

Advance XCMS Data Processing

H. Paul Benton

Reminder of what we're trying to do

Peak Detection

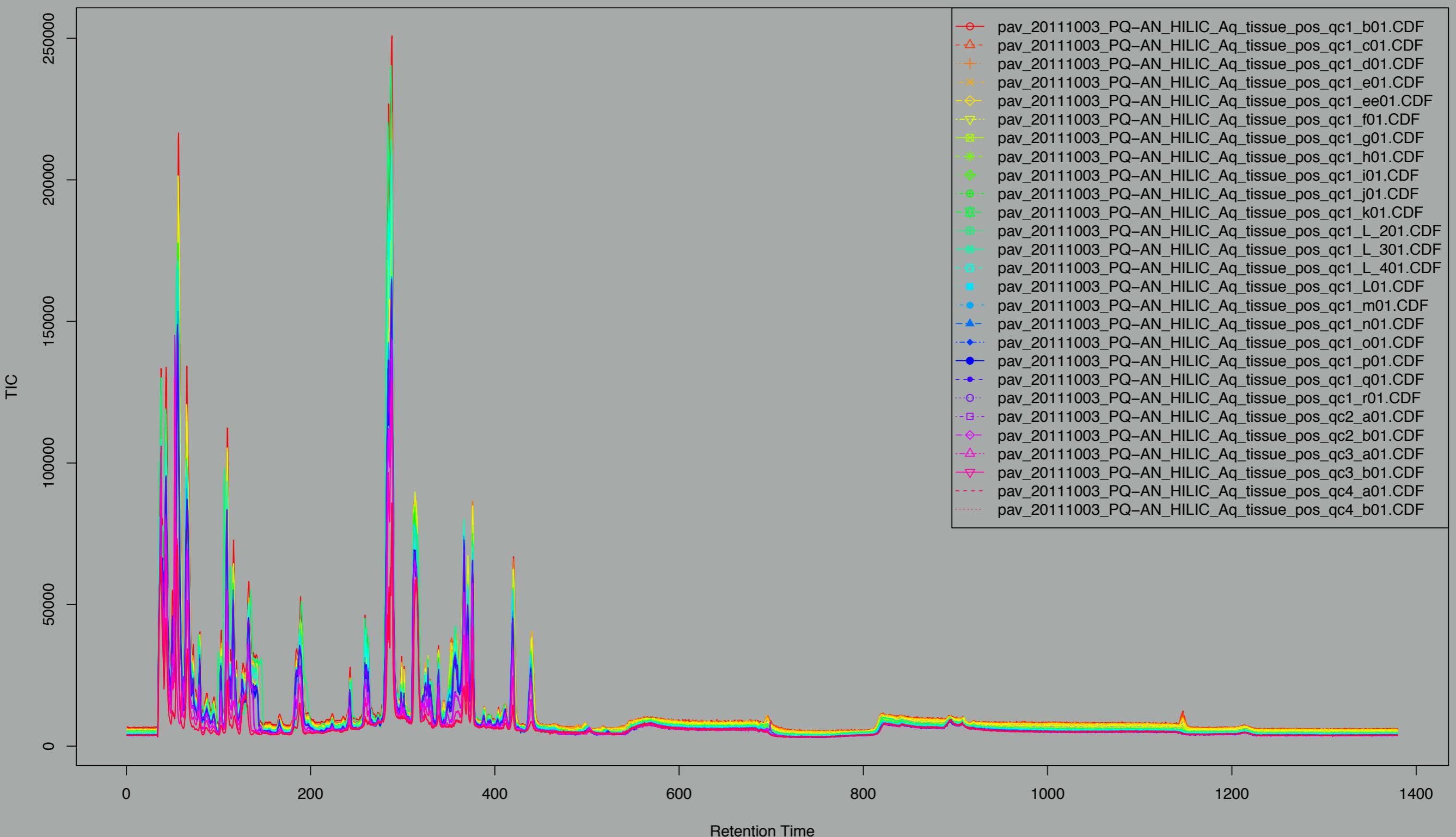


Grouping
Groups similar Peaks
across replicates

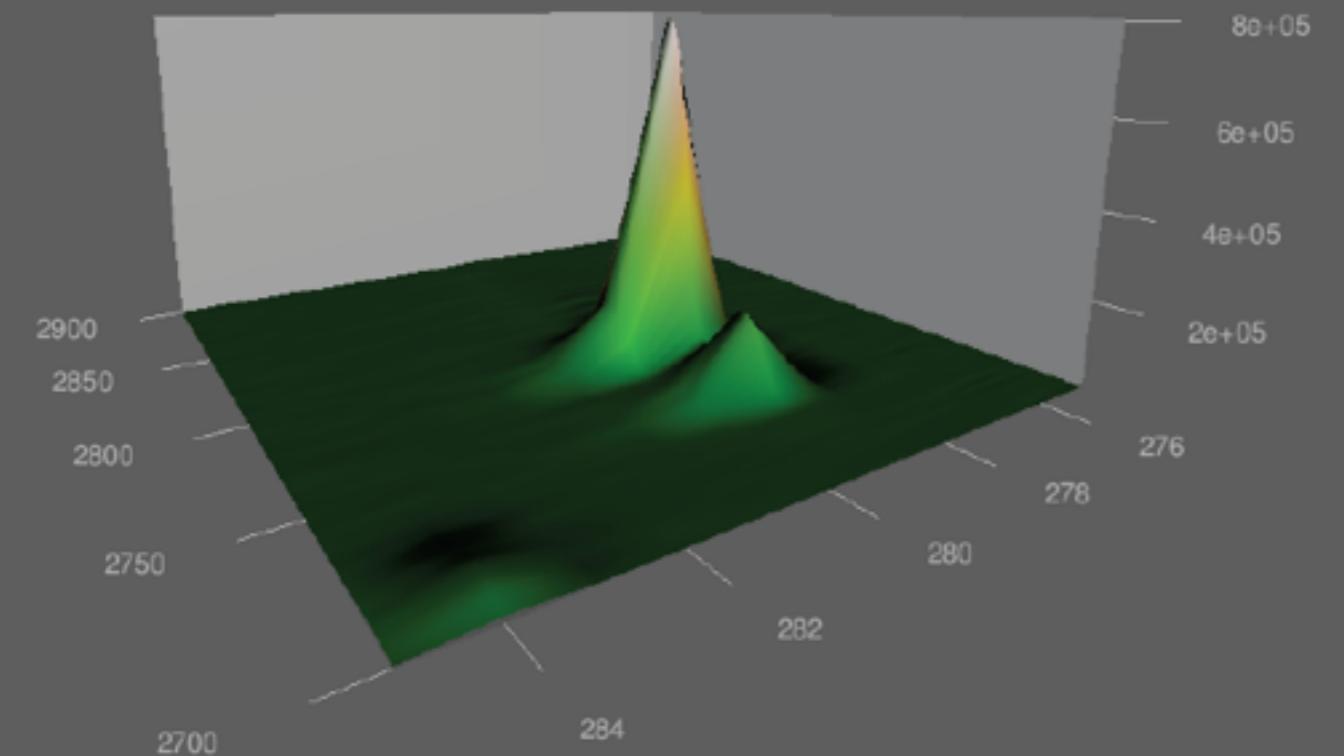
Retention Time
Alignment

Statistical Analysis
of Classes

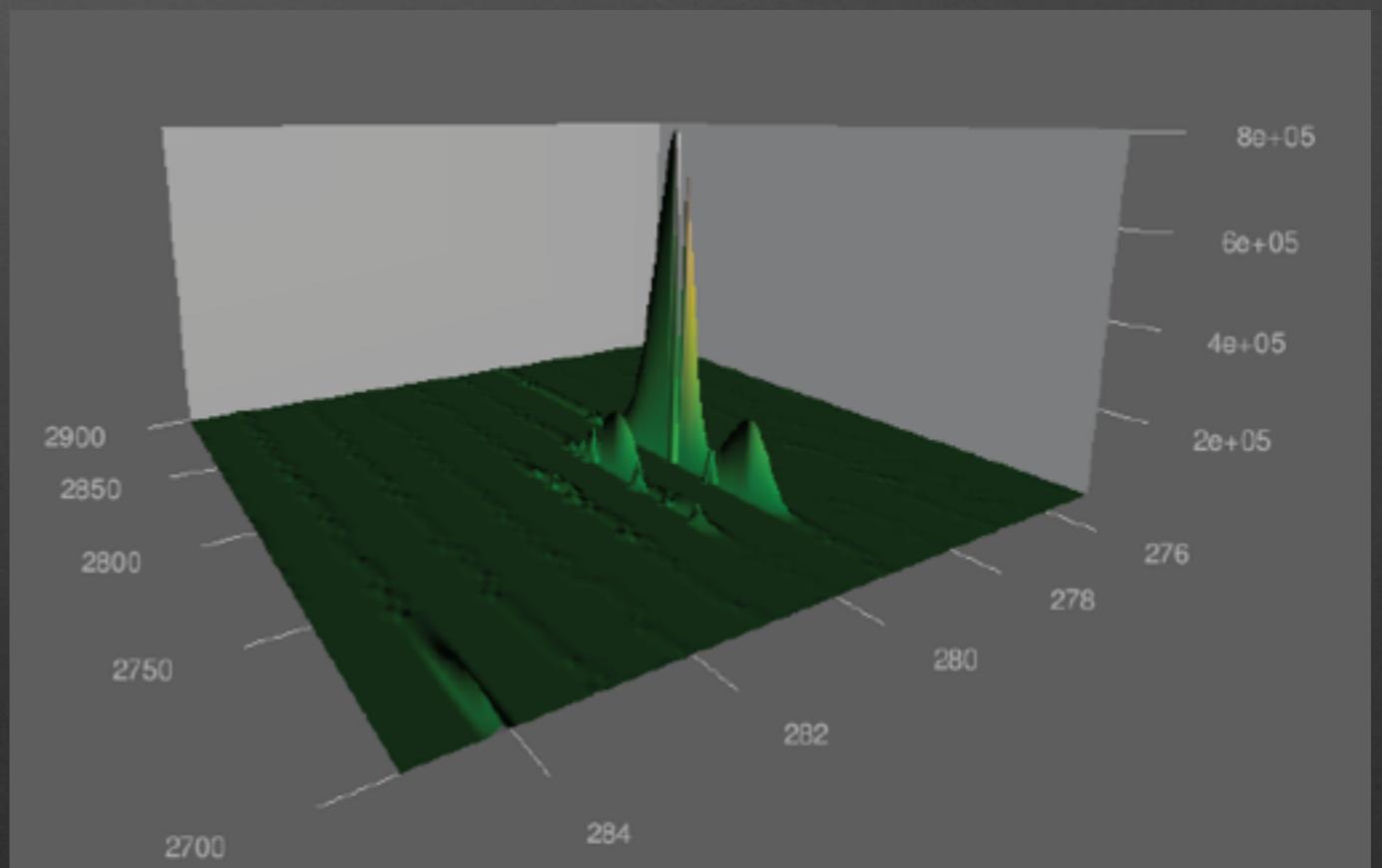
Total Ion Chromatograms



Parameters Matter !

