

Recount Meta-Analysis

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Contents

Analysis	1
Load libraries we will need	1
Get data sets	1
Load the data	2
Load the metadata file	2
Get additional geo data for these samples	2
Merge the data sets	3
Get just the normals	3
Do the analysis comparing tissue to	3
Compare CAT plots	7
Reproducibility	9

Analysis

Load libraries we will need

```
library('dplyr')
library('recount')
library('magrittr')
library('limma')
library('edgeR')
library('ffpe')
library('RskittleBrewer')
library('SummarizedExperiment')
library('devtools')
trop = RskittleBrewer::RskittleBrewer('tropical')
```

Get data sets

We identify two projects consisting of samples from colon SRP029880,SRP042228 and three that contain samples from blood SRP059039, SRP059172, SRP062966.

```
colon_proj <- c('SRP029880', 'SRP042228')
if(any(!file.exists(file.path(colon_proj, 'rse_gene.Rdata')))) {
  sapply(colon_proj, download_study)
}

blood_proj <- c('SRP059039', 'SRP059172', 'SRP062966')
if(any(!file.exists(file.path(blood_proj, 'rse_gene.Rdata')))) {
  sapply(blood_proj, download_study)
}
```

```
proj <- c(colon_proj, blood_proj)
```

Load the data

Now we load these data sets into R and calculate the number of genes and samples for each data set

```
dat <- lapply(proj, function(x) {
  load(file.path(x, 'rse_gene.Rdata'))
  return(rse_gene)
})
proj

## [1] "SRP029880" "SRP042228" "SRP059039" "SRP059172" "SRP062966"

sapply(dat, dim)

##      [,1] [,2] [,3] [,4] [,5]
## [1,] 23779 23779 23779 23779 23779
## [2,]   54   314   205   169   117
```

Load the metadata file

Now we load the metadata from the SRA samples

```
metadata <- all_metadata('sra')

## 2016-06-13 16:19:02 downloading the metadata to /var/folders/cx/n9s558kx6fb7jf5z_pgszgb80000gn/T//Rt...
```

Get additional geo data for these samples

Now we go through and collect geo information for the samples. We label them with their respective tissue and identify which samples are supposed to be normal.

```
if(!file.exists('charvec.Rdata')) {
  charvec <- vector('list', 5)
  dir.create('geoinfo', showWarnings = FALSE)
  for(i in 1:5){
    index <- match(colData(dat[[i]])$run, metadata$run)
    colData(dat[[i]])$geo <- metadata$geo_accession[index]
    info <- sapply(colData(dat[[i]])$geo, geo_info, destdir = 'geoinfo')
    charvec[[i]] <- sapply(info, geo_characteristics)
  }
  save(charvec, file = 'charvec.Rdata')
} else {
  load('charvec.Rdata')
}

## first data set - normals called 'normal-looking surrounding colonic epithelium'
colData(dat[[1]])$normal <- grepl('normal', unlist(charvec[1])[(1:54) * 2 - 1])
colData(dat[[1]])$tissue <- 'colon'

## second data set - normals called
colData(dat[[2]])$normal <- grepl('not ibd', tolower(unlist(charvec[[2]][5, ])))
```

```
colData(dat[[2]])$tissue <- 'colon'

## third data set - normals called Control
colData(dat[[3]])$normal <- grepl('Control', unlist(charvec[[3]][2, ]))
colData(dat[[3]])$tissue <- 'blood'

## fourth data set - normals called Control

colData(dat[[4]])$normal <- grepl('Control', unlist(charvec[[4]][1, ]))
colData(dat[[4]])$tissue <- 'blood'

## fifth data set - normals called healthy

colData(dat[[5]])$normal <- grepl('healthy', unlist(charvec[[5]][1, ]))
colData(dat[[5]])$tissue <- 'blood'
```

Merge the data sets

Now we merge the data sets into one ranged summarized experiment

```
mdat <- do.call(cbind, dat)
```

Get just the normals

Find out how many samples are normal in each study and subset to just the normal samples for further analysis.

```
table(colData(mdat)$normal, colData(mdat)$project)
```

```
##
##          SRP029880 SRP042228 SRP059039 SRP059172 SRP062966
##  FALSE          35         273         181         122          99
##   TRUE           19          41          24          47          18
```

