Expanding polygenic risk scores to include automatic genotype encodings and gene-gene interactions

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Authors

- Trang T. Le
 - © 0000-0003-3737-6565 · ♠ trang1618 · ❤ trang1618

Department of Biostatistics, Epidemiology and Informatics, Institute for Biomedical Informatics, University of Pennsylvania, Philadelphia, PA 19104

- Hoyt Gong
- · Patryk Orzechowski
 - **b** 0000-0003-3578-9809

Department of Biostatistics, Epidemiology and Informatics, Institute for Biomedical Informatics, University of Pennsylvania, Philadelphia, PA 19104

- · Elisabetta Manduchi
 - **1** 0000-0002-4110-3714

Department of Biostatistics, Epidemiology and Informatics, Institute for Biomedical Informatics, University of Pennsylvania, Philadelphia, PA 19104

- · Jason H. Moore[†]

Department of Biostatistics, Epidemiology and Informatics, Institute for Biomedical Informatics, University of Pennsylvania, Philadelphia, PA 19104 · Funded by National Institutes of Health Grant Nos. LM010098, LM012601, Al116794

- These authors contributed equally to this work.
- [†] Direct correspondence to jhmoore@upenn.edu.

Abstract

Polygenic Risk Scores (PRSs) are aggregation of genetic risk factors of specific diseases and have been successfully used to identify subgroups of individuals who are more susceptible to those diseases. While severall studies have focused on identifying the correct genetic variants to include in PRS, most existing statistical models focus on the marginal effect of the variants on the phenotypic outcome but do not account for the effect of gene-gene interactions. Here, we propose a novel calculation of the risk score that expands beyond marginal effect of individual variants on the phenotypic outcome. The Multilocus Risk Score (MRS) method effectively selects alternative genotype encodings and captures epistatic gene-gene interactions by utilizing an efficient implementation of the model-based Multifactor Dimensionality Reduction technique. On a diverse, unbiased collection of datasets, MRS outperforms the standard PRS in the majority of the cases, especially when at least two-way

interactions between genes are present. Our findings suggest that more precise models that take epistatic interactions into account are necessary and will yield greater utility for polygenic risk profiling.

Introduction

As the field of traditional genomics rapidly expands its sequencing technologies and translational abilities, novel applications of genomic data are starting to arise in addressing disease burden. Beginning with the completion of the Human Genome Project in 2003, increased interest in cataloguing genomic data spurred the innovation of massively parallel, chip-based genotyping arrays. Leveraging these technologies, early researchers were able to characterize and categorize gene variants across millions of individuals internationally. In particular, the advent of projects such as the International HapMap Project [1] and the 1000 Genomes Project sought to document haplotype [2] structure (i.e. gene variants) involved in specific diseases of the human genome. As such, the gross information of nucleotide polymorphisms within publicly available databases has rapidly increased in the beginning of the 21st century with the rise in omics sequencing capabilities. This genomic information, coupled with additional high resolution marks for other individual biological variants (e.g. transcripts, epigenetic marks, metabolites) has been touted to further drive precision medicine approaches using genetics.

Complementing the rapid growth in our understanding of gene variants in the human genome was the emergence of using statistical techniques, formalized as genome-wide association studies (GWAS), to identify gene variants associated with common human diseases. From a population perspective, GWA studies have sought to discern genetic connections to various phenotypes by studying genotypic variation at biallelic markers across the human genome [3,4,5]. Such non-candidate driven GWA studies consider gene variations (i.e. SNPs, deletions, intertions, CNVs) to resulting phenotype values to ultimately report allele frequency differences among a case and control group in the form of an odds ratio. This technical revolution in the field of genomic medicine fueled our progressing capabilities to map associations of gene variants with disease on an increasingly granular level to single nucleotide polymorphisms (SNPs).

