#### Contents

Basic DESeq2 results exploration	1
Introduction	1
Code setup	. 1
PCA	2
Sample-to-sample distances	3
MA plots	3
P-values distribution	6
Adjusted p-values distribution	7
Top features	8
Count plots top features	8
Reproducibility	19
Bibliography	21

## Basic DESeq2 results exploration

Project: PDF report.

#### Introduction

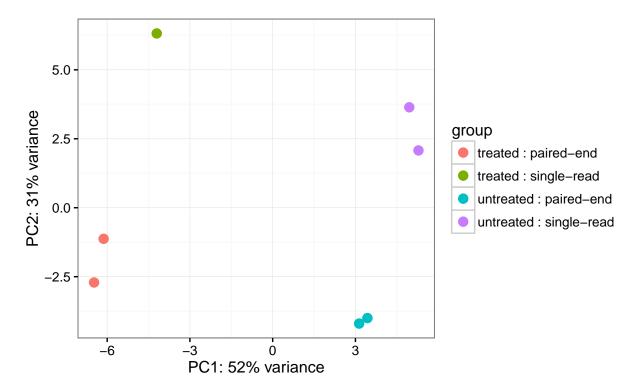
This report is meant to help explore DESeq2 (Love, Huber, and Anders, 2014) results and was generated using the regionReport (Collado-Torres, Jaffe, and Leek, 2015) package. While the report is rich, it is meant to just start the exploration of the results and exemplify some of the code used to do so. If you need a more in-depth analysis for your specific data set you might want to use the customCode argument. This report is based on the vignette of the DESeq2 (Love, Huber, and Anders, 2014) package which you can find here.

### Code setup

This section contains the code for setting up the rest of the report.

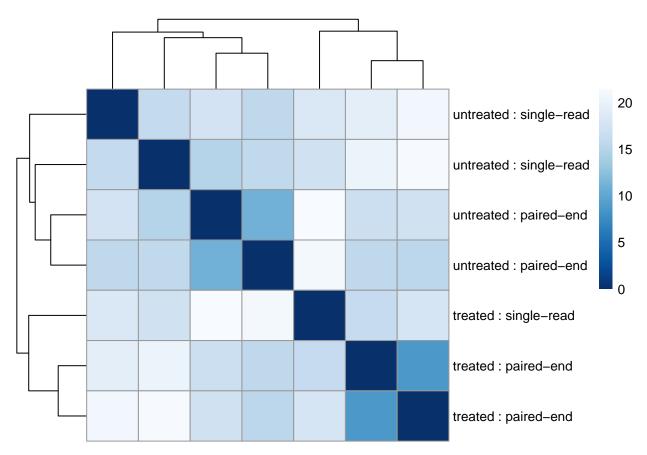
```
## knitrBoostrap and device chunk options
load_install('knitr')
opts_chunk$set(bootstrap.show.code = FALSE, dev = device)
if(!outputIsHTML) opts_chunk$set(bootstrap.show.code = FALSE, dev = device, echo = FALSE)
```

## PCA



The above plot shows the first two principal components that explain the variability in the data using the regularized log count data. If you are unfamiliar with principal component analysis, you might want to check the Wikipedia entry or this interactive explanation. In this case, the first and second principal component explain 52 and 31 percent of the variance respectively.

### Sample-to-sample distances

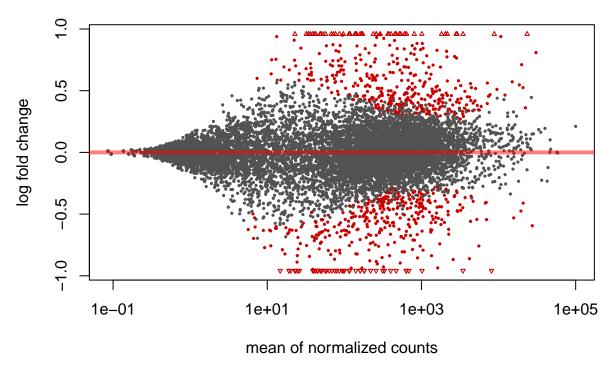


This plot shows how samples are clustered based on their euclidean distance using the regularized log transformed count data. This figure gives an overview of how the samples are hierarchically clustered. It is a complementary figure to the PCA plot.

