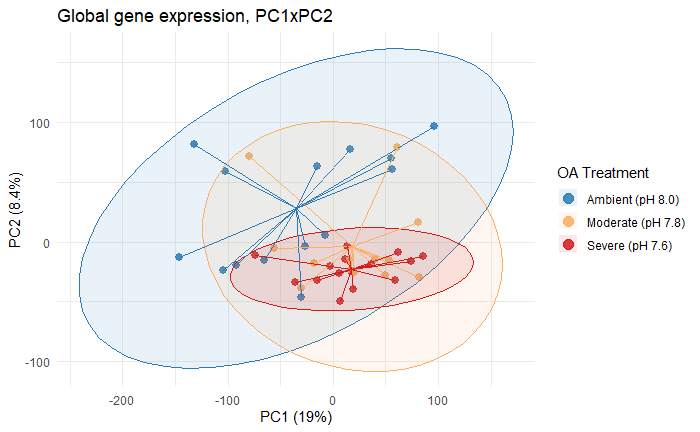
As with many rearing experiments, there was high larval mortality rate, which could possibly result in treatment-specific survival rates among the ~20 families and genotype-specific expression patterns. We therefore assessed whether there were genetic differences among treatments. Single nucleotide polymorphisms (SNPs) were extracted from RNA-Seq reads to examine the genetic composition of sampled crabs. Variants were identified using the GATK toolkit [(McKenna et al., 2010)](https://paperpile.com/c/bgbcYo/Y7MPS). Briefly, RNA-Seq reads were aligned to the draft red king crab genome [(Veldsman et al., 2021b)](https://paperpile.com/c/bgbcYo/DkoIn). The genome was first concatenated into 50 larger contigs, with 1000N separating each original contig, which was necessary to reduce the processing time in GATK. Alignment files were deduplicated using MarkDuplicates, reads spanning splicing events and CigarN reads were split, variants were called using HaplotypeCaller, then joint-genotyped using GenotypeGVCFs. SNPs were filtered using VariantFiltration to hard-filter loci with any of the characteristics FS>60, QD<2, QUAL<30, SOR>3, DP<15, DP>150, or AF<0.30. SNPs were then pruned with snpgdsLDpruning from the R package *SNPRelate* v1.30.1to remove those in linkage-disequilibrium and with >15% missing rate or <5% minor allele frequency. One sample from the ambient treatment was removed from the genetic analysis due to high missingness. From 227,781 candidate SNPs, 331 markers resulted from the above filtering and were retained for genetic analysis.

Differences in genetic composition among treatments was examined using multivariate analysis, estimates of diversity, and parentage analysis. Using *SNPRelate*, allele frequencies (major and minor) and per-SNP missing rate were calculated with *snpgdsSNPRateFreq()*, and PCA biplots of the first four principal components (PCs) were constructed with snpgdsPCA(). The first four PCs were selected as they explained over 25% of the total variance, and each additional PC explained less than 4% of the total variance. Pairwise Fst values [(Weir and Cockerham, 1984)](https://paperpile.com/c/bgbcYo/wFYFK) among treatments were calculated with stamppFst() from *STaMPP* v1.6.3 using 1000 bootstraps to generate 95% confidence intervals and *p*-values. Parentage analysis was performed using the 331 SNPs with *Colony* v2.0.6.6 for R [(Jones and Wang, 2010)](https://paperpile.com/c/bgbcYo/jbTqn), specifying polygamous males and females with three replicate medium-length runs using the full likelihood method with high precision. The parentage of samples with cluster probability < 0.5 were considered invalid (three samples from each treatment). The relationship between global expression patterns and genetic structure was assessed by regressing genotype PCA sample scores against the expression-derived PCA sample scores along the first two principal components, using OA treatment as a covariate.

## Results

##### RNA-Seq pre-processing for gene expression analysis

A total of ~2.65B paired-end reads (henceforth “fragments”) remained after discarding ~4.55M (0.17%) during initial quality-filtering, with a per-sample mean of 61.7M +/- 10.7M fragments. Across all samples ~2.14B reads were aligned to the *P. camtschaticus* draft genome [(Veldsman et al., 2021a, 2021b)](https://paperpile.com/c/bgbcYo/DkoIn+kuGmz), for a total alignment rate of 80.47%. The average per-sample alignment rate was 80.43%±2.15%, which included 43.19%±1.50% and 24.52%±0.89% of concordantly mapped fragments that mapped uniquely and multiple times, respectively. The multi-mapped fragments were assigned by Bowtie2 to the “best” location, and therefore counted once in the downstream analysis. Of the ~2.14B aligned fragments, ~1.27B were assigned to gene-coding regions of the *P. camtschaticus* draft genome (59.6%). The remainder were not included in the downstream analyses as they mapped to non-coding regions (22.3%), were assigned ambiguously (6.7%), were singletons (8.3%) or chimeras (3.1%). Initial examination of the gene counts using PCA identified one sample from the ambient pH treatment as an outlier, which was removed from the dataset and resulted in 14, 13, and 15 samples for the ambient, moderate OA, and severe OA treatments, respectively (Table 2). In total, we detected all 162,611 gene features that are in the draft *P. camtschaticus* genome [(Veldsman et al., 2021a)](https://paperpile.com/c/bgbcYo/kuGmz), but after removing low frequency genes (totalling 0.75% of fragments), 74,778 genes remained for analysis, 32,435 of which mapped to genes in the Uniprot/Swissprot database. The high number of genes reflects the large *P. camtschaticus* draft genome, which includes a high degree of repeat elements [(Veldsman et al., 2021b)](https://paperpile.com/c/bgbcYo/DkoIn). A one-way ANOVA indicated that the number of fragments retained for analyses did not differ among OA treatments (F(2,39)=0.22, *p*-value=0.80), and ranged from 15.4M to 38.2M per sample and averaged 29.4M±5.1M, mapped to on average 75K±80 genes.

**Figure 1**. PCA biplot of first the two principal components, constructed from all expressed genes. 

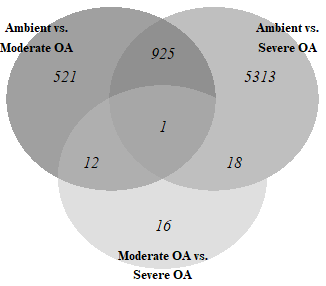
##### Global expression patterns

Global expression profiles of red king crabs reared in ambient conditions differed from those reared in either moderate or severe OA. Pairwise permANOVA tests detected significant differences in multivariate space among ambient conditions and OA treatments (moderate OA: F(1)=1.84, *p*-adj=0.042; severe OA: F(1)=2.70, *p*-adj=3.0e-3), but not between the two OA treatments (F(1)=1.20, *p*-adj=0.50), which is evident from the biplot of the first two principal components (PC1 & PC2) (Figure 1). Ambient-reared crabs were separated from those reared in OA treatments along PC1 (19.0%) and PC2 (8.4%), which combined explained 27.4% of variation in global expression. While the scree test indicated that PC3 and PC4 also explained a significant amount of variation (7.0% and 6.2%, respectively), there was no separation among treatments along those axes (Supplemental Figure 1).