In tandem with the movement towards precision medicine, the post-GWAS era strives to bring significant population-derived gene variants into individual level metrics actionable in health delivery settings. While GWA studies indeed capture gene variants associated with a phenotype of interest on a population level, translating such results to personalized individual metrics of risk requires additional granularity on aggregating contributions of many gene variants in the form of polygenic risk scores (PRS). Importantly, PRS provides an ability to explain inherited risk for disease in an individual by representing a weighted sum aggregate of risk alleles based on measured loci effect contributions derived from GWAS studies [6,7]. In quantifying the effect of particular combinations of genetic SNP variants towards risk prediction, PRS offers a probabilisitic susceptibility value of an individual to disease. Such genetic risk estimation scores are central to clinical decision-making, serving to reinforce individual health management in heritable disease detection and early prevention of various adult-onset conditions. The utility of PRS scores have been demonstrated in previous studies towards disease risk stratification across leading heritable causes of death in the developed world [10,11,8,9]

For each SNP i of an individual's genome of n possible SNPs for analysis, the PRS score is calculated via a summation across all significant SNPs as

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$$PRS = \sum_{i=1}^n \beta_i \cdot SNP_i$$

where /beta is the weighted risk contribution of the loci gene variant derived from risk score model parameters. Various approaches towards predicting risk of the same disease exist across PRS studies based on the above equation; models may vary according to the β weight according to the specific type of statistical model used to combine risk of individual variations, the n with respect to the specification of genetic variants considered, and the ability of the PRS to generalize to the entire population [6].

Historically, PRS models have previously characterized genomic architechture in a dichotimous division of Mendelian monogenic and polygenic inheritance, in which either one or many gene perturbations give rise to disease phenotypes in an individual, respectively [12,7]. Yet while such classification models arose from former sequencing technology and study design, a more realistic genetic archtechture of common adult-onset disease acknowledges dynamic interactions among a continuum of common low-risk to rare high-risk gene variants to cumultatively drive overall risk of an individual [13]. When only considering rare (minor allele frequency, MAF < 0.5%), high-risk gene variants, such genomic variation only contributes approximately 1-10% towards adult-onset disease incidence [14,7]. Often, more relevant and complete sources of genetic risk is captured from complex smaller interactions of both common (MAF > 5%) and low-frequency (MAF > 0.5% and < 5%) genetic variants each contributing individual, appreciable effects [15,16]. Existing standard multivariate categorical data analysis approaches fall short in handling such enormous possible gene interaction combinations with both linear and nonlinear effects. In this context, more robust and efficient methods towards a polygeneic risk calculation are necessary in capturing the overlap between context-dependent effects of both rare and common alleles on human genetic disorder.

With respect to better understanding the epistasis across an individual's genome, various statistical models have been designed with the intent of capturing high dimensional gene-gene (GxG) interactions. The Multifactor Dimensionality Reduction (MDR) method is one such nonparametric, model-free framework that addresses these challenges and has been extensively applied to detect nonlinear complex GxG interactions associated with individual disease [17]. By isolating a specific pool of genetic factors from all polymorphism and cross-valiating prediction scores averaged across identified high risk multi-locus genotypes, the original MDR approach is able to categorize multi-loci genotypes into two groups of risk based on some threshold value. While created with the primary intention towards GxG interaction detection, the MDR model has additionally demonstrated applicability as a risk score calculation model in constructing PRS scores [18].

Modifications built on top of the MDR framework have been proposed in order to better capture multiple significant epistasis models and potential missed interactions owning to limitations of the original model in the higher dimensions. Model-Based Multifactor Dimensionality Reduction (MB-MDR) was formulated as a flexible GxG detection framework for dichotomous traits and unrelated individuals [19]. Rather than a direct comparison against a threshold level in the original MDR method, MB-MDR merges multi-locus genotypes exhibiting significant High or Low risk levels through association testing and adds an additional 'No evidence of risk' categorization.

In the present work, we aim to reformulate the PRS using the MB-MDR approach to better capture epistatic gene interactions of individual disease risk in a novel Multilocus Risk Score (MRS). In observing prediction accuracy results on an evidence-based simulated dataset from HIBACHI, we demonstrate the improved performance of our epistasis enriched MRS towards characterizing more granular disease etiology.