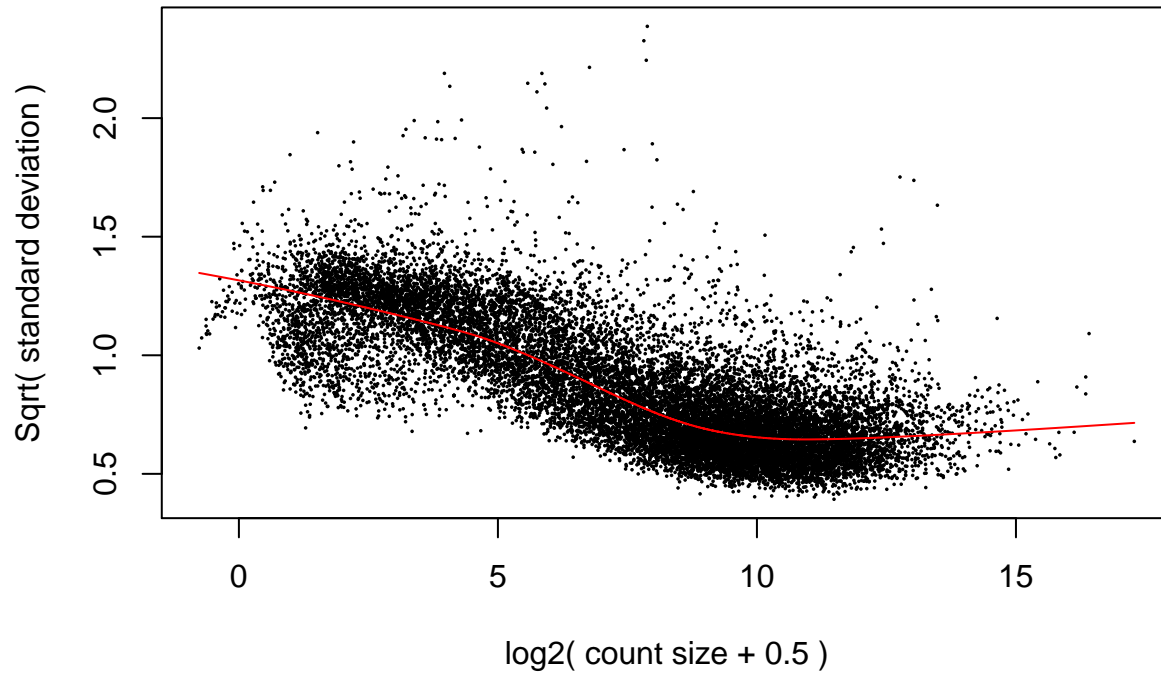
```
ndat <- mdat[, colData(mdat)$normal]
```

Do the analysis comparing tissue to

Here we do a differential expression analysis comparing blood to colon. We consider only the genes that have an average normalized count of at least 5 across the data set.

```
ndat <- scale_counts(ndat)
ndat_counts <- assays(ndat)[[1]]
keep <- rowMeans(ndat_counts) > 5
ndat_counts = ndat_counts[keep, ]
design <- model.matrix(~colData(ndat)$tissue)
dge <- DGEList(counts = ndat_counts)
dge <- calcNormFactors(dge)
v <- voom(dge, design, plot=TRUE)
```

voom: Mean-variance trend



```
fit <- lmFit(v, design)
fit <- eBayes(fit)
topTable(fit)
```

```
## Removing intercept from test coefficients
```

##	logFC	AveExpr	t	P.Value	adj.P.Val	B
## 7103	15.409396	0.4924358	79.43601	2.020597e-125	3.560898e-121	272.2428
## 10083	13.253543	-0.3745418	75.54867	3.368731e-122	2.968357e-118	265.4188
## 1909	7.783661	-2.5134139	75.11897	7.820641e-122	4.594105e-118	263.9306
## 56667	14.923360	0.6573342	73.53603	1.811337e-120	7.980298e-117	261.7541
## 25878	11.098638	-1.1683531	72.98350	5.507110e-120	1.941036e-116	260.6763
## 57381	8.703755	-1.8892183	70.64164	6.712078e-118	1.971449e-114	256.0142
## 133584	7.382474	-2.6339197	70.55688	8.008934e-118	2.016306e-114	255.2011
## 1441	-8.823323	8.1510433	-68.43403	7.140208e-116	1.143926e-112	254.6341
## 1015	14.208157	0.9677960	69.54431	6.710833e-117	1.314056e-113	254.2764
## 441094	6.910093	-2.8053456	69.62619	5.644920e-117	1.243505e-113	253.1423

GTEx analysis

Now we do the GTEx analysis comparing blood to colon.

```
## Download the GTEx data
if(!file.exists(file.path('SRP012682', 'rse_gene.Rdata'))){
  download_study('SRP012682')
}
load(file.path('SRP012682', 'rse_gene.Rdata'))

gtex_metadata <- all_metadata('gtex')
```