Global gene expression was less variable among crabs reared in OA treatments compared to those reared in ambient conditions. Using a test of multivariate homogeneity of group dispersions, we found that variation in global gene expression differed by treatment (F(2,39)=4.9, p=0.012). Pairwise comparisons indicated that variation differed between ambient and severe OA (*p*-valuepermuted=8.3e-3)(Figure 1), but did not differ between ambient and moderate OA (*p*-valuepermuted=0.32) or moderate and severe OA (*p*-valuepermuted=0.25).

##### Differential gene expression

Analysis in DESeq2 identified 6,806 genes that were differentially expressed among treatments (9.1% of all examined genes). The number of differentially expressed genes increased with OA severity: 1,459 genes differed between ambient and moderate OA (526 upregulated in moderate OA, 933 downregulated), and 6,257 genes differed between ambient and severe OA (2,350 upregulated in severe OA, 3,907 downregulated). Only 47 genes differed between moderate and severe OA treatments (22 upregulated in severe OA, 25 downregulated), indicating that the two OA treatments induced a similar transcriptional response (Figure 3, Supplemental Table 1).



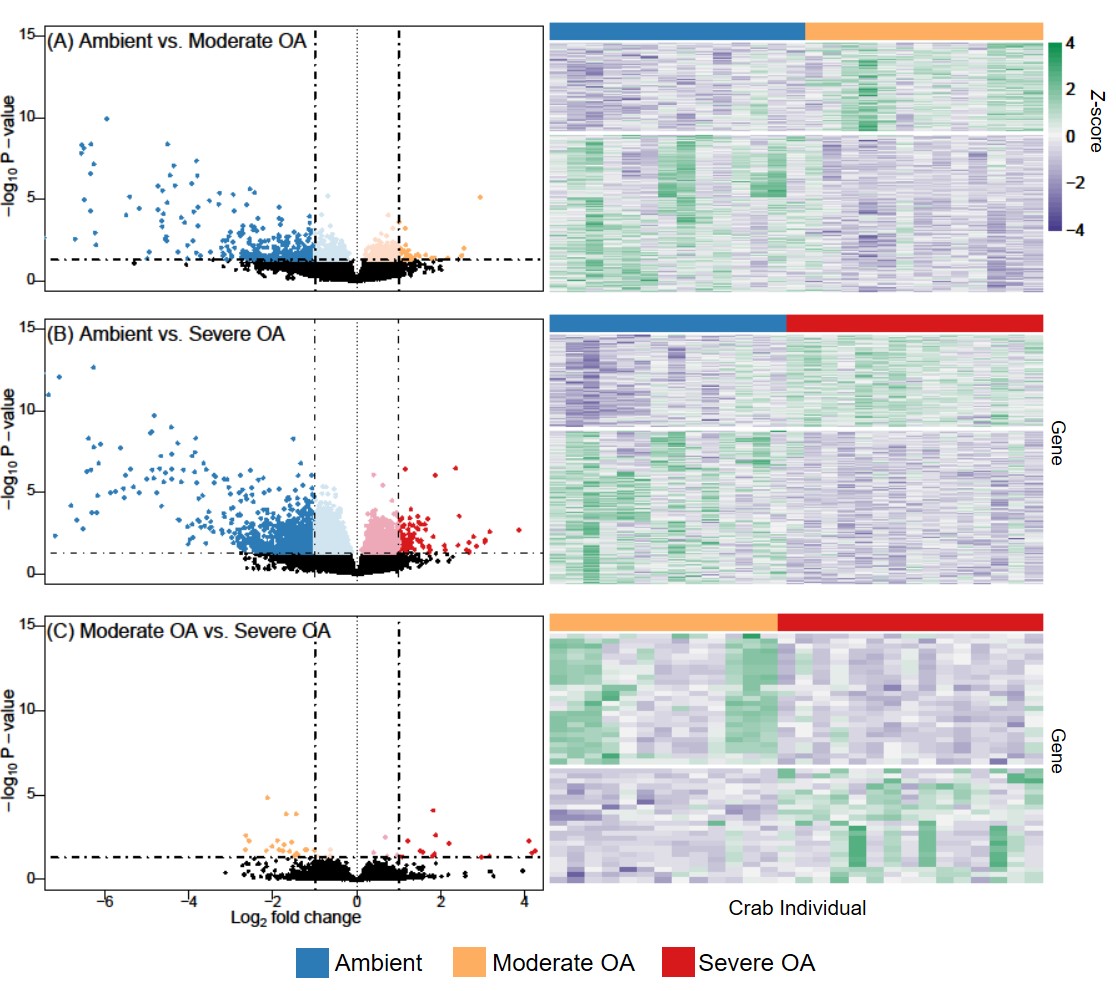
**Figure 2**. Venn diagram showing the number of differentially expressed genes among each pairwise OA treatment contrast, and the number that are shared among contrasts.

**Table 4**: Coefficient of variation mean ∓ SD for genes that were upregulated or downregulated relative to other treatments, or not differentially expressed (Non-DEG). Genes that were only differentially expressed among moderate and severe OA (n=16) were not included in these calculations.

|  |  |  |  |
| --- | --- | --- | --- |
|  | Ambient | Moderate OA | Severe OA |
| Up-regulated | 11.0%∓10.3% | 6.8%∓4.7% | 4.0%∓4.2% |
| Down-regulated | 5.5%∓4.1% | 11.2%∓9.5% | 8.5%∓7.2% |
| Non-DEG | 8.3%∓5.6% | 8.0%∓5.3% | 7.2%∓5.2% |

##### Co-expression network analysis

We performed a weighted gene co-expression network analysis (WGCNA) to identify groups of genes that were co-expressed (i.e. gene modules), and for which expression correlated with pCO2 concentration. The 74,778 examined genes were assigned to 41 modules, 14 of which had eigengenes that correlated significantly with pCO2 (Supplemental Table 2). For eight of the modules, which contained in total 22,537 genes, the eigengenes correlated negatively with pCO2 indicating that expression decreased as OA treatment became more severe (panels a-h in Figure 5). Six modules, containing 19,248 genes in total, correlated positively with pCO2, indicating higher expression in OA treatments (panels i-m in Figure 5). There was high overlap between the differentially expressed genes and WGCNA analysis, with 92.8% of DEG’s assigned to one of the 14 pCO2-correlated co-expression modules.



