RmirR

Marco Ortiz, Augusto Berrocal

Centro de Ciencias Genómicas Universidad Nacional Autónoma de México

October 16th, 2009

An analysis with RmiR start giving to the function read.mir the list of miRNAs and the gene's list with the right annotation With the default values the function searches integerScan and Pictar and prints only the couples of gene and miRNA present in bout databases.

Downloading the package I

```
source("http://bioconductor.org/biocLite.R") biocLite("RmiR") And we are going to download and annotation (could be a microarray data) biocLite("hgug4112a.db") biocLite("org.Hs.eg.db")
```

- > library(RmiR)
- > library(hgug4112a.db)
- > library(org.Hs.eg.db)

An analisys with RmiR start giving to the function read.mir the list of miRNAs and the gene's list with the right anotation

With the default values the function searches integerScan and Pictar and prints only the coupels of gene and miRNA present in bouth databases.

Datasets to work with I

```
> genes \leftarrow data.frame(genes = c("A_23_P171258",
     "A_23_P150053", "A_23_P150053",
    "A_23_P150053", "A_23_P202435",
      "A_24_P90097", "A_23_P127948"))
> genes expr <- c(1.21, -1.5, -1.34,
      -1.45, -2.41, -2.32, -3.03)
+
> mirna <- data.frame(mirna = c("hsa-miR-148b".
+
      "hsa-miR-27b", "hsa-miR-25",
+
      "hsa-miR-181a", "hsa-miR-27a",
    "hsa-miR-7", "hsa-miR-32",
      "hsa-miR-32", "hsa-miR-7"))
+
 mirna$expr <- c(1.23, 3.52, -2.42,
      5.2, 2.2, -1.42, -1.23, -1.2,
+
      -1.37)
```

> genes

```
genes expr
1 A_23_P171258 1.21
2 A_23_P150053 -1.50
3 A_23_P150053 -1.34
```

The basic function read.mir I

With read.mir we search genes and miRNAs on bout of the databases earlier mentioned, the function gives the average of the expression of different probes of the microarray identifying the same gene and it also computes the coefficient of variation (CV). If there is just one probe identifying a gene, no average cantbe done, so CV will be NA.

And what about if you have another identification than the probe id? I

If the result is annotated by another identification than the platform probe id we can specify the annotation identifiers with other parameter id, the possible values for this id's "probes", "genes", "alias", "ensemble", "unigene".

Some examples I

> genes.e <- genes

An example with entrez gene identifiers.

```
> genes.e$gene_id <- c(22, 59, 59,
+ 59, 120, 120, 133)
> genes.e <- genes.e[, c("gene_id",</pre>
     "expr")]
> genes.e
 gene_id expr
1 22 1.21
2 59 -1.50
> read.mir(genes = genes.e, mirna = mirna,
      annotation = "hgug4112a.db",
      id = "genes")
```

```
1 22 hsa-miR-148b 1.230
4 133 hsa-miR-181a 5.200
mirCV symbol geneExpr geneCV
1 NA ABCB7 1.21 NA
4 NA ADM -3.03 NA
```

gene_id mature_miRNA mirExpr

More examples I

```
> genes.a <- genes
> genes.a$alias <- c("ABCB7", "ADD3",
     "ADDL", "ADD3", "AAT6", "ACTA2",
+ "ADM")
> genes.a <- genes.a[, c("alias",</pre>
+ "expr")]
> genes.a
 alias expr
1 ABCB7 1.21
2 ADD3 -1.50
> read.mir(genes = genes.a, mirna = mirna,
+
   annotation = "hgug4112a.db",
+ id = "alias")
```

Marco Ortiz, Augusto Berrocal

More examples II

ADM -3.03

NA

NA

Results by individual probes I

Some times we do not need the average of the results, for example when we would like to test this probe separately, in this case we prefer the results as they are, it is to say that with each probe annotated individually.

```
> read.mir(genes = genes, mirna = mirna,
+
     annotation = "hgug4112a.db",
     at.least = 1, id.out = "probes")
   gene_id mature_miRNA mirExpr
1
        22 hsa-miR-148b
                          1.230
        59 hsa-miR-27a 2.200
        mirCV
                 probe_id geneExpr
           NA A_23_P171258 1.21
           NA A_23_P150053 -1.50
```

The regulation of target genes by miRNAs occure usually at three different ways.

- ► The miRNAs could promote the expression of genes
- ► They could repress expression
- ▶ May be we cannot see any difference

Looking at the correlation between different miRNAs targets couples we can obtain the correlated and the anti correlated couples.

This does not mean that there is short biological relevance but it could be some hints for farther investigation. To use the function RmiRtc we need two or more objects of the class read.mir

```
> data(RmiR)
> res1 <- read.mir(gene = gene1,
     mirna = mir1, annotation = "hgug4112a.db",
    verbose = TRUE)
In targetscan database there are 13 genes and 35
In pictar database there are 7 genes and 27 micro
> res2 <- read.mir(gene = gene2,
     mirna = mir2, annotation = "hgug4112a.db",
     verbose = TRUE)
```

In targetscan database there are 12 genes and 23 mi

Marco Ortiz, Augusto Berrocal

In pictar database there are 6 genes and 24 microRN

```
> res3 <- read.mir(gene = gene3,
```

In targetscan database there are 13 genes and 35 micr

In pictar database there are 7 genes and 27 microRNA

```
> res_tc <- RmiRtc(timeline = c("res1",
```

+ "res2", "res3"), timevalue =
$$c(12,$$

+ 24, 48))

And now we use the function and look for the correlation. We can decide to filter the object by a correlation andor a gene expression threshold

> res_fil

The function RmiRtc II

```
An object of class miRtcList
[[1]]
gene_id mature_miRNA
22     351 hsa-miR-20a
17     351 hsa-miR-20b
```

```
[[3]]
12 24 48
5 0.71 -0.95 -1.67
```

The function readRmiRtc filter the genes ad returns a list ranked by the number of the miRNA satisfying the thresholds.

```
cbind(res_fil$couples, res_fil$geneExpr,
     res_fil$mirExpr)[res_fil$couples$gene_id ==
+
     351 & res_fil$cor <= -0.9,
  gene_id mature_miRNA 12
                               24
22
      351 hsa-miR-20a 0.71 -0.95
17
      351 hsa-miR-20b 0.71 -0.95
      351 hsa-miR-93 0.71 -0.95
19
     48
          12 24 48
22 -1.67 0.32 1.73 2.12
17 -1.67 0.06 1.10 1.61
19 -1.67 0.30 1.25 1.19
```

```
Marco Ortiz,
Augusto Berrocal
```

```
> plotRmiRtc(res_fil, gene_id = 351,
+ legend.y = 0, legend.x = 30)
```

Plotting a time course experiment II

APP and its miRNAs expression trends