## MA plots

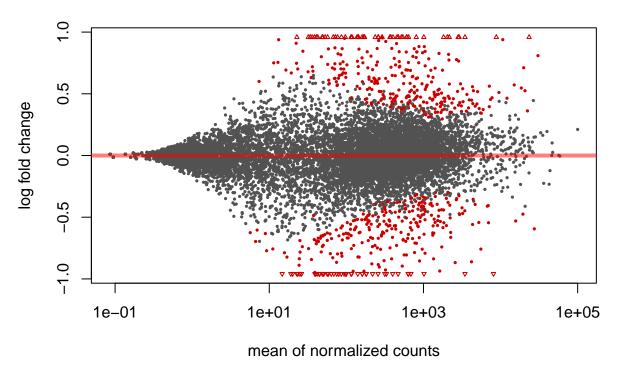
This section contains three MA plots (see Wikipedia) that compare the mean of the normalized counts against the log fold change. They show one point per feature. The points are shown in red if the feature has an adjusted p-value less than alpha, that is, the statistically significant features are shown in red.

## MA plot with alpha = 0.1



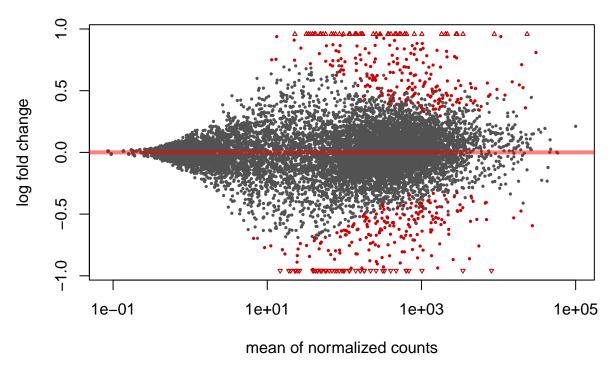
This first plot shows uses alpha = 0.1, which is the alpha value used to determine which resulting features were significant when running the function DESeq2::results().

## MA plot with alpha = 0.05



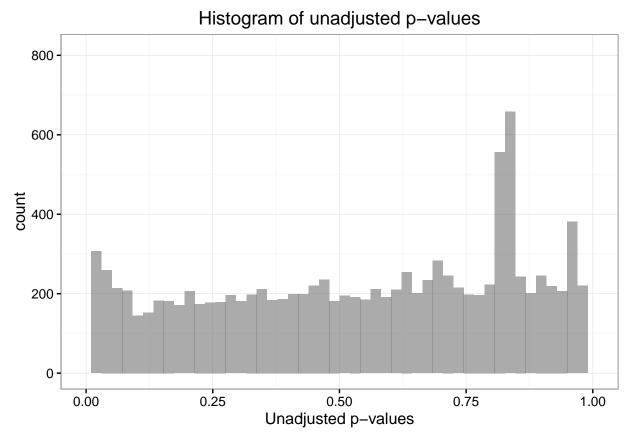
This second MA plot uses alpha = 0.05 and can be used agains the first MA plot to identify which features have adjusted p-values between 0.05 and 0.1.

# MA plot for top 500 features



The third and final MA plot uses an alpha such that the top 500 features are shown in the plot. These are the features that whose details are included in the top features interactive table.

### P-values distribution



This plot shows a histogram of the unadjusted p-values. It might be skewed right or left, or flat as shown in the Wikipedia examples. The shape depends on the percent of features that are differentially expressed. For further information on how to interpret a histogram of p-values check David Robinson's post on this topic.

```
## Min. 1st Qu. Median Mean 3rd Qu. Max. NA's ## 0.0000 0.2266 0.5367 0.5072 0.8091 0.9999 2649
```

This is the numerical summary of the distribution of the p-values.

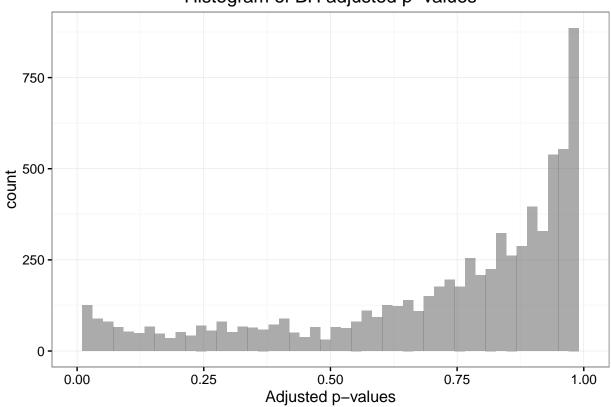
Cut	Count
0.0001	321
0.0010	467
0.0100	806
0.0250	1053
0.0500	1371
0.1000	1858
0.2000	2698
0.3000	3612
0.4000	4541
0.5000	5566
0.6000	6526
0.7000	7642
0.8000	8735
0.9000	10600

Cut	Count
1.0000	11821

This table shows the number of features with p-values less or equal than some commonly used cutoff values.