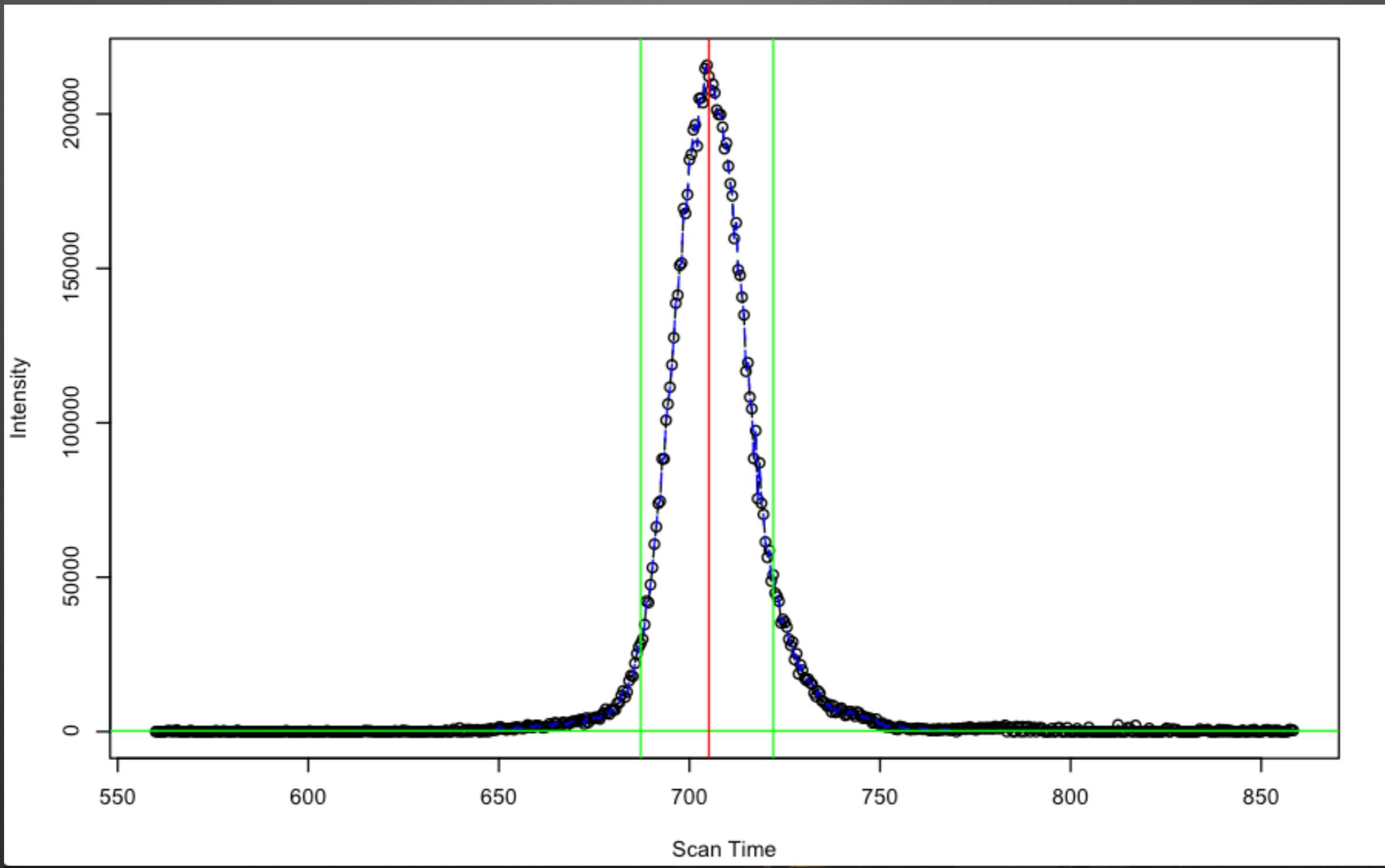


step = 1

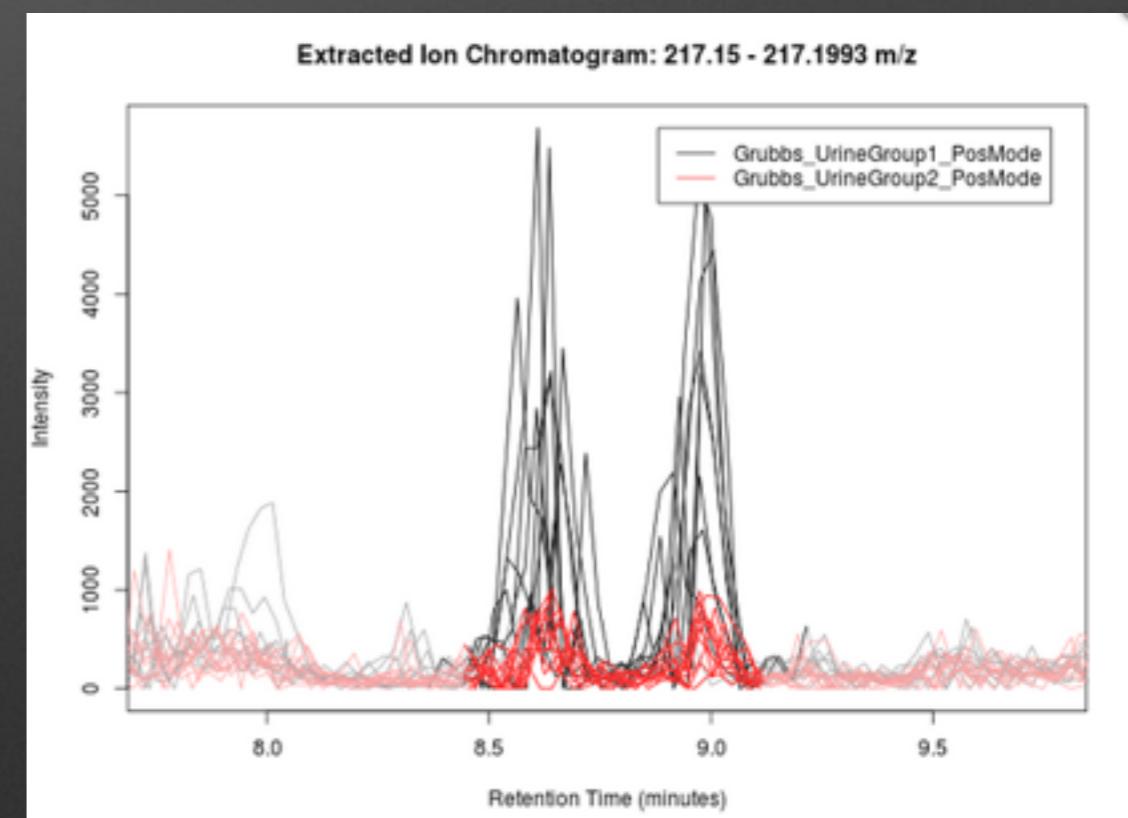
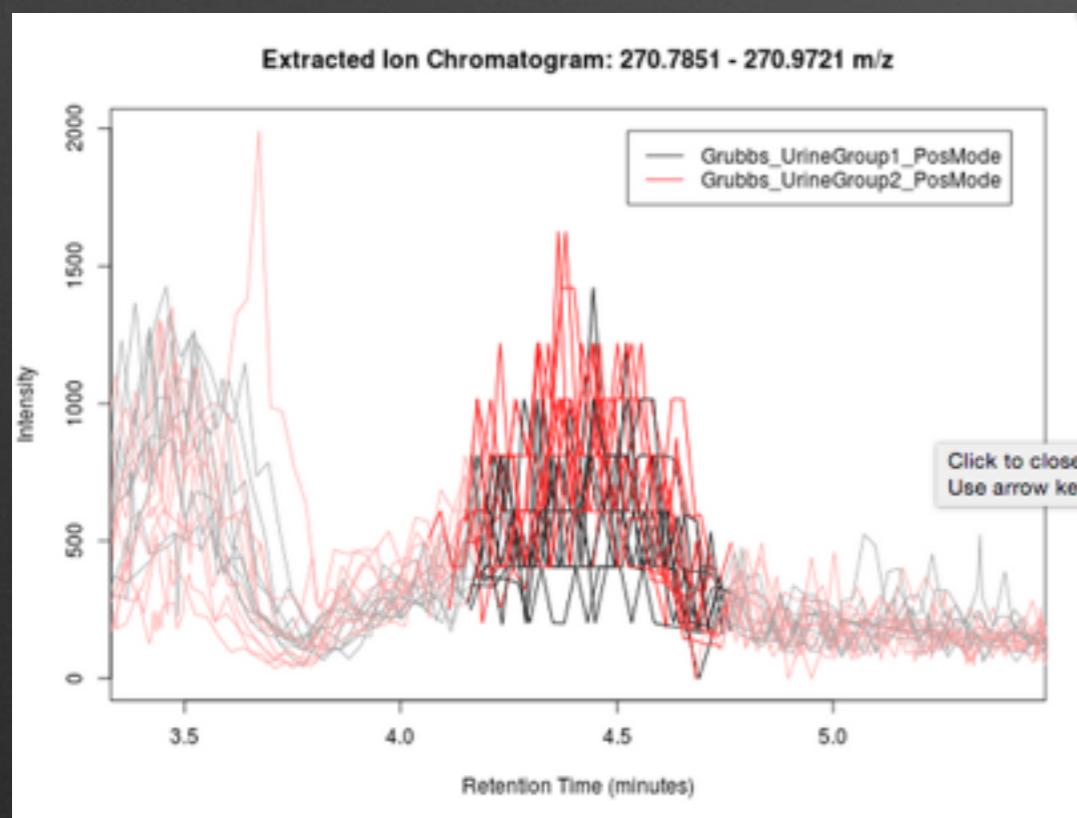
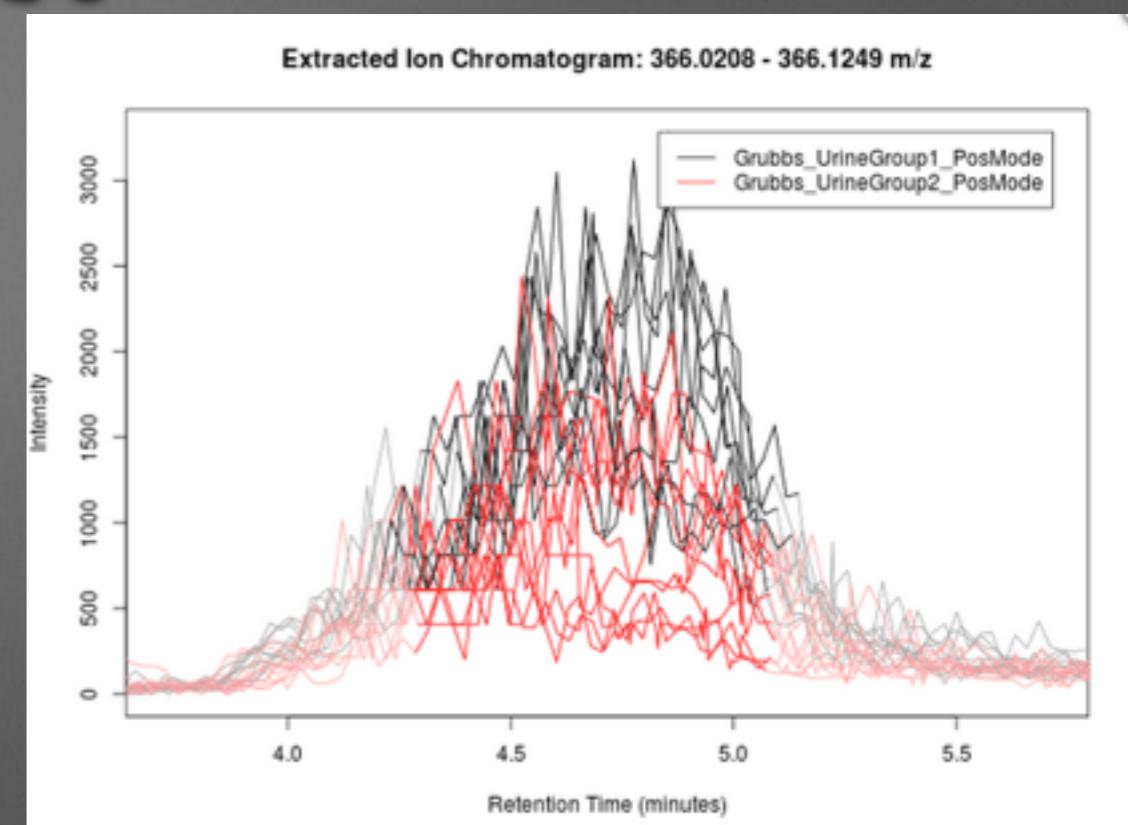
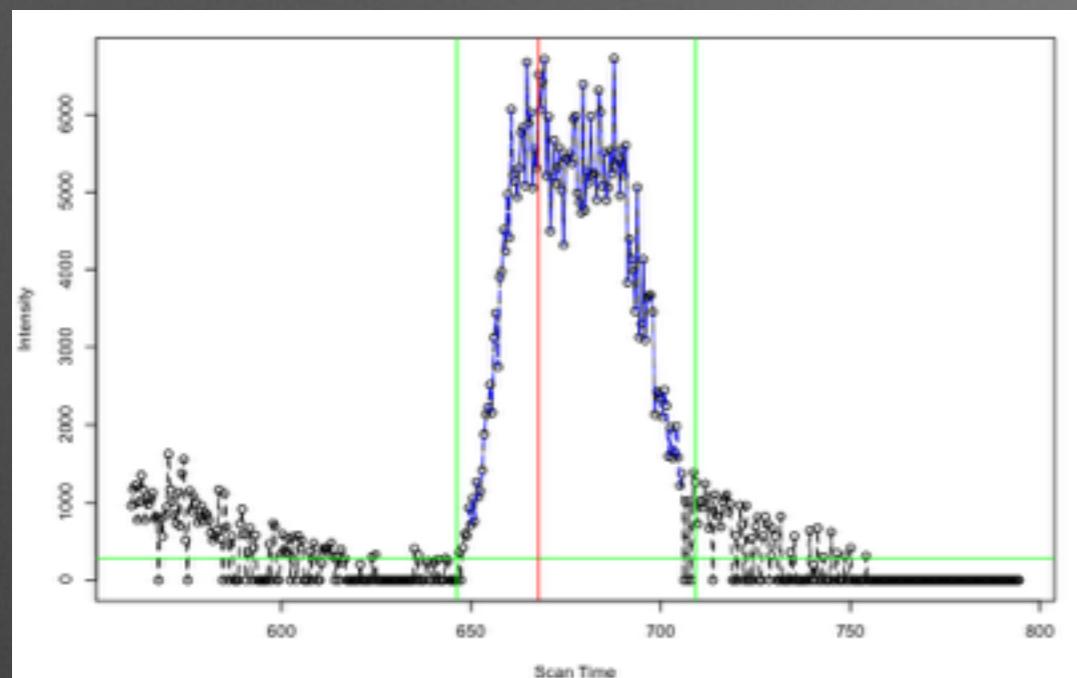


step = 0.1

Peak Detection... Easy !



Right?



Peak detection

- Data comes in two types in MS : centroid & profile
- Generally high resolution or low resolution ~ high mass accuracy or low mass accuracy
- Two main choices in XCMS
 - MatchedFilter - profile low res
 - CentWave - centroid high res

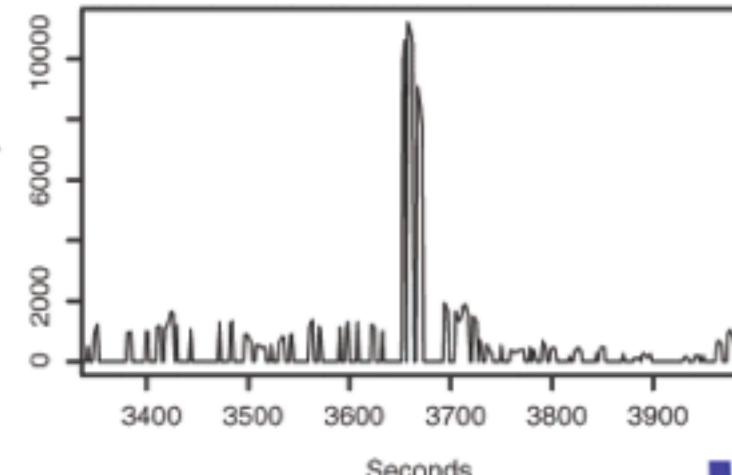
Hat fitting

- Different hat for different heads (& faces apparently)
- A hat has to fit well so it must be sized



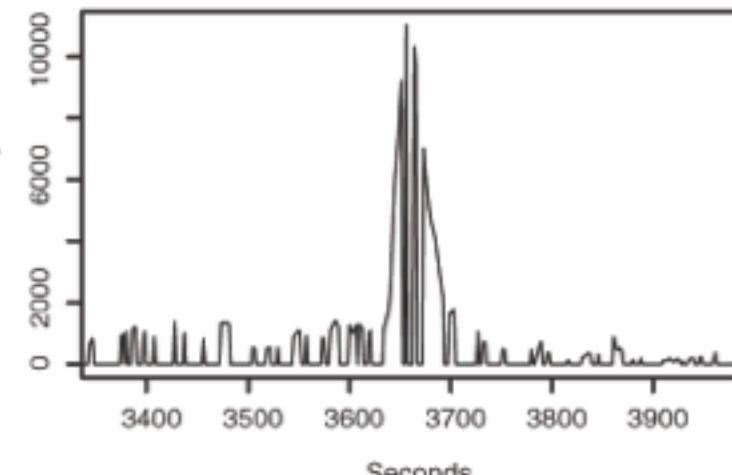
MatchedFilter

Extracted Ion Chromatogram: 268.1 m/z

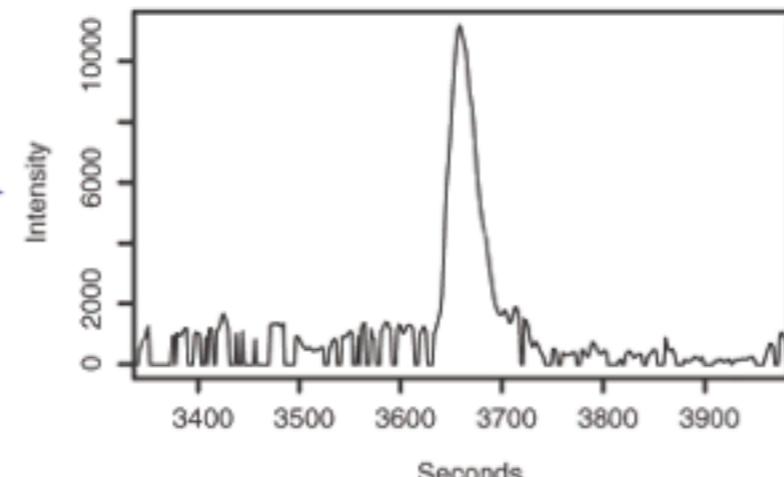


Combination

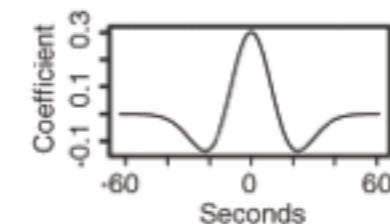
Extracted Ion Chromatogram: 268.2 m/z



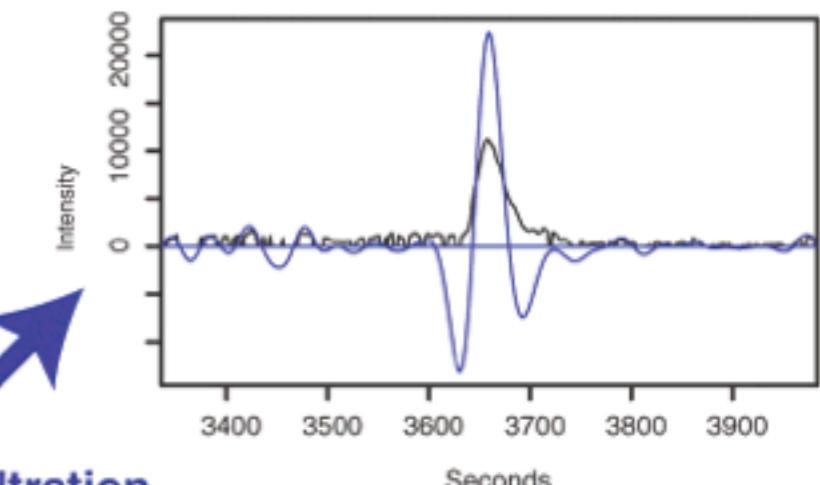
Combined Chromatogram: 268.1 – 268.2 m/z