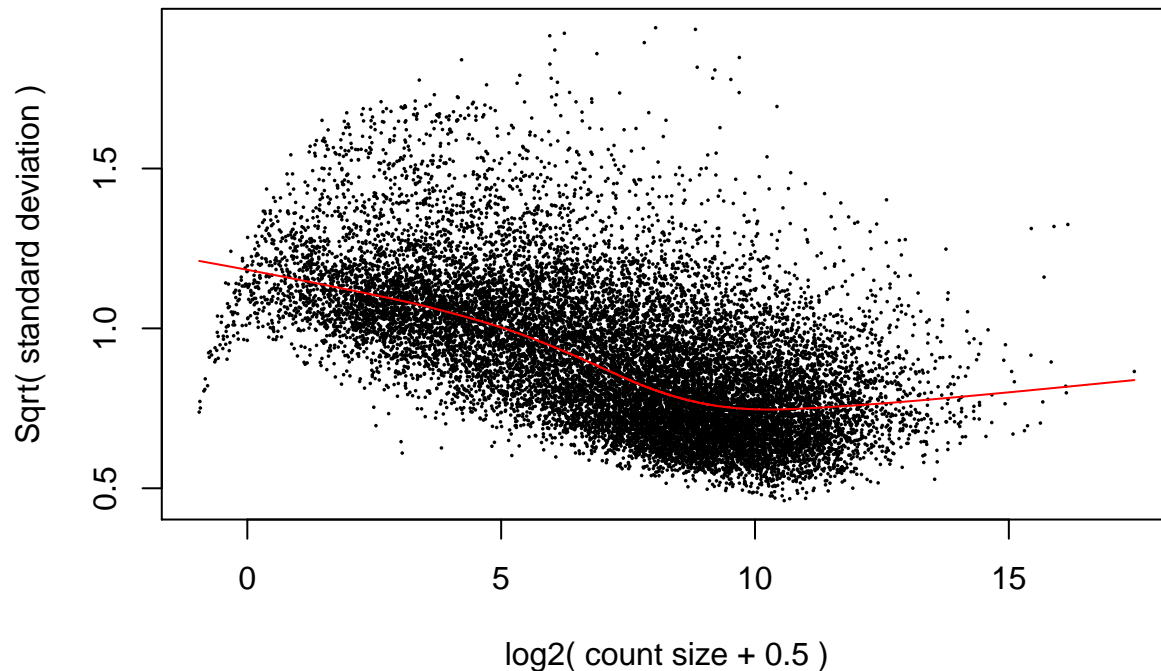
```
## 2016-06-13 16:19:31 downloading the metadata to /var/folders/cx/n9s558kx6fb7jf5z_pgszgb80000gn/T//Rtr
gtex_blood <- rse_gene[, subset(gtex_metadata, smtsd == 'Whole Blood')$run]
colData(gtex_blood)$tissue <- 'wholeblood'
gtex_colon <- rse_gene[, subset(gtex_metadata, smts == 'Colon')$run]
colData(gtex_colon)$tissue <- 'colon'

gtex_both <- do.call(cbind, list(gtex_blood, gtex_colon))
colData(gtex_both)$batch <- gtex_metadata[match(colData(gtex_both)$run,
  gtex_metadata$run), 'smgebtch']

gtex_both <- scale_counts(gtex_both)
gtex_both_counts <- assays(gtex_both)[[1]]
gtex_both_counts <- gtex_both_counts[keep, ]

design_gtex <- model.matrix(~colData(gtex_both)$tissue +
  colData(gtex_both)$batch)
dge_gtex <- DGEList(counts = gtex_both_counts)
dge_gtex <- calcNormFactors(dge_gtex)
v_gtex <- voom(dge_gtex, design_gtex, plot=TRUE)
```

voom: Mean-variance trend



```
fit_gtex <- lmFit(v_gtex, design_gtex)
fit_gtex <- eBayes(fit_gtex)
topTable(fit_gtex, coef = 2)
```

##	logFC	AveExpr	t	P.Value	adj.P.Val	B
## 6793	4.773601	6.779423	90.21488	0	0	874.9992
## 409	6.040177	7.723587	88.28947	0	0	861.2107
## 4542	7.490221	7.258075	84.59393	0	0	834.0351
## 4688	8.381934	6.579985	84.59066	0	0	833.9083

```
## 3936 7.980207 8.364888 84.02830 0 0 829.8006
## 1793 -5.244754 2.974639 -83.29415 0 0 823.9264
## 101 7.478347 6.054686 83.16901 0 0 823.2335
## 752 6.065599 7.218798 81.74749 0 0 812.4860
## 8514 4.617038 6.310878 80.81177 0 0 805.2694
## 51312 6.423430 7.944806 80.73634 0 0 804.6646
```