Multifactor Dimensionality Reduction (MDR) and model-based MDR (MB-MDR)

MDR is a nonparametric method that detects multiple genetic loci associated with a clinical outcome by reducing the dimension of a genotype dataset by pooling multilocus genotypes into high-risk and low-risk groups [17]. Extended from the original MDR algorithm, MB-MDR addresses existing limitations of MDR by increasing detectability of important interactions and decreasing bias by allowing O labels for individuals with no evidence for abberant risk. Several improvements have been made to MB-MDR since it was first introduced in 2009, and its current implementation efficiently and effectively detects multiple sets of significant gene-gene interactions in relation to a trait of interest while efficiently controlling type I error rates.

In addition to the test statistic and P values associated with each genotype combination, another important output of MB-MDR is the HLO matrices generated from the affected- and unaffected-subjects matrices (in the case of binary outcome). Briefly, for each genotype combination, an HLO matrix is a 3 x 3 matrix with each cell containing H (high), L (low) or O (no evidence), indicating risk of an individual whose genotype pairs fall into that cell [20]. For an example binary outcome problem, a genotype combination SNP_1 and SNP_2 will have a χ^2 value, an associated P value and an HLO matrix that looks like

	$SNP_1 = 0$	$SNP_1 = 1$	$SNP_1 = 2$
$SNP_2 = 0$	O	O	O
$SNP_2 = 1$	O	H	L
$SNP_2 = 2$	O	L	H

We discuss in the following subsection how these values were utilized in the formulation of the Multilocus Risk Score (MRS).

[More on the significance of SNP combination vs. significance of H/L/O here...]

Multilocus Risk Score (MRS)

We apply the MB-MDR software [20] v.4.4.1 to simulated datasets of n individuals, p SNPs to obtain the significance level of each combination of SNPs. We let k_d denote the number of significant combinations for a specific model dimension d (e.g. d=2 results in pairs of SNPs). In this study, no significance threshold is imposed at the SNP combination level and, thus, k_d reaches its maximum value of C_p^d (p choose d).

For each subject i ($i=1,2,\cdots,n$), the d-way interaction risk score is calculated as

$$MRS_d(i) = \sum_{i=1}^{k_d} \chi_j^2 \times HLO_j(X_{ij})$$

where χ_j^2 is the test statistic of each genotype combination j from a χ_j^2 test with one degree of freedom for the simulated binary trait, X_{ij} is the j^{th} genotype combinations of subject i and HLO_j represents the j^{th} recoded HLO matrix (1 = High, -1 = Low, 0 = No evidence). In this study, we selected the default multiple testing correction algorithms for an MB-MDR model where the χ_j^2 for each genotype combination is derived from the gammaMAXT algorithm for two tests: H versus LO and L versus HO. As an example, consider a pair $X_{*j} = (SNP_{j_1}, SNP_{j_2})$ with $\chi_j^2 = 8.3$ and corresponding HLO matrix of all O's except an L in the first cell. Then, all subjects' current risks would remain the same except the ones with $SNP_{j_1} = SNP_{j_2} = 0$ where their risks are subtracted by 8.3.

In this study, we consider 1-way and 2-way interactions and thus the combined risk is simply the total of the first two dimensions: $MRS = MRS_1 + MRS_2$. We will examine the combined risk MRS and also its components, MRS1 and MRS2, separately.

Mutual information and information gain

For a given simulated data set, we apply entropy-based methods to measure how much information about the phenotype is due to either marginal effects or the synergistic effects of the variants after subtracting the marginal effects. A dataset's main effect (i.e. marginal effect ME) can be measured as the total of mutual information between each genotype SNP_j and the phenotypic class y based on Shannon's entropy H [21]:

Shannon's entropy
$$H$$
 [21]:
$$ME = \sum_{j}^{k} I(SNP_{j}; y) = \sum_{j}^{k} H(y) - H(y|SNP_{j}).$$