**Figure 3**. Pairwise differential expression among OA treatments for (A) ambient (pH 8.0) vs. moderate OA (pH 7.8), (B) ambient vs. severe OA (pH 7.5), and (C) moderate OA vs. severe OA. Volcano plots (left panels) show Log2 fold change along the x-axes, with higher absolute values indicating larger differences among treatments, and -log10 *p*-values along the y-axes, with higher numbers indicating higher significance. Each point represents a unique gene, with differentially expressed genes (*p*-adj < 0.05) color coded to indicate those that are expressed at higher levels in ambient treatment (blue), moderate OA (orange), or severe OA (red). Points with darker colors indicate those that have |Log2FC| >0.5. Heatmaps (right panels) show expression of differentially expressed genes only (rows=genes) at per-sample resolution (columns=samples), with the purple-green gradient indicating the *z*-score of expression values standardized across samples for each gene, where green and purple indicate higher and lower expression, respectively.

#### Functional Analyses

##### Enrichment analysis of differentially expressed genes

GO enrichment analysis revealed 14 and 104 biological processes that were enriched in genes that were differentially upregulated and downregulated in an OA treatment compared to ambient, respectively (Supplemental Table 1). In moderate-OA reared crab, nine enriched processes in upregulated genes were involved in negative regulation of growth and cell death, transcription regulation, protein folding via heat shock proteins, and DNA integration via transposons (Figure 4). In severe-OA reared crab, five enriched processes in upregulated genes were involved in transposable element activity and regulation of transcription (Figure 4).

A variety of biological processes were downregulated in OA-reared crab compared to ambient-reared crab. Of the 23 processes that were enriched in genes down-regulated in moderate-OA reared crab, many were involved in the metabolism of various compounds (e.g., carbohydrates, lipids, fatty acids, estrogen), energy production (e.g., tricarboxylic acid cycle), and ceramide activity (biosynthesis, translocation, and metabolism), but also were involved in protein folding and N-linked glycosylation, toxin transport, and telomerase activity (Figure 4, see Supplemental Table 3 for full list). There were 81 enriched processes in genes down-regulated in severe-OA reared crab. Many processes that were enriched in moderate-OA reared crab were also enriched in genes downregulated in severe-OA reared crab (Figure 4). Enriched processes that were uniquely downregulated in severe-OA reared crab included translation (the most significantly enriched down regulated process, *p*-value=8.6e-14), and those related to DNA replication, microtubules, immune function, and aerobic respiration (Figure 4, see Supplemental Table 2 for full list).

A total of five biological processes were enriched in genes that were differentially expressed among crab reared in moderate vs. severe OA. Three processes involved in transposition were more active in the severe OA-reared crab, and two processes involved in molecular chaperone activity via the heat shock complex were more active in the moderate OA-reared crab.

##### Enrichment analysis of genes potentially critical in an OA environment

Of the 526 and 2,350 genes that were upregulated in moderate and severe OA relative to ambient treatment, respectively, 66 (13%) and 1,319 (56%) were expressed at consistent levels across individuals within treatments (CV < 3%, hereafter referred to as low-CV genes). These upregulated low CV genes are of interest as they may provide critical functions in the OA environment. Low CV genes upregulated in severe OA treatment were enriched for biological processes involved in transcription regulation, DNA integration (transposon activity), nervous system processes (neurogenesis, signal transduction, calcium ion-regulated exocytosis of neurotransmitter), developmental processes (multicellular organism development, keratinocyte differentiation, glycosaminoglycan biosynthesis), regulation of cardiac muscle cell contraction, and potassium ion transport (Supplemental Table 4). Low CV genes upregulated in moderate OA treatment were enriched for chromatin organization and transposition (Supplemental Table 4). In contrast, low-CV genes that were upregulated in ambient treatment relative to either OA treatment were primarily enriched for processes involved in protein biosynthesis, glycolysis, and ion transport (including hydrogen ion transport) (Supplemental Table 4).

##### Enrichment analysis of co-expressed genes that correlate with pCO2

Genes that decreased significantly with pCO2 (eight modules) were enriched for 193 biological processes (65 at FDR<10%), which were predominantly related to protein production, energy production, metabolic activity, tissue development, muscle activity, cell cycle, immune function, chaperone mediated protein folding, and telomere maintenance (Figure 5, Supplemental Table 5). Genes that increased significantly with pCO2 (six modules) were enriched for 48 processes (10 at FDR<10%), focused on transcription regulation and signaling (Figure 5, Supplemental Table 5). We also found that transposition and the related process DNA integration were enriched in both downregulated and upregulated modules (5 modules each).

**Table 5**: Percent of annotated differentially expressed genes that map to transposable elements. Colors indicate genes that were more active in ambient (blue), moderate OA (orange), and severe OA (red) treatments in each pairwise comparison. Values below the dotted line indicate the % of upregulated genes with low within-treatment variation (CV < 3%) that were TEs.

|  |  |  |  |
| --- | --- | --- | --- |
|  | Ambient | Moderate | Severe |
| Ambient |  | 51% | 65% |
| Moderate OA | 31% |  | 86% |
| Severe OA | 28% | 13% |  |
| Upregulated with low variability (CV< 3%) | 16% | 50% | 60% |

##### Transposable element composition

A large portion of the RKC transcriptome mapped to transposable elements (20,860 TEs), comprising 28% of the 74,778 analyzed genes, and 64% of the 32,435 annotated genes. These TEs mapped to 67 distinct Uniprot Species IDs, all of which were retroelements (Supplemental Table 6). Transposable element activity was high in all treatments, but activity increased with OA severity. Of the annotated genes that were upregulated in severe OA compared to ambient and moderate OA treatments, 65% and 86% mapped to TE’s, respectively, while 51% of genes upregulated in moderate OA compared to ambient were TE’s (Table 5, Figure 7). In contrast, 31% and 28% of genes that were more abundant in ambient treatment compared to moderate and severe OA mapped to TEs (Table 5, Figure 7). A large percentage of upregulated low-CV genes were also transposable elements, comprising 50% and 60% of those genes in moderate and severe OA-treated crab, respectively. In contrast, 16% of the low-CV upregulated genes in ambient-pH-reared crab were transposable elements (Table 5). The TE composition of co-expressed gene modules was similar – on average the percent of genes that increased and decreased with pCO2 was 73% and 51%, respectively (Figure 7).

##### 

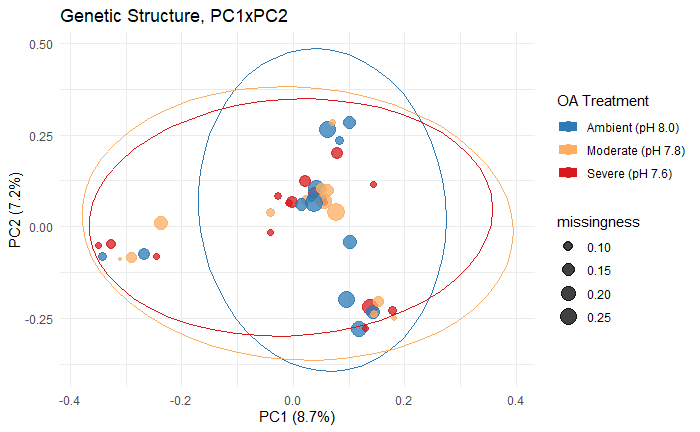
**Figure 4**: Enriched Gene Ontology biological processes in genes that are differentially expressed between the three treatment contrasts, indicating which processes were upregulated in ambient (blue), moderate OA (orange), and severe OA (red) in each contrast. Columns indicate pairwise contrasts, and rows indicate GO terms. For processes upregulated in ambient treatment (blue) only the top 20 processes by *p*-value were included in this figure. Point size indicates the -Log10 transformed *p*-value (all *p*-values < 0.05), such that larger points are more significantly enriched.

