

1 Resilience to sensory variance in the primary visual 2 cortex

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8 ABSTRACT

Our daily endeavors occur in a complex visual environment, whose intrinsic variability challenges the way we integrate information to make decisions. By processing myriads of parallel sensory inputs, our brain is theoretically able to compute the variance of its environment, a cue which is known to guide our behavior. Yet, the neurobiological and computational basis of such variance computations are still poorly understood. Here, we quantified the dynamics of sensory variance modulations of cat primary visual cortex neurons. We report two archetypal neuronal responses, one of which is resilient to changes in variance and can co-encode the sensory feature and its variance, significantly improving the population encoding of orientation. The existence of these variance-specific responses could be accounted for by a model of intracortical recurrent connectivity. We thus propose that local recurrent circuits process uncertainty as a generic computation, advancing our understanding of how the brain handles naturalistic inputs.

10 Introduction

Selectivity to the orientation of visual stimuli is an archetypal feature of the neurons in the mammalian primary visual cortex (V1)¹, which has been historically studied using low-complexity stimuli such as oriented gratings². While this approach offers a clear hypothesis as to what neurons are responding to, it only probes for neural selectivity to individual input parameters, such as orientation or spatial frequency. Natural vision, however, involves rich cortical dynamics³ integrating a mixture of multiple local parameters and global contextual information⁴. Hence, a majority of our understanding of V1 relies on neural responses to single inputs in orientation space, rather than naturalistic responses to multiple orientations.

This knowledge gap is not trivial, as the variance of distributions of sensory inputs is a fundamental cue on which our brain relies to produce coherent integration of sensory inputs and prior knowledge of the world^{5,6} in order to drive behavior⁷. According to Bayesian inference rules, low-variance inputs are processed through fast feedforward pathways, whereas higher sensory variance elicits a slower, recurrent integration⁸. How the brain performs computations on variance is not yet fully understood. In V1, it has been shown that single neurons undergo nonlinear tuning modulations as a function of their input's variance⁹ which can serve as a functional encoding scheme^{10,11}. These recent results align with earlier models of recurrent cortical activity of V1^{12,13} and also match psychophysical measurements in humans^{14–16}. While it seems that local interactions within V1 are sufficient to encode orientation variance¹⁷, the quantification of single neuron responses, their dynamics and their link to a functional population encoding of variance remains to be established.

Here, we investigate the neural basis of variance processes in V1 using stimuli matching the orientation content of natural images¹⁸. We present a quantitative analysis of single neurons' variance-tuning functions, as well as their dynamics, reporting heterogeneous modulations. Two archetypal response type emerge in V1, one of which relies on predominantly supragranular neurons that maintain robust orientation tuning despite high sensory variance, allowing them to co-encode orientation and variance, and enhancing V1's orientation distribution encoding. A well-established V1 intracortical recurrence model accounts for these “resilient neurons”, aligning with canonical Bayesian frameworks⁶ and suggesting uncertainty computations as a new generic function for local recurrent cortical connectivity.

33 Results

34 Single neuron response in V1 depends on input variance

We recorded neural activity from 249 anesthetized cat V1 neurons, and measured orientation-selective responses to naturalistic images called Motion Clouds¹⁸. These stimuli are band-pass filtered white noise textures and offer three advantages over both simple grating-like stimuli and complex natural images. First, they enable fine control of mean θ and variance, controlled by

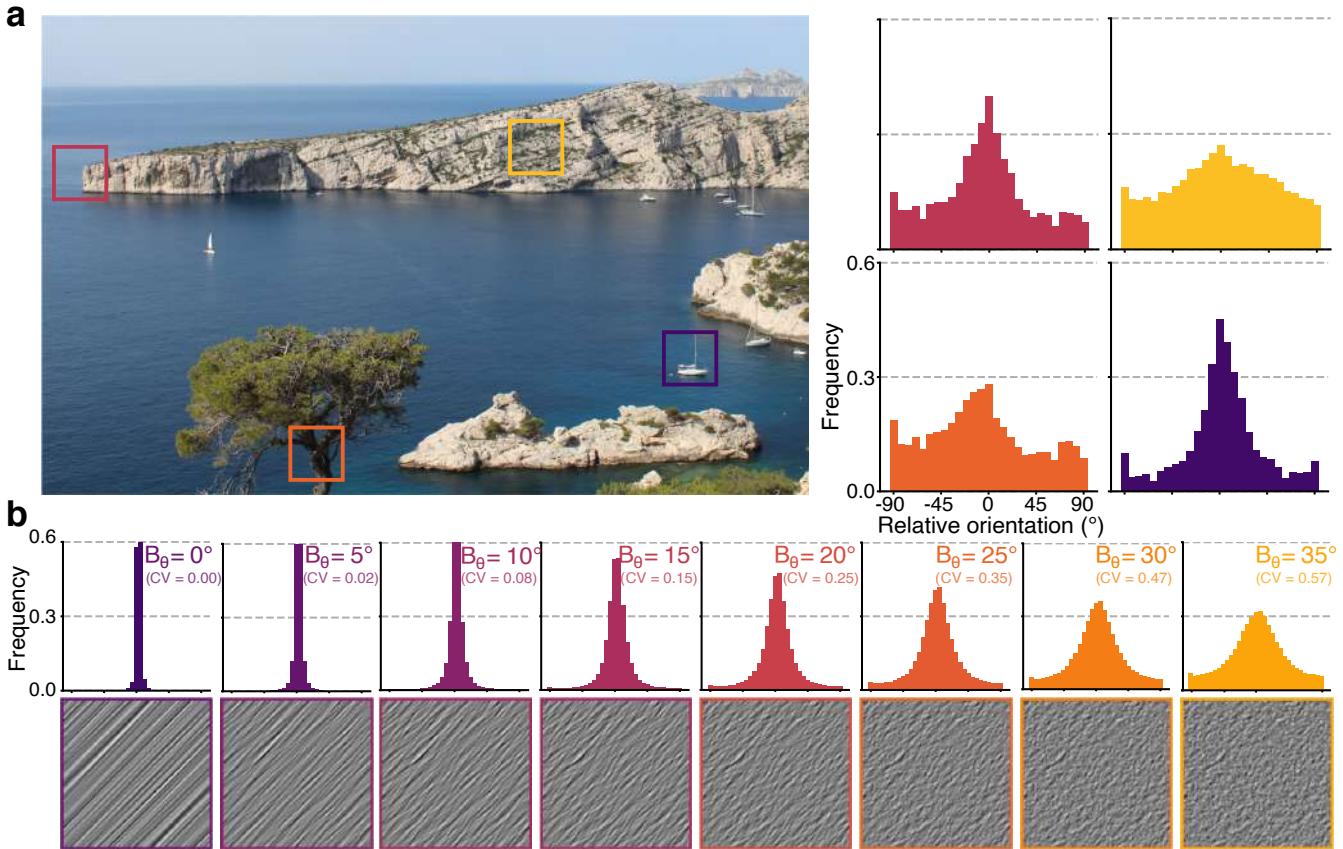


Figure 1. Variance of orientation distributions characterizes local regions of natural images. **(a)** Distributions of orientation from four 200x200 px regions of a natural image (picture taken by H.J.L.) obtained by an histogram of oriented gradients (32x32 px/cell), centered around the most frequent orientation. **(b)** Motion Clouds, naturalistic stimuli (bottom row) with mean orientation $\theta = 45^\circ$ and increasing variance (B_θ) from left to right. Distributions of orientation of the stimuli are shown on the upper row. Circular Variance (CV) of the distribution is shown for comparison.

38 B_θ , of orientation distributions through a generative model, thereby reproducing natural images' oriented content (Figure 1).
 39 Second, as they are stationary in the spatial domain, they only probe orientation space, excluding any second-order information
 40 exploitable by the visual cortex¹⁹. Third, by conforming to natural images' $1/f^2$ power spectrum distribution²⁰, they attain a
 41 desirable balance between controllability and naturalness²¹. We generated 96 Motion Clouds by varying mean orientation θ
 42 between 0° and 180° in 12 even steps and variance B_θ between $\approx 0^\circ$ and 35° in 8 evenly spaced steps.

43 All recorded neurons displayed orientation selectivity to Motion Clouds. Nearly all (98.8%, $p < 0.05$, Wilcoxon signed-rank
 44 test) units maintained their preferred orientation when variance B_θ increased, while peak amplitude of tuning curve diminished
 45 significantly (95.1% units, $p < 0.05$, Wilcoxon signed-rank test, 73.1% mean amplitude decrease for $B_\theta = 35^\circ$). Only 28.5%
 46 of the recorded units were still tuned for $B_\theta = 35.0^\circ$ stimuli ($p < 0.05$, Wilcoxon signed-rank test). Thus, increasing input
 47 variance reduces single neuron tuning, which manifests heterogeneously across neurons, as evidenced by two representative
 48 single units shown in Figure 2a. Neuron A illustrates single units which are no longer orientation-tuned when variance B_θ
 49 reaches 35° ($W = 171.0$, $p = 0.24$, Wilcoxon signed-rank test), unlike neuron B ($W = 22.5$, $p = 10^{-6}$) which exemplifies the
 50 aforementioned 28.5% variance-resilient units. These response types are characterized by functions relating B_θ to goodness of
 51 tuning (circular variance, CV), named here "variance-tuning functions" (VTF, Figure 2b). Such VTFs represent the input/output
 52 transformation in variance space, and are well-fitted with Naka-Rushton functions²² (Figure S 2a). This allows to summarize
 53 variance modulations using only three parameters: n , the VTF non-linearity; $B_{\theta50}$, the input variance level for the tuned-untuned
 54 state transition; and f_0 , the orientation tuning goodness for lowest-variance inputs. Overall, VTFs exposed diverse responses to
 55 variance among V1 neurons, with median values outlining a characteristic VTF that is slightly non-linear, with a changepoint at
 56 $B_\theta = 19.2^\circ$ (Figure 2c). In other words, most neurons tend to change abruptly in tuning when input variance reaches 19.2° ,
 57 after which the response become less sensitive to orientation. Alternative metrics were also calculated, including variance-half
 58 width at half height (HWHH) and variance-maximum response functions (Figure S 2b-e). Although HWHH displayed patterns

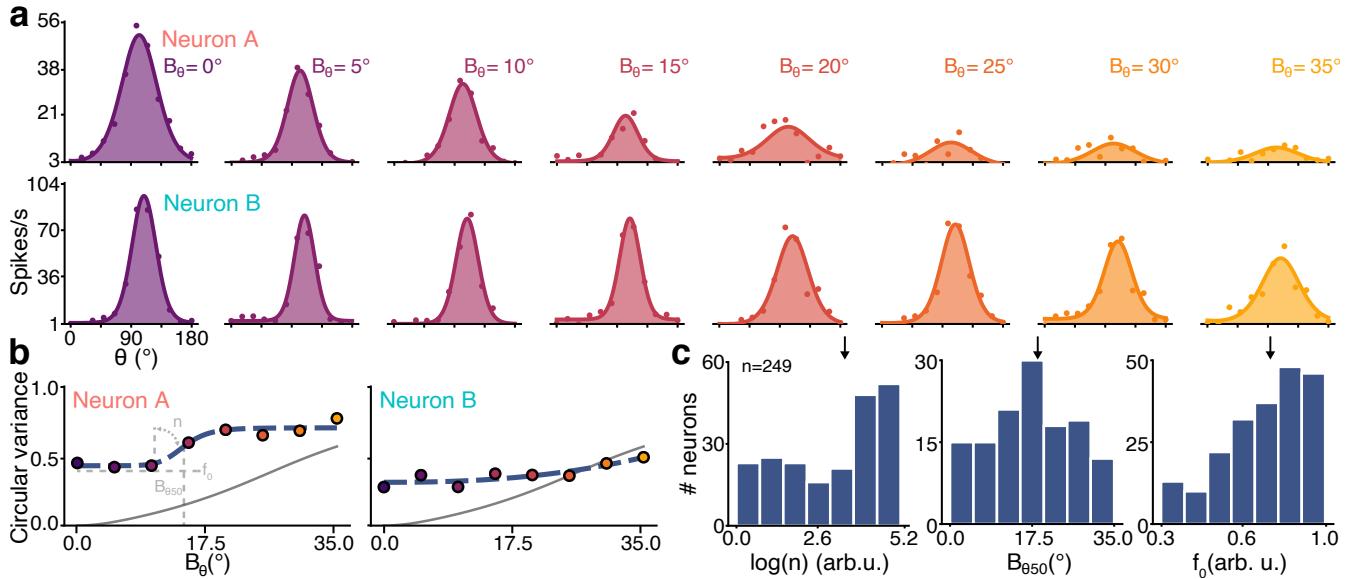


Figure 2. Single neuron tuning correlates with input variance. Additional examples are shown in Figure S 1. (a) Tuning curves of two neurons responding to Motion Clouds of increasing variance B_θ . Dots indicate the mean firing rate across trials (baseline subtracted, 300 ms average) and lines represent a fitted von Mises function. (b) Variance-tuning functions (VTF), relating the change of orientation tuning measured by the circular variance (CV, dots) as a function of input variance B_θ , fitted with a Naka-Rushton (NKR) function (dashed curves, parameters shown in light gray). Parameters of the VTF are $\log(n) = 8.4$, $B_{\theta50} = 14.7^\circ$, $f_0 = 0.4$ for neuron A and $\log(n) = 2.4$, $B_{\theta50} = 35.0^\circ$, $f_0 = 0.3$ for neuron B. The CV identity curve is shown in solid gray. (c) Histograms of the NKR parameters (in the [5%;95%] range of possible NKR fitting values) for the 249 recorded units. Median values are indicated by a black arrow ($\log(n) = 3.6$, $B_{\theta50} = 19.2^\circ$, $f_0 = 0.75$).

resembling VTFs, we elected to not use it, as its reliance on fits, its consequent susceptibility to fitting artifact, and its similarity with CV are not desirable properties. Since CV also inherently accounts for the firing rate at the preferred orientation (see Methods), we relied on this metric to describe both maximum amplitude and goodness of tuning in a single metric.

Orientation variance impacts not only orientation tuning, but also the dynamics of the response of V1 neurons (Figure 3). Interestingly, both effects are linked, as demonstrated by the two example VTFs: neuron B, which exhibited orientation-tuned responses for $B_\theta = 35^\circ$ inputs (Figure 2a), also had slower time-dependent change of goodness of tuning (relative min. of reduction of 42% of max. CV at 200 ms post-stimulation onset, $B_\theta = 0^\circ$) compared to neuron A (relative min. of 26% of max. CV at 90 ms post-stimulation onset, Figure 3b). These dynamical modulations were also heterogeneously distributed amongst the population, significantly more spikes emitted 200 ms after stimulation onset for $B_\theta = 35^\circ$ (Figure 3d, $U = 14936.0$, $p < 0.001$, Mann-Whitney U-test). In summary, orientation variance induces changes in both tuning and dynamics of V1 neurons, revealing two archetypal types of response: either fast in time and non-linear with respect to variance (neuron A) or slow in time and linear with respect to variance (neuron B).

Multiple types of variance responses are found in V1

To properly characterize the two aforementioned types of responses to variance, we separated the recorded neurons in two groups using K-means clustering the Principal Components (PC, Figure 4) of the neuronal responses. Clustering was performed on the VTFs (Figure 4b), tuning statistical measurements (Figure 4c,d) and response dynamics (Figure 4e,f). We used the first 2 PC for clustering the data, which accounted for 39.1% of cumulative variance (Figure S 4a), and chose two clusters based on the number of example responses and the empirical absence of an “elbow”²³ in the Within-Clusters-Sum-of-Squares (WCSS) curve (Figure S 4b). This splits the data into a cluster of 164 neurons, including neuron A, and another cluster of 85 neurons associated with neuron B’s response type. As neuron B displayed resilience to increased input variance (Figure 2a), its cluster was labeled “resilient neurons.” Conversely, neurons clustered with neuron A were labeled “vulnerable neurons” (blue and red colors, respectively, Figure 4a). Opting to categorize the data into two distinct response types facilitates a comprehensive understanding of the underlying continuum of behaviors. This approach has proven successful in the characterization of novel visual responses, such as V1 simple/complex cells²⁴ and MT pattern/component cells²⁵. To further enhance the analysis, a continuum-based analysis is provided in Figure 6f.