Filter Function



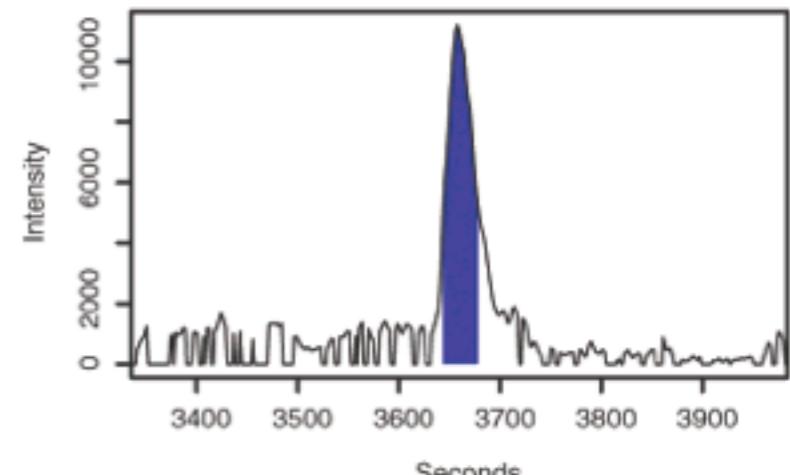
Filtered with Second Derivative Gaussian



Filtration

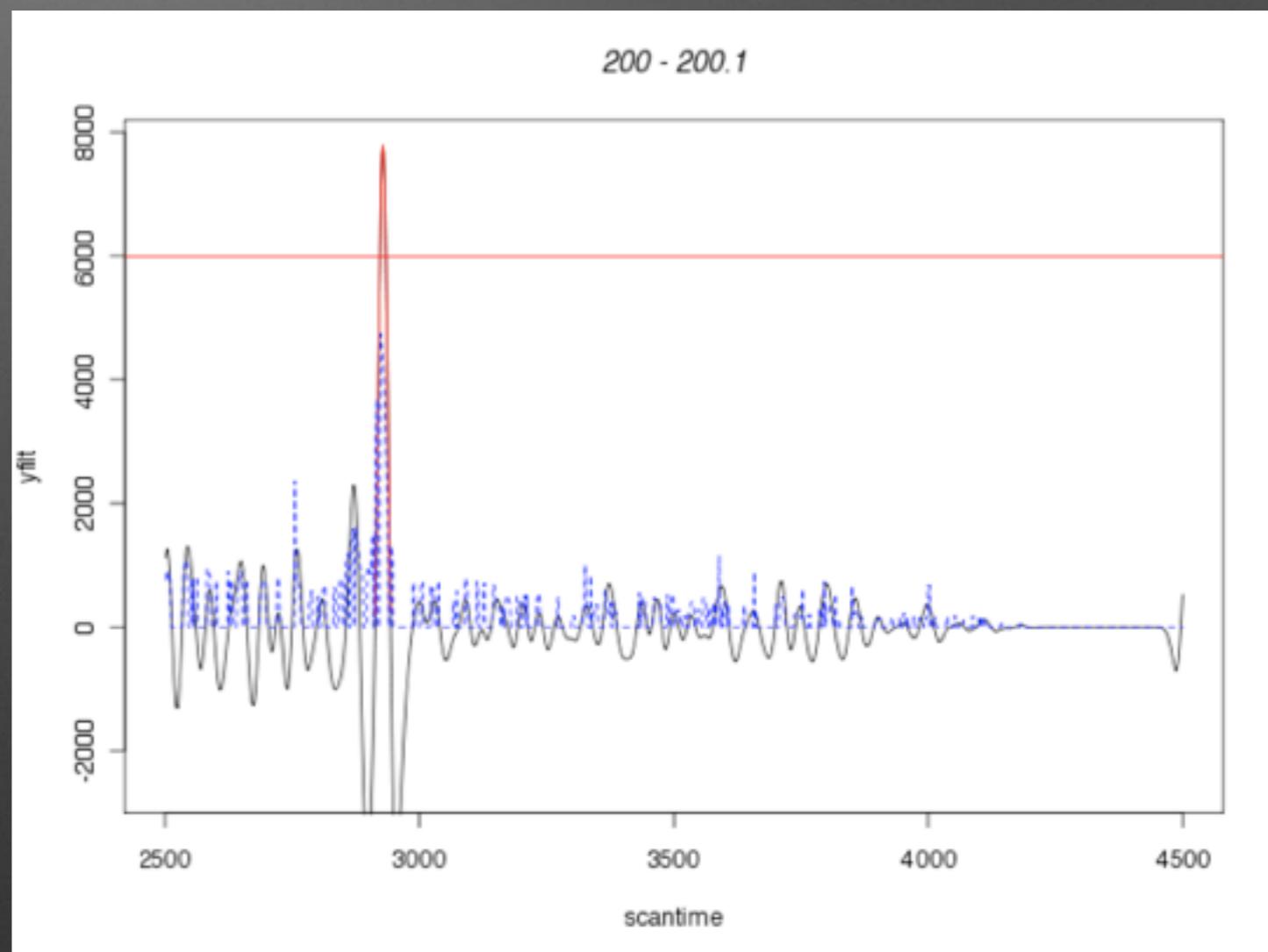
Integration

Peak Integrated between Zero Crossing Points



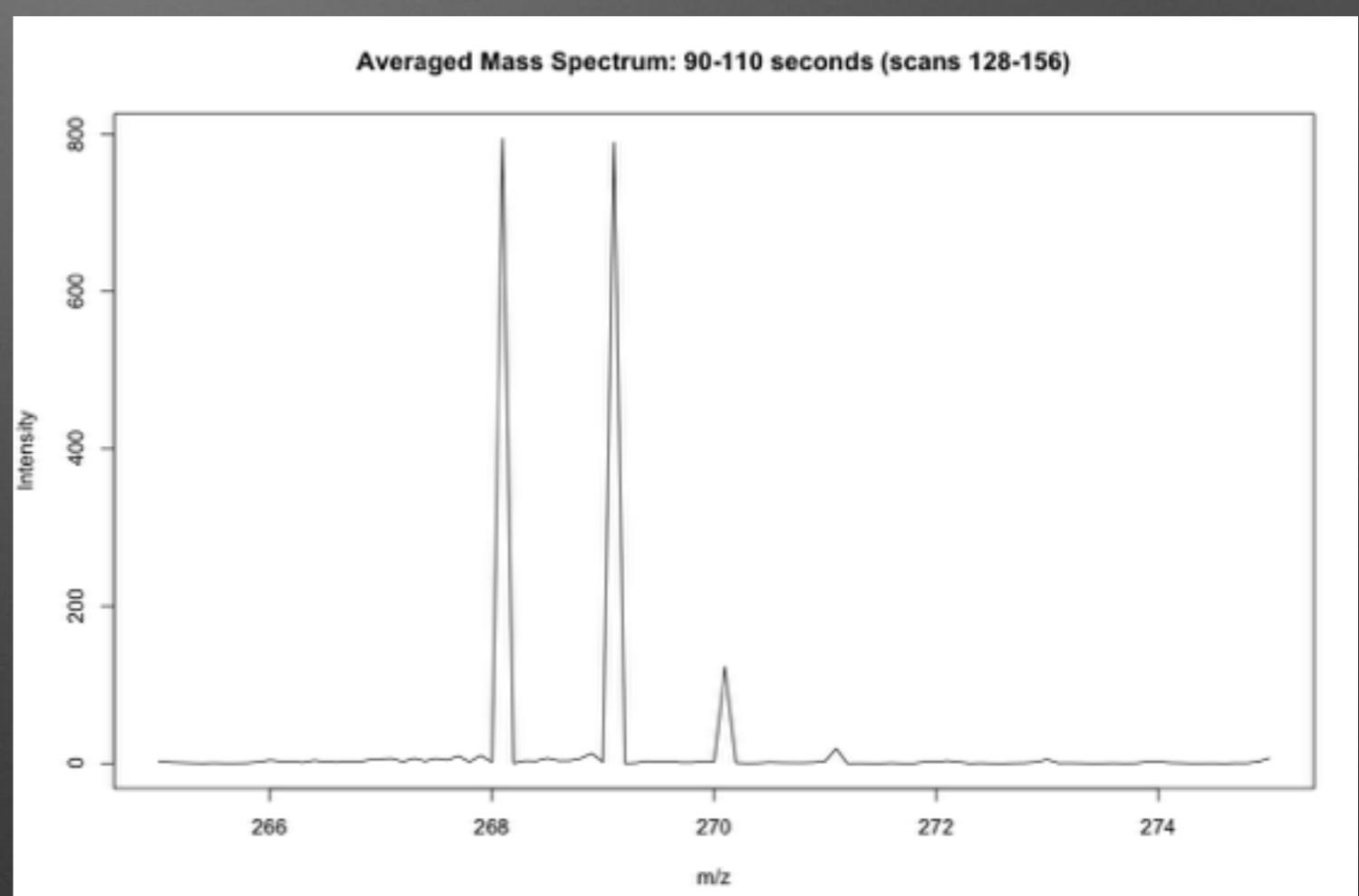
MatchedFilter : wearing hats

- Bin of each X m/z
- Apply a filter function to the data
- Any peak above a s/n ratio is selected
- Peak is selected to filter baseline



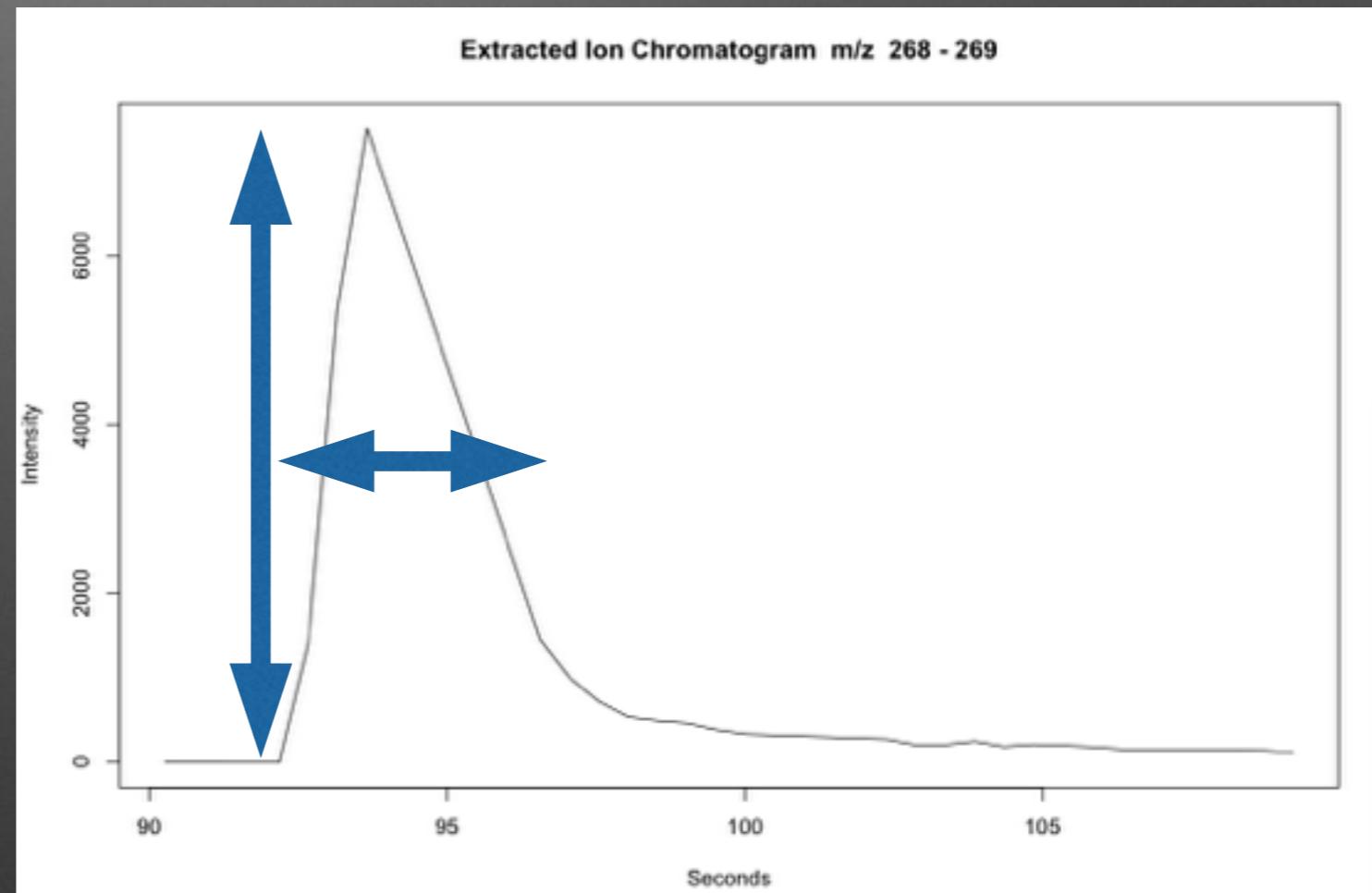
Matched Filter : Sizing the hats

- Profile Data - profMethod
 - binlinBase - profile data
 - bin - centroid data
- profStep - Bin Size
- Peak Width - FWHM



Matched Filter : Sizing the hats

- Profile Data - `profMethod`
 - `binlinBase` - profile data
 - `bin` - centroid data
- `profStep` - Bin Size
- Peak Width - FWHM

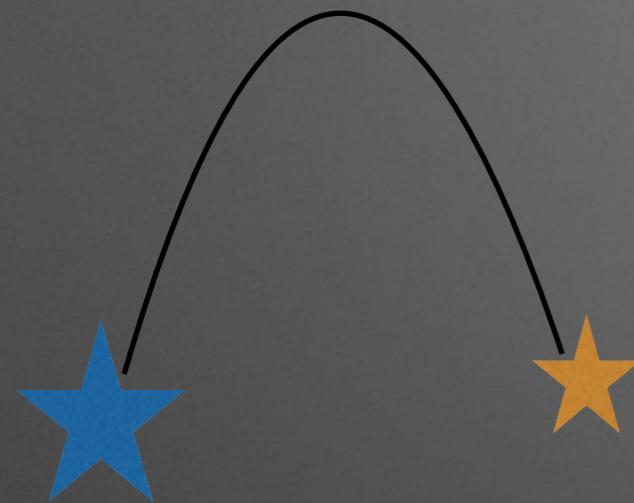


Missiles are like ions!