We measure the 2-way interaction information (i.e. degree of synergistic effects of genotypes on the phenotype) of each dataset by summing the pairwise information gain between all pairs of genetic attributes. Specifically, if we let X_j denote the j^{th} genotype combination (SNP_{j_1},SNP_{j_2}) , the total 2-way interaction gain (i.e. synergistic effects SE) is calculated as

2-way interaction gain (i.e. synergistic effects
$$SE$$
) is calculated as
$$SE = \sum_{j}^{k} IG(X_{j}; y) = \sum_{j}^{k} I(SNP_{j_{1}}, SNP_{j_{2}}; y) - I(SNP_{j_{1}}; y) - I(SNP_{j_{2}}; y),$$

where IG measures how much of the phenotypic class y can be explained by the 2-way epistatic interaction within the genotype combination X_j . We refer the reader to Ref. [22] for more details on the calculation of the entropy-based terms.

Simulated data

The primary objective of this data simulation process was to provide a comprehensive set of reproducible and diverse datasets for the current study. Each dataset, containing 1000 samples (rows) and 10 SNPs (columns), was generated in the following manner.

The values in the matrix were drawn using an uniform distribution among four potential options: with 1/2 probability of drawing '1' (representing heterozygous minor alleles aa), 1/4 probability of drawing '2' (representing a major homozygous allele AA) and 1/4 of drawing '0' (representing homozygous major allele AA). [There is a mistake in the sentence above.] The binary endpoint for the data was determined using a recently proposed evolutionary-based method for dataset generation called Heuristic Identification of Biological Architectures for simulating Complex Hierarchical Interactions (HIBACHI) [23]. This method uses Genetic Programming (GP) to build different mathematical and logical models resulting in a binary endpoint, such that the objective function called fitness is maximized. In this study, to arrive at a diverse collection of datasets, we aim to maximize the difference in predictive performance of all pairs of twelve pre-selected classifiers from the extensive library of scikit-learn [24] (Table 1). [but there are only 450 datasets? Shouldn't there be only 10 classifiers? We should also cite Max's PSB paper here] Therefore, we define the fitness function of HIBACHI as the accuracy difference between two classifiers. [Don't we have three objectives in total? We would also need to rephrase the sentence below.] In other words, the first classifier was supposed to perform as well as possible on the data while the second as bad as possible.

Table 1. Selected machine learning methods and their parameters.

Algorithm	Parameters	
GaussianNB	'priors': {None,2}, 'var_smoothing': {1e-9, 1e-7, 1e-5, 1e-3, 1e-1, 1e+1}	
BernoulliNB	'alpha': {1e-3, 1e-2, 1e-1, 1., 10., 100.},'fit_prior': {True, False}	
DecisionTree	'criterion': {"gini", "entropy"}, 'max_depth': [1, 11], 'min_samples_split': [2, 21], 'min_samples_leaf': [1, 21]	
ExtraTrees	'n_estimators': {50,100}, 'criterion': {"gini", "entropy"},'max_features': [0.1, 1], 'min_samples_split': range(2, 21,2),'min_samples_leaf': range(1, 21,2), 'bootstrap': {True, False}	
RandomForest	'n_estimators': {50,100}, 'criterion': {"gini", "entropy"},'max_features': [0.1, 1], 'min_samples_split': range(2, 21,4), 'min_samples_leaf': range(1, 21,5), 'bootstrap': {True, False}	
GradientBoosting	'n_estimators': {50,100},'learning_rate': {1e-3, 1e-2, 1e-1, 0.5, 1}, 'max_depth': {2,4,8}, 'min_samples_split': [2, 21], 'min_samples_leaf': [1, 21], 'subsample': [0.1, 1], 'max_features': [0.1, 1]	
KNeighbors	'n_neighbors': [1, 100], 'weights': {"uniform", "distance"} , 'p': {1, 2}	
LinearSVC	'penalty': {'l1','l2'}, 'C': {1e-4, 1e-3, 1e-2, 1, 1e2}, 'dual'=False	
LogisticRegression	'C': {1e-3, 1e-2, 1e-1, 1., 10.}, 'solver' : {'newton-cg', 'lbfgs', 'liblinear', 'sag'}	
XGBoost	'n_estimators'=100, 'max_depth': [2, 10], 'learning_rate': {1e-3, 1e-2, 1e-1, 0.5, 1.}, 'subsample': [0.05, 1], 'min_child_weight': [1, 20],	
SVC	'kernel'='rbf', 'C' : {1e-4, 1e-2, 1, 1e2}, 'gamma': {0.01, 0.1, 1, 10, 100},	
MLPClassifier	: 'hidden_layer_sizes': {(10),(11),(12),(13),(14),(15),(10,5)},	

