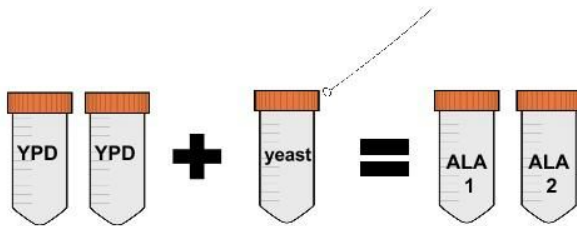


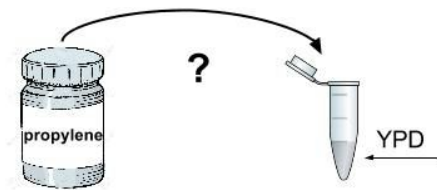
# Biological experiment

We want to observe the impact of the propylene glycol which is the main component of the liquid in e-cigarette on an organism to see if it is toxic. So we choose to study yeast because it is an organism which is relatively similar to humans. Indeed, the 2 organisms are eukaryotes. We put yeasts in culture with different concentration of propylene glycol to look if there is an impact on their growth.

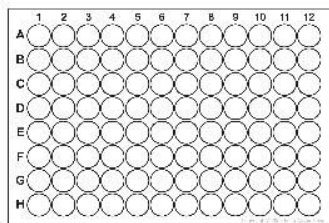
## 1 Yeast culture preparation



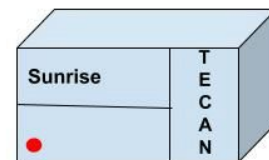
## 2 Propylene Concentration



## 3 Preparation of the 96 well plate



## 4 Optical Density Measurement



### Materials:

- yeasts solution
- YPD
- propylene glycol (8euros)
- falcons
- pipettes
- tips
- hood (to have sterile conditions)
- 96 well plate
- oil
- spectrophotometer (TECAN)

## Protocol:

I) The first step is the preparation of yeast culture:

- 1) Put 2mL ml of YPD into two 10ml falcon tube.
- 2) Add the yeast from yeast solution ( wild-type strain) into your falcons using a loop (one strain per falcon) under the hood. We obtain two yeast solution that we called ALA1 and ALA2
- 3) Put them at the incubator during 12h.

II) The second step is to determine the concentration of propylene glycol that we put with yeast. The LD50 is 20g / kg and the weight of one yeast is in range 3-10 picograms (we consider that the average weight is 5 picograms). In one well, we will put 2 microliters of yeast solution. Thus, we have to calculate the number of cells that are in this volume to next, find out the total weight of the sample.

- 1) Put 6,6 microliters of yeast solution in counting chambers.
- 2) Count the number of cells in one little square
- 3) Multiply this number by 90 to obtain the number of yeast in 1 microliters of solution (here you can directly multiply by 180 to have the cells number in 2 microliters)
- 4) Multiply this number by  $5 \times 10^{-12}$  to have the weight due to the yeast in your samples.
- 5) Repeat step 1-6 for the second yeast solution

In our case we have 720 cells per small square for ALA2 and 9 cells per small square for ALA1. So the weights are  $648\,000 \times 10^{-12}$  g for ALA2 and  $8100 \times 10^{-12}$  for ALA 1.

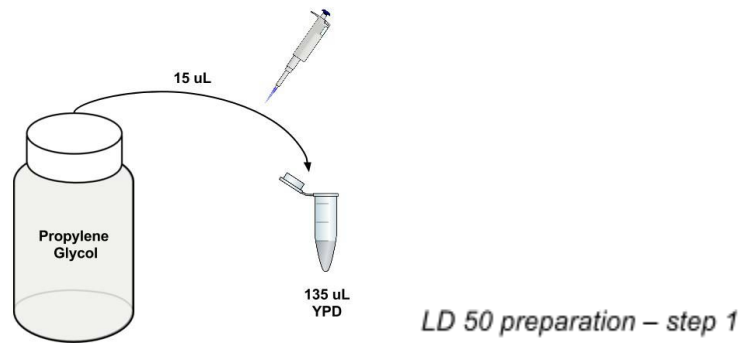
Now we have the yeast weight, we can deduce the concentration. As the density of PG is the same than water (1g/ml), the lethal dose is also equal to 20mL/ kg. By doing a rull of three, you can deduce simply the volume of PG as the LD50 dose.