# Adjusted p-values distribution





This plot shows a histogram of the BH adjusted p-values. It might be skewed right or left, or flat as shown in the Wikipedia examples.

##	Min.	1st Qu.	Median	Mean 3	3rd Qu.	Max.	NA's
##	0.000	0.543	0.816	0.693	0.944	1.000	6272

This is the numerical summary of the distribution of the BH adjusted p-values.

Cut	Count
0.0001	193
0.0010	272
0.0100	418
0.0250	521
0.0500	630
0.1000	796
0.2000	1050

Cut	Count
0.3000	1340
0.4000	1645
0.5000	1917
0.6000	2324
0.7000	2932
0.8000	3928
0.9000	5352
1.0000	8198

This table shows the number of features with BH adjusted p-values less or equal than some commonly used cutoff values.

## Top features

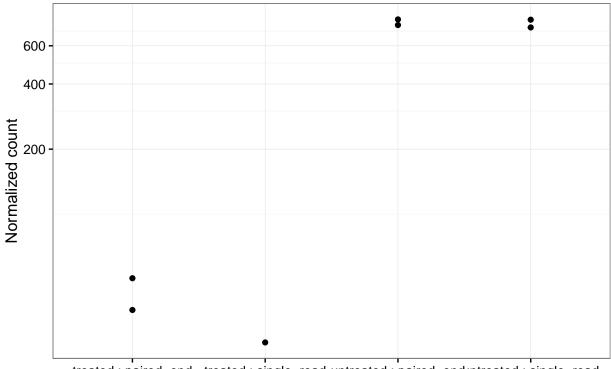
This table shows the top 500 features ordered by their BH adjusted p-values. Since the report is in PDF format, only the top 20 features are shown.

Feature	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
FBgn0039155	453.28	3.72	0.16	23.24	1.638802e-119	1.343490e-115
FBgn0029167	2165.04	2.08	0.10	20.29	1.433135e-91	5.874422e-88
FBgn0035085	366.83	2.23	0.14	16.33	6.382004 e-60	1.743989e-56
FBgn0029896	257.90	2.21	0.16	13.91	5.403276e-44	1.107402e-40
FBgn0034736	118.41	2.56	0.18	13.89	7.666232e-44	1.256955e-40
FBgn0040091	610.60	1.43	0.12	11.97	4.969184e-33	6.789562e-30
FBgn0000071	180.01	-2.13	0.18	-11.76	6.218219 e-32	7.282423e-29
FBgn0011260	140.36	-1.96	0.17	-11.60	3.961715 e-31	4.059767e-28
FBgn0034434	76.91	2.38	0.21	11.22	3.270683e-29	2.979229e-26
FBgn0001226	686.72	-1.53	0.14	-11.11	1.091294e-28	8.946427e-26
FBgn0038832	167.59	2.03	0.19	10.89	1.294031e-27	9.644060 e-25
FBgn0026562	23460.08	1.89	0.18	10.54	5.552988e-26	3.793616e-23
FBgn0031191+FBgn0027279	2043.81	0.96	0.09	10.45	1.514563e-25	9.551069 e-23
FBgn0033913	2816.22	1.10	0.11	10.38	2.958920 e-25	1.617149e-22
FBgn0051092	93.33	-1.95	0.19	-10.39	2.854880e-25	1.617149e-22
FBgn0023479	1823.54	1.37	0.13	10.30	6.881846 e-25	3.438274e-22
FBgn0035189	115.49	-2.11	0.21	-10.30	7.129867e-25	3.438274e-22
FBgn0033764	41.75	-2.22	0.22	-10.20	2.018303e-24	9.192247e-22
FBgn0003501	161.22	-1.76	0.18	-10.02	1.222950 e-23	5.276706e-21
FBgn0039419	806.81	1.11	0.11	9.85	6.544643e- $23$	2.682649 e-20

