
FACTORS INFLUENCING THE AMPHIBIAN MYCOBIOME, WITH A FOCUS ON KNOWN AND UNKNOWN CHYTRIDS

MRES PROJECT PROPOSAL

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1 Introduction

Chytridiomycosis, caused by the amphibian chytrid fungus *Batrachochytrium dendrobatidis* (*Bd*), has been causing extirpations and extinctions in frogs over the past decades, with a presumed 90 extinctions, and has been linked to the decline of over 500 more species (Scheele et al. 2019). *Batrachochytrium salamandrivorans* (*BSaI*), another chytrid fungus, is a more recently emerged, highly virulent pathogen that is the sister taxon to *Bd* (Martel et al. 2013). *BSaI* was discovered over 20 years after *Bd* and is driving declines in fire salamanders in Europe. Given the discovery of *BSaI* and the prediction that more than 92% of fungal species are yet to be described, it is likely that there are other undiscovered chytrids that parasitise amphibians (Hawksworth and Lücking 2017). These are likely to be phylogeographically constrained endemic species that become tomorrow's emerging infections. Conversely, evolutionarily-distinct lineages with superior competitiveness may provide protection against virulent strains, such as *Bd* and *BSaI*, through competitive exclusion (Hardin 1960).

In the past few years, there has been an increase in research on the amphibian skin microbiome, for example that by (Bates et al. 2018) and (Bates et al. 2019) but little work specialising on the fungal communities present on amphibian skin, the mycobiome, and there has been no study on the factors influencing the diversity of the amphibian mycobiome globally. This is mostly due to difficulties in sequencing the ITS-2 region in fungi compared to bacteria (Tipton, Ghedin, and Morris 2017). A global fungal composition analysis of the fungi that have evolved the ability to colonise, or even infect, amphibian skin would allow the association between different commensal fungi and *Bd* and *BSaI* to be determined for the first time, improving understanding of the interaction between amphibians' skin, *Bd*, *BSaI* and other fungi. Negatively associated species may be evidence of niche exclusion (Hardin 1960), and positively associated species may represent secondary infections as a consequence of chytridiomycosis-associated dysbioses. This evidence could provide important groundwork for future conservation and novel promycotic treatments if strong negative associations are found.

2 Proposed Questions

- Which factors influence the amphibian mycobiome on a global scale?
- What are the associations between *Bd*, *BSaI* and other fungal species in the amphibian mycobiome?
- Are novel chytrid species a key link to providing a current and future defence against virulent evolutionarily-distinct lineages or do they themselves pose a threat of becoming virulent in the future?

3 Proposed Methods

Over the past decade, amphibian skin swabs and environmental abiotic data have been collected for a variety of species and life stages from all over the world, allowing an analysis of the amphibian mycobiome to be done on a global scale for the first time. The dataset currently contains 10 runs from 16 countries over 4 continents, with 3,185 samples with 175 controls. There are still some sequencing data yet to be generated, which will be done using the Illumina MiSeq platform from the ITS-2 barcoding region (the accepted to be the universal fungi barcode marker) (Schoch et al. 2012). This data will then be processed using existing bioinformatics pipelines, determining Operational Taxonomic Unit (OTU) tables, which cluster the sequences by 97% (representing assumed species-level cutoffs) similarity, and BLASTED against the UNITE and NCBI databases to assign taxonomy.

Using this data, it can be determined which commensal fungal species are present on large numbers of amphibian species from different populations and whether we can also detect unknown fungal species that may be pathogens. After accounting for potential autocorrelation and multiple comparisons, patterns can be identified and tested using statistical analyses in terms of possible influencing biogeographic factors, such as altitude, temperature, longitude and latitude etc. to deduce which factors better influence the composition of fungal species in an amphibian's mycobiome. These models will then be used to generate plots and GIS maps of locations of mycobiome similarity. The spread of *Bd*, focusing on the novel lineages, will also be looked at using GIS. The R package *cooccur* (Charles J. Marsh Daniel M. Griffith 2016) will be used to analyse cooccurrence between fungal species and *Bd* to evaluate which fungi are positively and negatively associated with *Bd*. Some of the unidentified fungal species could be unknown species of chytrid. Novel chytrid lineages will be identified through sequence alignment against pre-existing datasets in NCBI and by building phylogenetic trees in order to compare the candidate sequences with those of other chytrids.

4 Anticipated Outputs and Outcomes

Statistically and biologically significant patterns in the mycobiome data will be determined in order to confidently predict which factors most influence the amphibian mycobiome. This will give an overview of the global amphibian mycobiome and help to understand why certain amphibians are affected by *Bd* and *BSaI*, both in terms of abiotic factors and in terms of the other fungal species that are present. Identification of previously unknown chytrid species that may act as promyotics, causing niche exclusion of a competing virulent species/lineage through prior occupancy, could contribute to pre-emptive conservation work against unknown virulent chytrids.

5 Project Feasibility

All data has already been collected using MiSeq 350bp sequencing and almost all has already been processed and is ready for analysis. The time period for analysis and write up should be adequate to generate possible answers to all three of the above research questions.

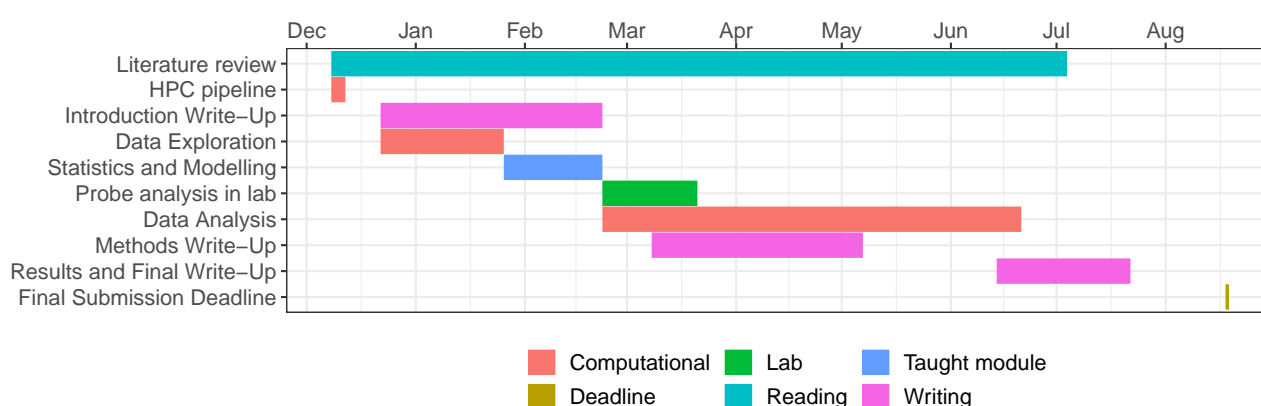


Figure 1: Proposed Project Timeline

Budgeted Item	Cost	Reasoning
Herpetofauna Workers Meeting 2020	£250	Herpetological conference cost and accommodation
qPCR Reagents	£175	To perform <i>Bd</i> qPCR in the laboratory
Laptop Adaptor	£75	To enable me to use USB sticks, HDMI etc. with my laptop
Total	£500	-

Table 1: Proposed Project Budget

References

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96 **6 Approval**

97 I have seen and approved the proposal and the budget.

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