### LD50

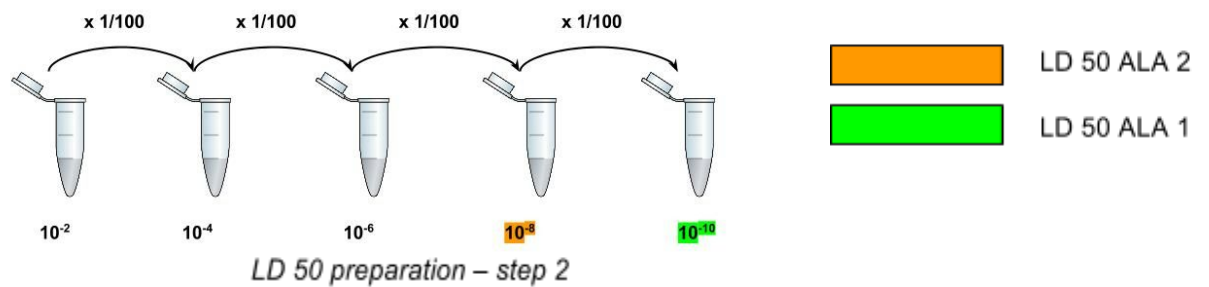
volume (mL)	masse (g)
20	1000
$1,2960 \times 10^{-8}$	$648\,000 \times 10^{-12}$
$1,62 \times 10^{-8}$	$8100 \times 10^{-12}$

So we used  $1.5 \times 10^{-8}$  mL for ALA2 and  $1.5 \times 10^{-10}$  mL for ALA1

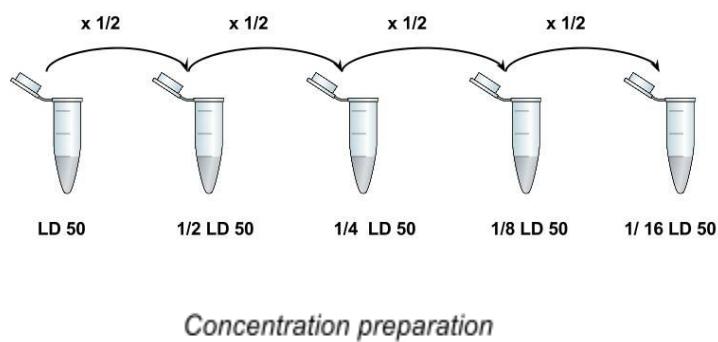
- 6) Take 15 microliters from the PG bottle using a P20 and put it into 135 microliters of YPD in an ependorf using a P1000



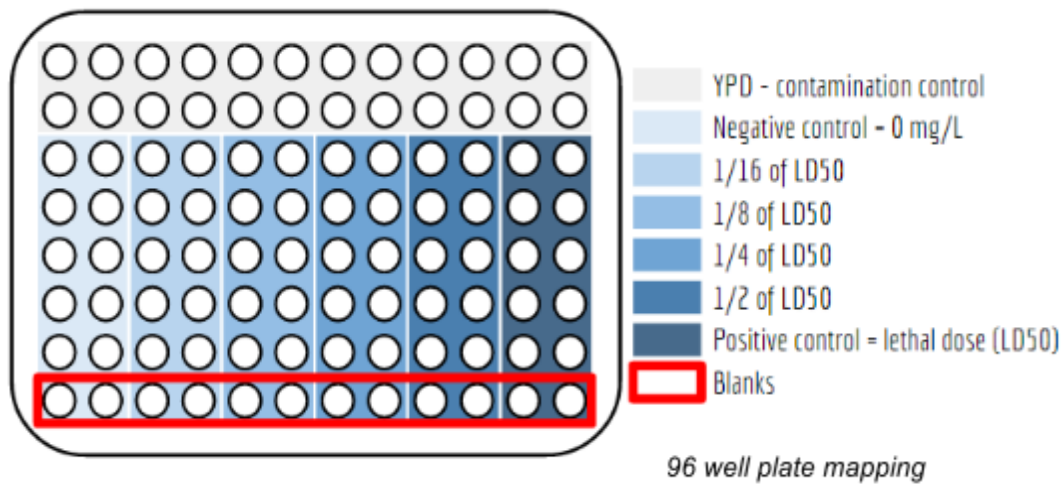
7) Do 4 serial dilutions by hundred from the solution prepared before (do the dilution in YPD and not water !! ) to obtain the LD50 concentration for ALA2 and do 5 serial dilutions for ALA1