##### 

##### **Figure 5:** Expression profiles of co-expressed gene modules that correlate with pCO2 treatment negatively (red, a-h) and positively (green, i-n), indicating those genes that decrease and increase with ocean acidification severity, respectively, where 370 µatm is ambient (pH 8.0), 700 µatm is moderate OA (pH 7.8), and 1400 µatm is severe OA (pH 7.5). Points indicate the mean eigengenes (i.e. first principal component) for each treatment within each module, with standard deviation among individuals within treatments indicated by error bars. Figure titles indicate the predominant biological functions that were enriched in each module. Detailed enrichment analysis results, number of genes in each module, and correlation statistics are in the supplemental materials. Transposition was omitted from figure titles as it was enriched in many modules.

##### Genetic relatedness analysis, integration with expression

Principal component analysis (PCA) constructed from SNPs (n=331) indicated genetic homogeneity among treatments (Figure 6). Individuals loosely aggregated into two or three clusters, primarily along PC1 and PC2 which explained 8.71% and 7.23% of variation, respectively, but no treatments were overrepresented in any of the clusters. Pairwise *F*ST values among samples from the three treatments did not differ from zero for any contrast (Table 6). Parentage analysis using Colony estimated the same number of mothers (n=9) and fathers (n=10) represented by surviving offspring in each treatment. No correlation was found among SNP-derived PCA scores and gene expression-derived PCA scores along the first two principal components (Supplemental Materials).



**Figure 6**: PCA biplot of the first two principal components, constructed from RNA-Seq derived SNPs (n=331). Points represent individual crabs that are color-coded by OA treatment, which do not indicate clustering by treatment, and sizes represent the percent of SNPs that are missing in each individual. The PCA and other genetic analyses (parentage, pairwise *F*ST) does not indicate that genetic composition of individuals surviving the 3-month exposure differed among treatments.

**Table 6: Pairwise *F*ST values**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| *Treatment 1* | *Treatment 2* | *F*ST | *CI Lower Bound* | *CI Upper Bound* | *p-value* |
| Ambient | Severe | -0.0015 | -0.0107 | 0.0073 | 0.67 |
| Ambient | Moderate | -0.0123 | -0.0207 | -0.0035 | 1.00 |
| Moderate | Severe | -0.0033 | -0.0125 | 0.0061 | 0.78 |

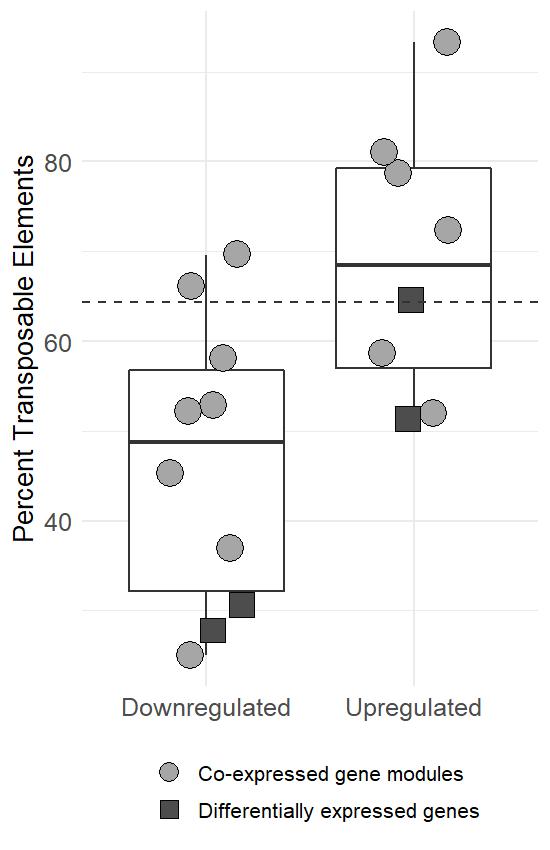
## Discussion

Calcifying marine species living at high latitudes may be particularly vulnerable to the impacts of ocean acidification due to more extreme changes projected to occur in those regions [(](https://paperpile.com/c/bgbcYo/9AcjK)Cooley et al. 2022[; Pilcher et al., 2022)](https://paperpile.com/c/bgbcYo/9AcjK). Crustaceans in some high latitude regions are likely already experiencing acidified conditions seasonally; at present the Bering Sea shelf drops to seasonal lows of around pH 7.5 (Mathis et al. 2014) and mean bottom pH is projected to drop by a further 0.3 units by 2100 (Pilcher et al. 2022). Red king crab, which is one of several valuable commercial fisheries in Alaskan waters, are sensitive to changes in ocean chemistry, resulting in high mortality and decreased growth [(Long et al., 2013; Coffey et al., 2017)](https://paperpile.com/c/bgbcYo/REazo+4gdpO). Understanding which aspects of red king crab physiology are altered in acidified conditions, and which are critical to their survival, is important for understanding their adaptive potential and predicting population dynamics. Here, we provide the first study to describe the molecular signatures of juvenile red king crab that were reared from the larval stage in acidified conditions.

Overall, RKC reared in OA are physiologically limited. There was a general reduction in transcriptional activity and inter-individual variability in OA-reared crab, suggesting that OA reduces the breadth of physiological functions compared to crabs reared in ambient seawater. Functional analysis indicates that energy production is depressed in OA conditions, which likely explains observed decreases in biosynthesis, the immune system, and myriad metabolic processes. The limited energy is shunted towards transcriptional regulation mechanisms, signaling systems, and control of growth. Transposable elements (TEs) in OA-reared crab were highly active, particularly in the most severe OA treatment, and were expressed at consistent levels, suggesting that TEs play a role in the OA-stress response. Expression patterns across multiple pCO2 treatments reveal that much of the RKC transcriptome responds to OA in a dose-dependent manner, with most differentially expressed genes correlating negatively (42%) or positively (35%) with pCO2. In the remainder of this section we expand on effects to transposable element activity, describe other processes that are triggered and suppressed by OA exposure during development in red king crab, and expand on effects of OA on the immune system. Finally, we discuss differences in gene expression variation observed among treatments, and consider whether there were significant genetic differences among treatment groups that may have contributed to the observed gene expression profiles.