The K-means clustering resulted in significant difference between the two groups’ VTF parameters (Figure 4b): resilient

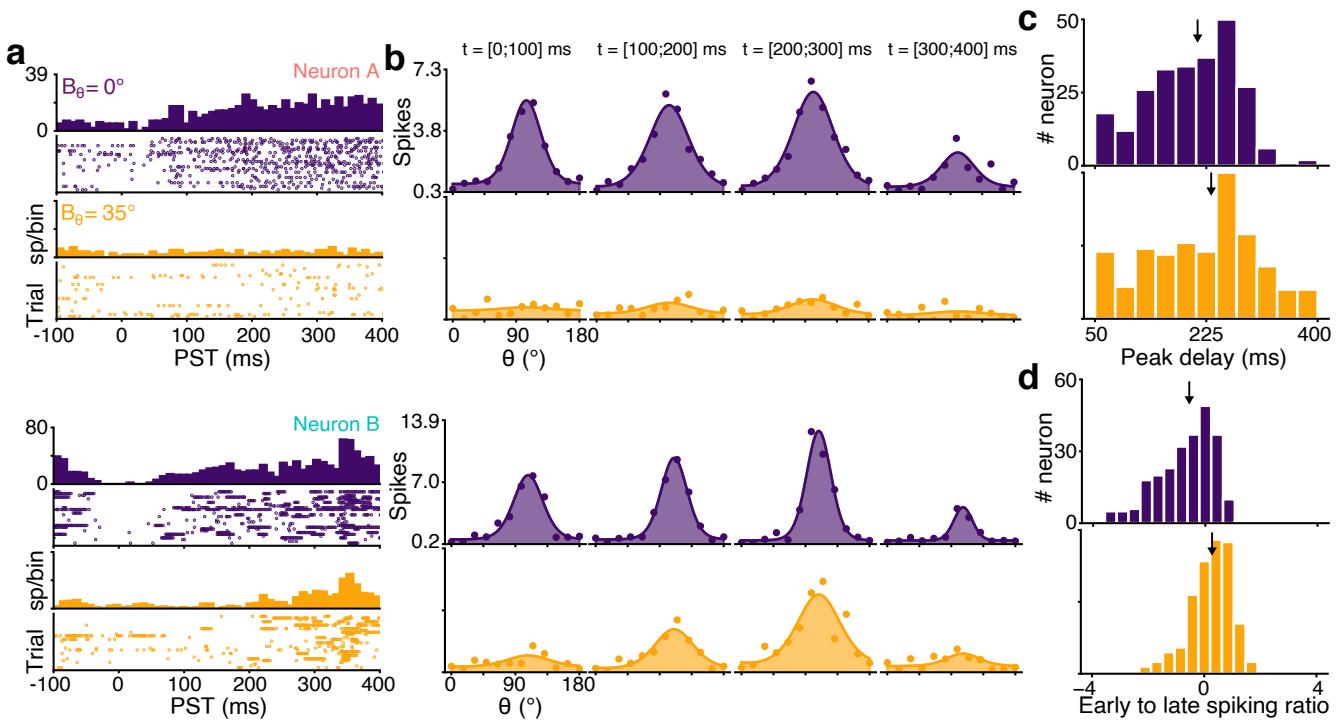


Figure 3. Neural dynamics depend on input variance. Additional examples are shown in Figure S 3. (a) Peristimulus time (PST) histogram and rasterplot for the two previous example neurons, at variance $B_\theta = 0^\circ$ (purple) and $B_\theta = 35^\circ$ (yellow). (b) Tuning curve dynamics in 100ms windows, starting at labeled times. (c) Delay to peak amplitude of tuning curves for $B_\theta = 0^\circ$ (purple, median = 210 ms) and $B_\theta = 35^\circ$ (yellow, median = 233 ms) for the population. Median values are indicated by a black arrow. (d) Log ratio of early (< 100 ms post-stimulation) and late (> 200 ms) spike counts for $B_\theta = 0^\circ$ (median = -0.54) and $B_\theta = 35^\circ$ (median = 0.27) for the population.

neurons had significantly more linear modulations ($\log(n)$, $U = 4029.0.0$, $p < 0.001$, Mann-Whitney U-test), higher change-points ($B_{\theta50}$, $U = 7854.0$, $p = 0.028$) and better tuning to low-variance inputs (f_0 , $U = 4992.0$, $p < 0.001$), which endows them with the ability to respond to orientation on a broader range of input variances^{26,27}. No significant differences in the variance-HWHH and variance-firing rate functions were observed, except for the non-linearity of the latter metric (Figure S 5). This is coherent with the clustering on the statistical measurement of orientation tuning, which showed that resilient neurons remained significantly tuned to higher values of B_θ ($B_{\theta\max}$, Figure 4c, $U = 9155.0$, $p < 0.001$). However, both groups of neurons had similar circular variance for $B_\theta = 35^\circ$ (Figure 4d). This suggests that both types of neurons were similarly poorly tuned for inputs of highest variance, but underwent different tuning changes between $B_\theta = 0^\circ$ and $B_\theta = 35^\circ$. In terms of dynamics, the two groups exhibited the same differences that characterized neurons A and B. Resilient neurons discharged significantly later than vulnerable neurons for $B_\theta = 0^\circ$ (Figure 4e, $U = 8455.5$, $p = 0.002$), but both groups were on par for inputs of $B_\theta = 35^\circ$ ($U = 7794.5$, $p = 0.063$). Interestingly, resilient neurons had significantly lower time to maximum amplitude of the tuning curve for $B_\theta = 0^\circ$ (Figure 4f, $U = 5542.5$, $p = 0.014$), which opposes the early/late ratio of spikes. Neither group showed variance-dependent modulation of the delay to maximum spike count ($U = 3058.0$, $p = 0.084$ and $U = 11545.5$, $p = 0.090$ for resilient and vulnerable, respectively), and both groups showed similar delay for $B_\theta = 35^\circ$ ($U = 6094.5$, $p = 0.158$).

The existence of these two groups of neurons could not be attributed to the integration of the drifting motion of the stimuli (direction selectivity index, unused in the clustering process, Figure 4g, $U = 7031.5$, $p = 0.910$). Instead, the location of the recorded units (unused in the clustering process) predominantly positioned the resilient neurons in supragranular layers, offering a mechanistic basis for their existence (Figure 4h). Moreover, resilient neurons have sharper orientation tuning and slower dynamics, which are distinctive features of supragranular neurons^{28,29}. This, however, does not establish a functional role for these two types of responses in V1.

Population level modulations of the orientation code

As the neuronal population has been separated in well-characterized groups, we wish to understand the functional role played by resilient and vulnerable neurons. To that end, we used a neuronal decoder that probes for population codes in V1, enabling us to

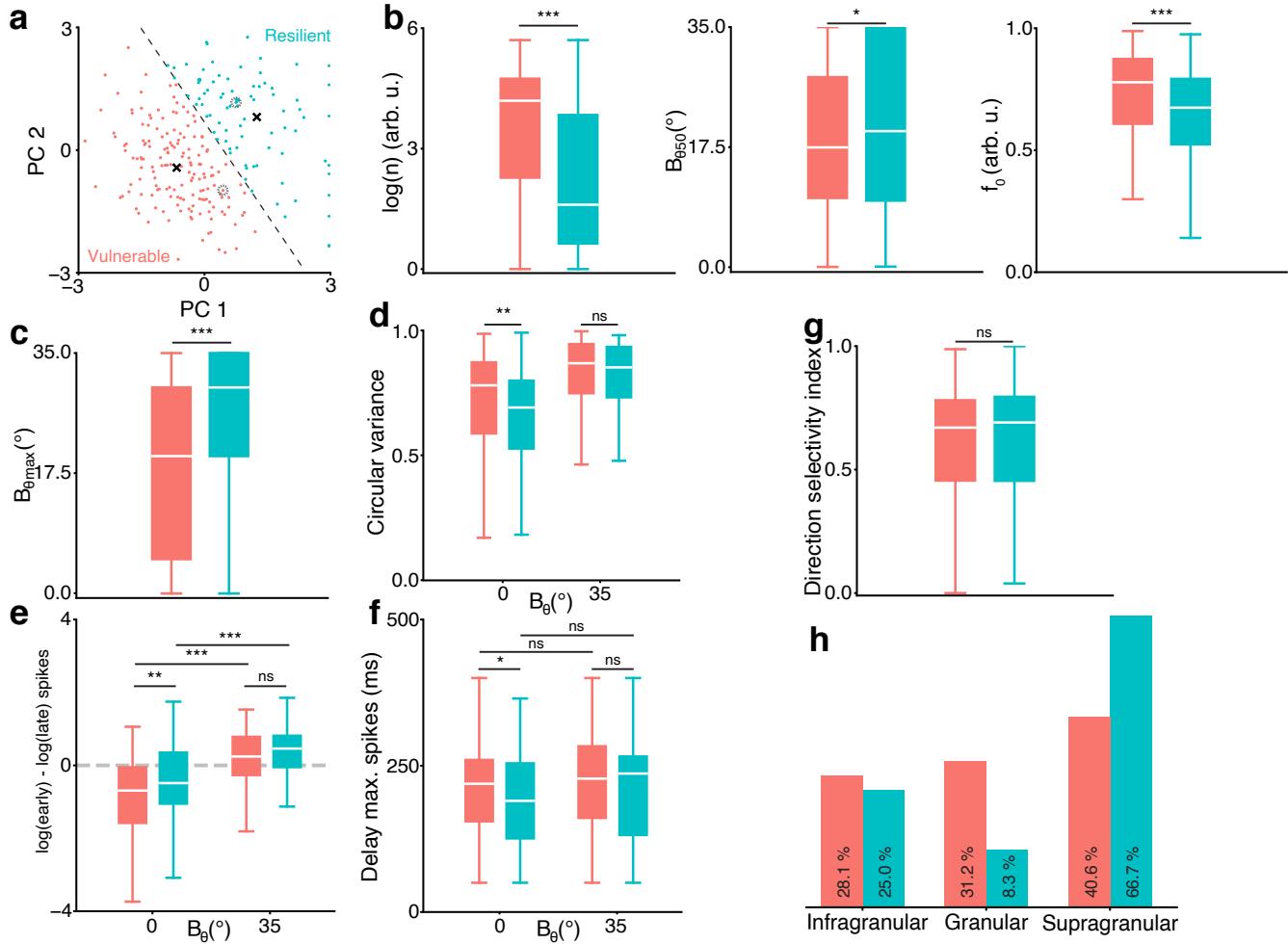


Figure 4. Responses to changes in variance fall into two categories. (a) Principal Components (PC) analysis of the data, K-Means clustered (2 clusters, centroids shown as black crosses and separatrix as dashed line). 9 resilient neurons with $PC_1 > 3$ are plotted at $PC_1 = 3$. Neuron A and B are shown as dashed circles. (b) Boxplot of the VTF parameters $\log(n)$, $B_{\theta50}$, f_0 (ns, not significant; *, $p < 0.05$; **, $p < 0.01$, ***, $p < 0.001$ Mann-Whitney U-test). (c) Maximum B_θ for significant orientation tuning curve. (d) Circular variance at $B_\theta = 0^\circ$ and $B_\theta = 35^\circ$. (e) Log ratio of the early (< 100 ms) and late (> 200 ms) spike counts at $B_\theta = 0^\circ$ and $B_\theta = 35^\circ$. (f) Delay to maximum peak amplitude of tuning curves at $B_\theta = 0^\circ$ and $B_\theta = 35^\circ$. (g) Direction selectivity index (unused in the clustering). (h) Laminar position (unused in the clustering).

seek what parameters of the stimuli each neuron group was encoding. We trained a multinomial logistic regression classifier³⁰, a probabilistic model that classifies data belonging to multiple classes (see Methods). This classifier received the firing rate of neurons in a sliding time window (100 ms) and learned, for each neuron, a coefficient that best predicts the class (i.e. the generative parameter θ , B_θ or $\theta \times B_\theta$) of the stimulus.

This decoder was first used to probe for representation of the stimuli's orientations θ in the population activity. For this purpose, the dataset of trials was separated for each variance, such that 8 independent, B_θ -specific, orientation decoders were learned, with optimal parametrization (Figure S 6). These orientation decoders were able to retrieve the correct stimulus' θ well above the chance level (1 out of 12 orientations, max. accuracy = 10.56 and 4.68 times chance level for $B_\theta = 0^\circ$ and $B_\theta = 35^\circ$, respectively) from the entire population recordings. The temporal evolution of these decoders' accuracy (Figure 5a) showed that maximally accurate orientation encoding correlates almost linearly with the stimulus's variance, as does the time to reach this accuracy (Figure 5e, black). These dynamics depend on the input's variance, exhibiting a rapid initial rise followed by a plateau for low-variance inputs, while steadily increasing linearly over time for high-variance inputs. Interestingly, the decoding accuracy remained stable for approximately 100 ms even after a stimulus was no longer displayed. Since the decoders are trained independently in each time window, this accumulative process occurs in the recordings themselves, and not in the decoder.

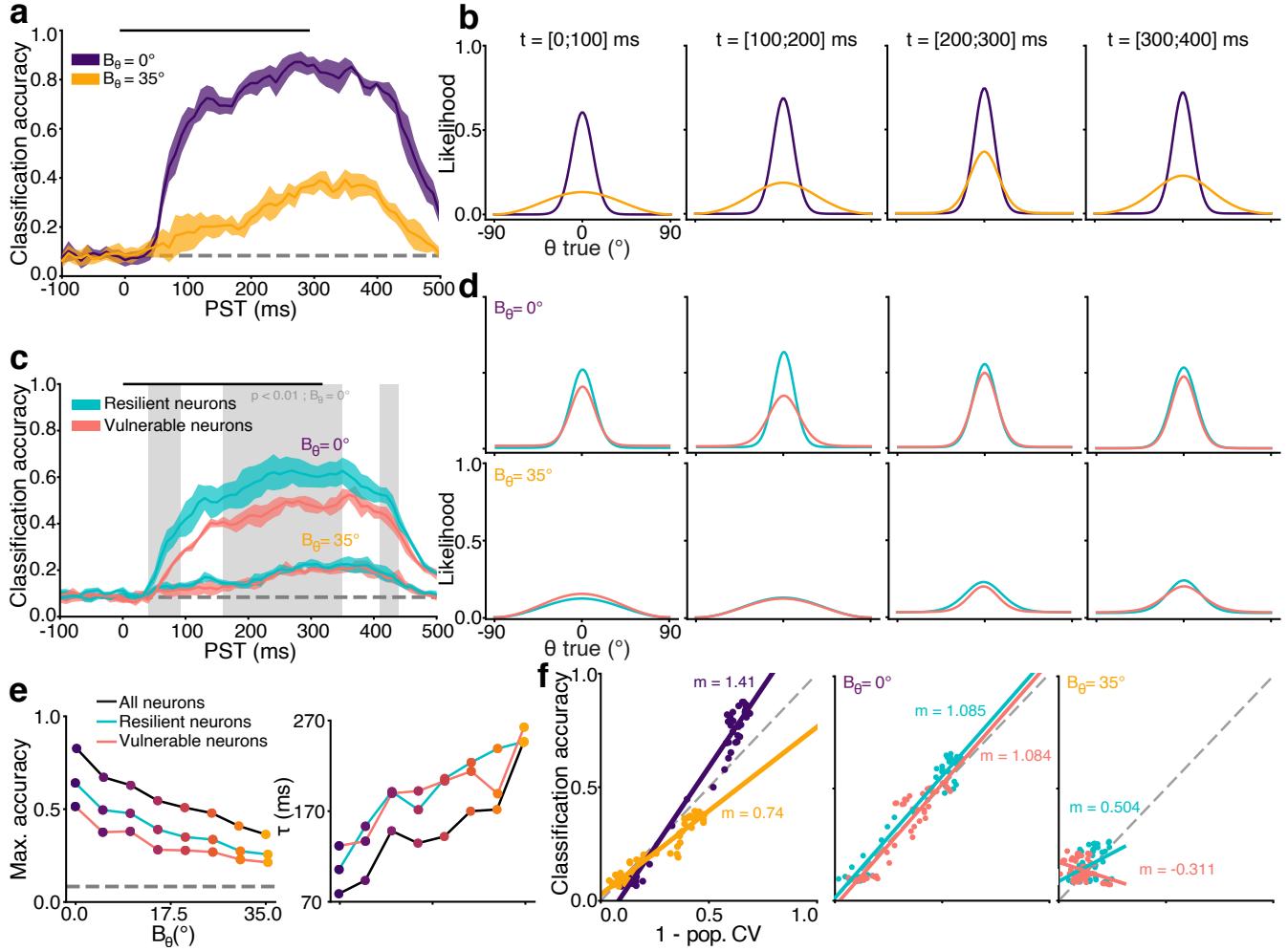


Figure 5. Input variance modulates orientation decoding in V1. (a) Time course of orientation θ decoding accuracy at two variances B_θ . Lines are the mean accuracy of 5 random resampling of 100 neurons and contour the SD. Significantly better decoding at chance level is represented by a gray dashed line and stimulation time by a black line. (b) Population tuning curves with a von Mises fit, showing the likelihood of decoding each θ in four time windows. (c) Same as (a) for the two groups of neurons. (d) Same as (b) for the two groups of neurons, with $B_\theta = 0^\circ$ (upper row) and $B_\theta = 35^\circ$ (lower row). (e) Time course parameters for three decoders at all B_θ , estimated by fitting a sigmoid up to PST = 300 ms. τ is the time constant. (f) Correlation between classification accuracy and population circular variance for the whole population (left), for both groups with $B_\theta = 0^\circ$ (middle) and $B_\theta = 35^\circ$ (right). Linear regression are shown as solid lines with slope m indicated (all significant, $p < 0.001$, Wald Test with t-distribution).