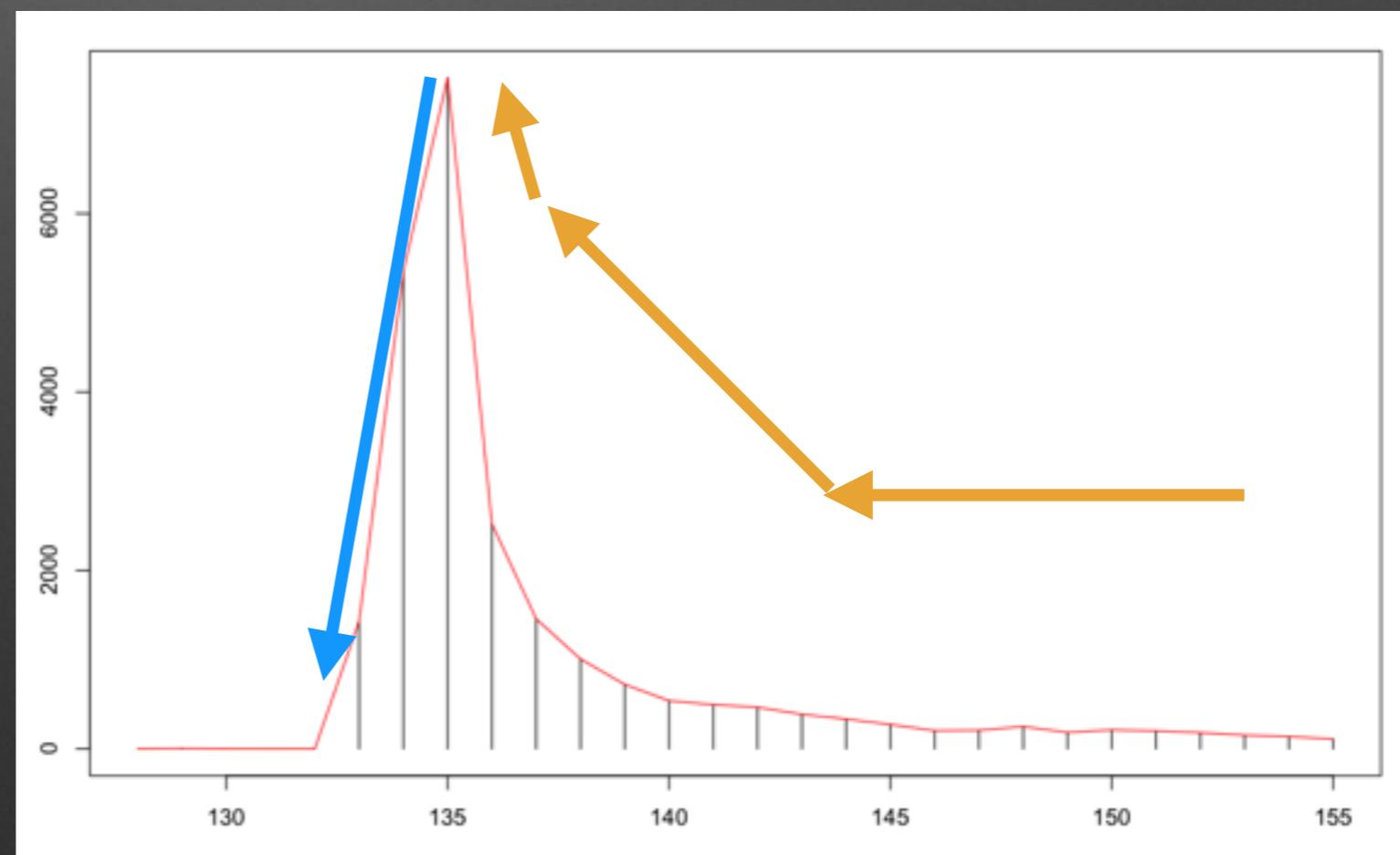


Kalman filtering

Tracking Missiles is
like tracking LC-MS traces



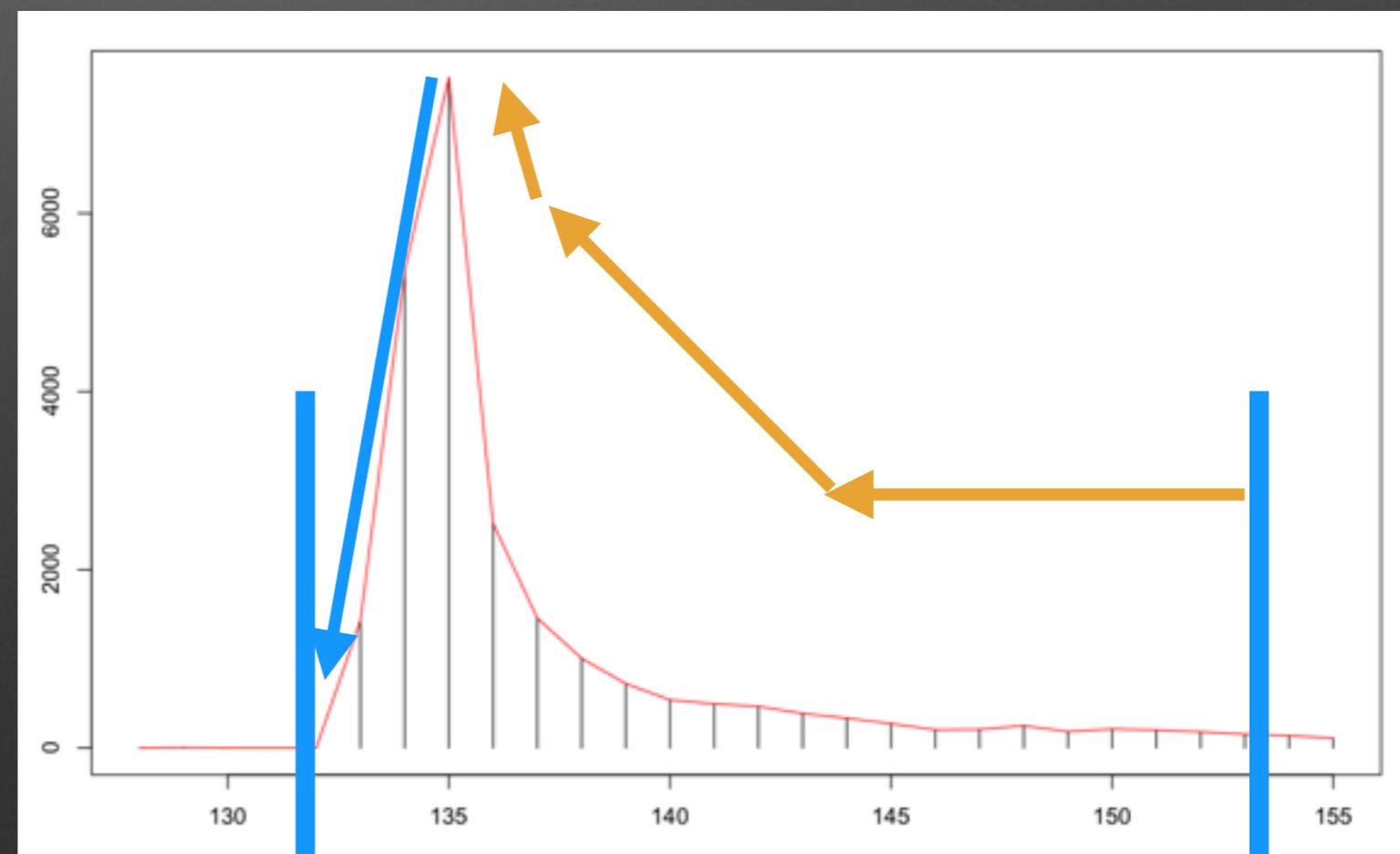
Trace backward along the trace
This will define the area of the 'bin'



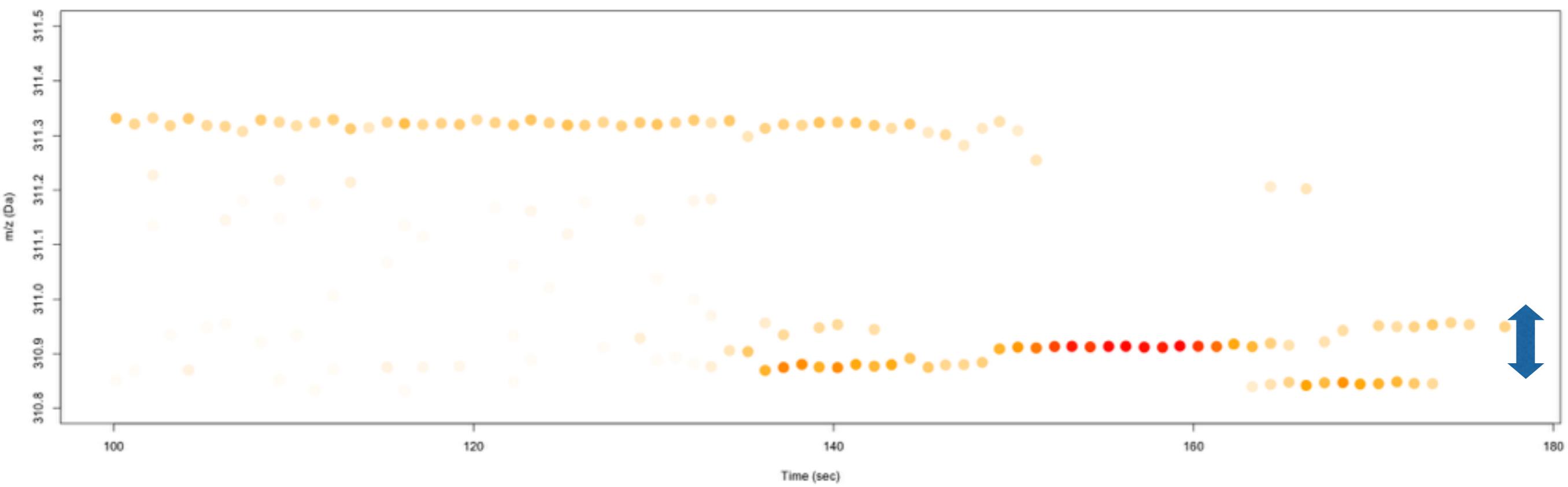
Tracking Missiles is
like tracking LC-MS traces



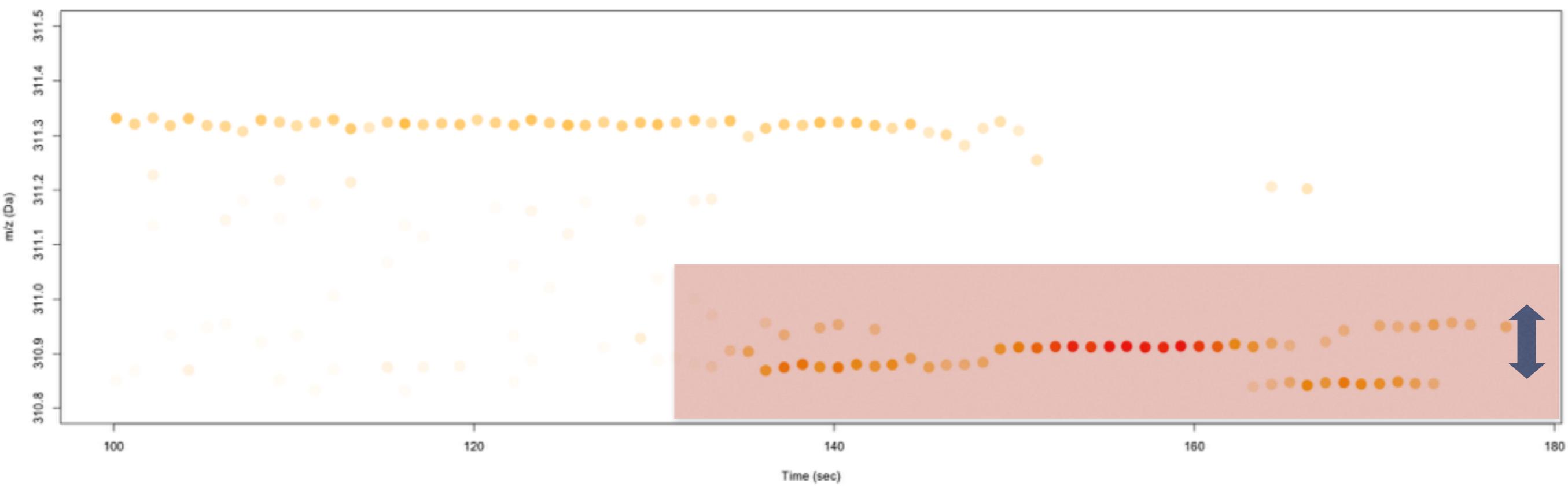
Trace backward along the trace
This will define the area of the 'bin'



