

# Package ‘iterbi’

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**Type** Package

**Title** A iteratively bifurcated clustering strategy for single-cell sequencing data.

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**Description** A iteratively bifurcated clustering strategy for single-cell sequencing data.

**License** GPL-3

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dataframe_to_vector	<i>transform dataframe to vector</i>
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**Description**

transform dataframe to vector

**Usage**

```
dataframe_to_vector(df)
```

**Arguments**

df	A dataframe from Seuratobj[["attr"]]
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**Value**

A vector with names

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draw_GO_barplot	<i>Draw barplot for GO annotation</i>
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**Description**

Draw barplot for GO annotation

**Usage**

```
draw_GO_barplot(barplot_df, cluster.chain)
```

**Arguments**

barplot_df	A GO annotation dataframe from clusterProfiler
cluster.chain	cluster.chain from iterbi

**Value**

A ggplot barplot

---

`draw_iterbi_cluster_tree`*Draw cluster tree/chain by clustree*

---

**Description**

Draw cluster tree/chain by clustree

**Usage**

```
draw_iterbi_cluster_tree(  
  seuratObj,  
  iterbi.cellMeta,  
  node_text_size = 7,  
  node_size = 8  
)
```

**Arguments**

seuratObj	A Seurat object
iterbi.cellMeta	iterbi.cellMeta from iterbi
node_text_size	see node_text_size in clustree function
node_size	see node_size in clustree function

**Value**

clustree object

---

`draw_marker_chain_dotplot`*Draw cluster tree/chain by clustree*

---

**Description**

Draw cluster tree/chain by clustree

**Usage**

```
draw_marker_chain_dotplot(  
  seuratObj,  
  iterbi.marker.chain,  
  rmDup = T,  
  top_n = 3,  
  rel_heights = c(0.5, 9, 1.5),  
  rel_widths = c(1, 4)  
)
```

**Arguments**

seuratObj	A Seurat object
iterbi.marker.chain	iterbi.marker.chain from iterbi
rmDup	remove duplicated genes or not
top_n	top n genes for dotplot (sort by P-value)
rel_heights	relative heights for plot_grid(), the first and last control the height of top and bottom margin, the middle one controls the height of tree
rel_widths	relative widths for plot_grid(), the first one controls the widths of tree, the second one controls the widths of dotplot

**Value**

A plot\_grid integrated tree and dotplot

---

draw\_marker\_chain\_heatmap

*Draw heatmap by ComplexHeatmap*

---

**Description**

Draw heatmap by ComplexHeatmap

**Usage**

```
draw_marker_chain_heatmap(
  seuratObj,
  iterbi.cellMeta,
  iterbi.marker.chain,
  compare_anno = "",
  known_markers = c()
)
```

**Arguments**

seuratObj	A Seurat object
iterbi.cellMeta	iterbi.cellMeta from iterbi
iterbi.marker.chain	iterbi.marker.chain from iterbi
compare_anno	select a annotation in seurat object for comparision (e.g. previous annotation)
known_markers	the genes to include in the heatmap (like known markers)

**Value**

ComplexHeatmap object

---

`find_bifurcation_resolution`*Find the best resolution for bifurcation in graph-based clustering*

---

**Description**

Find the best resolution for bifurcation in graph-based clustering

**Usage**

```
find_bifurcation_resolution(seuratObj, res_sets = 50)
```

**Arguments**

<code>seuratObj</code>	A Seurat object
<code>res_sets</code>	The number of resolution for searching

**Value**

The best resolution which can bifurcate all cells

---

`get.iterbi.colors`      *Colors for iterbi*

---

**Description**

Colors for iterbi

**Usage**

```
get.iterbi.colors()
```

**Value**

A color vector

---

```
get_initial_bifurcation_of_two_clusters
```

*Find initial bifurcation event of any two clusters in iterbi.cellMeta*

---

### Description

Find initial bifurcation event of any two clusters in iterbi.cellMeta

### Usage

```
get_initial_bifurcation_of_two_clusters(  
  iterbi.cellMeta,  
  cluster_1,  
  cluster_2,  
  balance_cells = T  
)
```

### Arguments

iterbi.cellMeta	iterbi.cellMeta from iterbi
cluster_1	cluster 1
cluster_2	cluster 2
balance_cells	output balanced cell number (T or F)

### Value

A list. subset\_seuratObj is the subset cells in cluster 1 and cluster 2 diff\_marker\_1 is the marker of cluster 1, diff\_marker\_2 is the marker of cluster 2