The full output of these decoders (see Methods) is a population tuning curve, which displays the likelihood of decoding all possible input classes (here, all θ , Figure 5b), rather than the proportion of correct decoding reported by the accuracy metric. The clear correlation between the sharpness of these population tuning curves (Figure 5f left) and the accuracy of the decoder shows that improvements of decoding accuracy rely directly on a population-level separation of features within orientation space³⁰, particularly at higher B_θ (Figure 5b, third panel). Overall, B_θ influences the temporality of the orientation code in V1, which echoes its influence on single-neuron dynamics (Figure 3). The short delay required to process precise inputs is congruent with the feedforward processing latency of V1³¹, while the increased time required to reach maximum accuracy for low precision oriented inputs suggests the involvement of a slower, recurrent mechanism.

We then sought to assert the role of the vulnerable and the resilient neural populations by decoding θ from either group. The number of neurons in each group was imbalanced (79 more vulnerable neurons), which influences the accuracy of the decoder (Figure S 6). Consequently, we randomly selected (with replacement) groups of 100 neurons from either population, repeating the selection 5 times. Using the same approach as with the global population decoding, we then trained B_θ -specific

136 orientation decoders on the activity of either group of neurons. Resilient neurons outperformed vulnerable ones in decoding
137 accuracy for 56% of the timesteps, mainly in the 160 – 330 ms period (Figure 5c). However, both groups exhibited similar
138 population tuning curves (Figure 5d) and time courses (Figure 5e). Despite the better tuning of resilient neurons to inputs
139 with higher variance (Figure 4), both groups have overall similar orientation encoding performances for $B_\theta = 35^\circ$. Therefore,
140 orientation can be decoded somewhat more effectively from the resilient neurons at the population level, but neither group
141 appears to have a clear or stable advantage over the other in this regard, especially at higher B_θ .

142 A subset of V1 neurons co-encode orientation and its variance

143 Given that orientation encoding did not reveal a fundamental difference in the respective contributions of resilient and vulnerable
144 neurons, we then investigated the encoding of the stimulus' variance B_θ . The same type of decoder previously used failed to
145 infer the variance B_θ (chance level = 1 out of 8 values of B_θ , max. accuracy = 1.91 times chance level) from the population
146 activity (Figure S 8a,b). This variance decoding also failed to reach more than twice the chance level (max. accuracy = 1.72
147 and 1.71 times chance level for resilient and vulnerable neurons, respectively) in both resilient and vulnerable neurons (Figure
148 S 8c,d). At the single neuron level, tuning curves flatten with increments of variance (Figure S 2a), which makes it difficult
149 to distinguish activity generated by stimuli with $B_\theta = 0.0^\circ$ and orthogonal orientation from the activity generated by stimuli
150 with $B_\theta = 35.0^\circ$ and preferred orientation. This limitation could potentially stem from the recording scale (249 neurons),
151 which is more than an order of magnitude smaller than the quantity of neurons a single V1 “biological decoder” can access³².
152 Thus, neither the decoding of variance B_θ nor the decoding of orientation θ accounts for a different role between resilient and
153 vulnerable neurons.

154 The decoding methods used so far have assumed that V1 encodes independently single input parameters. However, a more
155 realistic assumption is to consider the visual system’s natural inputs as distributions of information (Figure 1) that cortical
156 neurons must process from thalamic inputs³³ based on a probabilistic computational principle³⁴. Here, this implies that the
157 naturalistic form of processing for a V1 neuron would be co-encoding both the mean feature (θ) and its associated variance
158 (B_θ) to access the entire probability distribution.

159 We thus proceeded to train a decoder that retrieves both orientation and variance of the stimulus’ simultaneously, referred to
160 as a $\theta \times B_\theta$ decoder. This decoder correctly predicted orientation and variance with a maximum accuracy reaching 16.36 times
161 the chance level (1/96, Figure 6a, gray). The likelihood structure (Figure 6b, upper row) showed that the correct θ was decoded
162 with alongside multiple concurrent hypothesis over B_θ . The progressive increase of accuracy stems from the emergence of
163 a dominant encoding of θ at the correct B_θ , consequently diminishing the relative magnitude of representations over other
164 B_θ values over time. Interestingly, resilient neurons showed here a significantly different functional role from vulnerable
165 neurons, with markedly better co-encoding of B_θ and θ (max. accuracy = 11.0 and 9.0 times chance level for resilient and
166 vulnerable neurons, respectively, Figure 6a, blue, red). Both groups displayed ambiguity regarding B_θ (Figure 6b, lower row),
167 and correlated sharpening/accuracy ratios on the correct B_θ population curve (Figure 6c, left) or on the off-median population
168 curves (Figure 6c, right).

169 To understand the utility of this co-encoding, we marginalized the decoder over B_θ , creating an orientation-only encoder
170 that simultaneously learned both orientation and variance. Data from resilient neurons then provided significantly better
171 encoding of orientation than vulnerable neurons (max. accuracy = 6.0 and 5.4 times the 1/12 chance level for resilient and
172 vulnerable neurons respectively, Figure 6d, gray regions), demonstrating that the overall V1 orientation code improves with a
173 co-decoding of its variance. The distinction between resilient and vulnerable neurons is further emphasized by the decoder
174 coefficients, which represent the contributions of each type of neurons towards the overall $\theta \times B_\theta$ code (Figure 6e, for single
175 neuron examples see Figure S 9). Here, these coefficients are depicted as a polar plot, where the orientation θ (centered around
176 preferred orientation) is shown as the angle of each bin from the upper vertical and the variance B_θ is represented as the
177 eccentricity of each bin from the center. Visualizing the coefficients of the whole population decoder (i.e., trained on the 249
178 neurons, Figure 6a, gray) shows that the output learned from resilient neurons concurrently informs about both a wide range of
179 orientations and variances, as observed by the extent of the bins in the eccentricity (B_θ) axis (Figure 6e, bottom row). On the
180 other hand, the decoding process extracted orientation information on a very small range of B_θ from the activity of vulnerable
181 neurons (Figure 6e, top row). Even though the coefficients are learned independently at each time steps, the difference of
182 information between the two groups of neurons remains extremely stable through time.

183 Overall, orientation and its variance can be co-decoded simultaneously from resilient neurons, while only orientation can
184 be decoded from vulnerable neurons. This is confirmed by a continuous score-based decoding metric based on the K-means
185 parameters (Figure 6f) that correlates, for the entire population (i.e., without splitting in two groups), their maximum decoding
186 accuracy to a degree of vulnerability/resilience. After providing this functional rationale for resilient and vulnerable neurons,
187 we finally address the question of how both types of neurons can exist in V1.

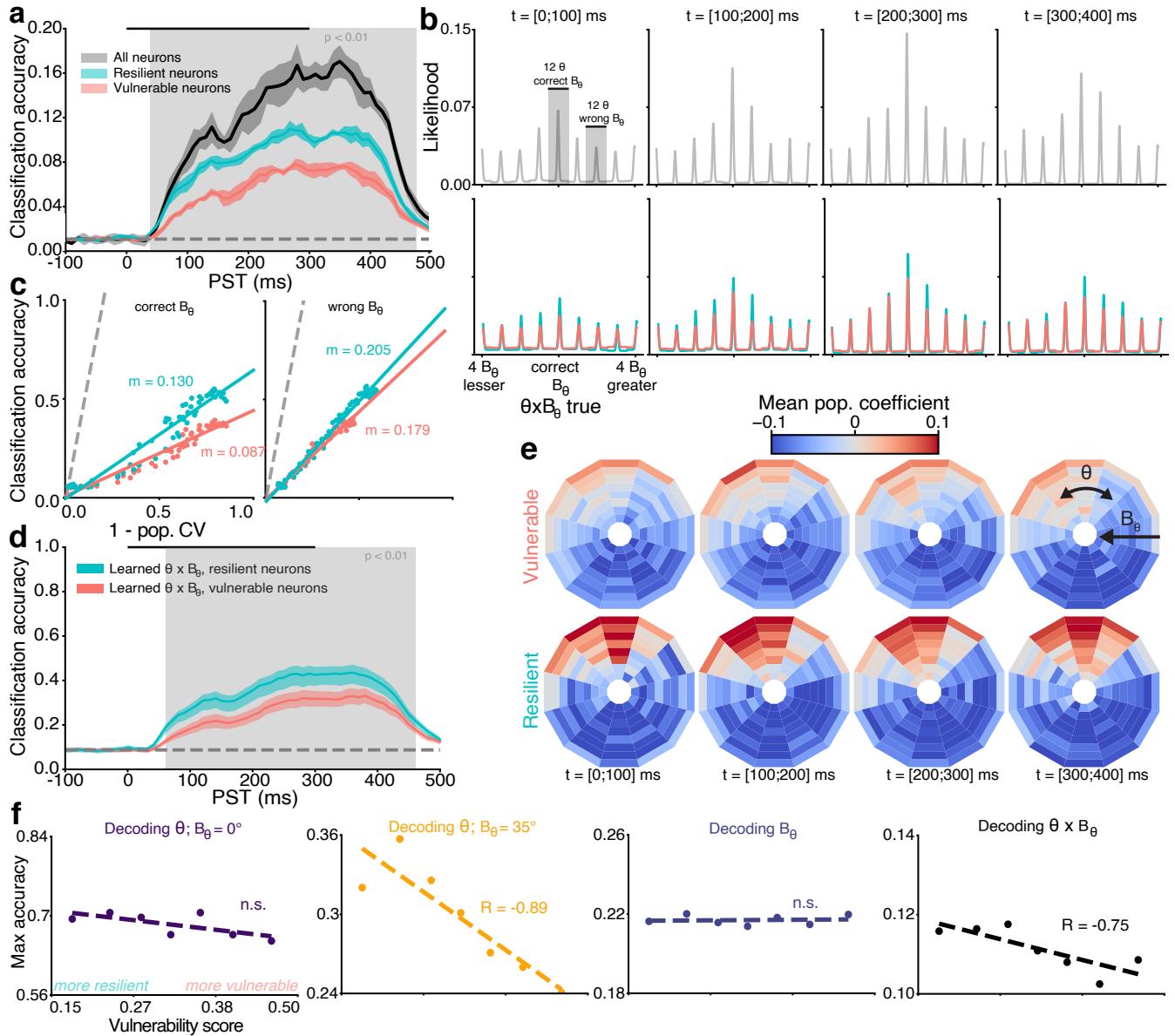


Figure 6. Orientation and its variance can be decoded from resilient neurons. **(a)** Time course of the accuracy for decoding $\theta \times B_\theta$ of Motion Clouds. Lines are the mean accuracy and contour the SD. Significantly better decoding from resilient neurons is shown as a gray overlay (Wilcoxon signed-rank test, threshold $p < 0.01$). Decoding at chance level (1/96) is represented by the gray dashed line. **(b)** Population tuning curves for the likelihood of decoding each $\theta \times B_\theta$ in four time windows, centered around the correct $\theta \times B_\theta$. **(c)** Correlation between classification accuracy and population circular variance for correct B_θ population tuning curves (left) and averaged across other B_θ tuning curves (right). Linear regression are shown as solid lines with slope m indicated (all significant $p < 0.001$, Wald Test with t-distribution). **(d)** Time course of the $\theta \times B_\theta$ decoder, marginalized over B_θ to produce θ -only outputs. **(e)** Mean decoding coefficients of the two groups yielded from the whole population $\theta \times B_\theta$ decoder. **(f)** Score-based decoding for θ (first and second columns), B_θ (third) and $\theta \times B_\theta$ (fourth). Raw scores (points) are fitted with a linear regression (dashed curve), with Spearman R shown in case of a significant correlation ($p < 0.05$).

188 Recurrent activity can explain the existence of neurons co-encoding orientation and variance

189 A notable difference between vulnerable and resilient neurons is their different location within the cortical layers (Figure 4h).
 190 This typically implies differences in local circuitry, particularly in the intra-V1 recurrent interactions between cortical columns,
 191 which are mostly confined to supragranular layers³⁵. Given that resilient neurons are predominantly found in these supragranular
 192 layers, we aimed to find a mechanistic rationale for the existence of the two groups of neurons based on local interactions in V1.