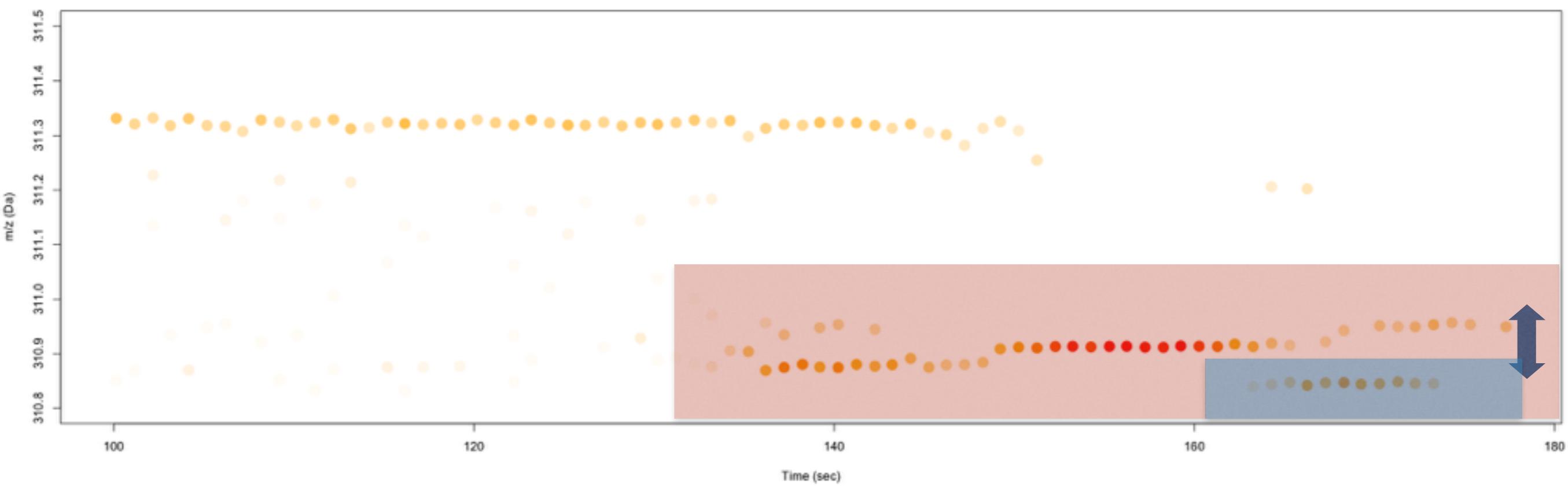
CentWave - ppm



CentWave - ppm

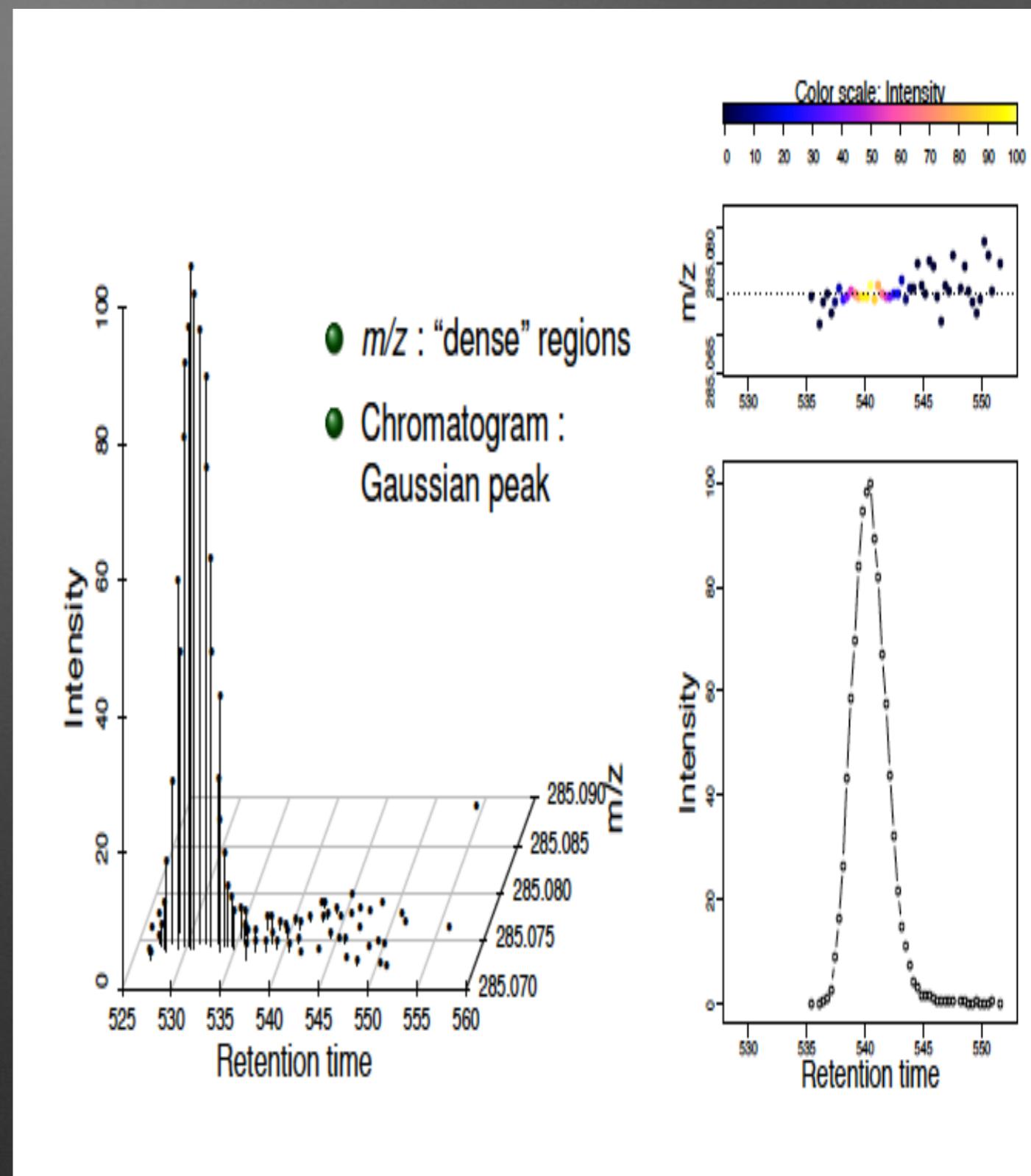


CentWave - ppm

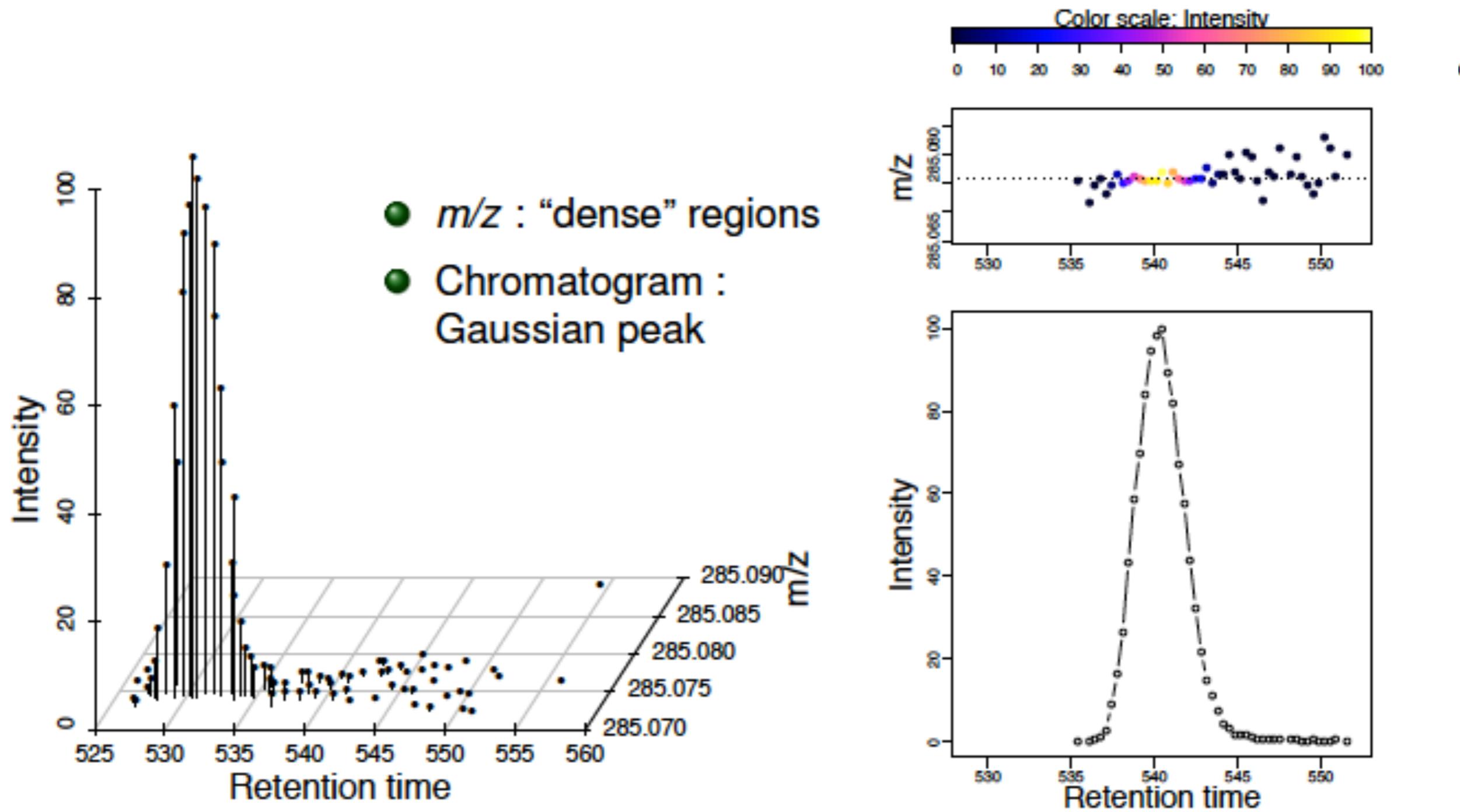


CentWave 2

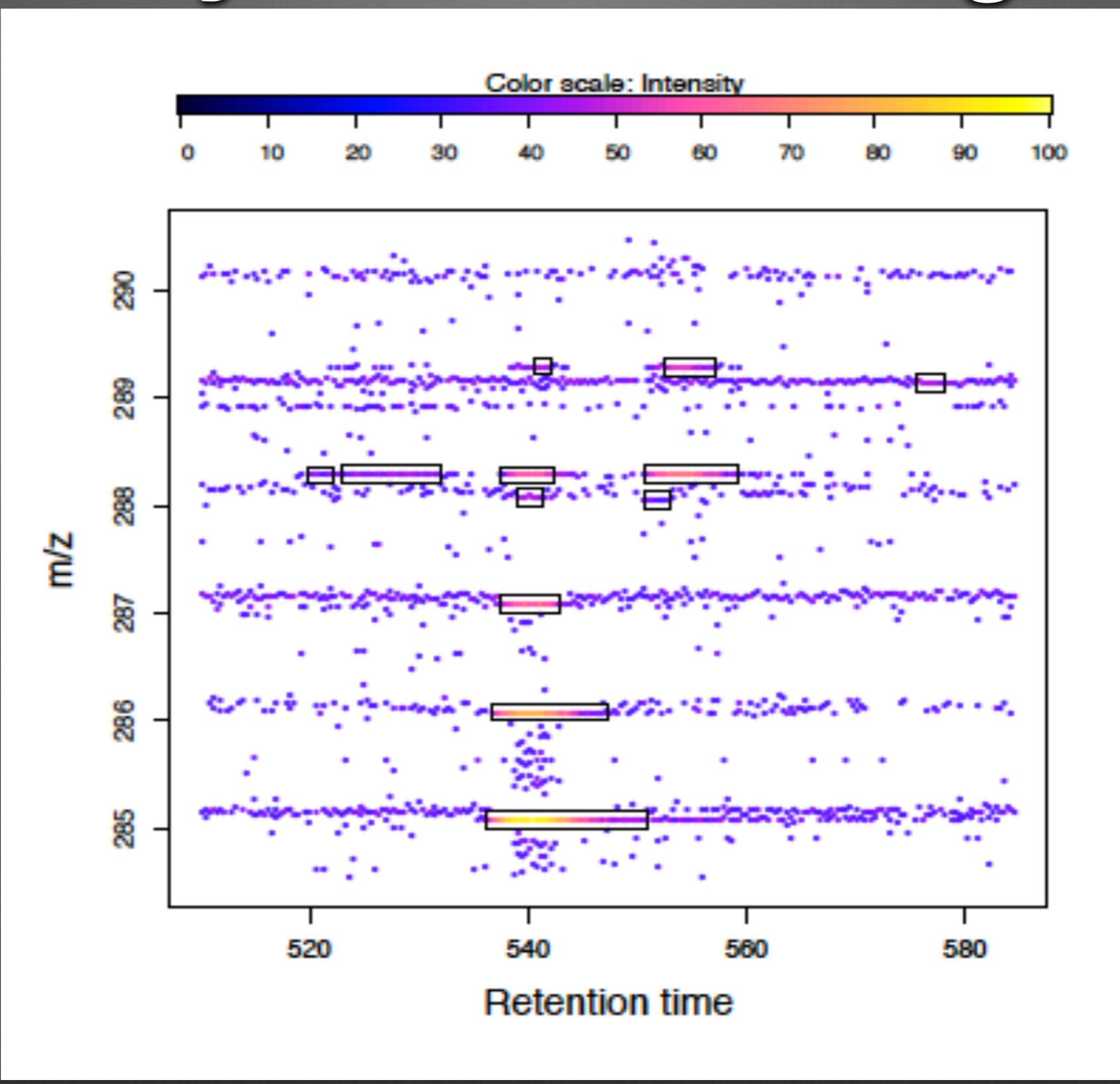
- **Regions of Interest (ROI)**
 - Found using Kalman Filter
 - Often over estimates



CentWave 2

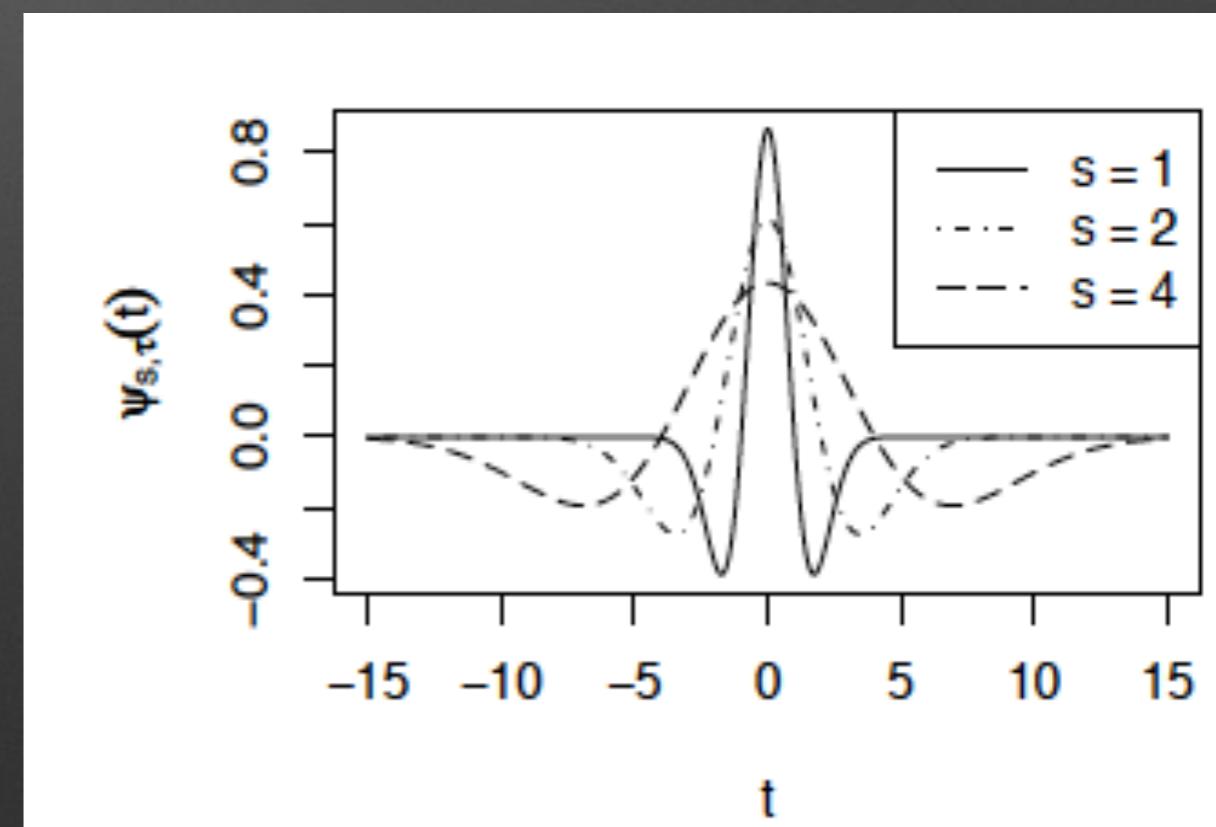
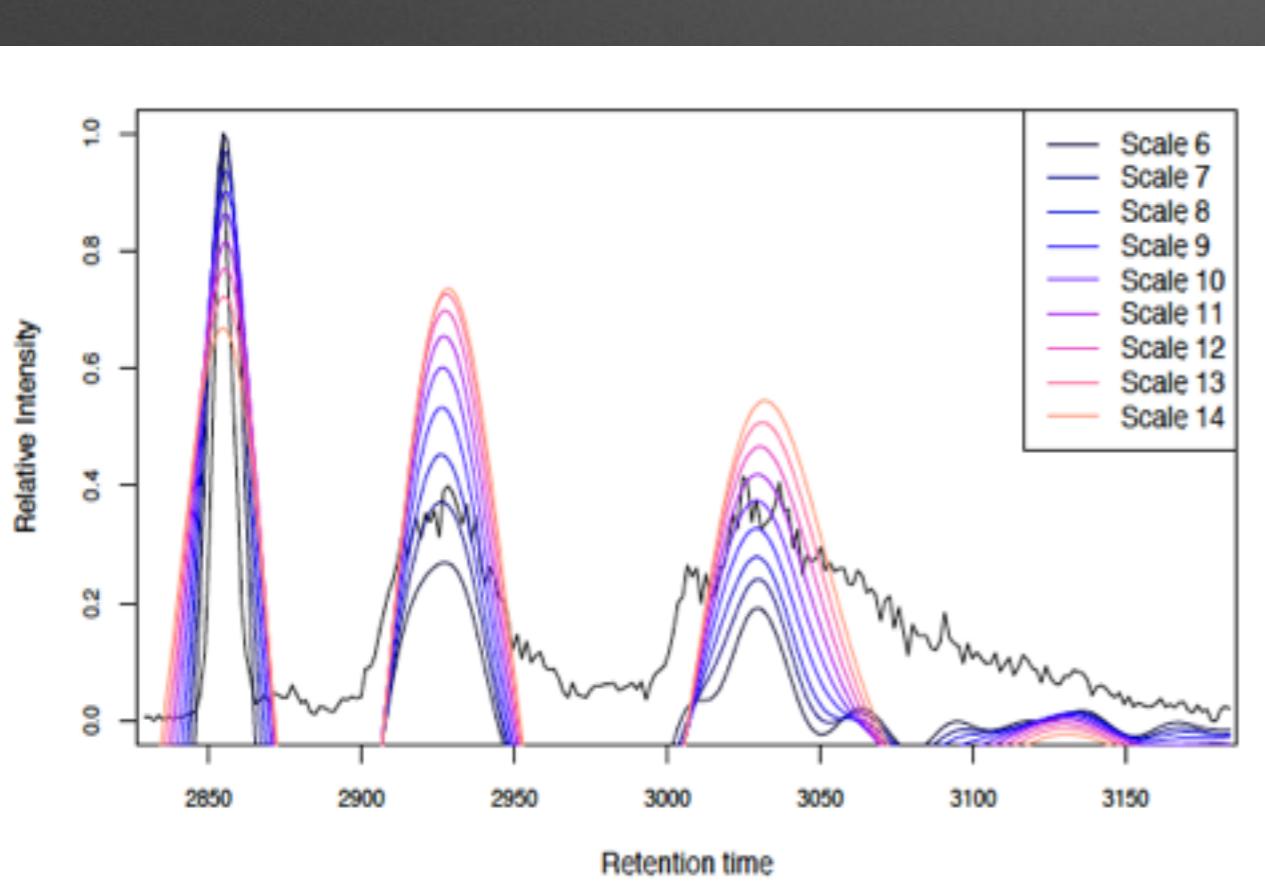


Dynamic Binning



Find and integrate the peak

- Wavelet formation are then used over the ROIs to find the peak
- Several passes of wavelets are used until the correction ‘fit’ is found (mexican hat wavelets)