GTEX analysis

Now we do the GTEX analysis comparing blood to lung

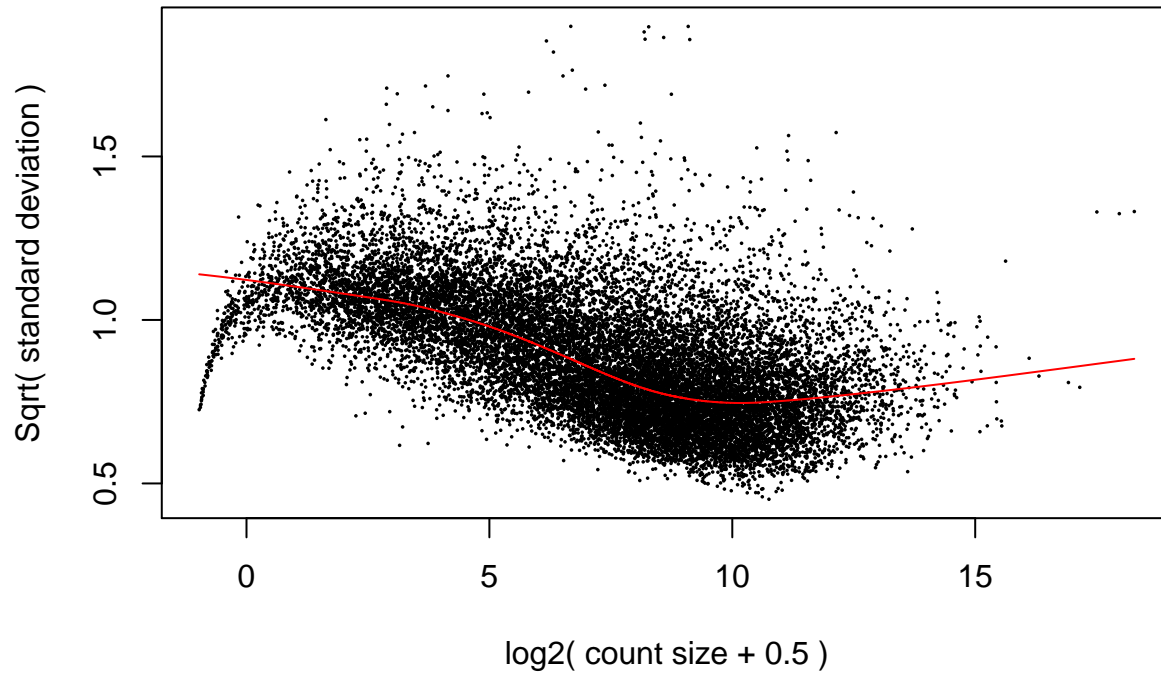
```
gtex_lung <- rse_gene[, subset(gtex_metadata, smts=='Lung')$run]
colData(gtex_lung)$tissue <- 'lung'

gtex_both_lung <- do.call(cbind, list(gtex_blood, gtex_lung))
colData(gtex_both_lung)$batch <- gtex_metadata[
  match(
    colData(gtex_both_lung)$run,
    gtex_metadata$run
  ), 'smgebtch']

gtex_both_lung <- scale_counts(gtex_both_lung)
gtex_both_lung_counts <- assays(gtex_both_lung)[[1]]
gtex_both_lung_counts <- gtex_both_lung_counts[keep,]

design_gtex_lung <- model.matrix(~colData(gtex_both_lung)$tissue +
  colData(gtex_both_lung)$batch)
dge_gtex_lung <- DGEList(counts = gtex_both_lung_counts)
dge_gtex_lung <- calcNormFactors(dge_gtex_lung)
v_gtex_lung <- voom(dge_gtex_lung, design_gtex_lung, plot = TRUE)
```

voom: Mean–variance trend



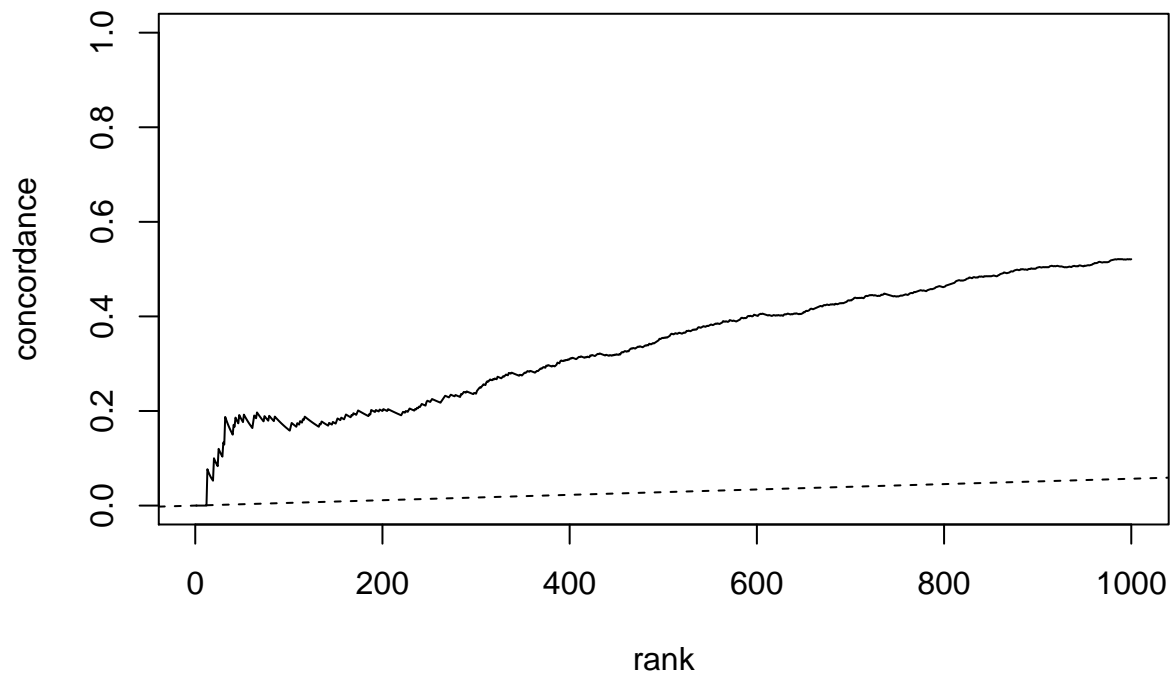
```
fit_gtex_lung <- lmFit(v_gtex_lung, design_gtex_lung)
fit_gtex_lung <- eBayes(fit_gtex_lung)
topTable(fit_gtex_lung, coef = 2)
```