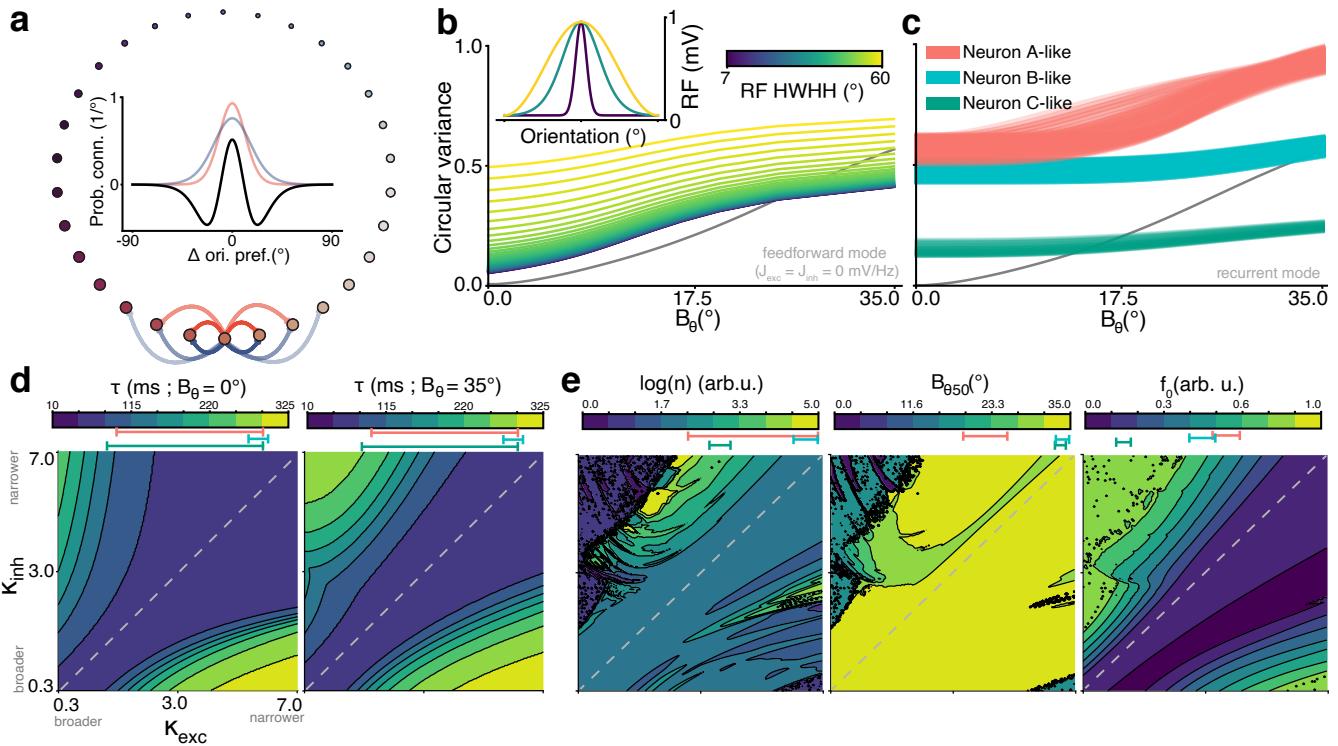


Figure 7. Recurrent interaction modeling can explain the existence of resilient and vulnerable neurons. **(a)** Ring topology of the network, with preferred orientation of each neuron. Inset: Recurrent connectivity profile for each neuron, computed as a difference (black) of excitatory (red) and inhibitory (blue) profiles, controlled by a measure of concentration κ_{exc} and κ_{inh} , respectively. **(b)** VTF without recurrent connectivity (i.e. only inputs convolved with receptive fields) and with varying RF's Half-Width at Half-Height (HWHH). The CV identity curve is shown in black. Inset shows examples of receptive fields RF. **(c)** VTF with recurrent connectivity, under two configurations retrieved by searching for VTF parameters close to those of neuron A, B (Figure 2b) and C (Figure S 1). **(d)** Delay to half maximum firing rate of the model (τ) for each connectivity profile, shown as a contour plot of κ_{exc} and κ_{inh} . [5%;95%] range of the parameters corresponding to the VTFs in (c) are displayed below the scale bars. **(e)** VTF parameters obtained from the model for each connectivity profile, shown as a contour plot of κ_{exc} and κ_{inh} . [5%;95%] range of the parameters of the VTFs shown in (c) are displayed below the scale bars.

We developed a neural network from a well-established computational model of recurrent connectivity in V1, originally used to account for the intracortical activity in cat V1³⁶ and later simplified as a center-surround filter in the orientation domain²⁹. This model has already accounted for an extensive range of emerging properties in cortical circuits^{37,38}. Briefly, it is built of orientation selective neurons tiling the orientation space and connected amongst themselves via recurrent synapses which follow an excitatory/inhibitory difference of von Mises distributions (Figure 7a). Here, we model inputs with higher variance as more spread in orientation space (Figure 1) and thus in model space, which hence drives the recurrent dynamics of the model based on B_θ (for a full description, see Methods).

Considering that feedforward connectivity with heterogeneous tuning can encode mixtures of orientations and natural images⁹, we first ran our model without recurrent synapses. We reproduced the heterogeneous selectivity by convolving the input with tuning curves of varying bandwidths (Figure 7b, inset). This feedforward mode of the network was only able to produce a limited number of responses (Figure 7b), in which increasing the bandwidth of the tuning curves increased the parameter f_0 of the VTF, but kept n and $B_{\theta50}$ constant.

Barring that explanation, we focused on the role of recurrent synapses and disabled the convolution of inputs. We varied the concentration parameters of the synaptic distributions κ_{inh} and κ_{exc} (Figure 7c,e) in 200 even steps ranging from 0.35 to 7, yielding 40,000 possible configurations of the model. This allowed to manipulate the VTF and to accurately reproduce those of single neurons recorded in V1 (neuron A, B in Figure 2b and C in Figure S 1, modeled in Figure 7c). Altering the type of recurrence between neurons with different orientation preference allowed to reproduce all VTF found in V1. The parameter spaces (Figure 7e) showed a trend for resilient VTFs (low n , high $B_{\theta50}$, low f_0) to be found mostly around the $K_{\text{exc}} ; K_{\text{inh}}$ identity line, thus produced by a balanced recurrent connectivity. Vulnerable VTFs (high n , low $B_{\theta50}$, high f_0) were, on the

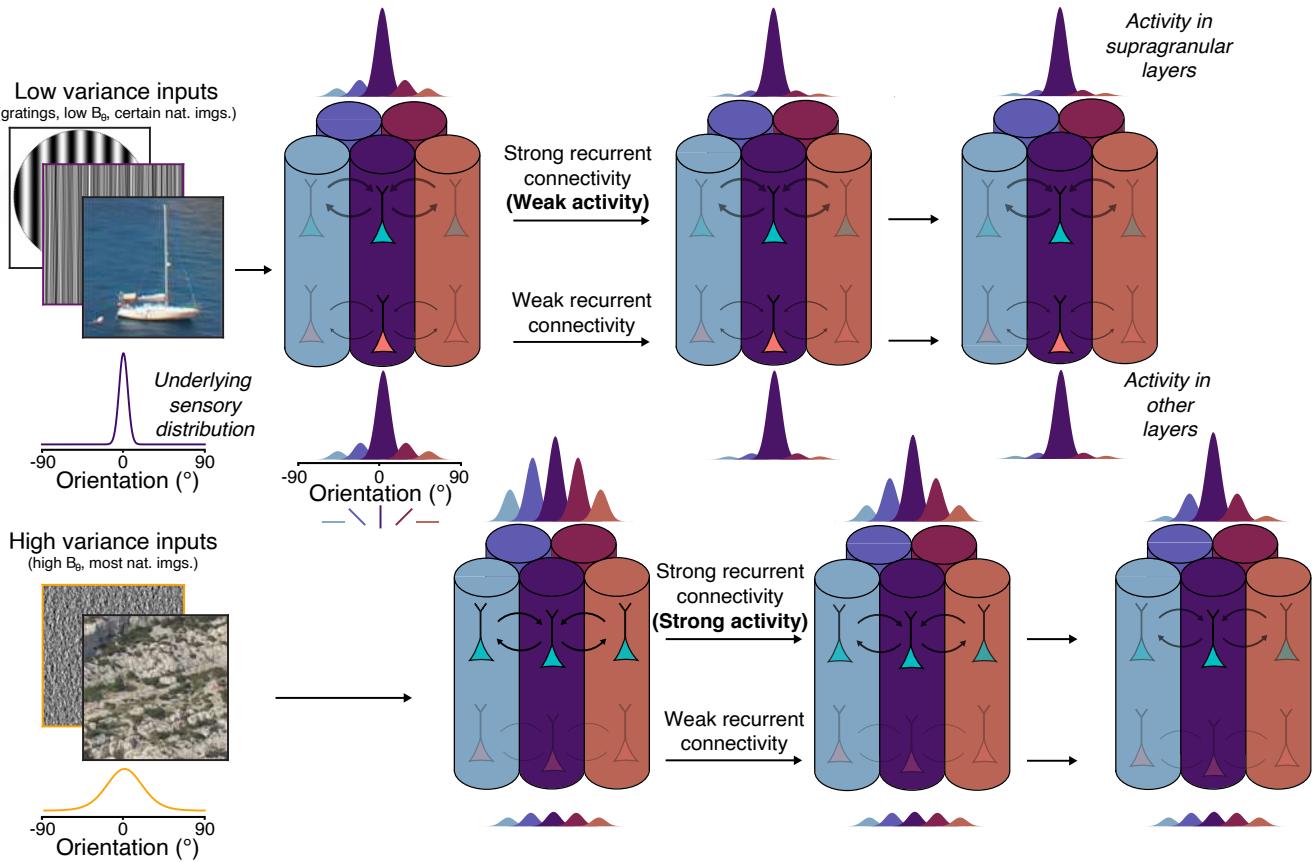


Figure 8. Summary of the findings. The top row is a representation of a set of orientation selective units (here, columns) processing low variance inputs, while the bottom row schematizes the processing of high variance inputs. In the case of low variance inputs to V1, the underlying sensory distribution is sharp in the orientation space, driving mostly a single orientation selective unit which processes orientation in fast feedforward manner. The feature encoded by this activity then stays stable through time (from left to right). For inputs of higher orientation variance, the sensory input is broadly distributed in the orientation space, which drives many dissimilarly tuned units, thus recruiting slow recurrent interactions. The quality of feature encoding progressively increases through time, as recurrent interactions perform computations to represent the most salient oriented feature in the input.

contrary, mostly found above the identity line, where the configuration of the network is dominated by excitation over inhibition. This is consistent with the range of parameters that yielded higher response latency (Figure 7d), which also occupied more parameter space when input variance increased. In summary, recurrence between V1 neurons seems to be sufficient to explain the existence of vulnerable and resilient neurons and, consequently, to account for the co-encoding of orientation and variance.

Discussion

The variance of oriented inputs to V1 impacts orientation selectivity⁹ and we have sought to understand how V1 could process this input parameter. We found that variance causes modulations in tuning (Figure 2) and dynamics (Figure 3) of single V1 neurons, which we have classified as either vulnerable or resilient (Figure 4). Decoding analysis revealed variance-dependent accumulative dynamics in the two groups of neurons (Figure 5) that are directly tied to a population-level separation of features within orientation space³⁰. Both groups can encode orientation but not variance (Figure S 8), and only resilient neurons are able to accurately co-encode orientation and variance of the input to V1 (Figure 6). Based on cortical layer position (Figure 4h) and on a computational approach (Figure 7), we propose that the processing input variance in V1 is supported by recurrent connectivity between local cortical populations (Figure 8). This not only improves the encoding of orientation in V1, but also links directly to canonical Bayesian frameworks, suggesting uncertainty computation as a new mechanism supported by local recurrent cortical connectivity.

Here, we restricted our approach to orientation space, rather than investigating the full extent of spatial relationships

which are present in natural images. Thus, full-field stimuli without second-order correlation were used, which compared to a purely ecological environment, have likely excluded end-stopped cells³⁹. Whilst this approach limited the responses to V1 and excluded higher-order cortical areas, there exists both neurobiological and computational evidences that V1 does not need to recruit other cortical areas to process orientation variance. For instance, the heterogeneous recurrent excitatory and inhibitory synaptic connectivity in V1^{40–43} sustains resilient orientation tuning⁴⁴ that can account for the diversity of single neurons' resilience under different connectivity profiles, as explored in our computational model (Figure 7). This is supported by the temporal scale of local recurrent connectivity, namely the slowly-conducted horizontal waves in an orientation map⁴⁵, which fit the view of variance processing as an iterative and accumulative computation implemented by local recurrent interactions between supragranular resilient neurons that are heavily connected through recurrent interactions with neighboring cortical columns^{28,29,35,45}. In this regard, our reported time scales may have been slightly affected by the use of anesthesia (halothane), which has a limited visible effect on V1^{46,47} and is less likely to cause modulations in this area compared to higher-order areas^{48–51}.

Computationally, most existing models support the idea that processing orientation variance can be achieved solely with local V1 computations¹⁰. For instance, Goris *et al.*⁹, reported that heterogeneously tuned V1 populations help encode the orientation distributions found in natural images, and that this functional diversity could be accounted for by a linear-nonlinear (L-NL) model. While this could explain the diversity of tuning in our data (Figure 2), we found that such a model failed to account for some types of modulations of the VTFs (Figure 7b). Therefore, we employed a model designed to replicate intracortical cat V1 data³⁸ and demonstrated that it reproduces various VTFs and dynamics observed in our recordings. The model used here pools activity from multiple orientation-tuned units into a single neuron, which we interpreted as a local recurrent model. While our results do not require contributions from extrastriate regions to explain the observed results, the possibility of recurrence involving neurons outside V1 cannot be entirely ruled out at this time⁵².

Our study confirms the findings in the anesthetized macaque literature⁹ by identifying single-neuron variance modulations that serve as the basis for decoding orientation variance at the population level in V1. This suggests that a common mechanism may underlie this neural mechanism in both felines and primates, which is a fundamental computational requirement for proper encoding of natural images in V1⁵³. Although gain/variance V1 functions have been previously reported¹⁷, we demonstrate a similar input-output relationship in the form of VTFs, that have the added benefit of characterizing and extrapolating variance modulations across the full dynamical range of V1 populations. Further, we finely analyzed the temporal component of the response that is absent from the literature. We propose that all these response properties can be linked to cortical layers, binding the idea that supragranular neurons with sharp tuning and slow dynamics^{28,29} support the co-encoding orientation and its variance.

This leads to an interesting tie to Bayesian inference, namely under the specific case of predictive coding³⁴, that canonically assigns (inverse) variance weighting of cortical activity to supragranular recurrent connectivity^{6,8}, without the need for extrastriate computations. This is an interesting perspective which opens up a general interpretation of our results into the broader context of processing variance/precision/uncertainty at different scales of investigations. Extending the present results to other cortical areas or other sensory modalities would be a simple process, given the generative stimulus framework used here¹⁸, which could yield pivotal new insights into our understanding of predictive processes in the brain.

Methods

Visual stimulation

Motion Clouds are generative model-based stimuli¹⁸ which allow for fine parameterized control over a naturalistic stimuli⁵⁴, which is a desirable trait when probing sensory systems under realistic conditions²¹. They are mathematically defined as band-pass filtered white noise stimuli, whose filters in Fourier space are defined as a parameterized distribution in a given perceptual axis (here, only orientation, but can be extended to speed⁵⁵ and scale⁵⁶). Thus, the Motion Clouds presently used are fully characterized by their mean orientation and their orientation variance, such that a given stimulus S can be defined as:

$$S = \mathcal{F}^{-1}(O(\theta, B_\theta)) \quad (1)$$

where \mathcal{F} is the Fourier transform and O the orientation envelope, characterized by its mean orientation θ and its orientation bandwidth B_θ . For $B_\theta < 45.0^\circ$, $B_\theta = 1/\sqrt{\kappa}$, where κ is the concentration parameter of a von Mises distribution, and hence approximates the standard deviation⁵⁷. It thus serves as a measure of the orientation variability in the pattern, and as such, we used the term "variance" to describe it throughout the text. A total of 96 different stimuli were generated, with 12 mean orientations θ ranging from 0 to π in even steps, and 8 orientation variance B_θ ranging from ≈ 0 to $\pi/5$ in even steps. The orientation envelope is a von Mises distribution:

$$O(\theta, B_\theta) = \exp \left\{ \frac{\cos(2(\theta_f - \theta))}{4 \cdot B_\theta^2} \right\} \quad (2)$$

277 where θ_f is the angle of the frequency components of the envelope in the Fourier plane, which controls the spatial frequency
 278 parameters of the stimuli, set here at 0.9 cycle per degree. The stimuli were drifting orthogonally in either direction with respect
 279 to the mean orientation θ at a speed of 10°/s, which is optimal to drive V1 neurons⁵⁸. For the range of values of B_θ considered
 280 here, the orientation envelope approximates a Gaussian distribution and B_θ is thus a measure of the variance of the orientation
 281 content of the stimuli.