##### Transposable elements are highly active in OA-reared crab

Many of the genes upregulated under OA conditions were TEs. While TEs were present in all gene sets, including those that were more active in ambient conditions, they comprised a much larger percentage of genes upregulated in OA-reared crab (Figure 7). TEs, or “jumping genes”, are DNA elements that move to new locations in the genome when activated, resulting in insertional mutations [(Bourque et al., 2018)](https://paperpile.com/c/bgbcYo/69KsC). Retrotransposons, the class of TEs detected in our study, are mobilized by a copy-and-paste mechanism where the DNA sequence is transcribed and an RNA intermediary is then reverse-transcribed into a cDNA copy before being integrated into the genome [(Casacuberta and González, 2013)](https://paperpile.com/c/bgbcYo/FUNT9). Increased transposable element activity in response to environmental stress is well documented in a variety of eukaryotes [(Horváth et al., 2017)](https://paperpile.com/c/bgbcYo/Sqm0O). There have been, however, only a few studies to detect stress-activated TEs in marine crustaceans [(de la Vega et al., 2007)](https://paperpile.com/c/bgbcYo/qANN1), and to our knowledge this is the first to do so in response to OA. While TEs comprise large portions of the genomes of arthropods [(Wu and Lu, 2019)](https://paperpile.com/c/bgbcYo/Lup5e), including crustaceans [(Tang et al., 2021; Veldsman et al., 2021b)](https://paperpile.com/c/bgbcYo/juS2Q+DkoIn), previous transcriptional characterization of stress-responses in crustaceans may have overlooked TE’s due to the common practice of masking repeat elements from genomes during analysis [(Lanciano and Cristofari, 2020)](https://paperpile.com/c/bgbcYo/c3rp1). Recent reports from a wide range of other taxa have also implicated TEs in the OA-response, including anemones [(Urbarova et al., 2019)](https://paperpile.com/c/bgbcYo/2Reuk), clams [(Lesser et al., 2019)](https://paperpile.com/c/bgbcYo/m9BSp), and diatoms [(Huang et al., 2018)](https://paperpile.com/c/bgbcYo/JmX0d). This breadth of taxa, which now includes crustaceans, suggests that TEs are a common response to exposure to acidified conditions.

Why TEs can become activated under stress is still under debate, as is whether they are detrimental or beneficial to an organism’s survival [(Casacuberta and González, 2013)](https://paperpile.com/c/bgbcYo/FUNT9). TEs may become more active because the mechanisms that suppress them are no longer functioning, which could negatively affect fitness due to pathological mutations. Alternatively, stress-activated TE’s could act as regulatory elements and/or increase diversity of proteins available to the organism, which could increase fitness through beneficial phenotypic variation. TEs may also be a component of the antiviral defense system, as they can stimulate the antiviral inflammatory response [(Macchietto et al., 2020; Hale, 2022)](https://paperpile.com/c/bgbcYo/KJd2h+19hO5). Determining how TE activity affected gene function and RKC tolerance of OA is beyond the scope of this study. However, many genes that were upregulated at consistent levels among OA-exposed individuals were TEs, suggesting that TE mobilization was a component of the RKC stress-response system, rather than simply the result of genomic instability. Given the poor performance of red king crab in previous OA studies, any added benefit of increased TE activity does not sufficiently mitigate negative effects, at least in the short-term. Yet, OA-induced TE activity may benefit RKC at an evolutionary scale, given that it could provide a mechanism for rapidly increasing genetic variability [(Horváth et al., 2017; Pimpinelli and Piacentini, 2020)](https://paperpile.com/c/bgbcYo/0yJpe+Sqm0O). It would be informative to examine whether the stress-induced mobilization of TE’s increases mutation rates by re-integration into the germline, resulting in heritable changes to the genome, which can occur in other species [(Ito et al., 2016)](https://paperpile.com/c/bgbcYo/JE41Y). If so, this could provide a mechanism by which OA exposure increases phenotypic diversity across generations.

**Figure 7**: The percent of genes in each co-expressed gene module and differentially expressed gene set (DEGs) that are transposable elements (TEs), categorized by whether genes were downregulated or upregulated in OA relative to ambient treatment. The proportion of all genes examined that were TEs (64%) is indicated by the dotted line. TEs were identified by searching within the protein names for the words transposon, transposable, LINE, retrotransposable element, transposable element, mobile element jockey, and pol polyprotein. Lines in the middle of each boxplot indicate the median percent TEs in downregulated and upregulated gene sets.

##### Processes that are suppressed in the OA-environment

OA resulted in a widespread downregulation of metabolic processes involved in multiple respiratory pathways, metabolism of a variety of compounds, and protein synthesis machinery. Reduced metabolic activity may be one way that RKCs mitigate acidosis [(Michaelidis et al., 2005; Small et al., 2010)](https://paperpile.com/c/bgbcYo/e2qFq+H1Ho6). Carbon dioxide is produced during respiration, therefore a decrease in metabolic activity reduces internal CO2 production. Decreased mitochondrial activity can also reduce oxidative cellular stress by reducing mitochondrial production of reactive oxygen species, which might otherwise increase due to environmental stressors [(Pörtner, 2010; Rato et al., 2017; Rivera-Ingraham and Lignot, 2017)](https://paperpile.com/c/bgbcYo/FZ4N7+OB39P+9jSi5). Decreased protein synthesis likely reflects energetic limitations in OA-reared crab, and likely contributed to the downregulation of other biosynthesis processes involved in tissue development, cell growth, and cell proliferation. Decreased growth is common for marine invertebrates exposed to OA conditions [(Melzner et al., 2019)](https://paperpile.com/c/bgbcYo/cLsPr), including RKC [(Long et al., 2013, 2019)](https://paperpile.com/c/bgbcYo/4gdpO+k5lvR), but much of the research has highlighted impacts to calcification [(Strader et al., 2020; Figuerola et al., 2021)](https://paperpile.com/c/bgbcYo/gUbIS+Sm63X). Our findings suggest that growth limitations are systemic, impacting even the most fundamental biosynthesis mechanisms.

Metabolic changes are quite common in response to ocean acidification [(Kelley and Lunden; Strader et al., 2020)](https://paperpile.com/c/bgbcYo/Tk03F+gUbIS). Ocean acidification can cause metabolic depression (and more generally, reduction in gene activity) in crustaceans and other marine invertebrates invertebrates [(Kaniewska et al., 2012; Evans and Watson-Wynn, 2014; Johnson and Hofmann, 2017; Kriefall et al., 2018; Bogan et al., 2020)](https://paperpile.com/c/bgbcYo/bjPzf+Ot6Yd+wLnlM+BZ6Vz+O3Sg2). In the present study, metabolic processes were downregulated in both the moderate and severe OA treatments, which likely reflects the sensitivity of RKC to changes in ocean chemistry, and suggests that both pH 7.8 and 7.5 are suboptimal conditions [(Sokolova et al., 2012)](https://paperpile.com/c/bgbcYo/P0Ezo). Given these findings, we might expect metabolic rate to decrease in response to OA in RKC. However, in previous respiration trials [(Long et al., 2019)](https://paperpile.com/c/bgbcYo/k5lvR) there was no change to metabolic rate measured by oxygen consumption in juvenile RKC exposed to the same OA conditions after 3 weeks. The decreased metabolic gene activity in the present study could reflect the duration and multi-stage exposure applied in this study - 3 months from hatching - which may influence the physiology of crabs in a way that short term/single life history stage- exposure does not.