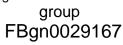
## Count plots top features

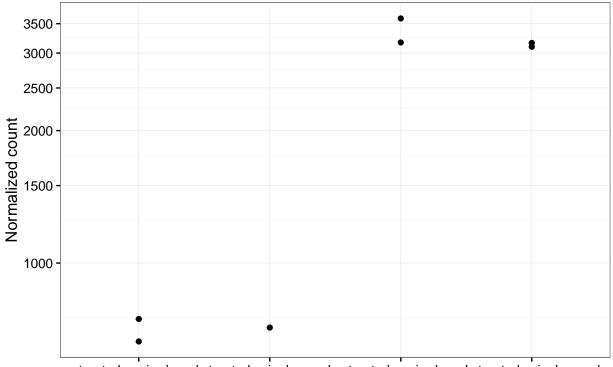
This section contains plots showing the normalized counts per sample for each group of interest. Only the best 20 features are shown, ranked by their BH adjusted p-values. The Y axis is on the log10 scale and the feature name is shown in the title of each plot.



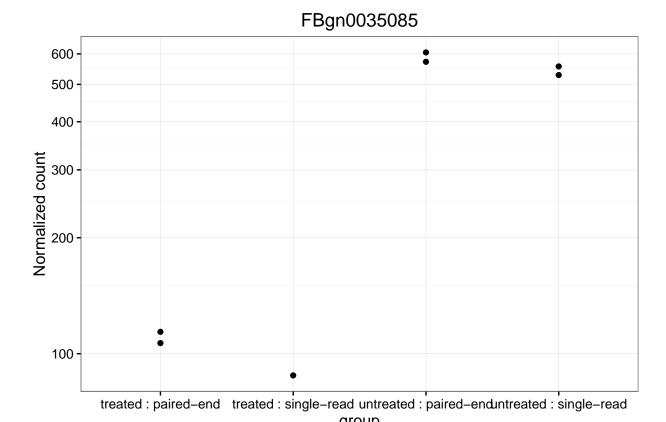


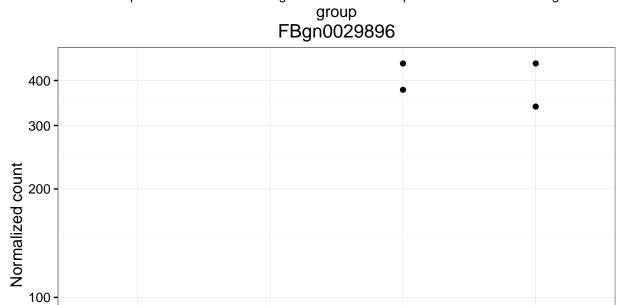
treated : paired-end treated : single-read untreated : paired-enduntreated : single-read



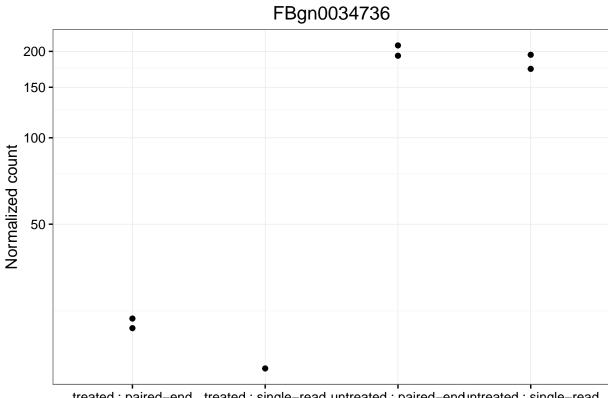


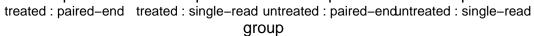
treated : paired-end treated : single-read untreated : paired-endintreated : single-read group