8) Prepare the concentration by doing 4 serial dilution of two from the LD 50 (ALA2 and ALA1)



**III)** The last step is the preparation of the 96 well plate (refer to the picture below):



1) Put 100 microliters of YPD using a P200 in the first two lines

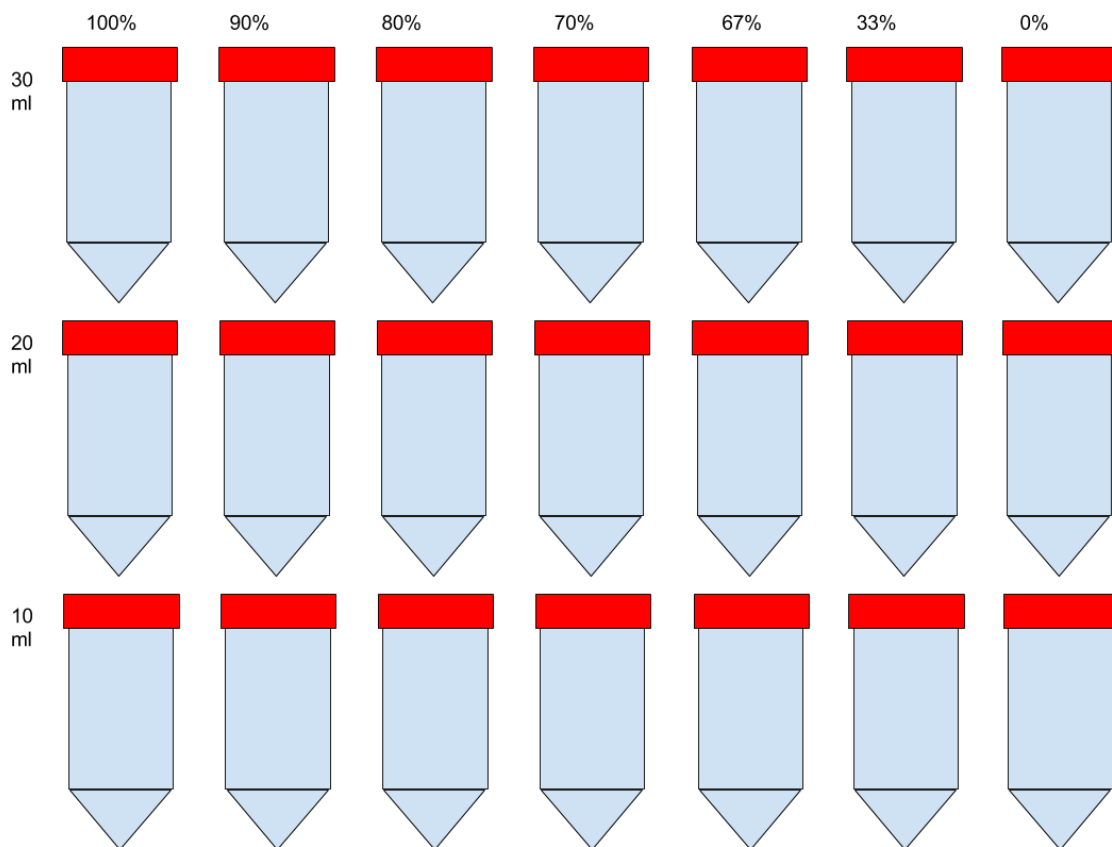
2) Put 98 microliters of PG diluted solutions (refer to the picture above) and 2 microliters of yeast in each well except for the last lines where you replace the yeast by 2 microliters of YPD.