##### Immune and stress-response processes are sensitive to OA

Ocean acidification affected the red king crab immune system. Genes involved in neuroinflammation (microglial cell activation), viral response, and the innate immune system decreased with pCO2. OA can negatively impact the immune system in crustaceans [(Hernroth et al., 2012, 2015; Meseck et al., 2016; McLean et al., 2018; Shields, 2019)](https://paperpile.com/c/bgbcYo/fyGG3+mRrW4+wxHvH+Tpagi+mAevT) and other marine invertebrates [(Bibby et al., 2008; Hernroth et al., 2011; Liu et al., 2016)](https://paperpile.com/c/bgbcYo/AmpaJ+tlx6j+U3RTB). The mechanisms by which ocean acidification alters immune function are not fully understood, and could relate to energy constraints and malfunction of immune-related enzymes and signaling/recognition pathways (among other mechanisms) [(Bibby et al., 2008; Liu et al., 2016)](https://paperpile.com/c/bgbcYo/AmpaJ+U3RTB). We find evidence that immunosuppression may in part be due to activation of the acute stress-response [(Adamo, 2012)](https://paperpile.com/c/bgbcYo/0t3JA). Genes coding for the octopamine receptors and a receptor for tyramine, the direct precursor of octopamine, were upregulated in OA. Octopamine is the invertebrate orthologue to norepinephrine, and is one of the key neurotransmitters that regulates the acute stress-response [(Adamo, 2008)](https://paperpile.com/c/bgbcYo/VzbYD). Increased expression of Orcokinin signaling molecules were also detected, which are involved in myostimulatory activity, but have also been implicated in the crustacean environmental stress-response [(Zhang et al., 2015; Buchberger et al., 2020)](https://paperpile.com/c/bgbcYo/18MWb+cpwYU). Increased expression of stress-response mediating neurotransmitters indicates that RKC juveniles may have maintained a state of physiological stress despite long-term OA exposure (3 months). Interestingly, the molecular chaperones (HSP70 and DNAJA1), which respond to a variety of stressors by refolding or facilitating the destruction of damaged proteins, increased in moderate OA but decreased substantially in severe OA, suggesting that either the crabs become too energetically exhausted to mount some cellular stress response processes or that some of those processes become ineffective as OA severity worsens. Further, it suggests that at moderate OA levels increased oxidative stress is damaging proteins, or that altered intracellular or extracellular pH affects folding patterns of proteins. Our findings suggest that OA likely induces the acute stress-response in RKC, and may cause high mortality indirectly by making them more vulnerable to microbial or viral infections. Furthermore, pH 7.5 appears to be beyond the level at which RKC is capable of mitigating cellular damage or fighting infection, indicating that the degree of acidification during rearing matters. Studies are needed that expose RKC to a range of pH levels alongside viral/bacterial challenges to understand the interaction between OA and infection.

##### Processes that are more active in the OA-environment

Gene activity that was upregulated in OA-reared crabs was more targeted, with only a handful of enriched processes (Figure 4). OA triggered a heavy investment in transcriptional regulation at multiple levels of transcriptional control. DNA methylation and histone demethylase activity was upregulated in OA-exposed crab, which are epigenetic mechanisms that control transcription through changes to chromatin structure and DNA accessibility [(Gibney and Nolan, 2010)](https://paperpile.com/c/bgbcYo/a56XM). A large percentage of upregulated genes (18%) were associated with the regulation of transcription from RNA polymerase II promoters. Increased expression was also detected in genes that code for post-translational gene control, such as Exportin-5, which is involved in mRNA silencing by microRNAs [(Yi et al., 2003)](https://paperpile.com/c/bgbcYo/qwHwS). Given that OA results in widespread downregulation of a variety of biological functions, we posit that a primary purpose of the transcriptional regulatory response of OA-reared RKC is to shut down (or dampen) less-critical metabolic and biosynthesis processes as part of the overall stress response. In addition to the broad transcriptional effects, these regulatory elements may be responsible for the unleashing of transposable element activity in response to ocean acidification [(Horváth et al., 2017)](https://paperpile.com/c/bgbcYo/Sqm0O).

Cell signaling and nervous system development was substantially more active in OA-exposed crab. Many upregulated genes were associated with signal transduction, and interacted with or spanned cell membranes (e.g. Teneurin-m, Semaphorin-1A, Ankyrin-2). Increased expression of these genes suggest that OA increases the need to transfer information between the external and internal environments, particularly across the plasma membrane, to regulate activity inside the cell. Investment in signaling molecules may also reflect negative effects of OA on transmembrane signaling, perhaps by damaging the membrane or cytoskeleton (e.g., due to oxidative stress, [(Pörtner, 2010)](https://paperpile.com/c/bgbcYo/9jSi5)), or by decreasing signaling molecule binding affinity [(Roggatz et al., 2016; Porteus et al., 2021)](https://paperpile.com/c/bgbcYo/vjyBj+2eBSn). Investment in the nervous system may also be necessary to fortify systems that monitor conditions in the environment. A gene coding for the protein “pinocchio”, which was upregulated in severe OA, is expressed in the antenna of some arthropods (e.g., fruit fly [(Rollmann et al., 2005)](https://paperpile.com/c/bgbcYo/6XRaX)), and acts as a chemosensory receptors [(Rollmann et al., 2005; Vizueta et al., 2020)](https://paperpile.com/c/bgbcYo/0yJtV+6XRaX). OA-associated carapace dissolution around neuritic canals is correlated with damaged setae in larval Dungeness crab or their underdevelop [(Bednaršek et al., 2020)](https://paperpile.com/c/bgbcYo/0huT0). Setae are important sensory structures which are innervated with chemo- and mechano-receptors [(Smolowitz, 2021)](https://paperpile.com/c/bgbcYo/zccch). Further, OA can alter the response of crabs to chemosensory cues suggesting a reduction in their ability to detect such cues (Draper and Weissburg 2022). The heavy investment in neurogenesis found here may be one way that OA-reared crab counteract the negative effects of OA to external sensory structures.