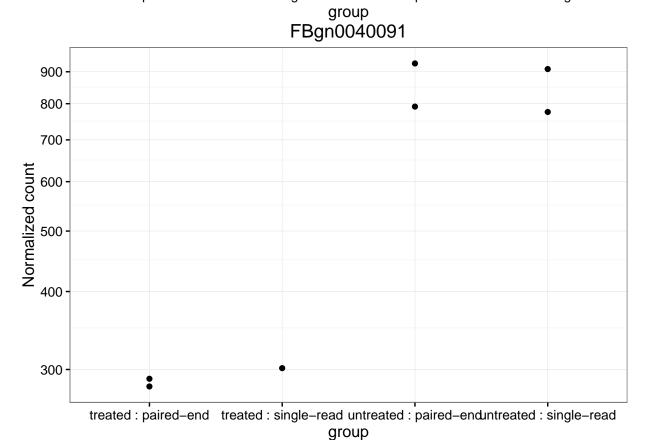


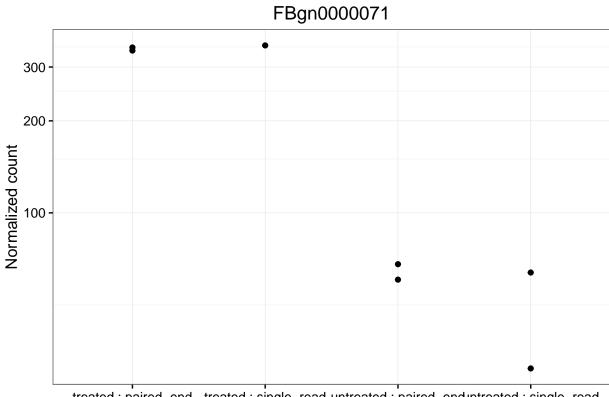


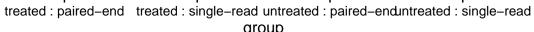
treated : paired-end treated : single-read untreated : paired-enduntreated : single-read group