282 All stimuli were generated using open-source Python code (see Additional information) and displayed using Psychopy⁵⁹.
 283 Monocular stimuli were projected with a ProPixed projector (VPixx Technologies Inc.) onto an isoluminant screen (Da-Lite[®])
 284 covering 104° × 79° of visual angle. All stimuli were displayed for 300 ms, interleaved with a mean luminance screen (25
 285 cd/m²) shown for 150 ms between each trial. Trials were fully randomized, and each stimulus (a unique combination of
 286 $\theta \times B_\theta \times$ drift direction) was presented 15 times. Stimuli were shown at 100% contrast, meaning that as B_θ increased, the
 287 amount of orientation energy at median orientation θ decreased, and conversely for off-median orientations (as illustrated in
 288 Figure 1b). This differs from manipulating the contrast, which would reduce the orientation energy at all orientations.

289 **Surgery**

290 Experiments were conducted on 3 adult cats (3.6 - 6.0 kg, 2 males). All surgical and experimental procedures were carried
 291 out in compliance with the guidelines of the Canadian Council on Animal care and were approved by the Ethics Committee
 292 of the University of Montreal (CDEA #20-006). Animals were initially sedated using acepromazine (Atravet®, 1 mg/kg)
 293 supplemented by atropine (0.1 mg/kg). Anesthesia was induced with 3.5% isoflurane in a 50:50 mixture of O₂:N₂O (v/v).
 294 Following tracheotomy, animals underwent artificial ventilation as muscle relaxation was achieved and maintained with an
 295 intravenous injection of 2% gallamine triethiodide (10 mg/kg/h) diluted in a 1:1 (v/v) solution of 5% dextrose lactated Ringer
 296 solution. Through the experiment, the expired level of CO₂ was maintained between 35 and 40 mmHg by adjusting the
 297 tidal volume and respiratory rate. Heart rate was monitored and body temperature was maintained at 37°C by means of a
 298 feedback-controlled heated blanket. Lidocaine hydrochlorine (2%) was applied locally at all incisions and pressure points
 299 and a craniotomy was performed over area 17 (V1, Horsley-Clarke coordinates 4-8P; 0.5-2L). Dexamethasone (4 mg) was
 300 administered intramuscularly every 12h to reduce cortical swelling. Eye lubricant was regularly applied to avoid corneal
 301 dehydration.

302 **Electrophysiological recordings**

303 During each recording session, pupils were dilated using atropine (Mydriacyl) while nictitating membranes were retracted
 304 using phenylephrine (Mydrin). Rigid contact lenses of appropriate power were used to correct the eyes' refraction. Anesthesia
 305 was changed to 0.5-1% halothane to avoid anesthesia-induced modulation of visual responses⁴⁷. Finally, small durectomies
 306 were performed before each electrode insertion and a 2% agar solution in saline was applied over the exposed cortical
 307 surface to stabilize recordings. Linear probes ($\approx 1 \text{ M}\Omega$, 1x32-6mm-100-177, Neuronexus) were lowered in the cortical tissue
 308 perpendicularly to the pia and extracellular activity was acquired at 30KHz using an Open Ephys acquisition board⁶⁰. Single
 309 units were isolated using Kilosort 2⁶¹ and manually curated using Phy⁶². Clusters with low amplitude templates or ill-defined
 310 margin were excluded from further analysis. Additional exclusion was performed if a cluster's was unstable (firing rate below
 311 5 spikes.s⁻¹ for more than 30 seconds), or if the neuron was not deemed sufficiently orientation selective ($R^2 < 0.75$ when
 312 fitted with a von Mises distribution). Passing that exclusion step, all remaining neurons responded to Motion Clouds. Laminar
 313 positions were determined by the depth of the recording site with respect to the pia, which was then cross-validated by the
 314 evoked Local Field Potential (LFP) using sink/source analysis^{63,64}.

315 **Single neuron analysis**

316 Orientation tuning curves were computed by selecting a 300 ms window maximizing spike-count variance⁶⁵. The firing rate
 317 was averaged across drift directions and a von Mises distribution⁵⁷ was fitted to the data:

$$f(\theta_k) = R_0 + (R_{\max} - R_0) \cdot \exp \{ \kappa \cdot (\cos(2(\theta_k - \theta_{\text{pref}})) - 1) \} \quad (3)$$

318 where θ_k is the orientation of the stimuli, R_{\max} is the response (baseline subtracted) at the preferred orientation θ_{pref} , R_0 the
 319 response at the orientation orthogonal to θ_{pref} and κ a measure of concentration. To control for direction selectivity when
 320 averaging tuning curves across drift direction, we computed a direction selectivity index:

$$D_s = \frac{R_{\text{pref}} - R_{\text{null}}}{R_{\text{pref}}} \quad (4)$$

321 where R_{pref} is the firing rate at the preferred direction (baseline subtracted) and R_{null} the firing rate at the preferred direction
 322 plus π . The quality of each tuning curve was assessed by computing a global metric, the circular variance (CV) of the unfitted

323 data, which varies from 0 for perfectly orientation-selective neurons to 1 for orientation untuned neurons²⁹. It is defined as:

$$CV = 1 - \left| \frac{\sum_k R(\theta_k) \cdot \exp\{2i\theta_k\}}{\sum_k R(\theta_k)} \right| \quad (5)$$

324 where $R(\theta_k)$ is the response of a neuron (baseline subtracted) to a stimulus of angle θ_k . The changes of CV as a function of B_θ
325 were fitted with a Naka-Rushton function²²:

$$f(B_\theta) = f_0 + f_{\max} \frac{B_\theta^n}{B_\theta^n + B_{\theta50}^n} \quad (6)$$

326 where f_0 is the base value of the function, $f_0 + f_{\max}$ its maximal value, $B_{\theta50}$ the stimulus' variance at half f_{\max} and n a strictly
327 positive exponent of the function.

328 The significance of the tuning to orientation was measured by comparing the unfitted firing rate at the preferred and
329 orthogonal orientations across trials, using a Wilcoxon signed-rank test correct for continuity, and the maximum value of
330 B_θ which yielded a significant result was designed as $B_{\theta\max}$ (i.e., the maximum variance at which a neuron is still tuned).
331 Shifts of the preferred orientation were evaluated as the difference of θ_{pref} between trials where $B_\theta = 0^\circ$ and $B_\theta = B_{\theta\max}$. The
332 significance of the variation of peak amplitude of the tuning curve was measured as by comparing the unfitted firing rate at the
333 preferred orientation between trials where $B_\theta = 0^\circ$ and $B_\theta = B_{\theta\max}$.

334 Population decoding

335 The parameters used to generate Motion Clouds were decoded from the neural recordings using a multinomial logistic regression
336 classifier³⁰. For a given stimulus, the activity of all the recorded neurons was a vector $X(t) = [X_1(t) \ X_2(t) \ \dots \ X_{249}(t)]$,
337 where $X_i(t)$ is the spike count of neuron i in a time window $[t; t + \Delta T]$. The onset of this window t was slid from -200 ms to 400
338 ms (relative to the stimulation time) in steps of 10 ms while ΔT was kept constant at 100 ms. It should be noted that merging
339 neural activity across electrodes or experiments is a common procedure^{66,67}, which we validated in our data by verifying that
340 the electrode or experiment which yielded the data could not be decoded from the neural activity (Figure S 7). Mathematically,
341 the multinomial logistic regression is an extension of the binary logistic regression³⁰ trained here to classify the spike vector
342 $X(t)$ between K classes. The probability of any such vector to belong to a given class is:

$$P(y = k | X(t)) = \frac{\exp\{\langle \beta_k, X(t) \rangle\}}{\sum_{k'=1}^K \exp\{\langle \beta_{k'}, X(t) \rangle\}} \quad (7)$$

343 where $\langle \cdot, \cdot \rangle$ is the scalar product over the different neurons, $k = 1, \dots, K$ is the class out of K possible values and β_k are the
344 coefficients learned during the training procedure of the classifier. Several decoders were trained with classification tasks:
345 decoding orientation θ ($K = 12$, Figure 5), decoding orientation variance B_θ ($K = 8$, Figure S 8) or both ($K = 12 \times 8 = 96$,
346 Figure 6). All meta-parameters were controlled, showing that the decoding performances stem mainly from experimental data
347 rather than fine-tuning of the decoder parameterization (Figure S 6). For all decoding experiments reported, we used integration
348 window size $\Delta T = 100$ ms, penalty type = ℓ_2 , regularization strength $C = 1$. and train/test split size = 0.15.

349 The performance of all decoders was reported as the average accuracy across all classes K , known as the balanced accuracy
350 score⁶⁸. The accuracy for each specific class k can also be reported in the form of a population tuning curve, in which the
351 likelihood of decoding each possible class K is given by equation 7. The significance of differences between two neuron
352 groups was reported only when two consecutive timesteps, i.e., 20ms or more, exhibited significant differences. To estimate the
353 timecourse of the decoders, they were fitted in the $[0; 300]$ ms range with a sigmoid function:

$$\sigma = \max_{\text{acc}} \left(\frac{1}{1 + e^{-k\tau}} \right) + \min_{\text{acc}} \quad (8)$$

354 where \max_{acc} and \min_{acc} are respectively the maximum and minimum accuracies of the decoder, k the steepness and τ the time
355 constant of the function. To perform decoding on the same number of vulnerable or resilient neurons, we randomly picked with
356 replacement groups of 100 neurons and bootstrapped this process 5 times.

As the neurons were clustered into two populations for comparison purposes (Figure 4), we also reported the decoding
accuracy based on a continuous vulnerability score (Figure 6f). This score was computed as a sum of neuronal responses
variables significantly different after the clustering, weighted by their mean Principal Component (PC-1 and PC-2) parameters:

$$\text{score} = 1 - W_1(B_{\theta50}) + W_2(1 - \log(n)) + W_3(1 - f_0) + W_4(B_{\theta\max}) + W_5(1 - CV) + W_6(\text{early/late ratio}) + W_7(\text{delay}) \quad (9)$$

357 where W_i is a parameter yielded by the Principal Component Analysis corresponding to its associated neuronal response
358 variable. Each variable is normalized, yielding a scalar score that varies between 0 (most resilient) to 1 (most vulnerable
359 neuron). This score-based decoding was performed on groups of 100 neurons sorted by descending score, and repeated a total
360 of 7 times on increasingly more vulnerable neurons (thus with an overlap of 20 neurons).

361 **Computational model**

362 We used a recurrent network of orientation-tuned neurons to model responses to increasing orientation variance B_θ . The model
 363 presently used was first used to account for the intracortical activity in the cat primary visual cortex³⁶, although it was presently
 364 simplified as a center-surround filter in the orientation domain²⁹. Notably, this network has been able to account for numerous
 365 experimental findings, including learning and adaptation of cortical neurons^{37,38}, whose implementations are similar to ours.

366 The model consisted of N orientation-tuned neurons, evenly tiling the orientation space between $-\pi$ and π . Each neuron is
 367 modeled as a single passive unit whose membrane potential obeys the equation:

$$\tau \delta V / \delta T + V = V_{\text{ff}} + V_{\text{exc}} - V_{\text{inh}} \quad (10)$$

368 where τ is the membrane time constant and V_{ff} , V_{exc} , V_{inh} are the synaptic potentials coming from the feedforward input,
 369 recurrent excitatory and recurrent inhibitory connectivity, respectively. The firing rate R at time t of each neuron is computed as
 370 an instantaneous quantity modulated by a gain α :

$$R(t) = \alpha \cdot \max(V(t), 0) \quad (11)$$

371 For computational simplicity, the neurons had no spontaneous firing rate and V was measured relative to the firing threshold.
 372 Each neuron could send mixed excitatory and inhibitory synaptic potentials to its neighbour, although this specific model has
 373 been reported to achieve similar behaviour with separate units³⁸. For each stimulus of main orientation θ , the input to a cell
 374 with preferred orientation θ_{pref} is:

$$V_{\text{ff}}(\theta_{\text{pref}}) = J_{\text{ff}} \frac{e^{\kappa_{\text{ff}} \cdot \cos(2(\theta - \theta_{\text{pref}}))}}{2\pi I_0(\kappa_{\text{ff}})} \quad (12)$$

375 where J_{ff} is the strength of the input and I_0 is the modified Bessel function of order 0. The right-hand side of the equation
 376 describes a von Mises with mean θ_{pref} and concentration κ_{ff} . This latter parameter is related to the orientation variance B_θ ,
 377 which was varied to yield a model's TVF B_θ/CV curves:

$$B_\theta = \sqrt{\frac{0.5 \arccos((\log(0.5) + \kappa_{\text{ff}})/\kappa_{\text{ff}})}{2 \log(2)}} \quad (13)$$

378 a total of 20 B_θ spanning the same range used in the experiments were used, each with 32 different θ tiling a $[-75^\circ; 75^\circ]$
 379 orientation space. The recurrent connectivity profile for excitatory (C_{exc}) and inhibitory (C_{inh}) synapses was controlled by
 380 separate von Mises distributions over the orientation space Θ :

$$C_{\text{exc}}(\theta_{\text{pref}}) = \frac{e^{\kappa_{\text{exc}} \cdot \cos(2(\theta - \theta_{\text{pref}}))}}{2\pi I_0(\kappa_{\text{exc}})} \quad (14)$$

381

$$C_{\text{inh}}(\theta_{\text{pref}}) = \frac{e^{\kappa_{\text{inh}} \cdot \cos(2(\theta - \theta_{\text{pref}}))}}{2\pi I_0(\kappa_{\text{inh}})} \quad (15)$$

382 which are both used to describe an overall connectivity kernel:

$$C_{\text{tot}}(\theta_{\text{pref}}) = J_{\text{exc}} C_{\text{exc}} - J_{\text{inh}} C_{\text{inh}} \quad (16)$$

383 which followed a typical Ricker wavelet (or Mexican hat) shape (Figure 7d). The overall activity of the network is then a
 384 weighted sum of the firing rates of all the neurons:

$$V_{\text{exc}} - V_{\text{inh}}(t) = \sum_{\Theta} C_{\text{tot}}(\theta_{\text{pref}}) \cdot R(t) \quad (17)$$

385 Parameterization of the model was done to match single V1 neuron recordings of anesthetized cats, in an experimental setup
 386 similar to the one used here⁶⁹. The computational procedure to match experimental data was entirely done in a previous
 387 publication³⁸. Briefly, it consisted in scanning a range of possible values for each parameters, then find all possible combinations
 388 using a metric of likeliness to single grating response, time-to-peak, peak response and tuning width. The parameters yielded
 389 by this procedure were $\tau = 10.8$ ms; $\alpha = 10.6$ Hz/mV; $J_{\text{ff}} = 9.57$ mv/Hz; $J_{\text{exc}} = 1.71$ Hz/mV; $J_{\text{inh}} = 2.0178$ Hz/mV. For the

390 feedforward mode of the model (Figure 7b), J_{exc} and J_{inh} were set to 0 Hz/mV and the input was convolved with a “receptive
391 field”:

$$\text{RF} = \frac{e^{\kappa_{\text{RF}} \cdot \cos(2(\theta - \Theta_{\text{pref}}))}}{2\pi I_0(\kappa_{\text{RF}})} \quad (18)$$

392 of which we reported the Half-Width at Half-Height, given by⁷⁰:

$$\text{HWHH} = 0.5 \arccos\left(\frac{\log(0.5) + \kappa}{\kappa}\right) \quad (19)$$

393 For the recurrent mode (Figure 7c-e), the concentration measures of the recurrent connectivity profiles κ_{exc} and κ_{inh} were
394 both varied from 0.35 to 7, in 200 even steps, and the input was not convolved with a receptive field.

395 Statistics and reproducibility

396 All data was analyzed using custom Python code. Statistical analysis was performed using non-parametric tests. Wilcoxon
397 signed-rank test with discarding of zero-differences was used for paired samples and Mann-Whitney U test with exact
398 computation of the U distribution was used for independent samples. Due to the impracticality of using error bars when plotting
399 time series, colored contours are used to represent standard deviation values (unless specified otherwise), with a solid line
400 representing mean values. For boxplots, the box extends from the lower to upper quartile values, with a solid white line at the
401 median value. The upper and lower whiskers extend to respectively $Q1 - 1.5 * IQR$ and $Q3 + 1.5 * IQR$, where $Q1$ and $Q3$ are
402 the lower and upper quartiles and IQR is the inter-quartile range.