##	logFC	AveExpr	t	P.Value	adj.P.Val	B
## 221395	-7.675466	3.4109146	-106.91629	0	0	996.3729
## 6943	-8.792146	1.6860275	-101.86151	0	0	964.1776
## 5754	-5.150188	2.7899033	-97.54954	0	0	935.9110
## 5420	-6.092697	4.0348220	-97.15255	0	0	933.2634
## 6909	-8.130718	2.9581907	-97.16373	0	0	933.1846
## 599	-2.835088	4.1595326	-96.50413	0	0	928.9989
## 10418	-6.396727	3.2092320	-94.48530	0	0	914.9644
## 9368	3.712945	6.1694693	94.43085	0	0	914.8108
## 207107	-8.226899	-0.5998609	-93.51892	0	0	908.0309
## 10160	-5.323208	3.0386054	-93.39156	0	0	907.3923

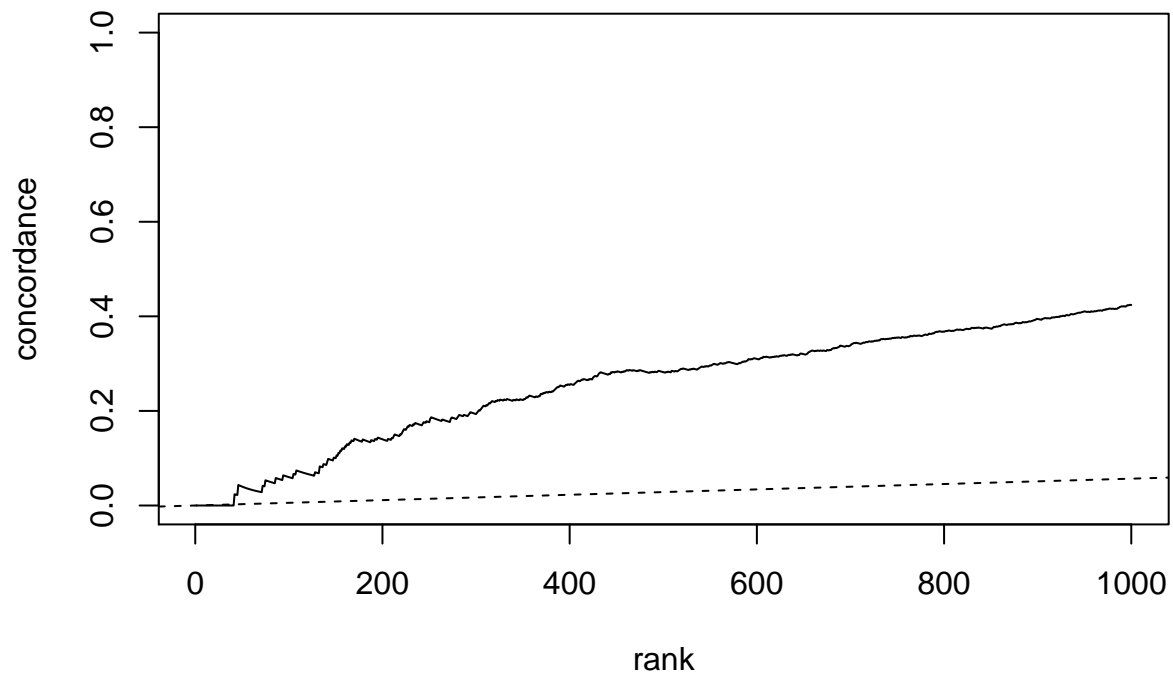
Compare CAT plots

Make CAT plots and compare different analyses.

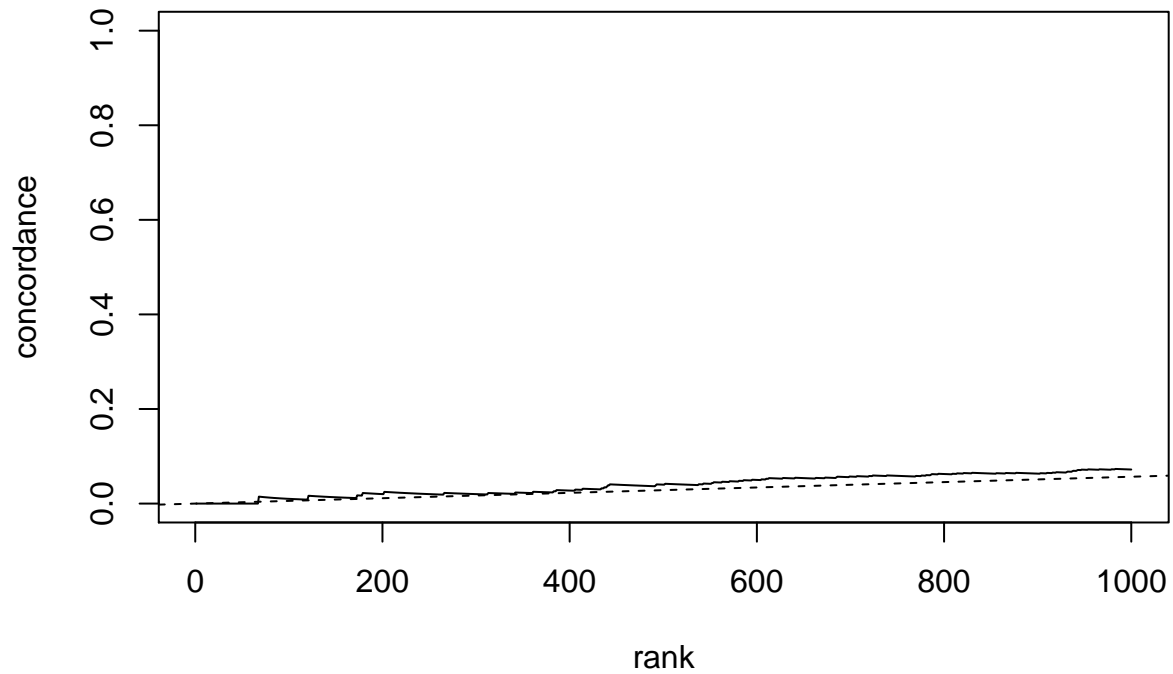
```
cat_sra_gtex <- CATplot(
  -rank(fit$coefficients[, 2]),
  -rank(-fit_gtex$coefficients[, 2]), maxrank = 1000, ylim = c(0,1))
```