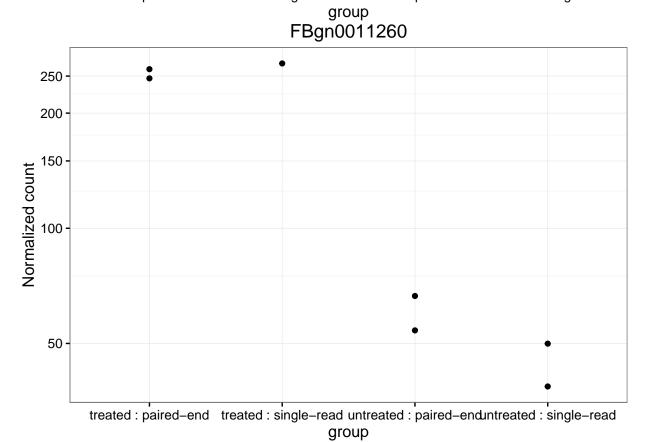


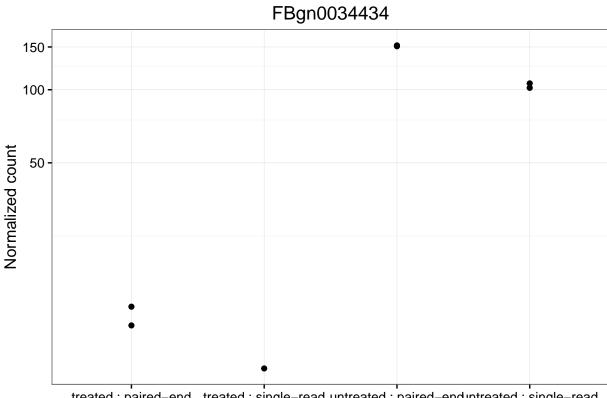


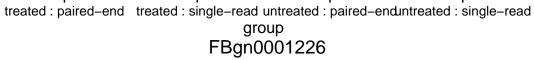


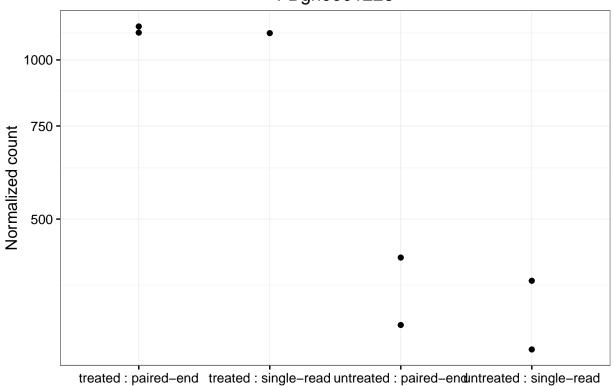






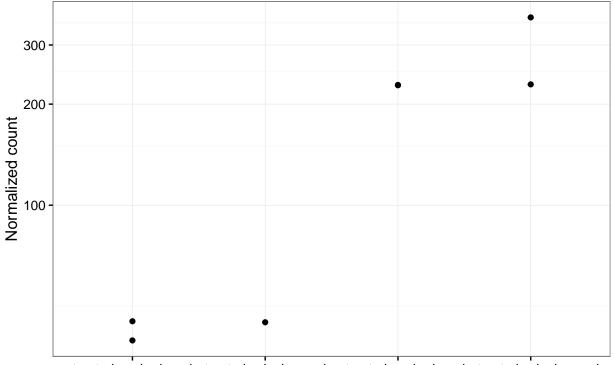






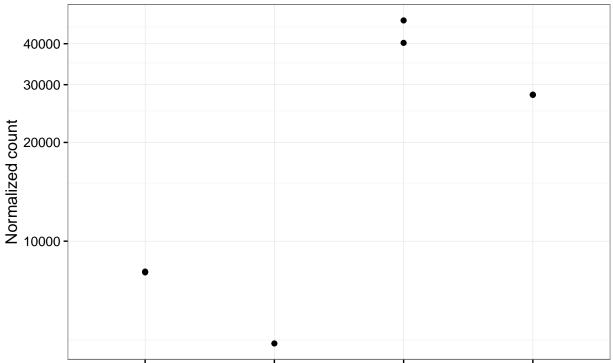
treated: paired-end treated: single-read untreated: paired-enduntreated: single-read group





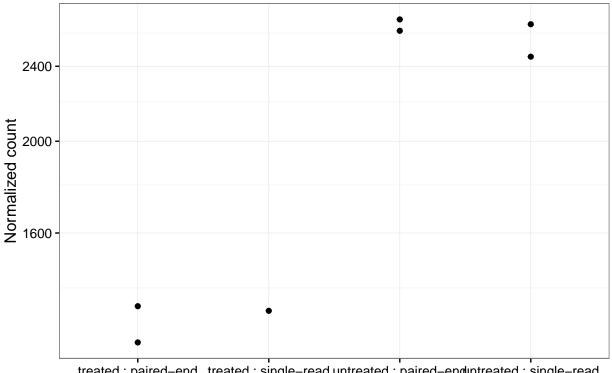
treated : paired-end treated : single-read untreated : paired-enduntreated : single-read



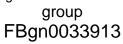


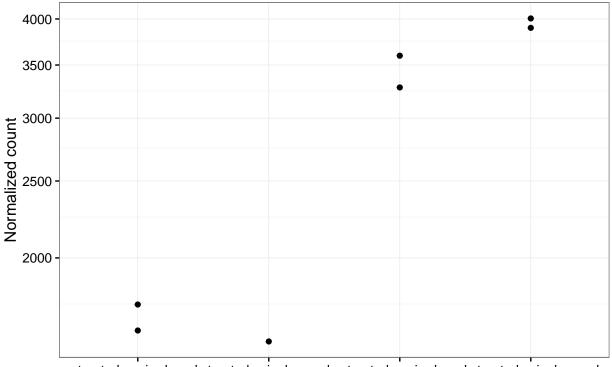
treated : paired-end treated : single-readuntreated : paired-end ntreated : single-read group

# FBgn0031191+FBgn0027279

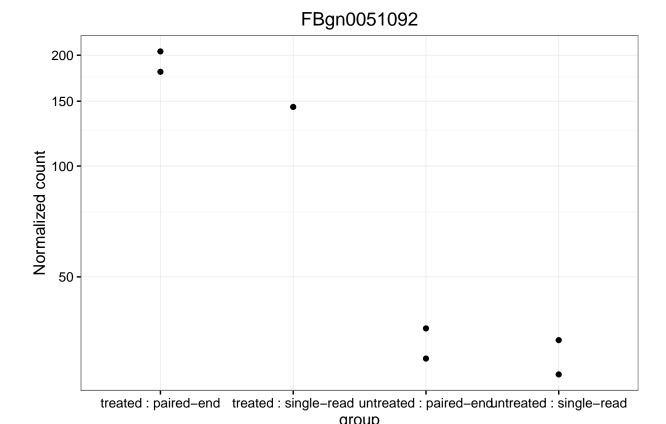


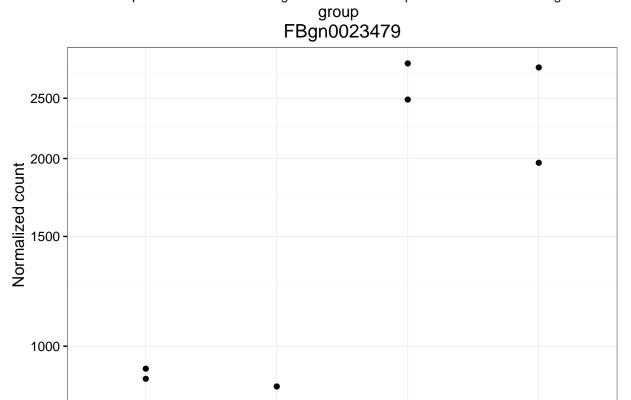
treated: paired-end treated: single-read untreated: paired-endintreated: single-read