General Principles

Peak Detection



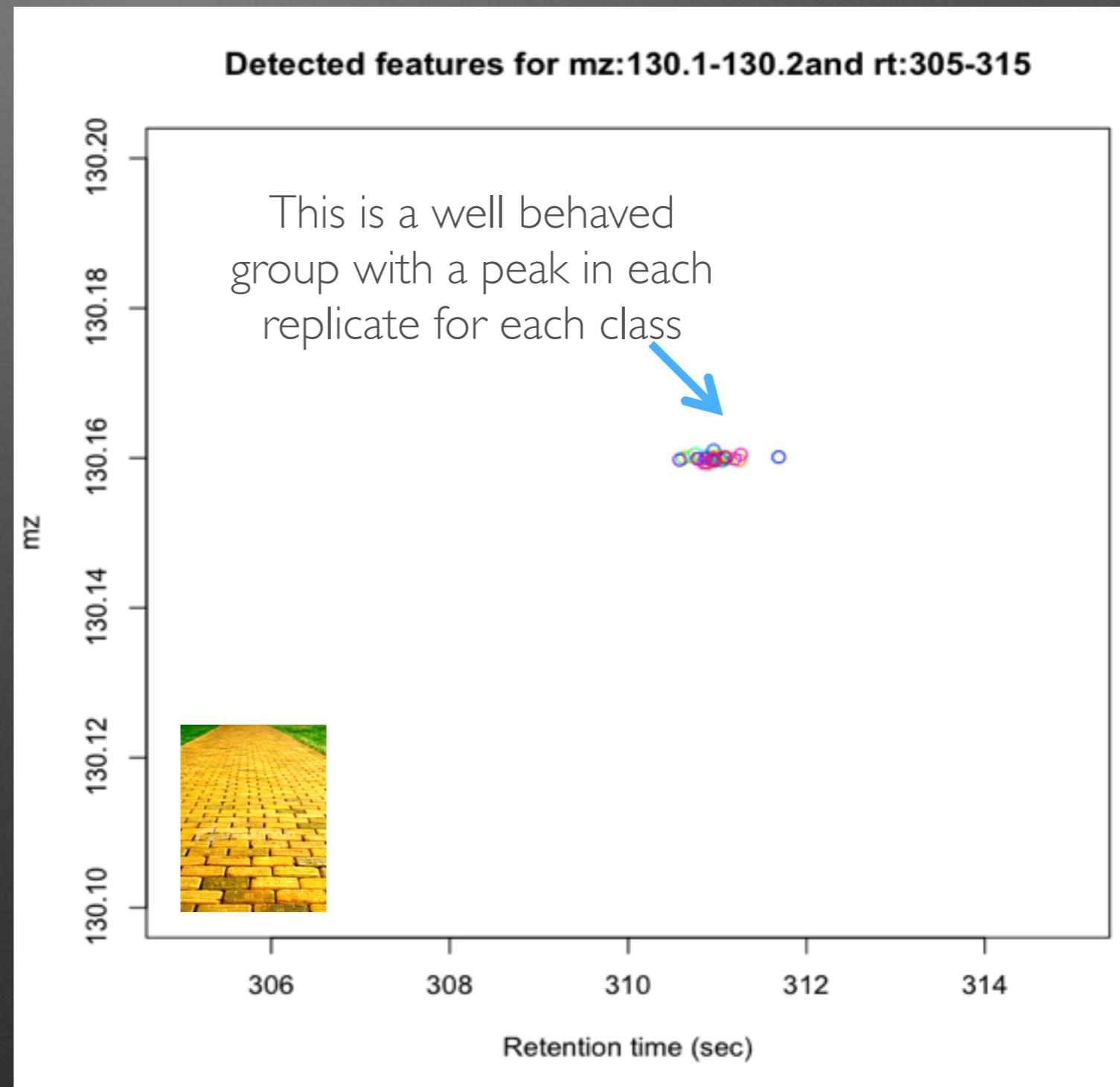
Grouping
Groups similar Peaks
across replicates

Retention Time
Alignment

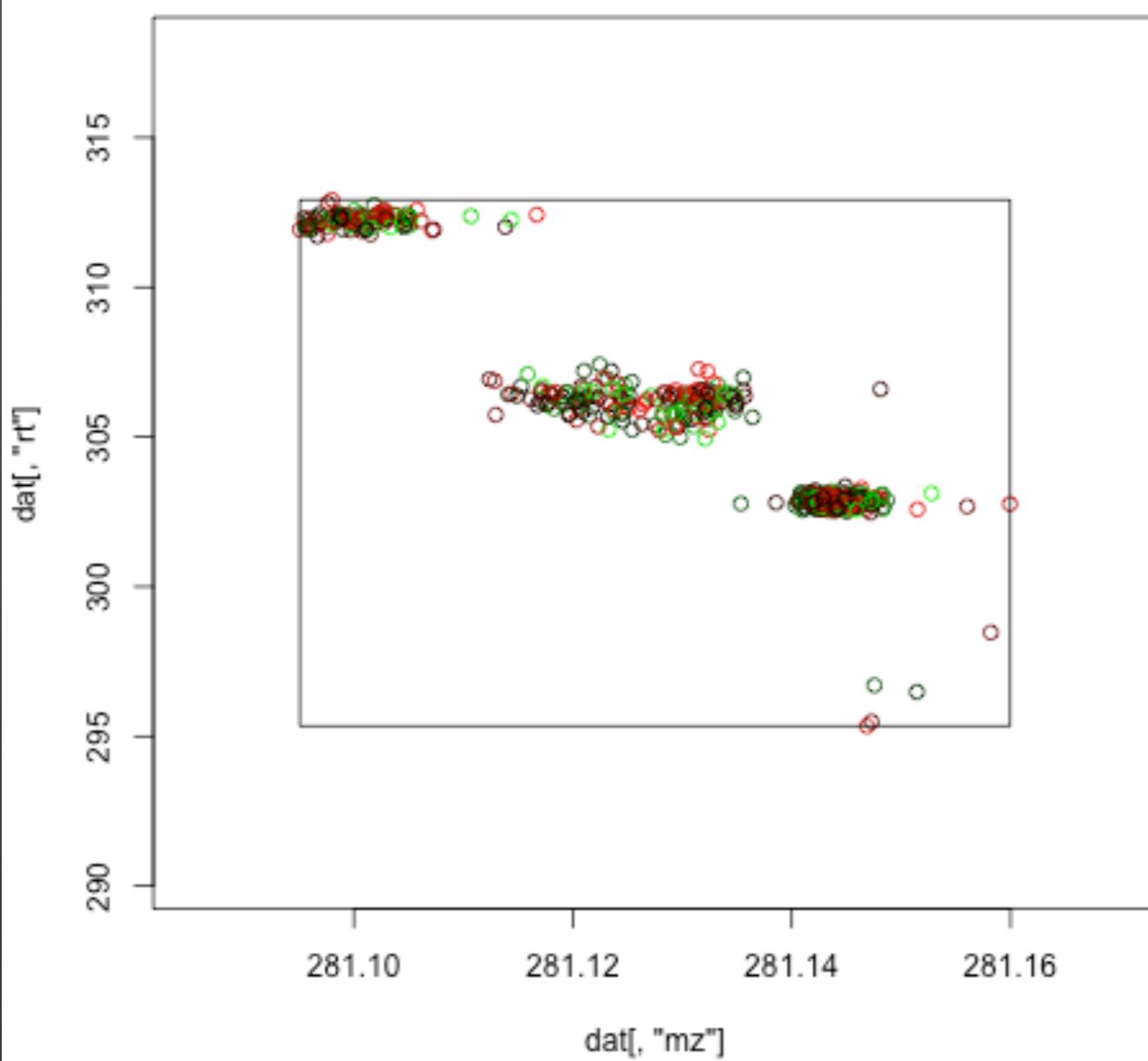
Statistical Analysis
of Classes

Grouping

- First time using all of the files
- Looks for closely clustered/dense peaks across multiple files.
- Once peaks are grouped they're known as a group or feature



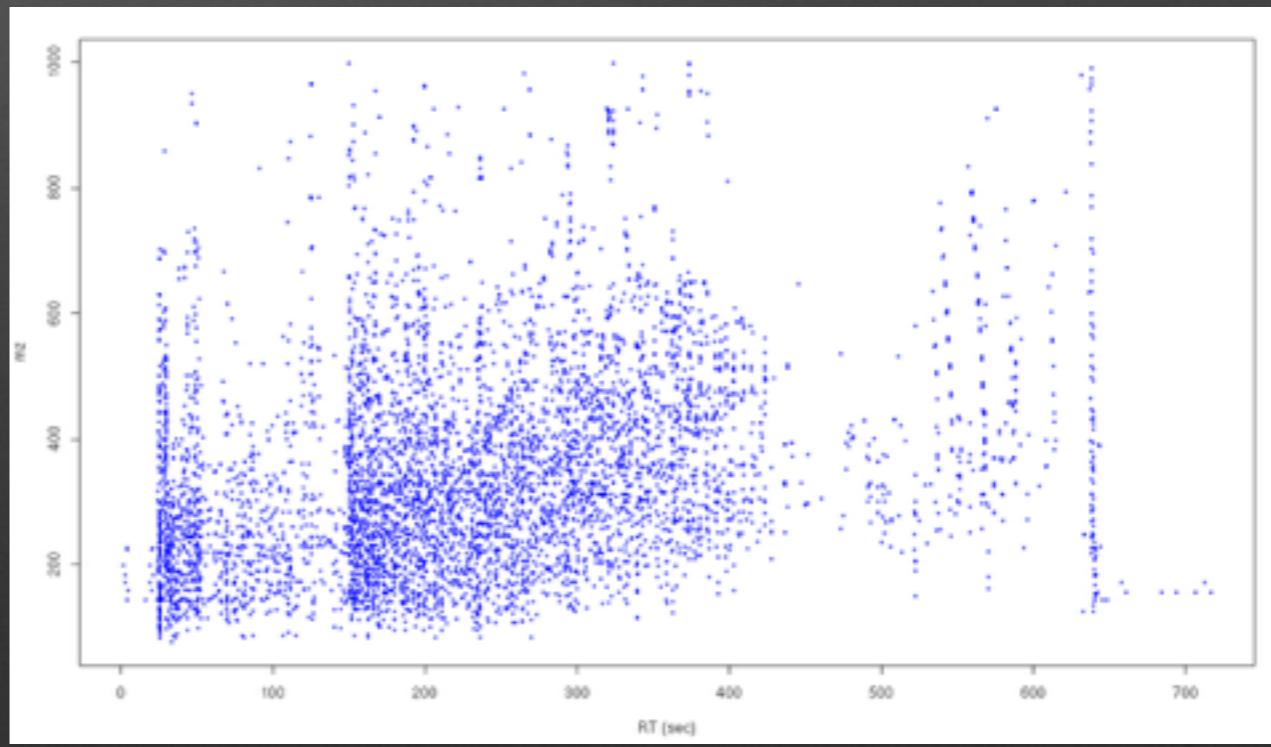
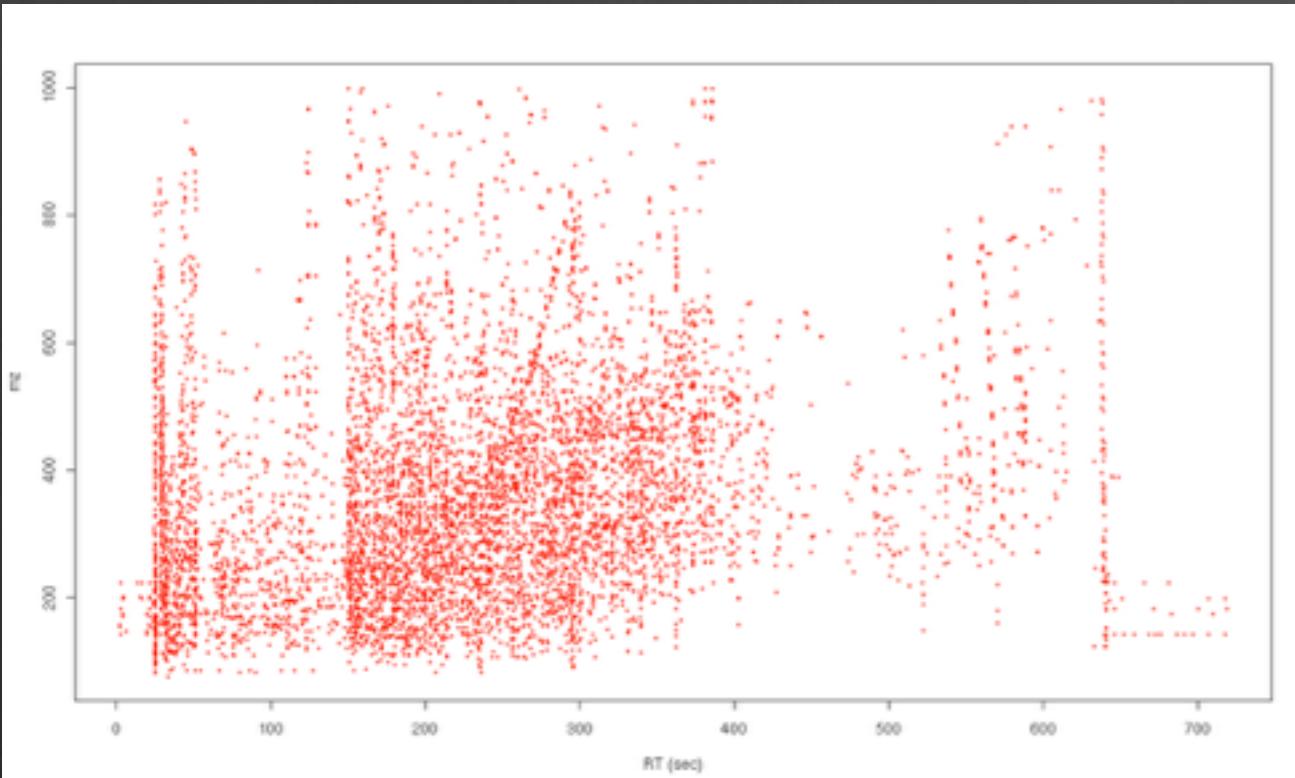
Bupropion@ACN RT:303.140162903349 sec



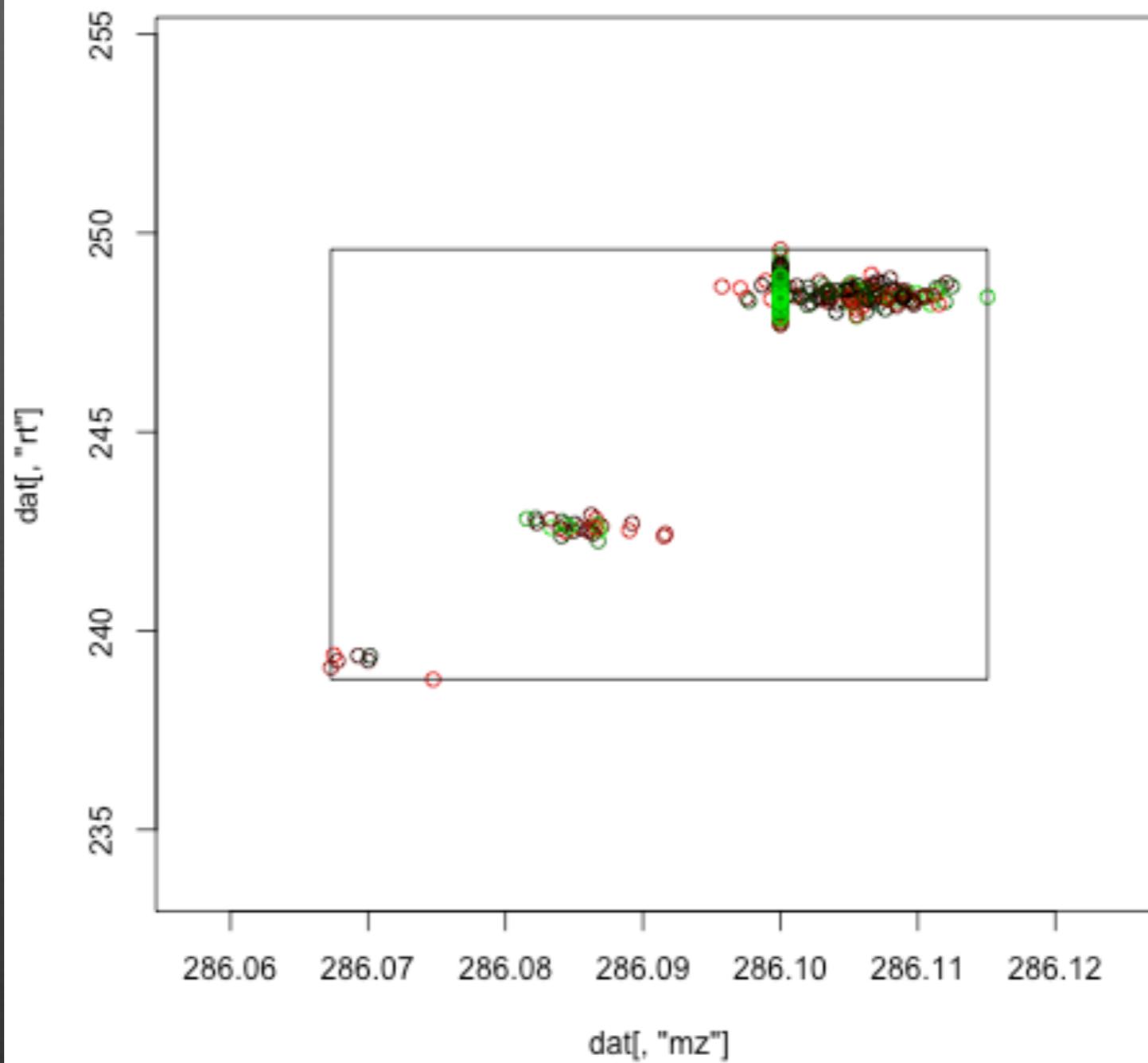
This would have 'npeak' >
number of samples
You could play with
parameters settings
Global parameters :(
Use different method :-)

Grouping = Nearest

- Based on mzMine grouping/alignment algorithm
 - Uses nearest neighbor estimation.