403 Data availability

404 Preprocessed data and Python code used in the present study are publicly available at <https://github.com/hugoladret/variance-processing-V1>.

406 References

- 407 1. Hubel, D. H. & Wiesel, T. N. Receptive fields of single neurones in the cat’s striate cortex. *The J. physiology* **148**, 574
408 (1959).
- 409 2. Priebe, N. J. Mechanisms of orientation selectivity in the primary visual cortex. *Annu. review vision science* **2**, 85–107
410 (2016).
- 411 3. Fiser, J., Chiu, C. & Weliky, M. Small modulation of ongoing cortical dynamics by sensory input during natural vision.
412 *Nature* **431**, 573–578 (2004).
- 413 4. Simoncelli, E. P. & Olshausen, B. A. Natural image statistics and neural representation. *Annu. review neuroscience* **24**,
414 1193–1216 (2001).
- 415 5. Helmholtz, H. v. *Treatise on physiological optics, 3 vols.* (Optical Society of America, 1924).
- 416 6. Friston, K. A theory of cortical responses. *Philos. transactions Royal Soc. B: Biol. sciences* **360**, 815–836 (2005).
- 417 7. Barthélémy, S. & Mamassian, P. Evaluation of objective uncertainty in the visual system. *PLoS computational biology* **5**,
418 e1000504 (2009).
- 419 8. Bastos, A. M. et al. Canonical microcircuits for predictive coding. *Neuron* **76**, 695–711 (2012).
- 420 9. Goris, R. L., Simoncelli, E. P. & Movshon, J. A. Origin and function of tuning diversity in macaque visual cortex. *Neuron*
421 **88**, 819–831 (2015).
- 422 10. Orbán, G., Berkes, P., Fiser, J. & Lengyel, M. Neural variability and sampling-based probabilistic representations in the
423 visual cortex. *Neuron* **92**, 530–543 (2016).
- 424 11. Festa, D., Aschner, A., Davila, A., Kohn, A. & Coen-Cagli, R. Neuronal variability reflects probabilistic inference tuned to
425 natural image statistics. *Nat. communications* **12**, 1–11 (2021).
- 426 12. Keeble, D., Kingdom, F., Moulden, B. & Morgan, M. Detection of orientationally multimodal textures. *Vis. Res.* **35**,
427 1991–2005 (1995).
- 428 13. Beaudot, W. H. & Mullen, K. T. Orientation discrimination in human vision: Psychophysics and modeling. *Vis. Res.* **46**,
429 26–46 (2006).

- 430 14. Phillips, G. C. & Wilson, H. R. Orientation bandwidths of spatial mechanisms measured by masking. *JOSA A* **1**, 226–232
431 (1984).
- 432 15. Heeley, D., Timney, B., Paterson, I. & Thompson, R. Width discrimination for band-pass stimuli. *Vis. Res.* **29**, 901–905
433 (1989).
- 434 16. Heeley, D. W. & Buchanan-Smith, H. M. The influence of stimulus shape on orientation acuity. *Exp. Brain Res.* **120**,
435 217–222 (1998).
- 436 17. Hénaff, O. J., Boundy-Singer, Z. M., Meding, K., Ziembka, C. M. & Goris, R. L. Representation of visual uncertainty
437 through neural gain variability. *Nat. communications* **11**, 1–12 (2020).
- 438 18. Leon, P. S., Vanzetta, I., Masson, G. S. & Perrinet, L. U. Motion clouds: model-based stimulus synthesis of natural-like
439 random textures for the study of motion perception. *J. Neurophysiol.* **107**, 3217–3226 (2012).
- 440 19. Johnson, A. P. & Baker, C. L. First-and second-order information in natural images: a filter-based approach to image
441 statistics. *JOSA A* **21**, 913–925 (2004).
- 442 20. Field, D. J. Relations between the statistics of natural images and the response properties of cortical cells. *JOSA A* **4**,
443 2379–2394 (1987).
- 444 21. Rust, N. C. & Movshon, J. A. In praise of artifice. *Nat. neuroscience* **8**, 1647–1650 (2005).
- 445 22. Naka, K. & Rushton, W. A. S-potentials from colour units in the retina of fish (cyprinidae). *The J. physiology* **185**, 536–555
446 (1966).
- 447 23. Thorndike, R. L. Who belongs in the family. In *Psychometrika* (Citeseer, 1953).
- 448 24. Hubel, D. H. & Wiesel, T. N. Receptive fields, binocular interaction and functional architecture in the cat's visual cortex.
449 *The J. physiology* **160**, 106 (1962).
- 450 25. Movshon, J. A. The analysis of moving visual patterns. *Exp. Brain Res.* 117–151 (1985).
- 451 26. Laughlin, S. A simple coding procedure enhances a neuron's information capacity. *Zeitschrift für Naturforschung c* **36**,
452 910–912 (1981).
- 453 27. Kinouchi, O. & Copelli, M. Optimal dynamical range of excitable networks at criticality. *Nat. physics* **2**, 348–351 (2006).
- 454 28. Ringach, D. L., Hawken, M. J. & Shapley, R. Dynamics of orientation tuning in macaque primary visual cortex. *Nature*
455 **387**, 281–284 (1997).
- 456 29. Ringach, D. L., Shapley, R. M. & Hawken, M. J. Orientation selectivity in macaque V1: diversity and laminar dependence.
457 *J. Neurosci.* **22**, 5639–5651 (2002).
- 458 30. Bishop, C. M. *Pattern recognition and machine learning* (springer, 2006).
- 459 31. Berens, P. *et al.* A fast and simple population code for orientation in primate V1. *J. Neurosci.* **32**, 10618–10626 (2012).
- 460 32. Chavane, F., Perrinet, L. U. & Rankin, J. Revisiting horizontal connectivity rules in V1: from like-to-like towards like-to-all.
461 *Brain Struct. Funct.* **227**, 1279–1295 (2022).
- 462 33. Roelfsema, P. R., Engel, A. K., König, P. & Singer, W. Visuomotor integration is associated with zero time-lag synchronization
463 among cortical areas. *Nature* **385**, 157–161 (1997).
- 464 34. Aitchison, L. & Lengyel, M. With or without you: predictive coding and bayesian inference in the brain. *Curr. opinion
465 neurobiology* **46**, 219–227 (2017).
- 466 35. Douglas, R. J., Martin, K. A. & Whitteridge, D. A canonical microcircuit for neocortex. *Neural computation* **1**, 480–488
467 (1989).
- 468 36. Somers, D. C., Nelson, S. B. & Sur, M. An emergent model of orientation selectivity in cat visual cortical simple cells. *J.
469 neuroscience* **15**, 5448–5465 (1995).
- 470 37. Teich, A. F. & Qian, N. Learning and adaptation in a recurrent model of V1 orientation selectivity. *J. Neurophysiol.* **89**,
471 2086–2100 (2003).
- 472 38. del Mar Quiroga, M., Morris, A. P. & Krekelberg, B. Adaptation without plasticity. *Cell reports* **17**, 58–68 (2016).
- 473 39. Hubel, D. H. & Wiesel, T. N. Receptive fields and functional architecture in two nonstriate visual areas (18 and 19) of the
474 cat. *J. Neurophysiol.* **28**, 229–289 (1965).
- 475 40. Jia, H., Rochefort, N. L., Chen, X. & Konnerth, A. Dendritic organization of sensory input to cortical neurons in vivo.
476 *Nature* **464**, 1307–1312 (2010).

- 477 41. Chen, X., Leischner, U., Rochefort, N. L., Nelken, I. & Konnerth, A. Functional mapping of single spines in cortical
478 neurons in vivo. *Nature* **475**, 501–505 (2011).
- 479 42. Iacaruso, M. F., Gasler, I. T. & Hofer, S. B. Synaptic organization of visual space in primary visual cortex. *Nature* **547**,
480 449–452 (2017).
- 481 43. Scholl, B., Wilson, D. E. & Fitzpatrick, D. Local order within global disorder: synaptic architecture of visual space.
482 *Neuron* **96**, 1127–1138 (2017).
- 483 44. Monier, C., Chavane, F., Baudot, P., Graham, L. J. & Frégnac, Y. Orientation and direction selectivity of synaptic inputs in
484 visual cortical neurons: a diversity of combinations produces spike tuning. *Neuron* **37**, 663–680 (2003).
- 485 45. Chavane, F. *et al.* Lateral spread of orientation selectivity in V1 is controlled by intracortical cooperativity. *Front. systems
486 neuroscience* **5**, 4 (2011).
- 487 46. Uhl, R. R., Squires, K. C., Bruce, D. L. & Starr, A. Effect of halothane anesthesia on the human cortical visual evoked
488 response. *Anesthesiology* **53**, 273–276 (1980).
- 489 47. Villeneuve, M. Y. & Casanova, C. On the use of isoflurane versus halothane in the study of visual response properties of
490 single cells in the primary visual cortex. *J. neuroscience methods* **129**, 19–31 (2003).
- 491 48. Martinez-Conde, S. *et al.* Effects of feedback projections from area 18 layers 2/3 to area 17 layers 2/3 in the cat visual
492 cortex. *J. Neurophysiol.* **82**, 2667–2675 (1999).
- 493 49. Wang, C., Waleszczyk, W., Burke, W. & Dreher, B. Modulatory influence of feedback projections from area 21a on
494 neuronal activities in striate cortex of the cat. *Cereb. Cortex* **10**, 1217–1232 (2000).
- 495 50. Huang, L., Chen, X. & Shou, T. Spatial frequency-dependent feedback of visual cortical area 21a modulating functional
496 orientation column maps in areas 17 and 18 of the cat. *Brain research* **998**, 194–201 (2004).
- 497 51. Hudetz, A. G., Vizuete, J. A., Pillay, S. & Mashour, G. A. Repertoire of mesoscopic cortical activity is not reduced during
498 anesthesia. *Neuroscience* **339**, 402–417 (2016).
- 499 52. Carandini, M. *et al.* Do we know what the early visual system does? *J. Neurosci.* **25**, 10577–10597 (2005).
- 500 53. Olshausen, B. A. & Field, D. J. Emergence of simple-cell receptive field properties by learning a sparse code for natural
501 images. *Nature* **381**, 607–609 (1996).
- 502 54. Vacher, J., Meso, A. I., Perrinet, L. U. & Peyré, G. Biologically inspired dynamic textures for probing motion perception.
503 *Adv. neural information processing systems* **28** (2015).
- 504 55. Simoncini, C., Perrinet, L. U., Montagnini, A., Mamassian, P. & Masson, G. S. More is not always better: adaptive gain
505 control explains dissociation between perception and action. *Nat. neuroscience* **15**, 1596–1603 (2012).
- 506 56. Ravello, C. R., Perrinet, L. U., Escobar, M.-J. & Palacios, A. G. Speed-selectivity in retinal ganglion cells is sharpened by
507 broad spatial frequency, naturalistic stimuli. *Sci. reports* **9**, 1–16 (2019).
- 508 57. Swindale, N. V. Orientation tuning curves: empirical description and estimation of parameters. *Biol. cybernetics* **78**, 45–56
509 (1998).
- 510 58. Movshon, J. A., Thompson, I. & Tolhurst, D. Spatial and temporal contrast sensitivity of neurones in areas 17 and 18 of
511 the cat's visual cortex. *The J. physiology* **283**, 101–120 (1978).
- 512 59. Peirce, J. *et al.* Psychopy2: Experiments in behavior made easy. *Behav. research methods* **51**, 195–203 (2019).
- 513 60. Siegle, J. H. *et al.* Open ephys: an open-source, plugin-based platform for multichannel electrophysiology. *J. neural
514 engineering* **14**, 045003 (2017).
- 515 61. Pachitariu, M., Steinmetz, N. A., Kadir, S. N., Carandini, M. & Harris, K. D. Fast and accurate spike sorting of high-channel
516 count probes with kilosort. In *Advances in neural information processing systems*, 4448–4456 (2016).
- 517 62. Rossant, C. *et al.* Spike sorting for large, dense electrode arrays. *Nat. neuroscience* **19**, 634–641 (2016).
- 518 63. Katzner, S. *et al.* Local origin of field potentials in visual cortex. *Neuron* **61**, 35–41 (2009).
- 519 64. Maier, A., Adams, G. K., Aura, C. & Leopold, D. A. Distinct superficial and deep laminar domains of activity in the visual
520 cortex during rest and stimulation. *Front. systems neuroscience* **4**, 31 (2010).
- 521 65. Smith, M. A., Majaj, N. J. & Movshon, J. A. Dynamics of motion signaling by neurons in macaque area mt. *Nat.
522 neuroscience* **8**, 220–228 (2005).
- 523 66. Quiroga, R. Q., Reddy, L., Koch, C. & Fried, I. Decoding visual inputs from multiple neurons in the human temporal lobe.
524 *J. Neurophysiol.* **98**, 1997–2007 (2007).

- 525 67. Guitchounts, G., Masis, J., Wolff, S. B. & Cox, D. Encoding of 3D head orienting movements in the primary visual cortex.
526 *Neuron* **108**, 512–525 (2020).
- 527 68. Brodersen, K. H., Ong, C. S., Stephan, K. E. & Buhmann, J. M. The balanced accuracy and its posterior distribution. In
528 *2010 20th International Conference on Pattern Recognition*, 3121–3124 (IEEE, 2010).
- 529 69. Felsen, G. *et al.* Dynamic modification of cortical orientation tuning mediated by recurrent connections. *Neuron* **36**,
530 945–954 (2002).
- 531 70. Swindale, N. V., Grinvald, A. & Shmuel, A. The spatial pattern of response magnitude and selectivity for orientation and
532 direction in cat visual cortex. *Cereb. cortex* **13**, 225–238 (2003).

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539 **Author contributions**

540 L.U.P., C.C. F.C., N.C., and H.J.L. designed the study. H.J.L., N.C. and L.K. collected the data. H.J.L. and L.U.P. analyzed the
541 data. H.J.L. and L.U.P. wrote the original draft of the manuscript. All authors reviewed and edited the manuscript.