Genes involved in negative regulation of growth and cell proliferation were also upregulated in OA conditions (Figure 4). Decreased growth often occurs when invertebrates are exposed to OA (references). Here, we find that controlling growth is one of only a handful of processes that are upregulated in OA-reared RKC, which suggests that it is a critical survival mechanism. Differentially upregulated genes include negative growth regulators Menin, Brain tumor protein, and Forkhead box protein O (FOXO). FOXO is a transcription factor that specifically inhibits growth in response to cellular stressors, including oxidative stress and nutrient deprivation [(Furukawa-Hibi et al., 2005)](https://paperpile.com/c/bgbcYo/QDVmL), and may therefore be one regulatory mechanism connecting oxidative stress with OA exposure [(Melzner et al., 2019)](https://paperpile.com/c/bgbcYo/cLsPr) and decreased growth rate in RKC and related species [(Long et al., 2013, 2017, 2019)](https://paperpile.com/c/bgbcYo/4gdpO+k5lvR+uWfS2). As slower growth is associated with OA resilience in other invertebrates [(Waldbusser et al., 2016; Swezey et al., 2020)](https://paperpile.com/c/bgbcYo/ydm9J+ZHZ17), a deeper investigation into these and similar genes and their variants across genotypes may help to identify populations that are less sensitive to OA.

Interestingly, a previous study reported that OA-exposed juvenile RKC predominantly upregulated genes related to the cuticle [(Stillman et al., 2020)](https://paperpile.com/c/bgbcYo/rVCv0), which we did not see. On the contrary, five genes coding for cuticle proteins (CP1158 & CPAM119) were downregulated in severe OA-reared crab (but cuticle processes were not enriched). The Stillman et al. (2020) experimental design was similar to the present study in many ways (similar laboratory & OA conditions), but the treatments were shorter in duration (~3 weeks) and occurred only during the juvenile stage. The transcriptional response of juvenile RKC may therefore depend on whether they are OA-naive, or if they were previously exposed to OA conditions. Exposure to OA and other stressors during the larval development stage may be particularly impactful, as it can alter an organism’s physiological trajectory [(Pechenik, 2006; Block et al., 2008)](https://paperpile.com/c/bgbcYo/gEQQh+h2nKt). For instance, larval exposure to OA carries over to affect growth and shell strength later in life in the Olympia oyster [(Hettinger et al., 2012, 2013; Sanford et al., 2014)](https://paperpile.com/c/bgbcYo/bwwIB+IdChG+AVuzU). Another possible factor influencing the transcriptional response to OA is when RKC are exposed relative to their molt cycle, which greatly influences gene expression [(Chang and Mykles, 2011; Mykles and Chang, 2020)](https://paperpile.com/c/bgbcYo/c7Zx9+mCzHw). We sampled juveniles early in the intermolt stage and, because of the design, know that all crabs were sampled at the same stage in the molt cycle (at ~5 days post molt), while the crabs sampled by Stillman et al. (2020) were not standardized by where they were in the molt cycle, thus possibly capturing some effects of OA on crabs in the early pre-molt stage when the new exoskeleton is being synthesized. A time-series analysis of the transcriptome in varying OA environments, particularly across multiple stages of the molt-cycle, would provide a more integrated view of the crustacean OA response.

##### OA decreases gene expression variability, which is not explained by genetic differences

The variability in gene expression was much lower among individuals reared in OA treatments than those reared in ambient pH. This is evident in the PCA constructed from genome-wide expression data (Figure 1), which reveals tighter clustering as OA treatment severity increases. Genes upregulated in OA-reared crabs were also expressed at more consistent levels, particularly in the severe OA-reared crab (Figure 2b, Table 3), suggesting the need for highly controlled levels of transcript that perform critical functions. Given the possibility for genotype-specific gene expression influencing our transcriptional results, we investigated whether genetic composition differed among OA treatments (i.e., did larvae from only a few families survive in OA treatments). We found no evidence of genetic differences among treatments: samples did not cluster by treatment in the PCA, and no families were over- or under-represented in any treatment. The constricted gene expression pattern observed in juveniles reared in OA treatments therefore is not attributed to genotype-specific expression. Rather, molecular signatures described above are plastic responses to OA conditions across many families. This may reflect a lack of standing genetic variability that is needed to fuel rapid adaptation to ocean acidification [(Bitter et al., 2019)](https://paperpile.com/c/bgbcYo/jTRCr), as transcriptional variability and other measures of phenotypic plasticity in response to OA may be associated with the tolerance of a species or population [(Kenkel and Matz, 2017)](https://paperpile.com/c/bgbcYo/yuCqw), or their potential for adaptive selection [(Crispo, 2008)](https://paperpile.com/c/bgbcYo/01UIE). As the OA literature continues to mature, closer attention should be paid to transcriptional variability, and how it relates to the species’ tolerance.

## Conclusion

RKC are quite sensitive to OA, even when compared to closely related species [(Coffey et al., 2017; Long et al., 2019)](https://paperpile.com/c/bgbcYo/REazo+k5lvR). Our findings reveal that RKC reared in OA conditions had a constellation of gene expression patterns indicative of a response to environmental stress. They were physiologically constrained, resulting in downregulated metabolic processes and immune function [(Sokolova et al., 2012)](https://paperpile.com/c/bgbcYo/P0Ezo). OA may therefore impact RKC growth in part through metabolic depression, and survival indirectly by compromising their ability to fight infection or respond to secondary stressors. Only a handful of processes are upregulated in OA, indicating a critical need for a more active and developed nervous system, and tight regulation of transcription and growth. Given the universal and invariable upregulation of transposable elements in OA-exposed crab, TE activity may serve as an effective stress response by producing novel or cryptic transcripts, but these effects may only be fully realized in future generations. Multistressor studies are needed, with a focus on the immunological effects of OA, the role (and vulnerability) of the nervous system in an acidified world, and the potential role of TE’s in fueling adaptation.

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# Supplemental - *still in progress*

**Supplemental Table 1**: Number of differentially expressed genes and enriched GO terms by pairwise treatment contrast. Colors indicate the number of DEGs and GO terms that were upregulated and downregulated in the more severe OA treatment, where ambient = pH 8.0, moderate = pH 7.8, and severe = pH 7.5.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **OA treatment contrast** | No. of DEGs | No. of enriched Biological Processes | No. of enriched Molecular Functions | No. of enriched Cellular Components | % Transposable Elements |
| Ambient vs Moderate | 526 ⬆  933 ⬇ | 9 ⬆  23 ⬇ | 7 ⬆  18 ⬇ | 2 ⬆  16 ⬇ | 51.4% ⬆  30.6% ⬇ |
| Ambient vs Severe | 2,350 ⬆  3,907 ⬇ | 5 ⬆  81 ⬇ | 10 ⬆  47 ⬇ | 1 ⬆  29 ⬇ | 64.5% ⬆  27.7% ⬇ |
| Moderate vs Severe | 22 ⬆  25 ⬇ | 3 ⬆  2 ⬇ | 5 ⬆  0 ⬇ | 0 ⬆  0 ⬇ | 85.7% ⬆  12.5% ⬇ |

**Supplemental Table 2:**  Statistics for modules with eigengenes that correlate significantly with pCO2 concentration. Modules in red and green indicate modules for which gene expression decreases and increases with pCO2, respectively. The percent of genes that are transposable elements are also included for each module. Modules are randomly assigned color names, and the letters correspond to Figure 5 panels in the main text.