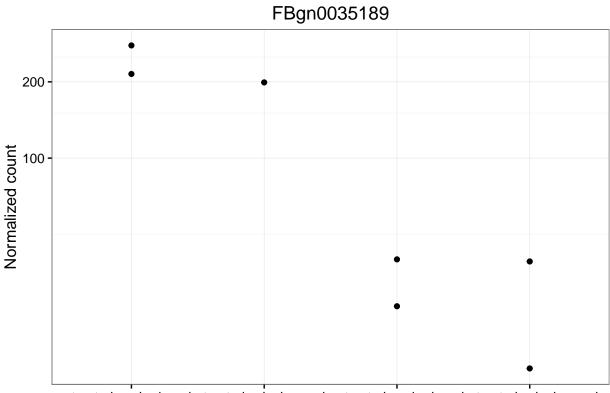


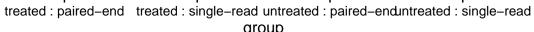
treated: paired-end treated: single-read untreated: paired-enduntreated: single-read group

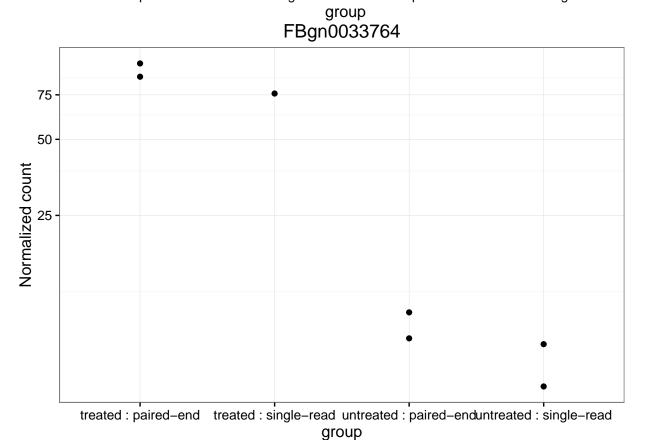


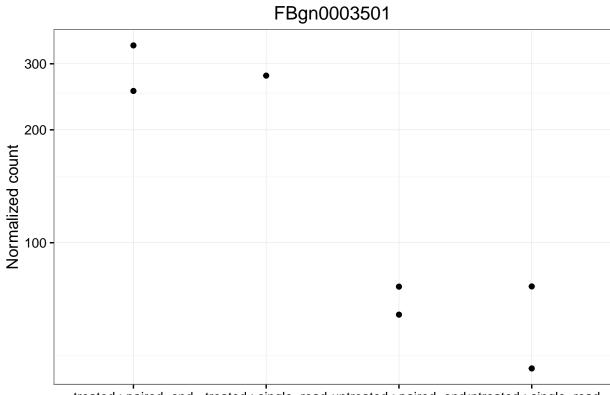


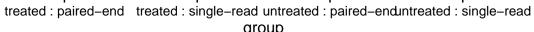
treated : paired-end treated : single-read untreated : paired-endintreated : single-read group