At each iteration, five randomly chosen settings of each machine learning method (Table 1) were chosen among all potential options and served as hyperparameters for that method. Each data model was analyzed using 5-fold cross-validation and the best hyperparameter set of the method pair was considered. Each experiment in which one machine learning method was to outperform the other was repeated five times. [more here? Do we select the best out of five times?]

For each simulated dataset, after randomly splitting the entire data in two smaller sets (80% training and 20% holdout), we built the MRS model on training data to obtain the χ^2 coefficients and the HLO matrix, and then we calculated risk score for each individual in the holdout set. We assess the performance of the MRS by comparing the area under the Receiving Operator Characteristic curve (auROC) with that of the standard PRS method on the holdout set.

Manuscript drafting

This manuscript is collaboratively written using the Manubot software which supports open paper writing via GitHub with Markdown [25]. Manubot uses continuous integration to monitor changes and automatically update the manuscript. As a result, the latest version of this manuscript is always available at https://lelaboratoire.github.io/rethink-prs-ms/.

Availability

Detailed simulation and analysis code needed to reproduce the results in this study is available at https://github.com/lelaboratoire/rethink-prs/.

Results

Datasets

[Patryk Orzechowski]

[simulated datasets of n=1000 individuals, p=10 SNPs]

MRS outperforms standard PRS in the majority of simulated datasets

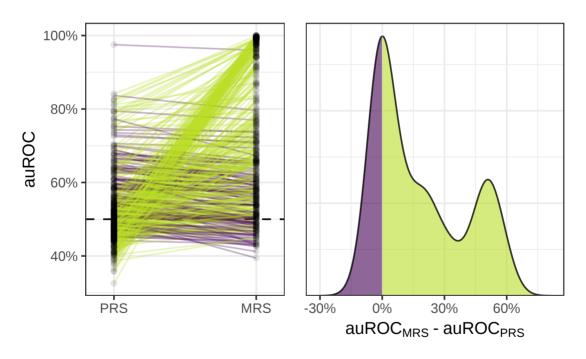


Figure 1: MRS produces improved auROC in the majority (335 green lines) of the 450 simulated datasets (each line represents a dataset). In many datasets, the standard PRS method performs poorly (auROC < 60%) while the new method yields auROC over 90%. This improvement in performance can be seen at the second peak (~50% auROC increase) in the density of the difference between the auROCs from the two methods (right).

In 335 out of 450 simulated datasets, MRS produces higher auROC compared to PRS (green lines, Fig. 1). In 363 datasets where the standard PRS method performs poorly (auROC < 60%), MRS performs particularly well (auROC > 90%) in 102 datasets. When MRS yields smaller auROC, the difference is small (3.3% \pm 2.8%, purple lines/areas). Across all datasets, the improvement of MRS over PRS is significant (P < 10^{-15}) according a Wilcoxon signed rank test. To assess whether this improvement in performance correlates with the amount of interaction effects [contained] in each dataset, in the

following section, we untangled the two components of MRS and test for the correlation between the difference in auROC and two entropy-based measures, main and interaction effect, of each dataset.

Assess improvement in performance

Individually, MRS1 and MRS2 both significantly outperformed the standard PRS method (both P values $< 10^{-15}$) according a Wilcoxon signed rank test. As the amount of main effects increases (Fig. 2 left column), MRS1 increasingly performs better than PRS, which is likely because encodings are inferred (top left). Meanwhile, MRS2's accuracy remain mostly similar to that of PRS (middle left). On the other hand, when the amount of interaction effects increases (Fig. 2 right column), MRS1 performs mostly on par to PRS while MRS2 increasingly performs better than PRS. Combining the gain from both MRS1 and MRS2, MRS's performance progressively increases compared to the standard PRS.