542 **Competing interests**

543 The authors declare no competing interests.

544 **Supplementary information**

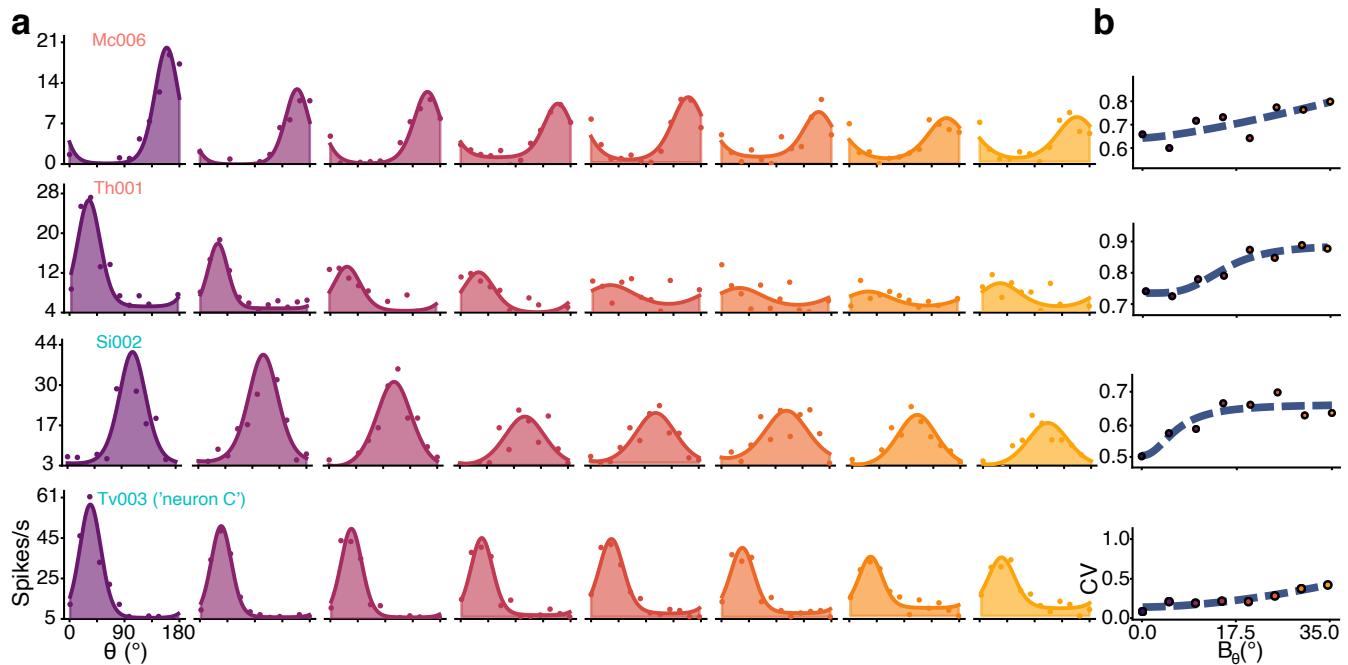


Figure S 1. Additional examples of single neuron tuning curves and VTF. **(a)** Tuning curves of four more neurons, stimulated with Motion Clouds of increasing variance (controlled by B_θ) from left to right. Colored dots represent the mean firing rate across trials (baseline subtracted, 300 ms average) and solid lines a fitted von Mises function. **(b)** Variance-tuning functions (VTF), measuring the changes of orientation tuning measured by the circular variance (CV, colored dots) as a function of stimuli variance, fitted with a Naka-Rushton (NKR) function (dashed curves, parameters shown in light gray).

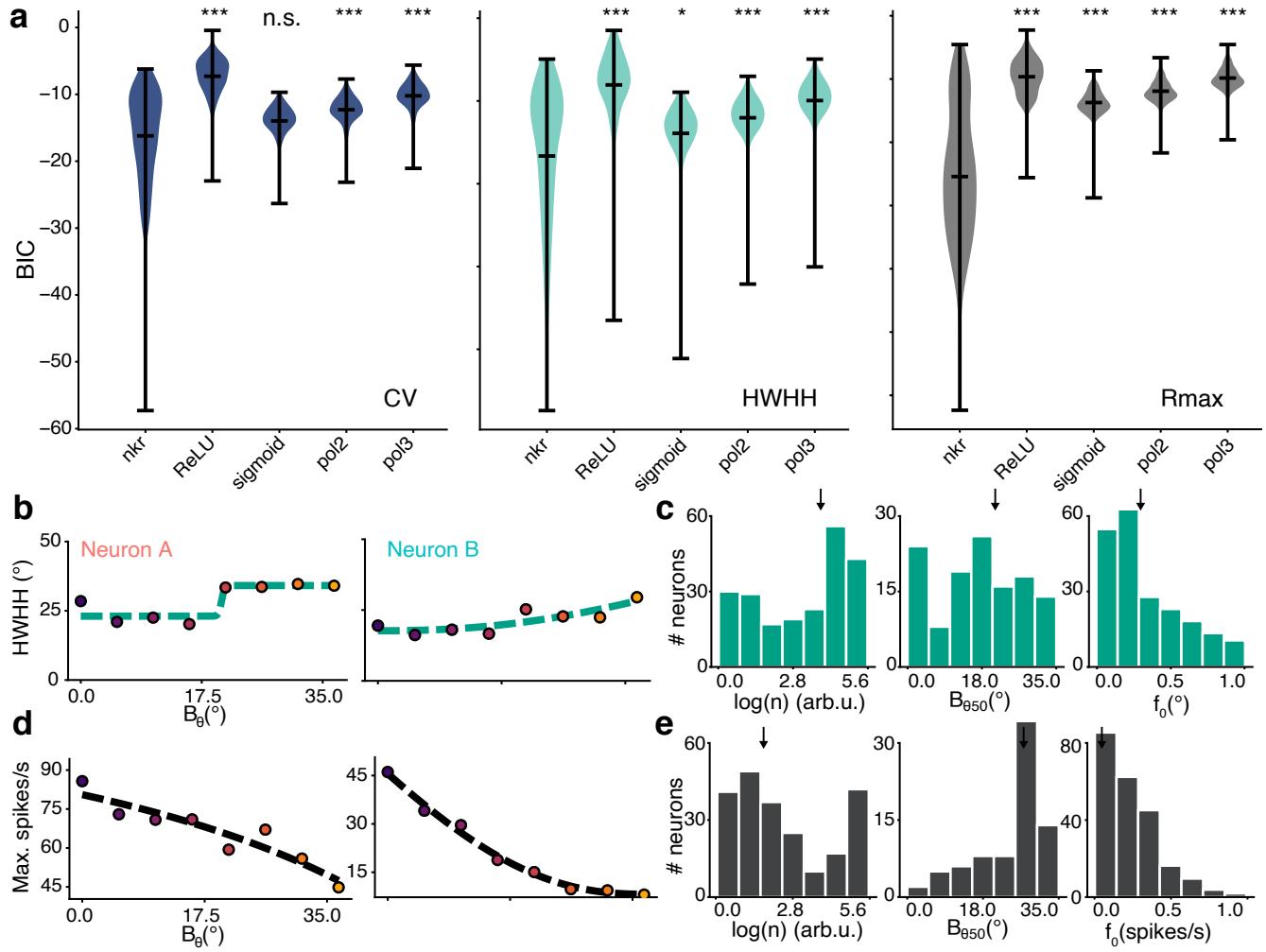


Figure S 2. Naka-Rushton function fitting of VTFs. (a) Violin plot of the Bayesian Information Criterion (BIC) of the CV curves of all recorded neurons. Each violin plot represents a different type of fitted equation, respectively: Naka-Rushton (nkr, see Methods) ; Rectified Linear Unit (ReLU, $f(x) = \max(0, x)$) ; logistic function (sigmoid, $f(x) = \frac{e^x}{e^x + 1}$) ; second degree polynomial function (pol2, $f(x) = ax^2 + bx + c$) and third degree polynomial function (pol3, $f(x) = ax^3 + bx^2 + cx + d$). A lower BIC indicates less information lost in the fitting process, hence a better fitting model. Naka-Rushton curves were chosen over sigmoid functions for the skewness of the BIC distributions towards negative values, as well as the explainability of their parameters. n.s., not significant; *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$ (Kruskal-Wallis H-test, post-hoc Dunn Pairwise test, Bonferroni corrected) (b) Variance-HWHH function, fitted with a Naka-Rushton function. (c) Histograms of the NKR parameters (in the [5%;95%] range of possible NKR fitting values) for the 249 recorded units. Median values are indicated by a black arrow ($\log(n) = 3.8$, $B_{\theta 50} = 16.5^{\circ}$, $f_0 = 0.25$). Parameter f_0 is given in normalized HWHH values. (d) Same as (b), with maximum firing rate as a function of input variance. (e) Same as (c), for maximum firing rate. Median values are indicated by a black arrow ($\log(n) = 2.15$, $B_{\theta 50} = 28.3^{\circ}$, $f_0 = 0.09$). Parameter f_0 is given in normalized firing rate values

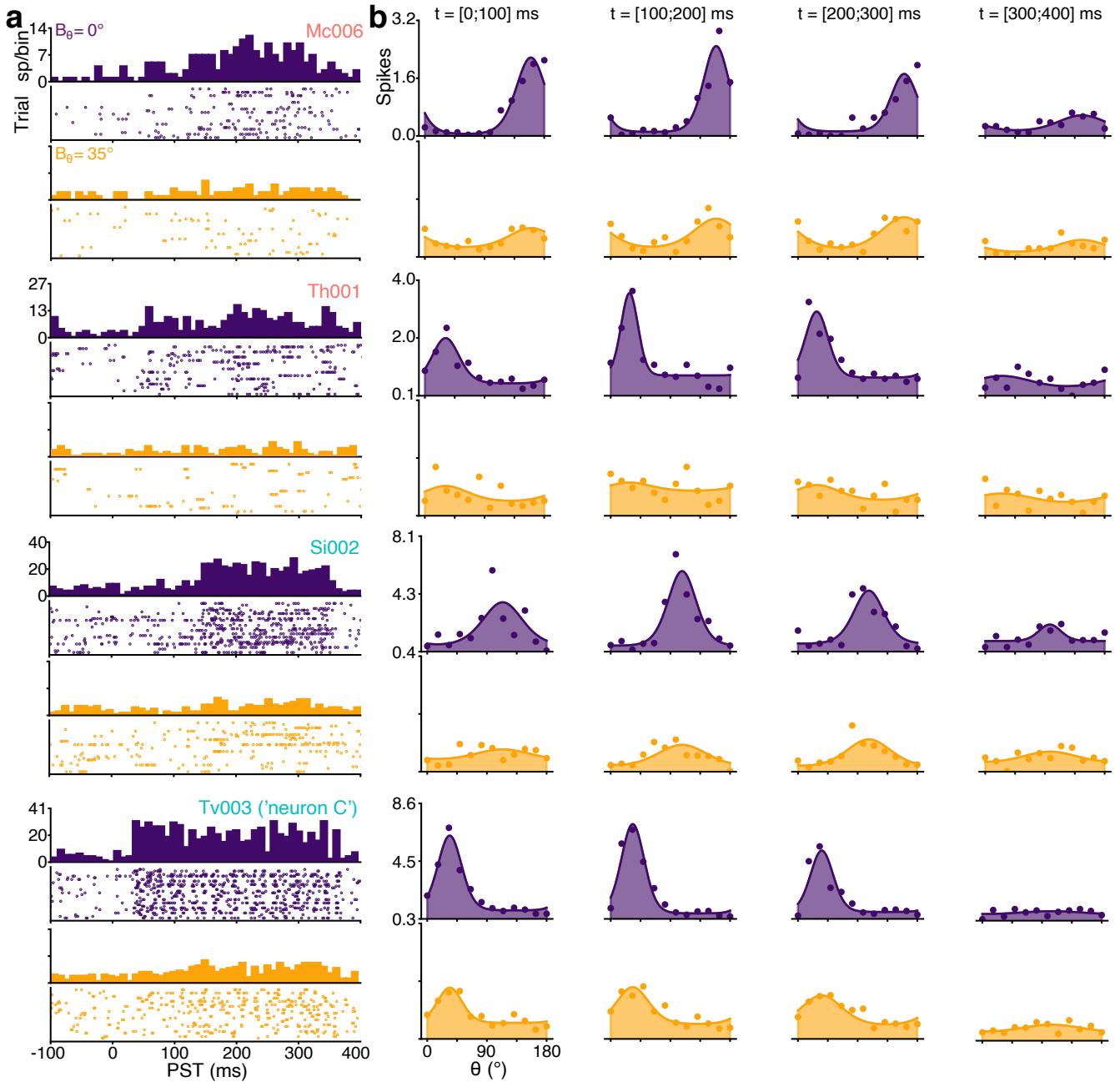


Figure S 3. Additional examples of dynamical properties of neurons. **(a)** Peristimulus time (PST) histogram and associated rasterplot of the four additional example neurons, for Motion Clouds with lowest ($B_\theta = 0^\circ$, purple) and highest ($B_\theta = 35^\circ$, yellow) variance. **(b)** Dynamics of the tuning curves shown in Figure 2 in 100 ms windows, starting at the time labelled atop of each column..

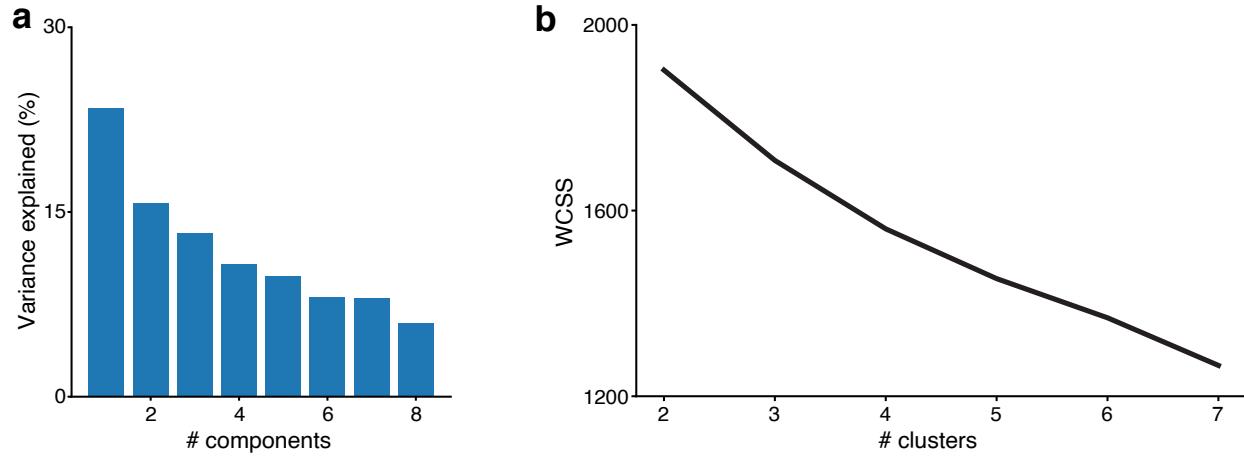


Figure S 4. Graphical reports of clustering analysis. (a) Fraction of variance explained as a function of the number of components used in the Principal Component Analysis. (b) Within-Cluster-Sum-of-Squares (WCSS) of the K-means clustering, expressed as a function of the number of clusters. An empirical way to select the number of clusters is to typically take the “elbow” of such a curve, if existent.

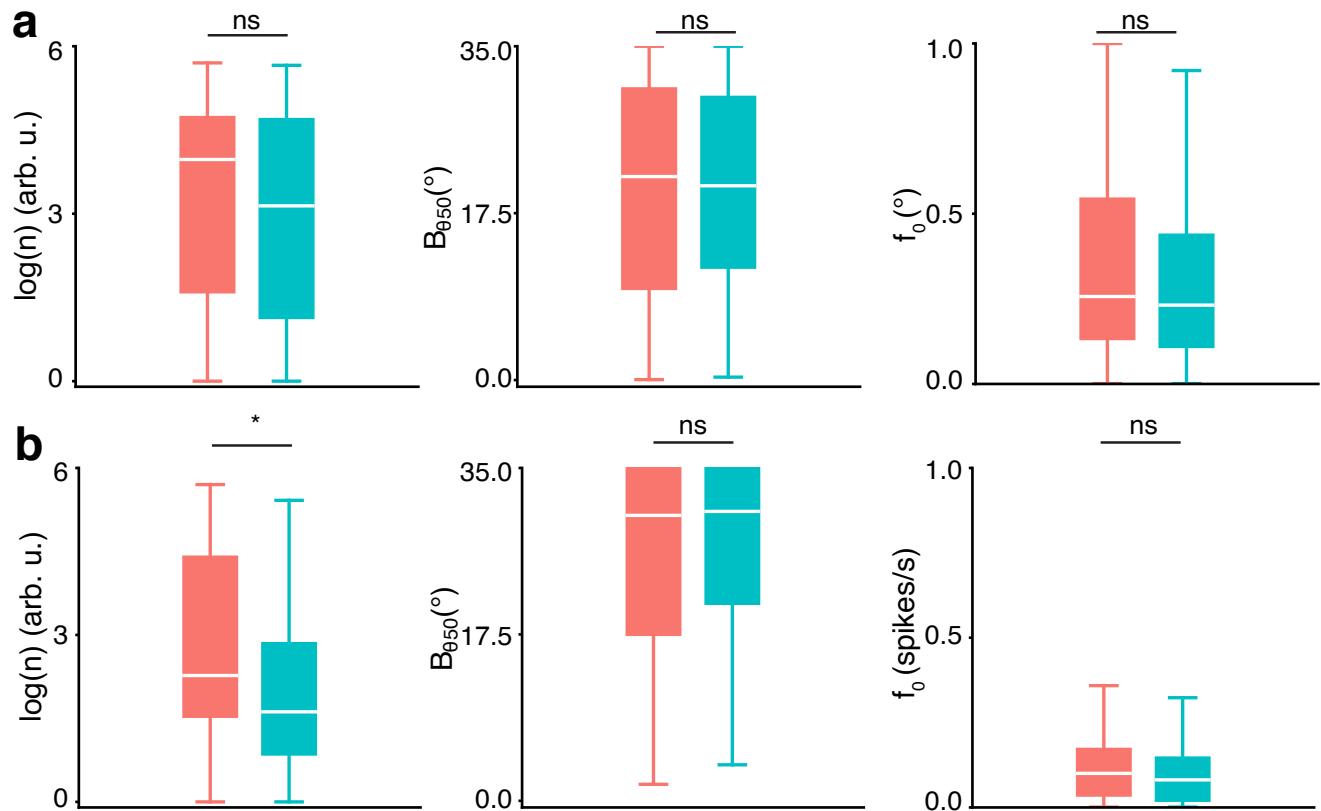


Figure S 5. Additional clustering variables. (a) Boxplot of the VTF parameters for HWHH functions. (b) Boxplot of the VTF parameters for maximum response functions.