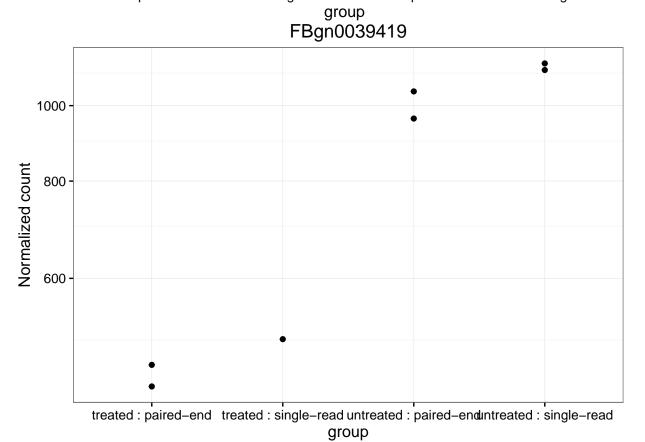












### Reproducibility

## BiocParallel

## biomaRt

## BSgenome

## Biostrings
## bitops

## bumphunter

## checkmate

## cluster

## codetools

The input for this report was generated with DESeq2 (Love, Huber, and Anders, 2014) using version 1.11.42 and the resulting features were called significantly differentially expressed if their BH adjusted p-values were less than alpha = 0.1. This report was generated in path /Users/lcollado/Dropbox/JHSPH/Code/regionReportSupp using the following call to DESeq2Report():

```
## DESeq2Report(dds = dds, project = "PDF report", intgroup = c("condition",
     "type"), outdir = "DESeq2-example", output = "DESeq2Report",
##
     device = "pdf", theme = theme_bw(), output_format = "pdf_document")
Date the report was generated.
## [1] "2016-04-12 07:32:12 EDT"
Wallclock time spent generating the report.
## Time difference of 25.555 secs
R session information.
setting value
##
  version R version 3.3.0 alpha (2016-03-23 r70368)
## system x86_64, darwin13.4.0
          X11
## ui
## language (EN)
## collate en_US.UTF-8
          America/New_York
## tz
          2016-04-12
   date
package
                    * version date
                                      source
  acepack
                      1.3-3.3 2014-11-24 CRAN (R 3.3.0)
##
## annotate
                     1.49.1
                             2016-02-06 Bioconductor
## AnnotationDbi
                     1.33.8
                             2016-04-10 Bioconductor
## backports
                     1.0.2
                             2016-03-18 CRAN (R 3.3.0)
## bibtex
                     0.4.0
                             2014-12-31 CRAN (R 3.3.0)
## Biobase
                    * 2.31.3
                             2016-01-14 Bioconductor
## BiocGenerics
                    * 0.17.4
                             2016-04-07 Bioconductor
```

2016-03-23 Bioconductor

2016-01-14 Bioconductor

2013-08-17 CRAN (R 3.3.0) 2016-02-21 Bioconductor

2016-03-29 Bioconductor

2016-04-08 CRAN (R 3.3.0)

2015-07-21 CRAN (R 3.3.0)

2015-07-15 CRAN (R 3.3.0)

2.39.12 2016-02-21 Bioconductor

1.5.21

2.27.2

1.0-6

1.39.4

1.11.5

1.7.4

2.0.3

0.2-14

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                                     2014-09-24 CRAN (R 3.3.0)
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                                     2016-03-23 Bioconductor
##
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                                     2016-01-14 Bioconductor
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                          * 0.1
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                                     2015-04-07 CRAN (R 3.3.0)
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                                     2016-03-24 Github (jimhester/knitrBootstrap@cdaa4a9)
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##
    RSQLite
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                            1.0.0
##
    rtracklayer
                            1.31.10
                                     2016-04-07 Bioconductor
##
    S4Vectors
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                                     2016-04-07 Bioconductor
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                                     2016-02-26 CRAN (R 3.3.0)
                            1.0-1
                                     2015-10-22 CRAN (R 3.3.0)
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##
    stringr
                            1.0.0
                                     2015-04-30 CRAN (R 3.3.0)
##
    SummarizedExperiment * 1.1.23
                                     2016-04-06 Bioconductor
    survival
                                     2015-07-02 CRAN (R 3.3.0)
                            2.38 - 3
   VariantAnnotation
                                     2016-04-07 Bioconductor
##
                            1.17.23
   XML
                            3.98-1.4 2016-03-01 CRAN (R 3.3.0)
##
##
  xtable
                            1.8 - 2
                                     2016-02-05 CRAN (R 3.3.0)
   XVector
                            0.11.8
                                     2016-04-06 Bioconductor
##
    yaml
                                     2014-06-12 CRAN (R 3.3.0)
                            2.1.13
                                     2016-03-19 Bioconductor
    zlibbioc
                            1.17.1
```

Pandoc version used: 1.17.0.3.

## **Bibliography**

This report was created with regionReport (Collado-Torres, Jaffe, and Leek, 2015) using rmarkdown (Allaire, Cheng, Xie, McPherson, et al., 2016) while knitr (Xie, 2014) and DT (Xie, 2015) were running behind the scenes. pheatmap (Kolde, 2015) was used to create the sample distances heatmap. Several plots were made with ggplot2 (Wickham, 2009).

Citations made with knitcitations (Boettiger, 2015). The BibTeX file can be found here.

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