---

```
get_marker_GO_chain      Get GO chain corresponding to a cluster chain
```

---

### Description

Get GO chain corresponding to a cluster chain

### Usage

```
get_marker_GO_chain(iterbi.GO.anno, cluster.chain, top_n = 3)
```

### Arguments

iterbi.GO.anno	A GO annotation dataframe from clusterProfiler
cluster.chain	cluster.chain from iterbi
top_n	top n GO terms for barplot (sort by P-value)

### Value

A ggplot barplot showed GO chain

---

get_test_data	<i>Download raw single-cell matrix for testing</i>
---------------	--

---

**Description**

Download raw single-cell matrix for testing

**Usage**

```
get_test_data()
```

**Value**

downloaded single-cell dataset

---

get_unique_marker	<i>filter uniquely expressed markers by expression percentage</i>
-------------------	---

---

**Description**

filter uniquely expressed markers by expression percentage

**Usage**

```
get_unique_marker(
  seuratObj,
  iterbi.marker.chain,
  assay = "RNA",
  slot = "data",
  min_cluster_pct = 0.3,
  max_bcg_pct = 0.1,
  min_diff = 0.3
)
```

**Arguments**

seuratObj	A Seurat object
iterbi.marker.chain	iterbi.marker.chain dataframe contains all the markers
assay	Assay used for prediction
slot	slot used for prediction
min_cluster_pct	Minimal expression percentage of target cluster
max_bcg_pct	Maximum expression percentage of the background cells (non-target cells)
min_diff	Minimal difference (expression percentage) between target cluster and background cells

**Value**

Uniquely expressed iterbi.marker.chain

---

hello	<i>Hello, World!</i>
-------	----------------------

---

### Description

Prints 'Hello, world!'.

### Usage

```
hello()
```

### Examples

```
hello()
```

---

identify_binary_markers	<i>Identify binary markers</i>
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---

### Description

Identify binary markers

### Usage

```
identify_binary_markers(
  seuratObj,
  p_value = 0.01,
  min_avg_logFC = 0.1,
  min_correlation = 0.3
)
```

### Arguments

seuratObj	A Seurat object
p_value	The number of resolution for searching
min_avg_logFC	Minimal threshold for logFC to pick markers
min_correlation	Minimal threshold for correlation to pick markers

### Value

A list. b1 is the positive marker of cluster 0, b2 is the positive marker of cluster 1.



---

`iterbi.bifucation.graph`*Bifurcation based on graph-based clustering*

---

**Description**

Bifurcation based on graph-based clustering

**Usage**

```
iterbi.bifucation.graph(seuratObj, res_sets = 50)
```

**Arguments**

<code>seuratObj</code>	A Seurat object
<code>res_sets</code>	The number of resolution for searching

**Value**

A bifurcated Seurat object (see `active.ident`).

---

`iterbi.bifucation.hclust`*Bifurcation based on hierarchical clustering*

---

**Description**

Bifurcation based on hierarchical clustering

**Usage**

```
iterbi.bifucation.hclust(seuratObj, method = "euclidean")
```

**Arguments**

<code>seuratObj</code>	A Seurat object
<code>method</code>	The distance measurement method "euclidean or correlation"

**Value**

A bifurcated Seurat object (see `active.ident`).

---

```
iterbi.bifucation.kmeans
```

*Bifurcation based on K-means clustering*

---

### Description

Bifurcation based on K-means clustering

### Usage

```
iterbi.bifucation.kmeans(seuratObj, method = "euclidean")
```

### Arguments

seuratObj	A Seurat object
method	The distance measurement method "euclidean or correlation"

### Value

A bifurcated Seurat object (see active.ident).