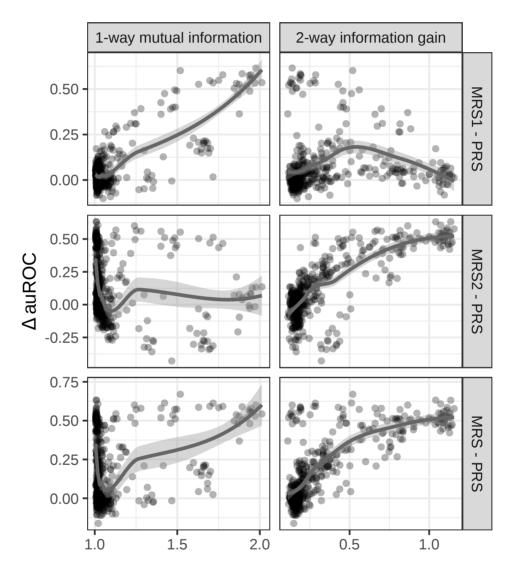


Figure 2: Combining 1-way (MRS1) and 2-way (MRS2) risk scores, MRS shows increasing outperformance to standard PRS as dataset contains more main and interaction effects.

Discussion

PRS holds much promise for development of new precision medicine approaches by identifying high risk individuals who may benefit from prioritized interventions. Nevertheless, one of its current limitation is the model simplicity [7]. As a first step towards addressing this issue and increasing comprehensiveness of risk profiling models, in this study, we developed a new model that enables automatic genotype encodings and takes into account gene-gene interactions. Utilizing the efficient

implementation of MB-MDR, MRS automatically infers the genotype encodings and simultaneously computes the risk matrices of pairs of variants.

We showed that the MRS method outperformed standard PRS in many of the simulated datasets, highlighting the importance of genotype encodings and consideration of epistasis. We further examined the association between this improvement and the amount of two-way epistatic effect induced in the binary phenotypic outcome. Appropriate phenotype encodings are important for improving the accuracy when there is a large amount of main effects of the variants on the phenotypic outcome. Meanwhile, inclusion of epistatic terms significantly increases the accuracy from PRS, especially when two-way interactions are present in the data. Although we only considered up to two-way interactions, it is straightforward to incorporate higher order interactions (e.g. three-way, four-way) into MRS. However, preliminary analyses on the simulated datasets did not show much improvement from the current MRS (results not shown). We also recommend estimating the computational expense prior to implementing high order interactions, especially for larger datasets encountered in practice.

We acknowledge a few limitations of the current study. First, MRS has not been applied to real-world data. Although we compensated the lack of real data with a diverse, unbiased set of simulated datasets, a future study analyzing will prove beneficial to quantify the new MRS model's utility in practice. Second, accounting for epistasis, in principle, is largely more computationally expensive compared to investigating solely main effects. Therefore, even with fast and efficient softwares, preselecting the variants (e.g. based on specific pathways or prior knowledge) will prove beneficial for accurate MRS computing when analyzing datasets containing a larger number of variants. However, we hope the promising preliminary results will open the doors to future approaches that encompass main and interaction effects and improve scalability.

Finally, we caution that a risk score model should be evaluated based on not only sensitivity and specificity but also with respect to potential clinical efficacy, and any genetic risk should be interpreted in aggregate with other risk factors. Future works focusing on gene-environment interactions with time-dependent risk factors will be crucial in order to communicate risk properly for preventive interventions.

In conclusion, MRS enhances the predictive capacity of current risk profiling model for complex diseases with polygenic architectures. While there is much work left to do to improve the personal and clinical utility of general risk profiling framework, we highlight that more comprehensive models that infer proper genotype encodings and account for epistatic effects greatly improve the prediction efficiency and affords new opportunities for more accurate clinical prevention.

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