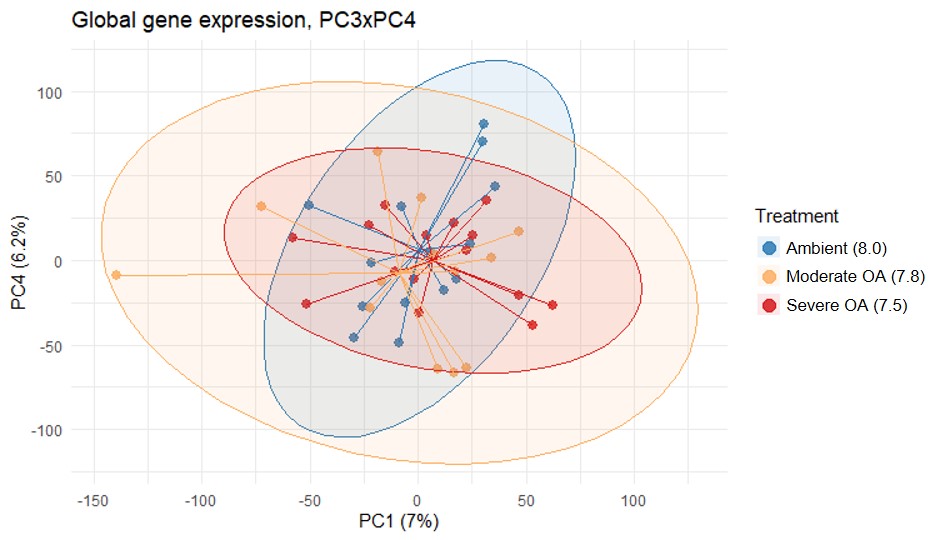
|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Module** | **No. of genes in module** | **No. of genes in module that are also differentially expressed** | **Weighted Pearson correlation with pCO2 (P-value)** | **% Transposable Elements** | **No. of enriched biological process GO terms at EASE<0.05**  **(FDR 10%)** |
| (a) magenta | 6,602 | 1,280 | -0.52 (4.5e-4) | 45% | 43 (11) |
| (b) lightcyan | 4,169 | 1,171 | -0.45 (2.9e-3) | 39% | 60 (31) |
| (c) firebrick4 | 1,573 | 191 | -0.39 (9.7e-3) | 60% | 4 (0) |
| (d) darkviolet | 92 | 17 | -0.37 (0.016) | 23% | 6 (4) |
| (e) royalblue | 1,245 | 197 | -0.36 (0.019) | 32% | 19 (3) |
| (f) coral1 | 162 | 21 | -0.34 (0.028) | 40% | 5 (0) |
| (g) plum1 | 449 | 40 | -0.31 (0.042) | 53% | 6 (1) |
| (h) pink | 8,245 | 980 | -0.30 (0.049) | 63% | 50 (15) |
| (i) blue2 | 96 | 16 | 0.36 (0.018) | 92% | 0 |
| (j) lightsteelblue1 | 306 | 20 | 0.42 (5.4e-3) | 53% | 2 (0) |
| (k) ivory | 301 | 84 | 0.44 (3.4e-3) | 43% | 13 (0) |
| (l) lightgreen | 7,080 | 515 | 0.45 (2.7e-3) | 76% | 13 (4) |
| (m) purple | 2,862 | 328 | 0.46 (2.1e-3) | 77% | 7 (4) |
| (n) green | 8,603 | 1,453 | 0.50 (6.6e-4) | 68% | 13 (2) |

**Supplemental Table 3**: [Enrichment results from DEG analysis](https://docs.google.com/spreadsheets/d/1Jo5gOZgxBS92_PTSmZVJhdQFfyr35j7Xrzvw_hiHVfk/edit?usp=sharing) - Tab “DEGs”

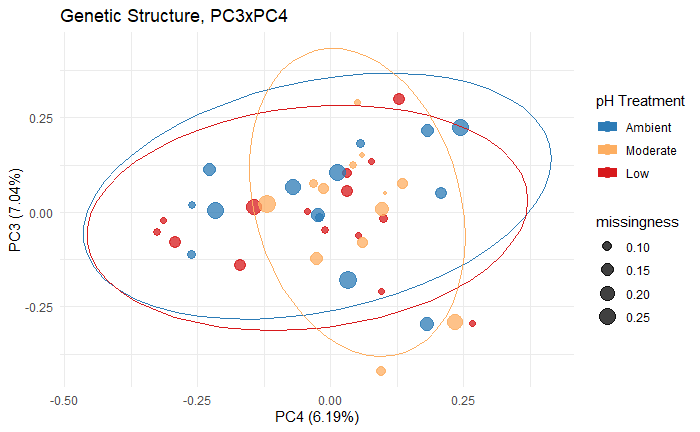
**Supplemental Table 4**: [Enrichment results from low-CV genes](https://docs.google.com/spreadsheets/d/1Jo5gOZgxBS92_PTSmZVJhdQFfyr35j7Xrzvw_hiHVfk/edit?usp=sharing) - Tab “Upregulated Low CV genes”

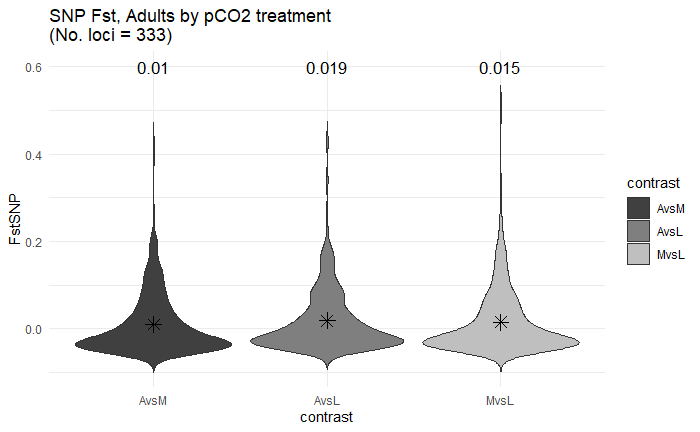
**Supplemental Table 5**: [Enrichment results from WGCNA modules](https://docs.google.com/spreadsheets/d/1Jo5gOZgxBS92_PTSmZVJhdQFfyr35j7Xrzvw_hiHVfk/edit?usp=sharing) - Tab “WGCNA modules”

**Supplemental Table 6:** Transposable elements detected, list of 67 unique Uniprot IDs



**Supplemental Figure 1.** P31 x PC4 constructed from all genes.





**Supplemental Figure 2**: Pairwise Fst distributions among OA treatments

## 

**Supplemental Figure 3**: Pairwise Fst distributions among OA treatments