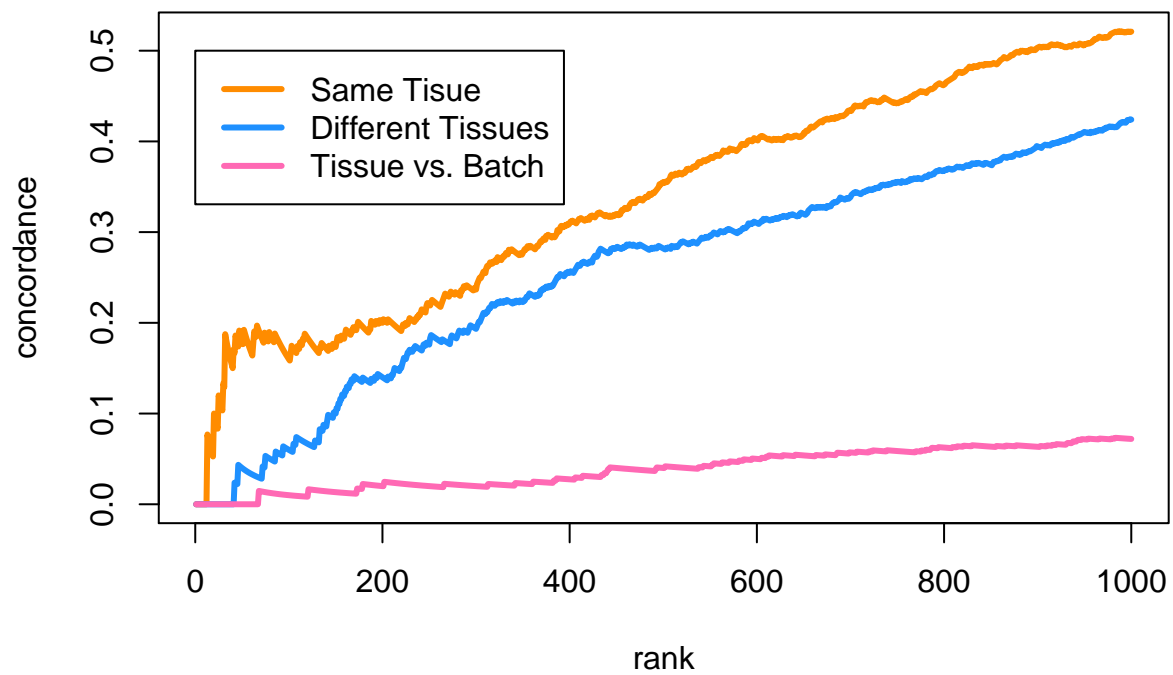
```
cat_sra_gtex_lung = CATplot(
  -rank(fit$coefficients[, 2]),
  -rank(-fit_gtex_lung$coefficients[, 2]), maxrank = 1000, ylim = c(0,1))
```



```
cat_sra_gtex_batch = CATplot(
  -rank(fit$coefficients[, 2]),
  -rank(-fit_gtex_lung$coefficients[, 3]), maxrank = 1000, ylim = c(0,1))
```

```
plot(cat_sra_gtex, type = 'l', col = trop[1], lwd = 3)
lines(cat_sra_gtex_lung, type = 'l', col = trop[2], lwd = 3)
lines(cat_sra_gtex_batch, type = 'l', col = trop[3], lwd = 3)
legend(0, 0.5, legend=c('Same Tissue', 'Different Tissues',
  'Tissue vs. Batch'), col = trop[1:3], lwd = 3)
```



Reproducibility

This analysis report was made possible thanks to:

- R (R Core Team, 2016)
- *BiocStyle* (Oleś, Morgan, and Huber, 2016)
- *derfinder* (Collado-Torres, Nellore, Frazee, Wilks, et al., 2016)
- *devtools* (Wickham and Chang, 2016)
- *dplyr* (Wickham and Francois, 2015)
- *edgeR* (Robinson, McCarthy, and Smyth, 2010)
- *ffpe* (Waldron, L, Ogino, Shuji, Hoshida, Yujin, Shima, Kaori, et al., 2012)
- *knitcitations* (Boettiger, 2015)
- *magrittr* (Bache and Wickham, 2014)
- *recount* (Collado-Torres and Leek, 2016)
- *rmarkdown* (Allaire, Cheng, Xie, McPherson, et al., 2016)
- *RSkittleBrewer* (Frazee, 2016)
- *SummarizedExperiment* (Morgan, Obenchain, Hester, and Pagès, 2016)
- *limma* (Law, Chen, Shi, and Smyth, 2014)