---

```
iterbi_GO_annotation
```

*Perform GO annotation of iterbi.marker.chain dataframe*

---

### Description

Perform GO annotation of iterbi.marker.chain dataframe

### Usage

```
iterbi_GO_annotation(  
  iterbi.marker.chain,  
  organism = "hs",  
  pvalueCutoff = 0.05,  
  min_count = 5  
)
```

### Arguments

iterbi.marker.chain	iterbi.marker.chain dataframe contains all the markers
organism	"hs" for Homo sapiens, or "mm" for Mus musculus
pvalueCutoff	cut off P-value for GO annotations
min_count	Minimal count of genes for GO annotations

### Value

GO annotation dataframe labeled with cluster

---

ora.go.kegg.clusterProfiler

*GO KEGG ORA analysis by clusterProfiler*


---

**Description**

GO KEGG ORA analysis by clusterProfiler

**Usage**

```
ora.go.kegg.clusterProfiler(geneList = markerList, organism = "hs")
```

**Arguments**

geneList	a gene list
organism	"hs" for Homo sapiens, or "mm" for Mus musculus

**Value**

a list with GO and KEGG annotation result (clusterProfiler format)

---

prepare\_expression\_matrix\_heatmap

*Prepare expression matrix for heatmap visualization, estimate the overall expression of marker modules*


---

**Description**

Prepare expression matrix for heatmap visualization, estimate the overall expression of marker modules

**Usage**

```
prepare_expression_matrix_heatmap(
  seuratObj,
  iterbi.marker.chain,
  assay = "RNA",
  slot = "scale.data",
  known_markers = c()
)
```

**Arguments**

seuratObj	A Seurat object
iterbi.marker.chain	iterbi.marker.chain from iterbi
assay	Assay used for prediction
slot	slot used for prediction
known_markers	the genes to include in the heatmap (like known markers)

**Value**

A expression matrix contain the average expression of marker modules and known markers

---

remove_duplicated_GO	<i>remove duplicated GO terms based on overlaps of genes</i>
----------------------	--

---

**Description**

remove duplicated GO terms based on overlaps of genes

**Usage**

```
remove_duplicated_GO(tmp.GO.df, max.overlap = 0.6)
```

**Arguments**

tmp.GO.df	GO annotation dataframe from clusterProfiler package
max.overlap	Maximum overlapped percentage of genes

**Value**

GO annotation dataframe without duplications

---

remove_duplicate_marker_chain	<i>Remove duplicated markers from last level</i>
-------------------------------	--

---

**Description**

Remove duplicated markers from last level

**Usage**

```
remove_duplicate_marker_chain(iterbi.marker.chain)
```

**Arguments**

iterbi.marker.chain	iterbi.marker.chain from iterbi
---------------------	---------------------------------

**Value**

iterbi.marker.chain without duplicated genes

---

rename.iterbi	<i>rename the clusters inside the iterbi result</i>
---------------	---

---

**Description**

rename the clusters inside the iterbi result

**Usage**

```
rename.iterbi(iterbi.result)
```

**Arguments**

iterbi.result    A result file from run.iterbi() function

**Value**

A renamed list. cellMeta contains the final bifurcation for each level marker\_chain contains all the significant markers for each cluster bifurcation contains the bifurcation details (parent, child1, child2)

---

run.iterbi	<i>The main function to perform iteratively bifurcation clustering</i>
------------	--

---

**Description**

The main function to perform iteratively bifurcation clustering

**Usage**

```
run.iterbi(
  seuratObj,
  method = "graph",
  min.marker.num = 100,
  max.level.num = 20,
  min.cell.count = 50,
  res_sets = 30,
  verbose = T
)
```

**Arguments**

seuratObj	A Seurat object
method	The method to perform bifurcation clustering "graph (default), hclust or kmeans"
min.marker.num	Minimal number of markers to confirm a bifurcation
max.level.num	Maximum number of level for bifurcation
min.cell.count	Minimal number of cells to perform bifurcation (must bigger than PC number: 50)
res_sets	The number of resolution for searching
verbose	Print detail processing messages

**Value**

A list. cellMeta contains the preliminary bifurcation for each level marker\_chain contains all the significant markers for each cluster bifurcation contains the bifurcation details (parent, child1, child2)

---

```
subset_seuratObj_bycells
```

*Subset seurat object by cell names*

---

**Description**

Subset seurat object by cell names

**Usage**

```
subset_seuratObj_bycells(seuratObj, tmp.cells)
```

**Arguments**

seuratObj	A Seurat object
tmp.cells	cells for subsetting

**Value**

A subset of seurat object

---

```
write_iterbi_to_seurat
```

*Write iterbi result to Seurat object*

---

**Description**

Write iterbi result to Seurat object

**Usage**

```
write_iterbi_to_seurat(seuratObj, iterbi.result)
```

**Arguments**

seuratObj	A Seurat object
iterbi.result	iterbi.result

**Value**

A Seurat object contains iterbi.result. iterbi will be stored in assay data region of Seurat object ("seuratObj@assays\$iterbi")

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