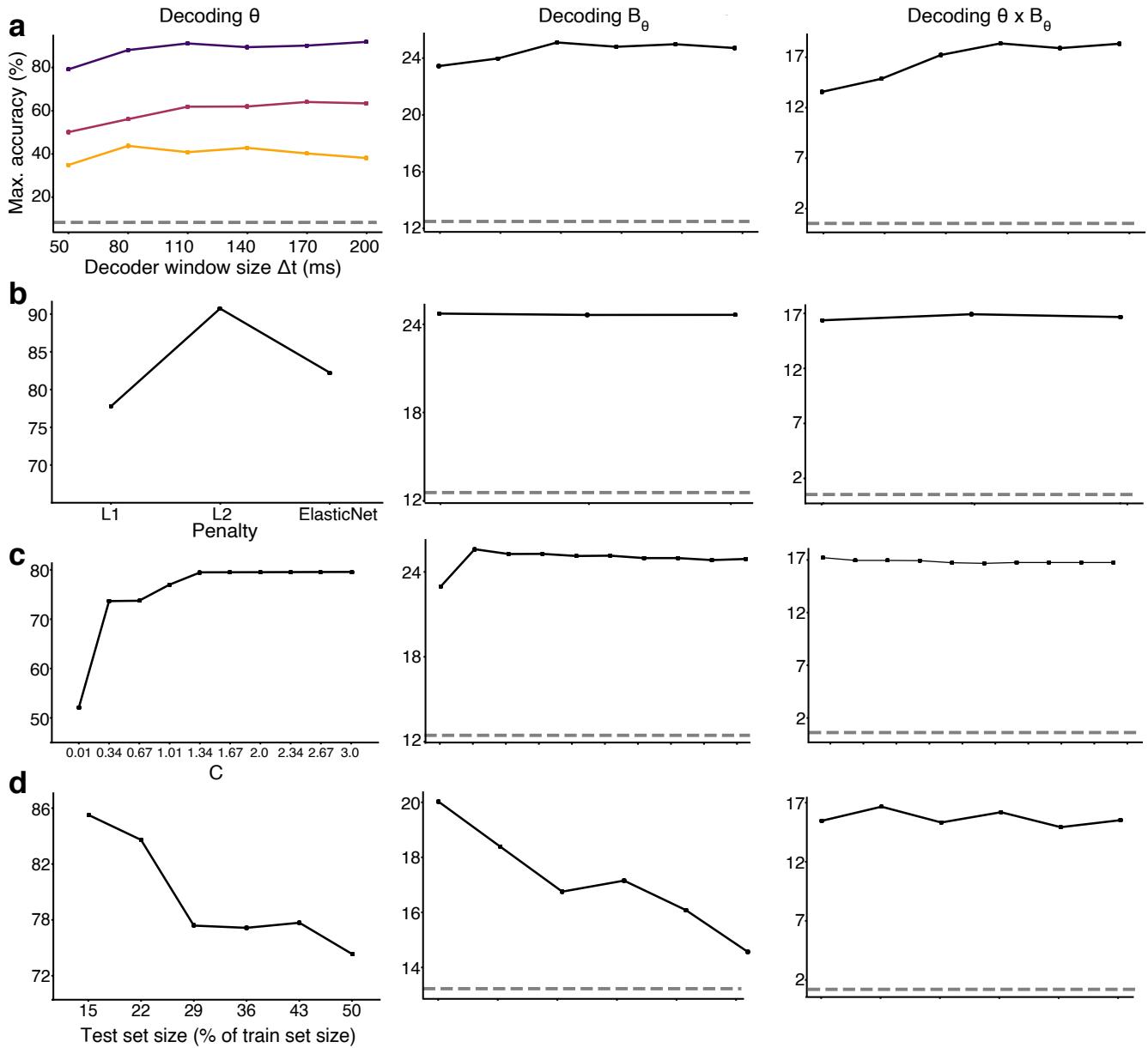


Figure S 6. Parameters evaluation of the decoders. **(a)** Optimization of the length of the time window ΔT , which controls the length of the integration time of the decoder (see Methods), for orientation decoders (left column), variance decoders (middle column) and orientation x variance decoders (right column). **(b)** Optimization of the penalization norm applied to the decoder, which controls the metric by which the classification error is minimized. l_1 and l_2 are defined as $\sum_i |x_i|$ and $\sqrt{\sum_i |x_i|^2}$, where x is a set of features fed to the classifier. The penalty norm “ElasticNet” linearly combines these two norms, here with equal weighting applied to both norms. **(c)** Optimization of the parameter C controls the regularization strength, which measure the magnitude of the penalty applied to large parameters, in order to prevent overfitting on reduced sets of data. **(d)** Optimization of the percentage of data kept out of the training set to evaluate the decoder’s accuracy.

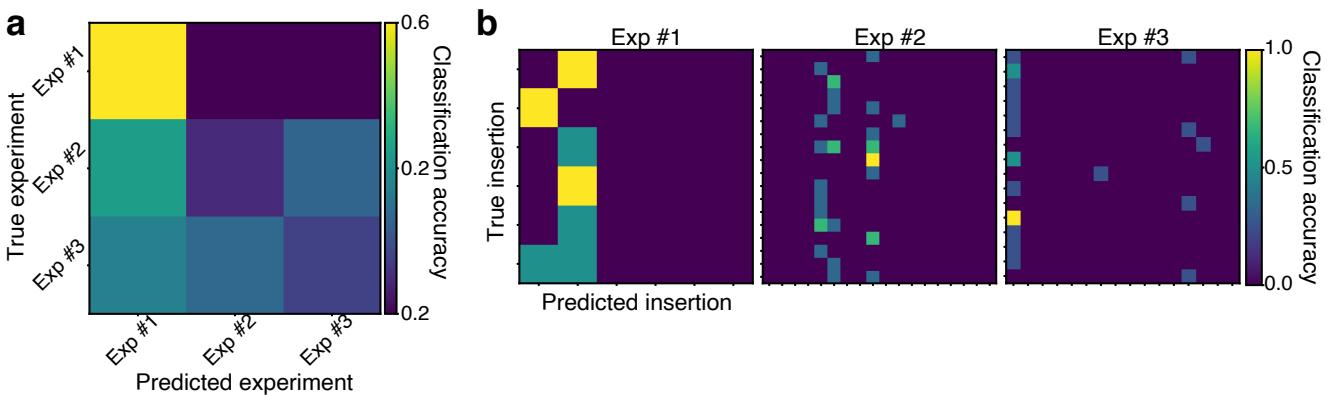


Figure S 7. Decoding the experiment's identity (the specific experiment which yielded the spikes) or insertion's identity (the specific insertion which yielded the spikes) of neurons does not produce any significant result, thereby validating the fusion of multiple datasets in the decoding process. **(a)** Confusion matrix of a decoder trained to retrieve the experiment identity using three groups of 30 neurons (bootstrapped 1000 times). **(b)** Confusion matrices of three decoders trained to retrieve the insertion identity of the neurons recorded in each individual experiment.

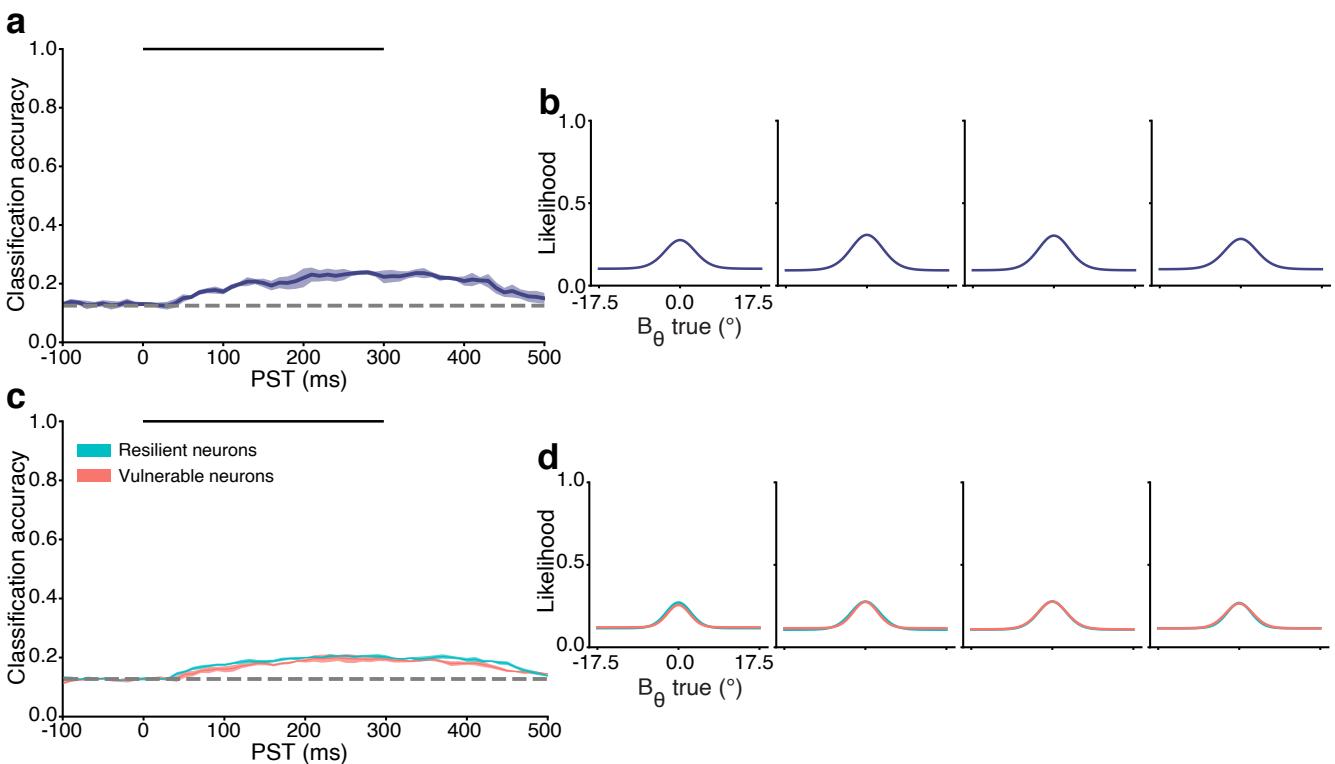


Figure S 8. Orientation variance cannot be accurately decoded from the population activity. **(a)** Time course of a decoder trained to retrieve the variance B_θ of Motion Clouds. Solid dark line represents the mean accuracy of a 5-fold cross validation and filled contour the SD. Decoding at chance level (here, 1/8) is represented by a gray dashed line. **(b)** Population tuning curve of the decoder, representing the likelihood of decoding as a function of error on variance. **(c)** Same as Figure S8a, trained with spikes from either resilient or vulnerable neurons. **(d)** Same as Figure S8b, trained with spikes from either resilient or vulnerable neurons.

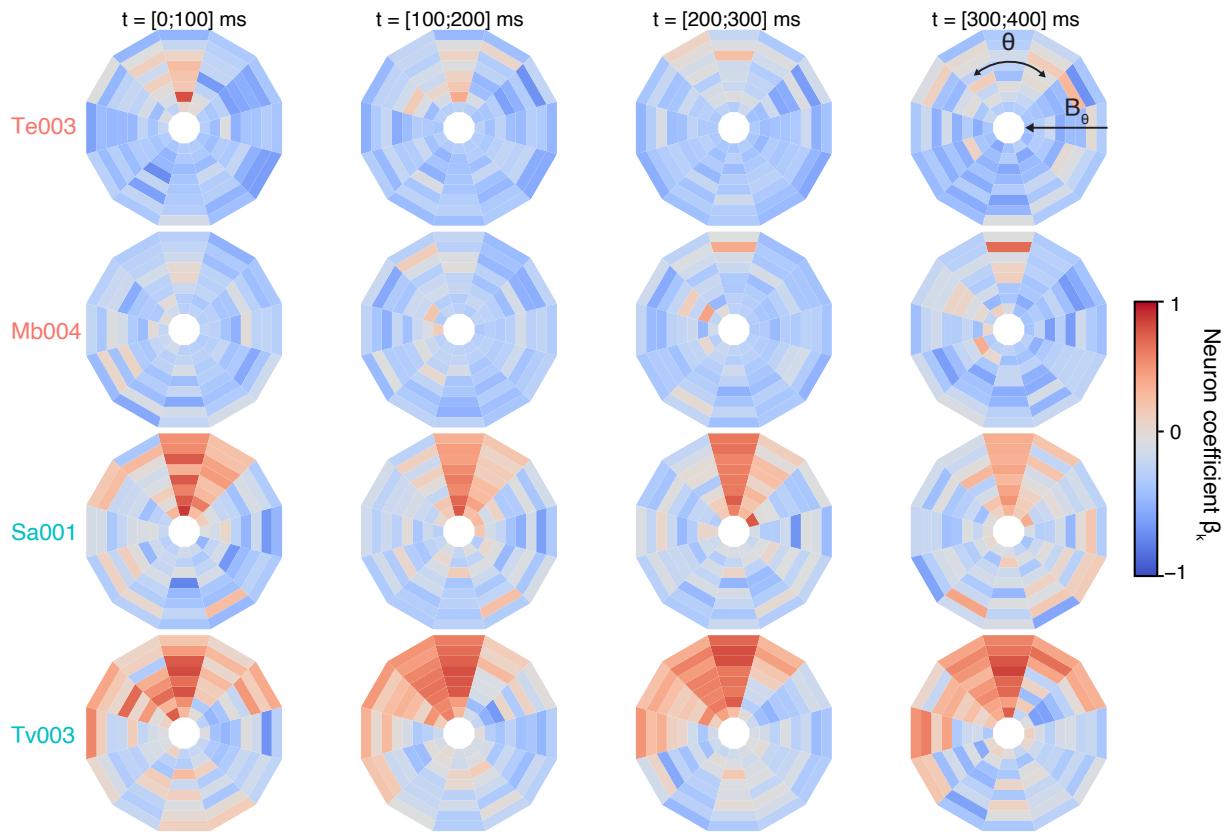


Figure S 9. Polar plot of coefficient matrices from two single vulnerable neurons (top rows) and two single resilient neurons (bottom rows). As in Figure 6e, the angle of each bin represents the error on the θ identity of the stimulus, Δ_θ , and the eccentricity corresponds to the coefficient for each B_θ (highest variance at the center). The temporal evolution of the coefficients from the decoder is normalized for each neuron.