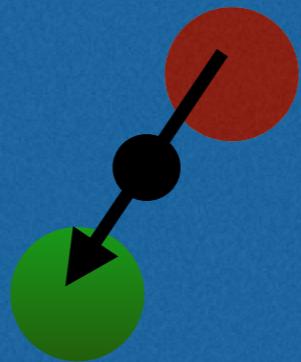


Uridine@ACN RT:248.378204723691 sec



Group.nearest

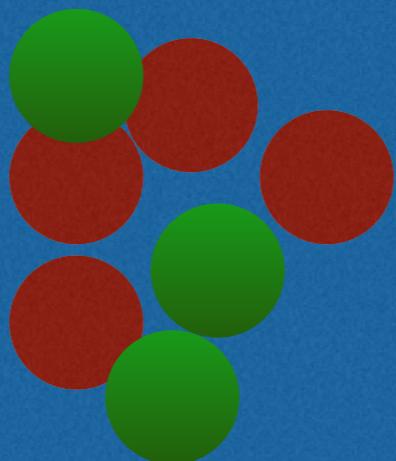
RT



m/z

Group.nearest

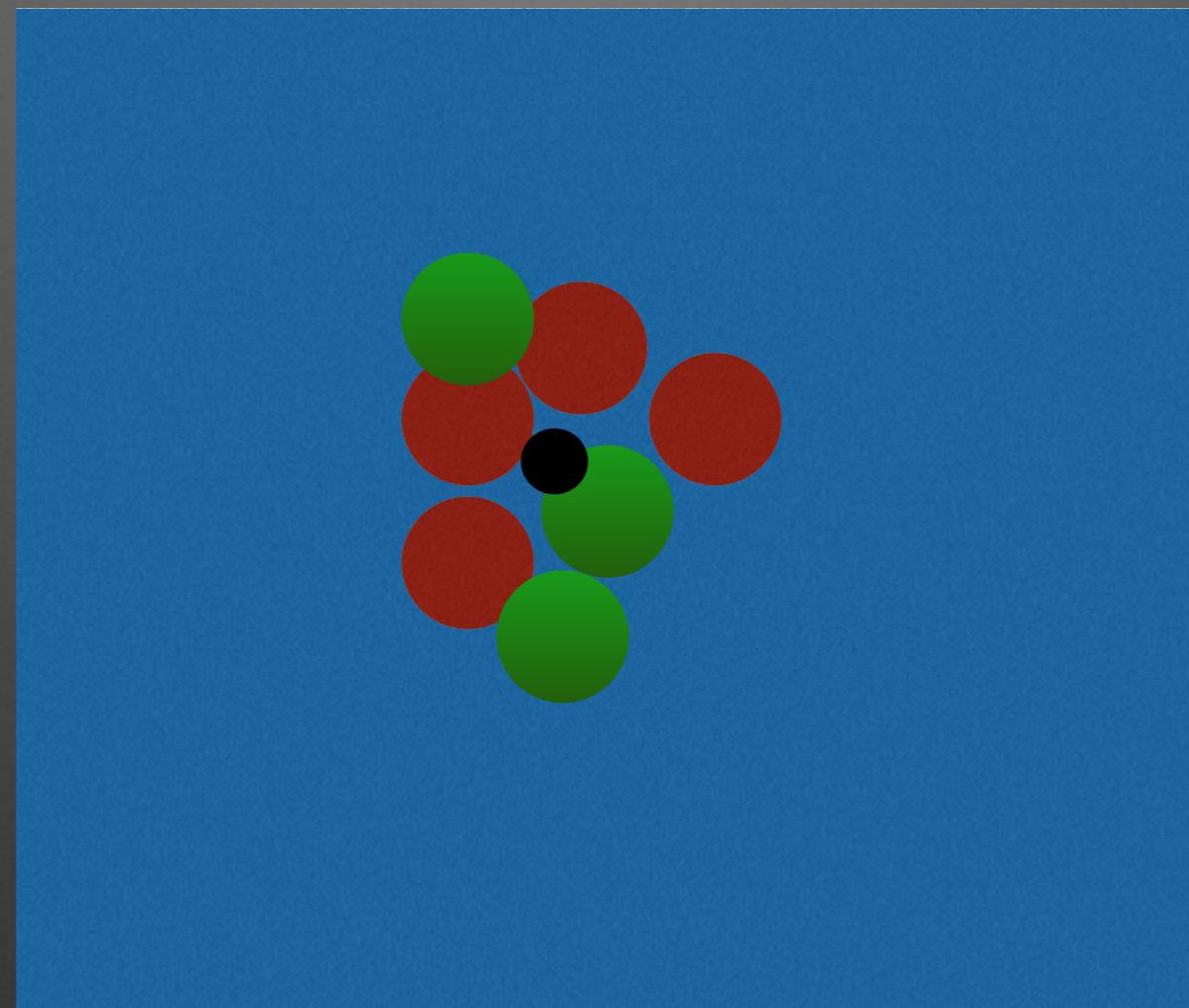
RT



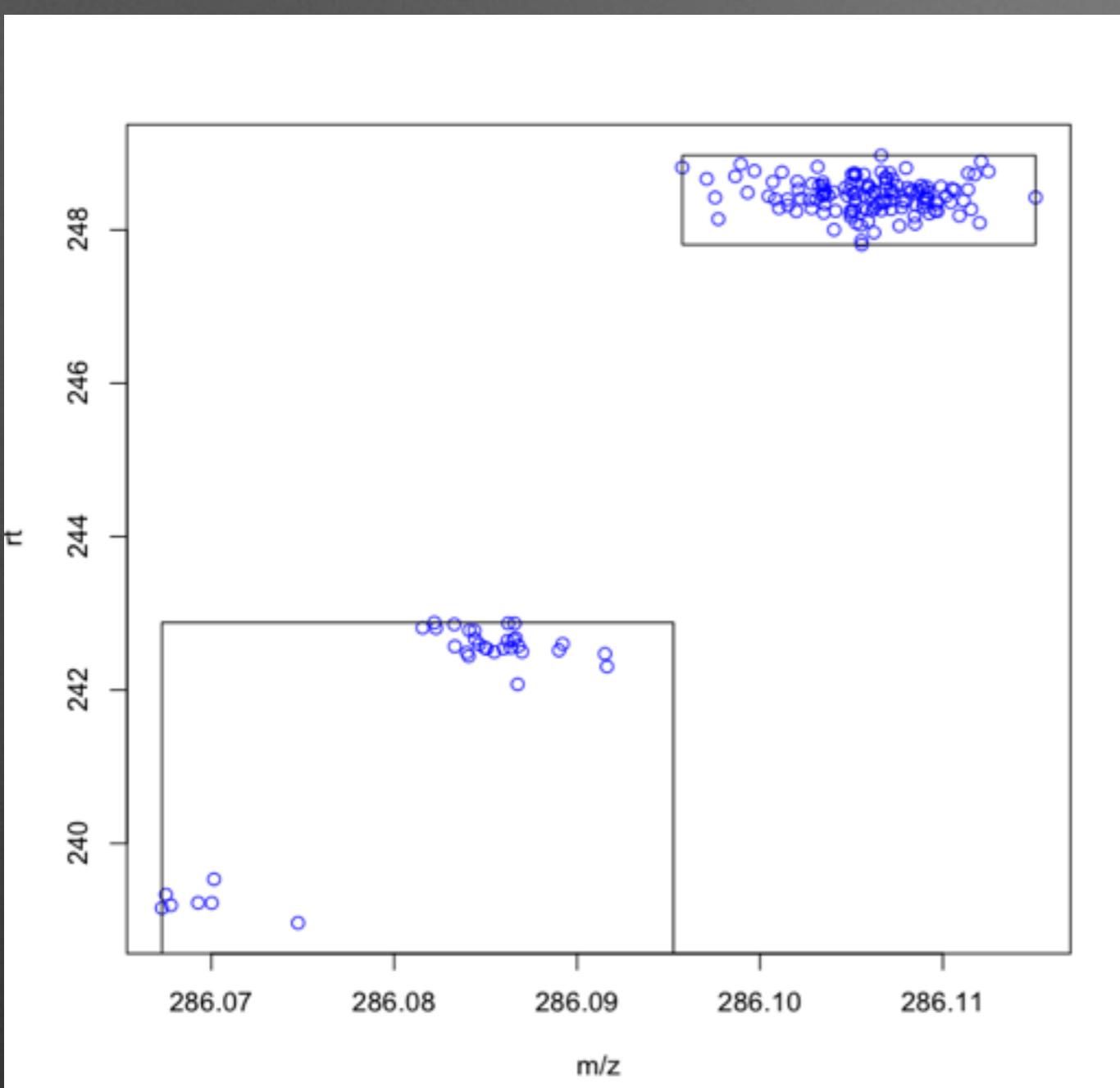
m/z

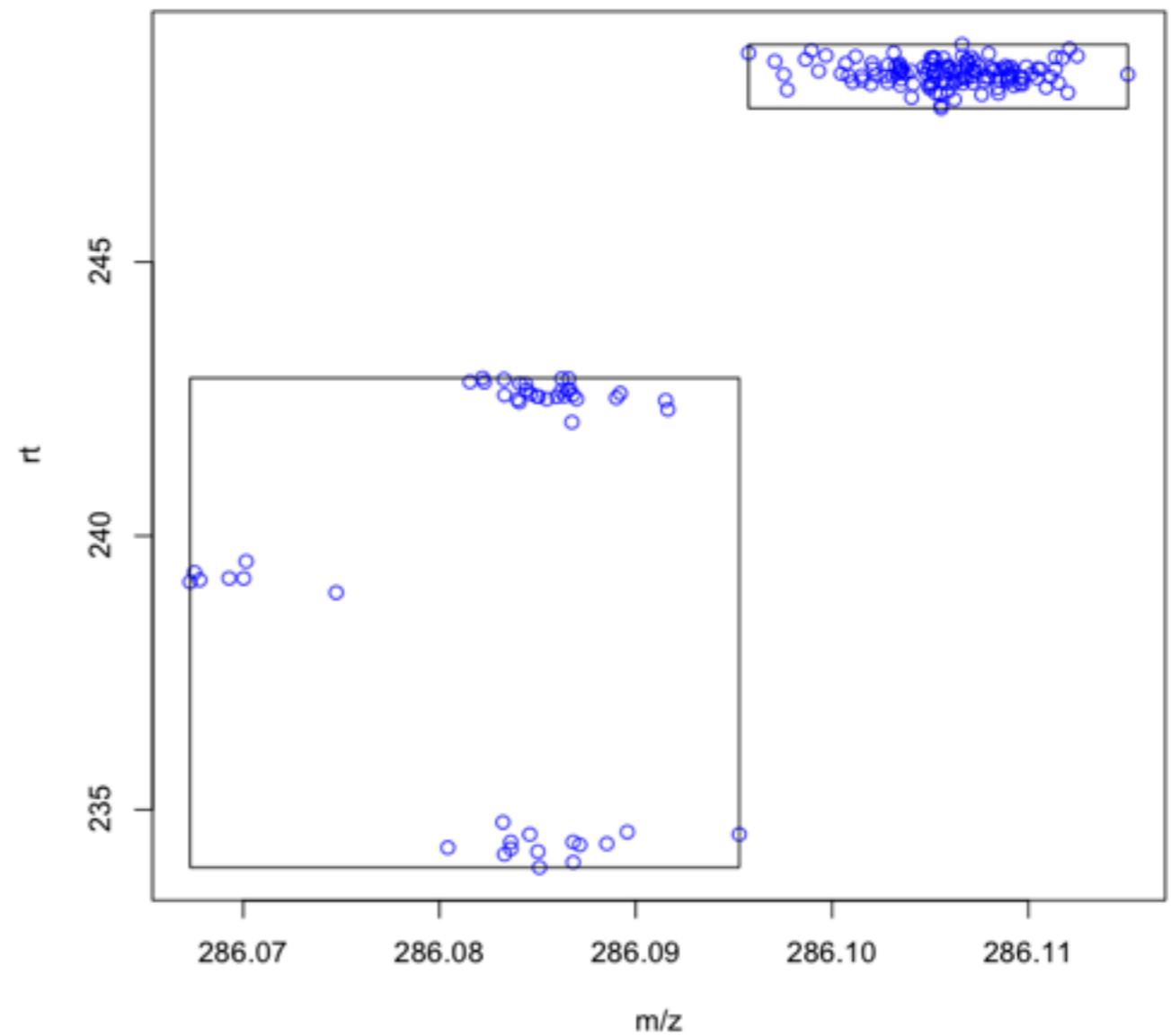
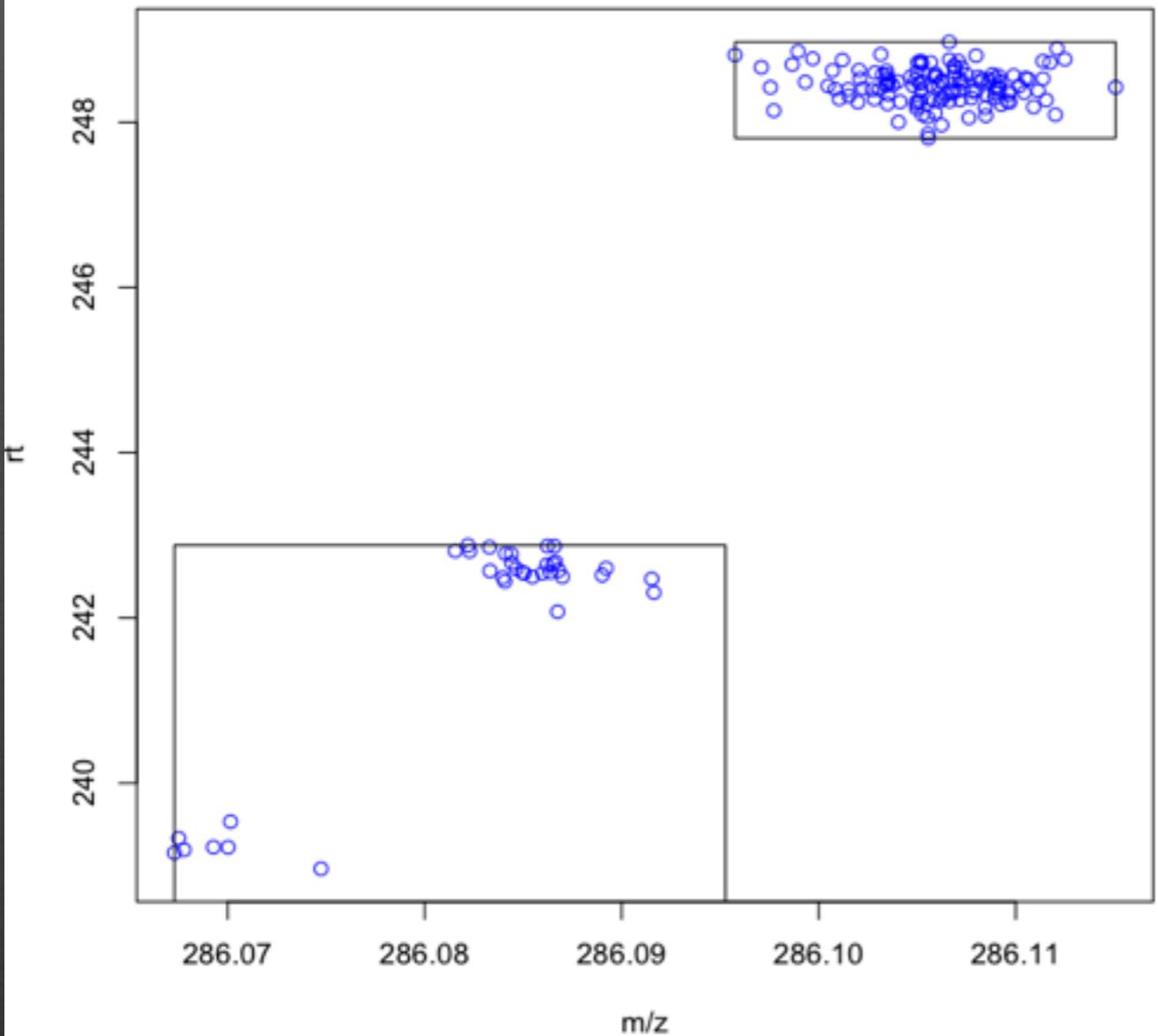
Group.nearest