Bibliography file

- [1] J. Allaire, J. Cheng, Y. Xie, J. McPherson, et al. *rmarkdown*: Dynamic Documents for R. R package version 0.9.6. 2016. URL: <https://CRAN.R-project.org/package=rmarkdown>.
- [2] S. M. Bache and H. Wickham. *magrittr*: A Forward-Pipe Operator for R. R package version 1.5. 2014. URL: <https://CRAN.R-project.org/package=magrittr>.
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- [11] M. D. Robinson, D. J. McCarthy and G. K. Smyth. “edgeR: a Bioconductor package for differential expression analysis of digital gene expression data”. In: *Bioinformatics* 26 (2010), pp. -1.
- [12] Waldron, L, Ogino, Shuji, Hoshida, Yujin, Shima, Kaori, et al. “Expression profiling of archival tumors for long-term health studies”. In: *Clinical Cancer Research* 18.22 (2012). PMID: 23136189, pp. 6136–6146. DOI: 10.1158/1078-0432.CCR-12-1915.
- [13] H. Wickham and W. Chang. *devtools*: Tools to Make Developing R Packages Easier. R package version 1.11.1. 2016. URL: <https://CRAN.R-project.org/package=devtools>.
- [14] H. Wickham and R. Francois. *dplyr*: A Grammar of Data Manipulation. R package version 0.4.3. 2015. URL: <https://CRAN.R-project.org/package=dplyr>.

```
## Time spent creating this report:
diff(c(timestart, Sys.time()))
```

```

## Time difference of 13.37282 mins
## Date this report was generated
message(Sys.time())

## 2016-06-13 16:32:23
## Reproducibility info
options(width = 120)
devtools::session_info()

## Session info -----
##   setting  value
##   version  R version 3.3.0 RC (2016-05-01 r70572)
##   system   x86_64, darwin13.4.0
##   ui       X11
##   language (EN)
##   collate  en_US.UTF-8
##   tz       America/New_York
##   date     2016-06-13

## Packages -----
##   package      * version  date      source
##   affy          1.51.0   2016-05-27 Bioconductor
##   affyio        1.43.0   2016-05-27 Bioconductor
##   annotate      1.51.0   2016-05-05 Bioconductor
##   AnnotationDbi 1.35.3   2016-05-27 Bioconductor
##   assertthat    0.1      2013-12-06 CRAN (R 3.3.0)
##   base64        2.0      2016-05-10 CRAN (R 3.3.0)
##   beanplot      1.2      2014-09-19 CRAN (R 3.3.0)
##   bibtex        0.4.0    2014-12-31 CRAN (R 3.3.0)
##   Biobase       * 2.33.0   2016-05-05 Bioconductor
##   BiocGenerics  * 0.19.1   2016-06-11 Bioconductor
##   BiocInstaller 1.23.4   2016-05-27 Bioconductor
##   BiocParallel  1.7.2    2016-05-20 Bioconductor
##   BiocStyle     * 2.1.6    2016-06-11 Bioconductor
##   biomaRt       2.29.2   2016-05-30 Bioconductor
##   Biostrings    2.41.2   2016-06-08 Bioconductor
##   bitops        1.0-6    2013-08-17 CRAN (R 3.3.0)
##   bumphunter    1.13.0   2016-05-05 Bioconductor
##   chron         2.3-47   2015-06-24 CRAN (R 3.3.0)
##   codetools     0.2-14   2015-07-15 CRAN (R 3.3.0)
##   colorout      * 1.1-2    2016-05-05 Github (jalvesaq/colorout@6538970)
##   colorspace    1.2-6    2015-03-11 CRAN (R 3.3.0)
##   data.table    1.9.6    2015-09-19 CRAN (R 3.3.0)
##   DBI           0.4-1    2016-05-08 CRAN (R 3.3.0)
##   devtools     * 1.11.1   2016-04-21 CRAN (R 3.3.0)
##   digest        0.6.9    2016-01-08 CRAN (R 3.3.0)
##   doRNG         1.6      2014-03-07 CRAN (R 3.3.0)
##   dplyr         * 0.4.3    2015-09-01 CRAN (R 3.3.0)
##   edgeR         * 3.15.0   2016-05-27 Bioconductor
##   evaluate      0.9      2016-04-29 CRAN (R 3.3.0)
##   ffpe         * 1.17.0   2016-05-27 Bioconductor
##   foreach       1.4.3    2015-10-13 CRAN (R 3.3.0)
##   formatR      1.4      2016-05-09 CRAN (R 3.3.0)