3) Put 30 microliters of oil in each well to avoid dessication

**IV)** Measure the optical density of each well using a TECAN spectrophotometer. Take measure every 30 minutes during 12h.

# Electronic experiment

With this experiment, we want to evaluate also the toxicity of the propylene glycol. So we know that the propylene glycol is a liquid which absorbs the air humidity. Thus, we go to measure the air humidity with a DHT 22 captor at different volumes of propylene glycol and with or not YPD.

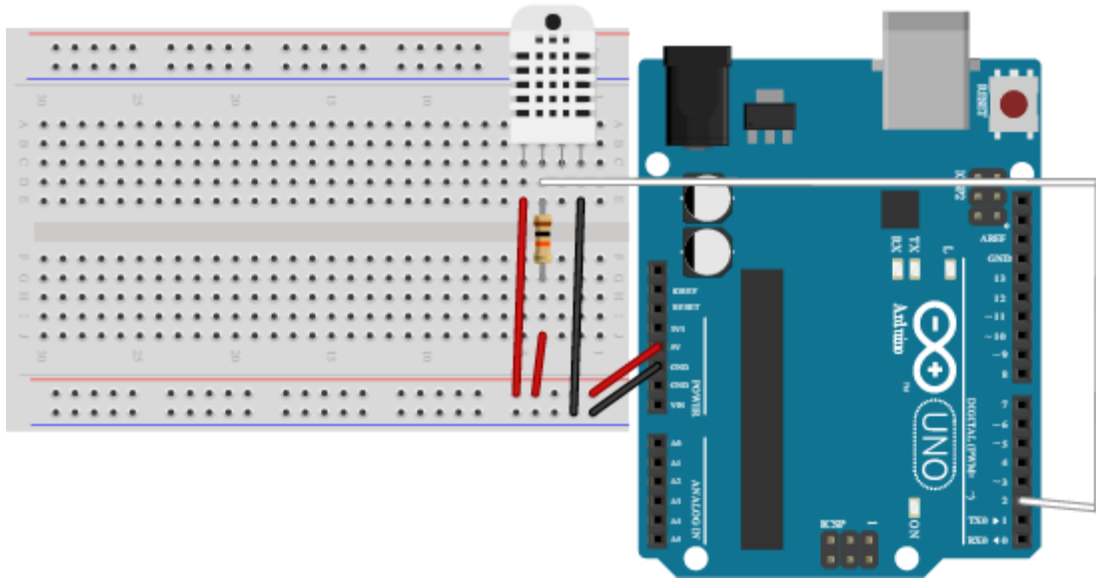


## Materials:

- 1 humidity captor DHT 22 (4euros)
- propylene glycol (8euros)
- YPD
- falcons
- 1 arduino
- thread of connections
- parafilm
- scotch
- A needle or one pulls cork

### Protocol:

- 1) Realize the montage to can use the humidity captor. We have 4 connections on the humidity captor. In this order, connect the 5V, digital 2, nothing and the ground.



- 2) If you can make welding, it is better for the measures with the captor.
- 3) Use this [link](#) to download the library to use the captor.
- 4) Take a falcon and drill the cork three times to pass each connections. We put on the cork parafilm and scotch to have a better insulation.
- 5) Put you solution in the falcon.
- 6) Put the humidity captor inside the falcon.
- 7) Close the falcon.
- 8) Measure the humidity all the 2 secondes during 10min.