```

##	genefilter	1.55.2	2016-05-27	Bioconductor
##	GenomeInfoDb	* 1.9.1	2016-05-13	Bioconductor
##	GenomicAlignments	1.9.2	2016-06-13	Bioconductor
##	GenomicFeatures	1.25.12	2016-05-21	Bioconductor
##	GenomicRanges	* 1.25.4	2016-06-10	Bioconductor
##	GEOquery	2.39.3	2016-05-20	Bioconductor
##	htmltools	0.3.5	2016-03-21	CRAN (R 3.3.0)
##	httr	1.1.0	2016-01-28	CRAN (R 3.3.0)
##	illuminaio	0.15.0	2016-05-27	Bioconductor
##	IRanges	* 2.7.6	2016-06-10	Bioconductor
##	iterators	1.0.8	2015-10-13	CRAN (R 3.3.0)
##	KernSmooth	2.23-15	2015-06-29	CRAN (R 3.3.0)
##	knitcitations	* 1.0.7	2015-10-28	CRAN (R 3.3.0)
##	knitr	1.13	2016-05-09	CRAN (R 3.3.0)
##	lattice	0.20-33	2015-07-14	CRAN (R 3.3.0)
##	limma	* 3.29.7	2016-06-13	Bioconductor
##	locfit	1.5-9.1	2013-04-20	CRAN (R 3.3.0)
##	lubridate	1.5.6	2016-04-06	CRAN (R 3.3.0)
##	lumi	2.25.0	2016-05-27	Bioconductor
##	magrittr	* 1.5	2014-11-22	CRAN (R 3.3.0)
##	MASS	7.3-45	2016-04-21	CRAN (R 3.3.0)
##	Matrix	1.2-6	2016-05-02	CRAN (R 3.3.0)
##	matrixStats	0.50.2	2016-04-24	CRAN (R 3.3.0)
##	mclust	5.2	2016-03-31	CRAN (R 3.3.0)
##	memoise	1.0.0	2016-01-29	CRAN (R 3.3.0)
##	methyllumi	2.19.3	2016-06-03	Bioconductor
##	mgcv	1.8-12	2016-03-03	CRAN (R 3.3.0)
##	minfi	1.19.2	2016-05-27	Bioconductor
##	multtest	2.29.0	2016-05-27	Bioconductor
##	nleqslv	3.0.1	2016-05-02	CRAN (R 3.3.0)
##	nlme	3.1-128	2016-05-10	CRAN (R 3.3.0)
##	nor1mix	1.2-1	2015-07-27	CRAN (R 3.3.0)
##	openssl	0.9.3	2016-05-04	CRAN (R 3.3.0)
##	pkgmaker	0.22	2014-05-14	CRAN (R 3.3.0)
##	plyr	1.8.3	2015-06-12	CRAN (R 3.3.0)
##	preprocessCore	1.35.0	2016-05-27	Bioconductor
##	quadprog	1.5-5	2013-04-17	CRAN (R 3.3.0)
##	R6	2.1.2	2016-01-26	CRAN (R 3.3.0)
##	RColorBrewer	1.1-2	2014-12-07	CRAN (R 3.3.0)
##	Rcpp	0.12.5	2016-05-14	CRAN (R 3.3.0)
##	RCurl	1.95-4.8	2016-03-01	CRAN (R 3.3.0)
##	recount	* 0.99.10	2016-06-12	Github (leekgroup/recount@7a7ea73)
##	RefManager	0.10.13	2016-04-04	CRAN (R 3.3.0)
##	registry	0.3	2015-07-08	CRAN (R 3.3.0)
##	reshape	0.8.5	2014-04-23	CRAN (R 3.3.0)
##	RJSONIO	1.3-0	2014-07-28	CRAN (R 3.3.0)
##	rmarkdown	* 0.9.6	2016-05-01	CRAN (R 3.3.0)
##	rngtools	1.2.4	2014-03-06	CRAN (R 3.3.0)
##	Rsamtools	1.25.0	2016-05-05	Bioconductor
##	RSkittleBrewer	* 1.1	2016-06-13	Github (alyssafrazee/RSkittleBrewer@230d1d0)
##	RSQLite	1.0.0	2014-10-25	CRAN (R 3.3.0)
##	rstudioapi	0.5	2016-01-24	CRAN (R 3.3.0)
##	rtracklayer	1.33.5	2016-06-13	Bioconductor
##	S4Vectors	* 0.11.4	2016-06-11	Bioconductor

##	sfsmisc	1.1-0	2016-02-23	CRAN (R 3.3.0)
##	siggenes	1.47.0	2016-05-27	Bioconductor
##	stringi	1.0-1	2015-10-22	CRAN (R 3.3.0)
##	stringr	1.0.0	2015-04-30	CRAN (R 3.3.0)
##	SummarizedExperiment *	1.3.4	2016-06-10	Bioconductor
##	survival	2.39-4	2016-05-11	CRAN (R 3.3.0)
##	TTR	* 0.23-1	2016-03-21	CRAN (R 3.3.0)
##	withr	1.0.1	2016-02-04	CRAN (R 3.3.0)
##	XML	3.98-1.4	2016-03-01	CRAN (R 3.3.0)
##	xtable	1.8-2	2016-02-05	CRAN (R 3.3.0)
##	xts	0.9-7	2014-01-02	CRAN (R 3.3.0)
##	XVector	0.13.0	2016-05-05	Bioconductor
##	yaml	2.1.13	2014-06-12	CRAN (R 3.3.0)
##	zlibbioc	1.19.0	2016-05-05	Bioconductor
##	zoo	1.7-13	2016-05-03	CRAN (R 3.3.0)