RT



m/z





General Principles

Peak Detection



Grouping
Groups similar Peaks
across replicates

**Retention Time
Alignment**

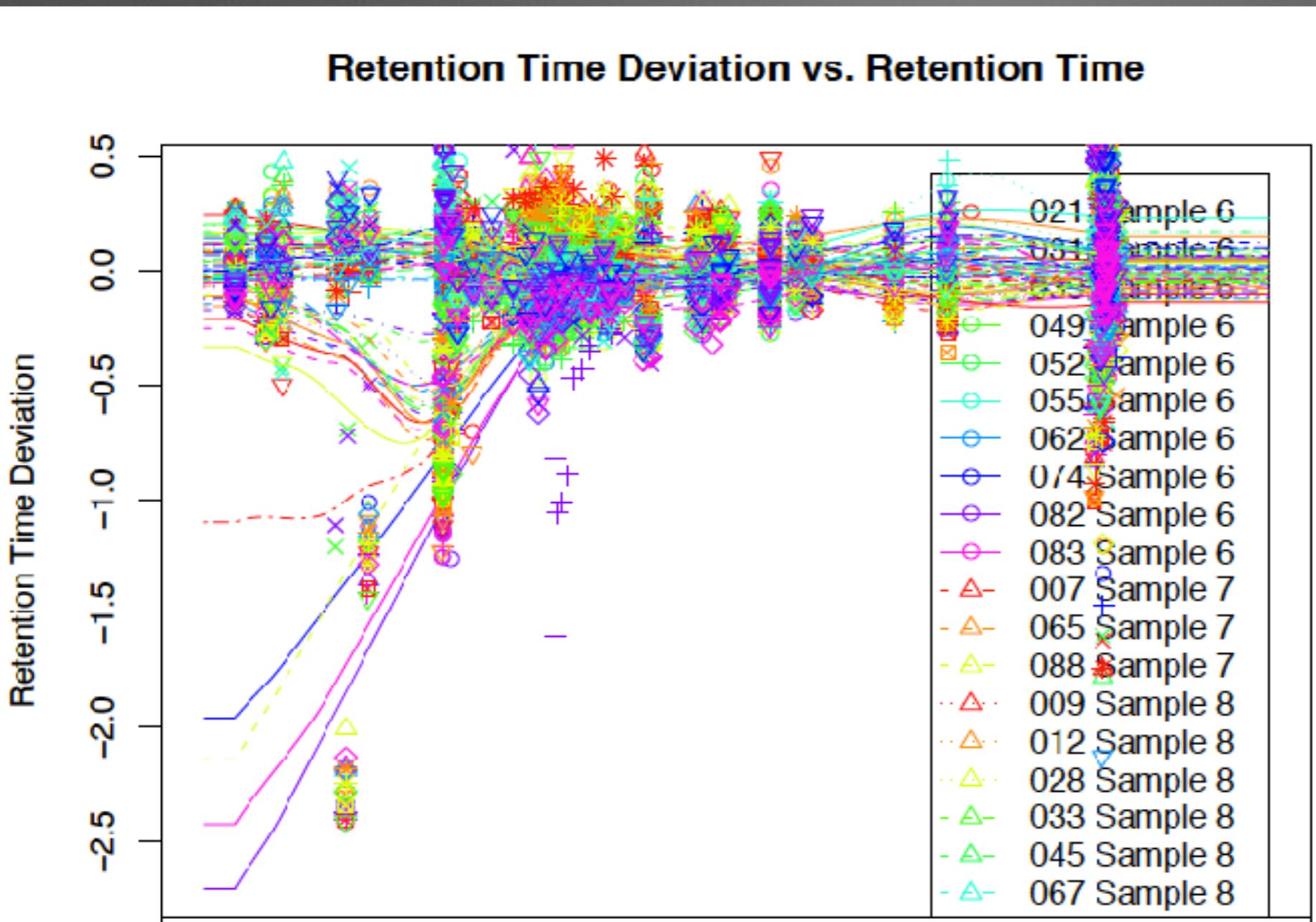
Statistical Analysis
of Classes

Retention Time alignment

- XCMS finds ‘well behaved groups’
 - These include group that have missing peaks, extra peaks or perfect groups (parameters)
 - Missing < n/2 !!
 - Median found for each group
 - Local regression used for each sample to find the deviation profile

Retention time alignment - loess

Retention Time Deviation vs. Retention Time



median rt of each
'well behaved'
group
vs
rt of each file

A good spread of anchors/'well behaved peak groups'

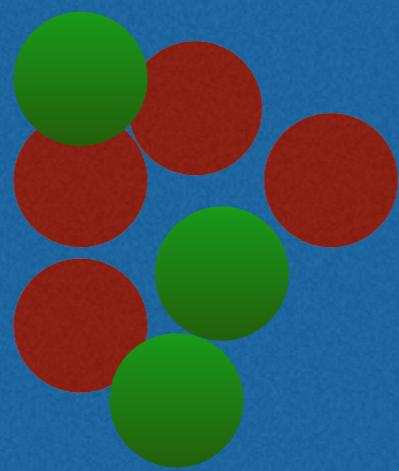
Alignment

- Parameters:
 - **missing** = number of peaks removed from a ‘well behaved peak group’
 - **extra** = Number of additional peak in a ‘well behaved peak group’
 - **span** = Amount of smoothing in regression fitting !
Very sensitive! ~ smaller value more local alignment,
larger more global alignment.

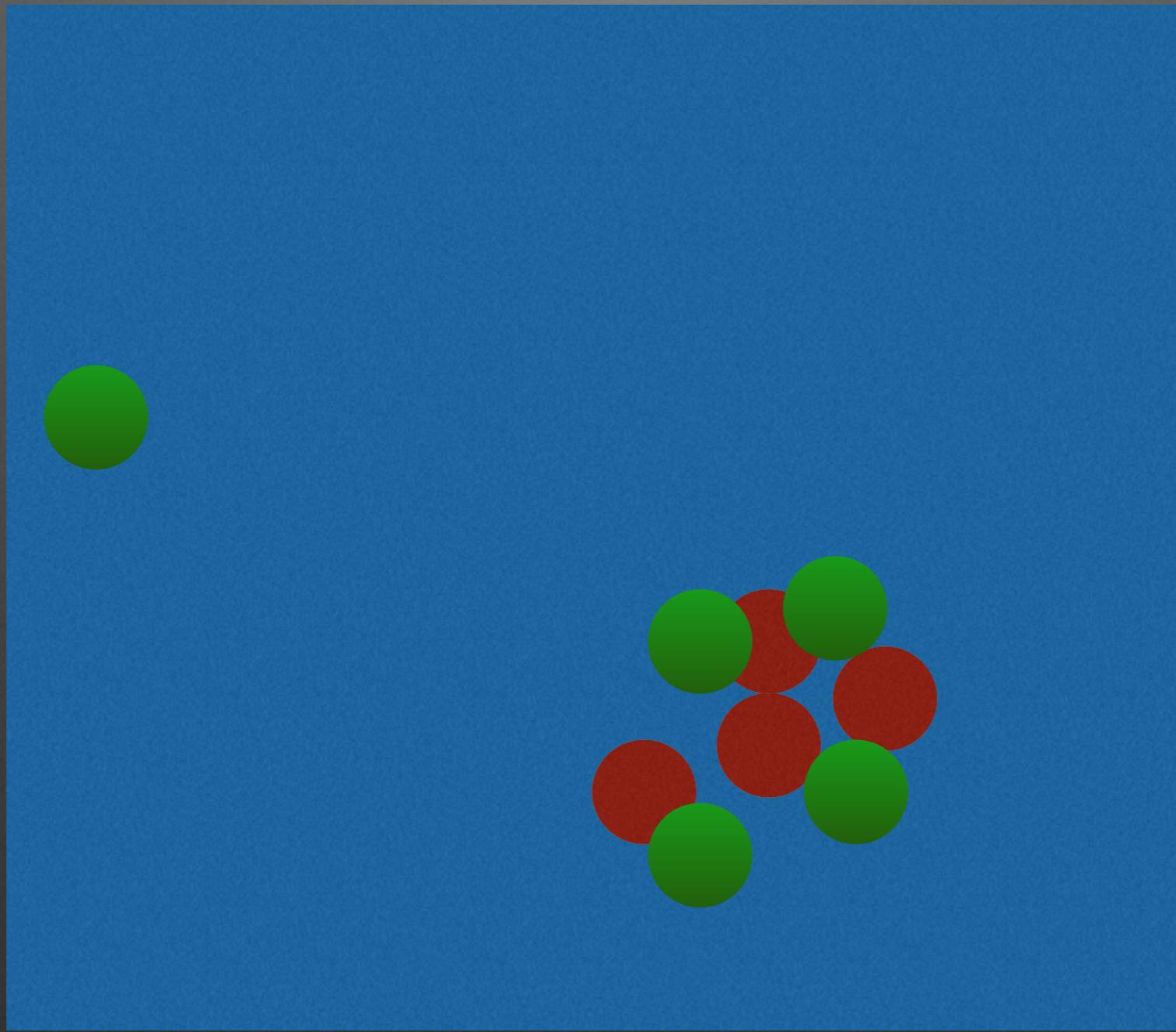
Missing = 1



4 samples
each



Extra = 1



Retention time alignment obiwarp

John T. Prince and Edward M. Marcotte
Chromatographic Alignment of ESI-LC-MS Proteomics
Data Sets by Ordered Bijective Interpolated Warping
Analytical Chemistry, 2060 78 (17), 6140-6152

- obi-warp.sourceforge.net - original program
- Retention time correction based on spectra similarity
- Doesn't rely on detected feature ~~ sort of
- No initial grouping needed

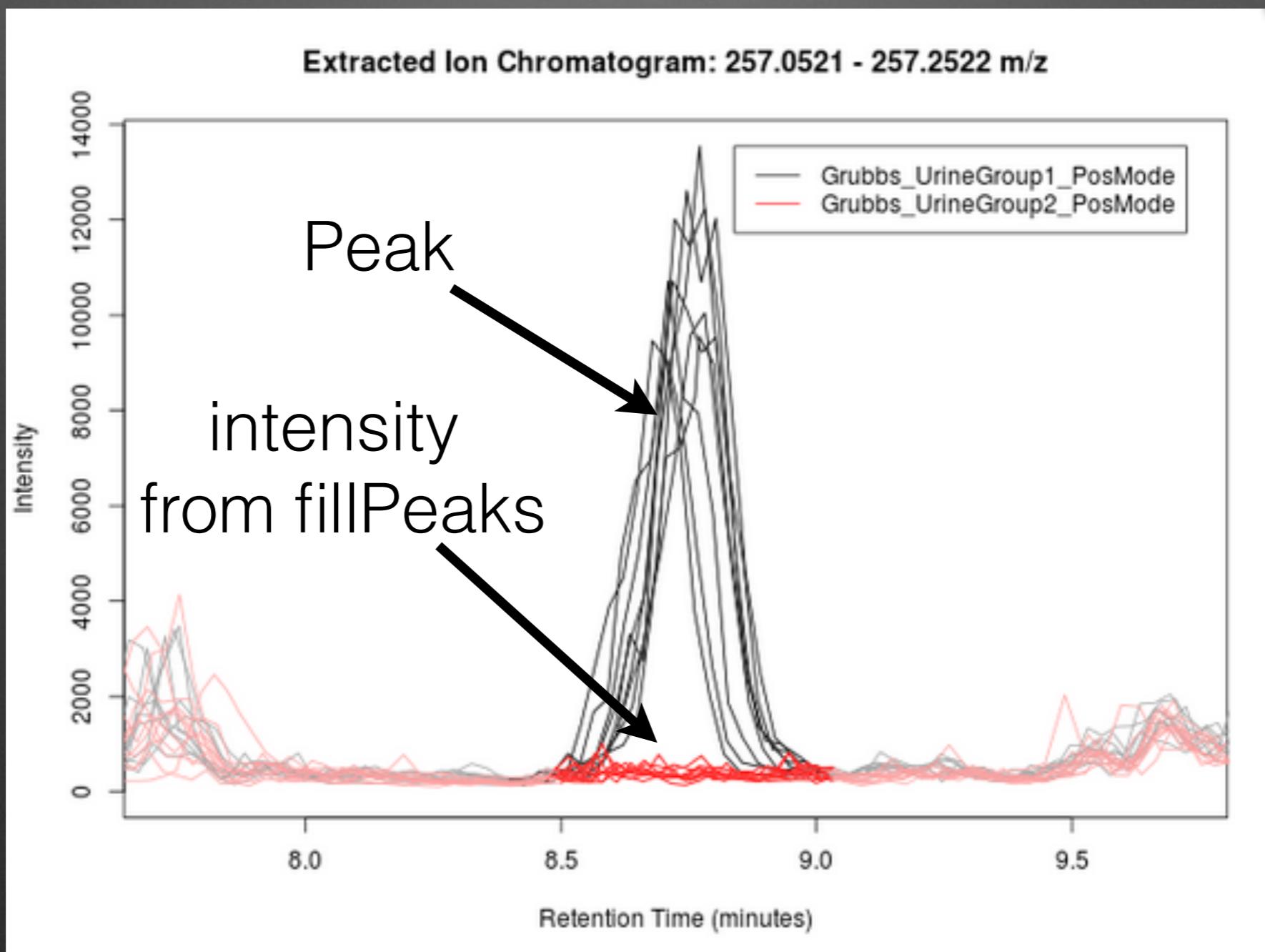
Retention time alignment obiwarp

- Uses a warping technique to warp data to a median chromatogram.
 - This acts as a mold which other spectra are warped to
- Uses a dynamic programming to find path of greatest similarity between median chromatogram and current chromatogram



FillPeaks

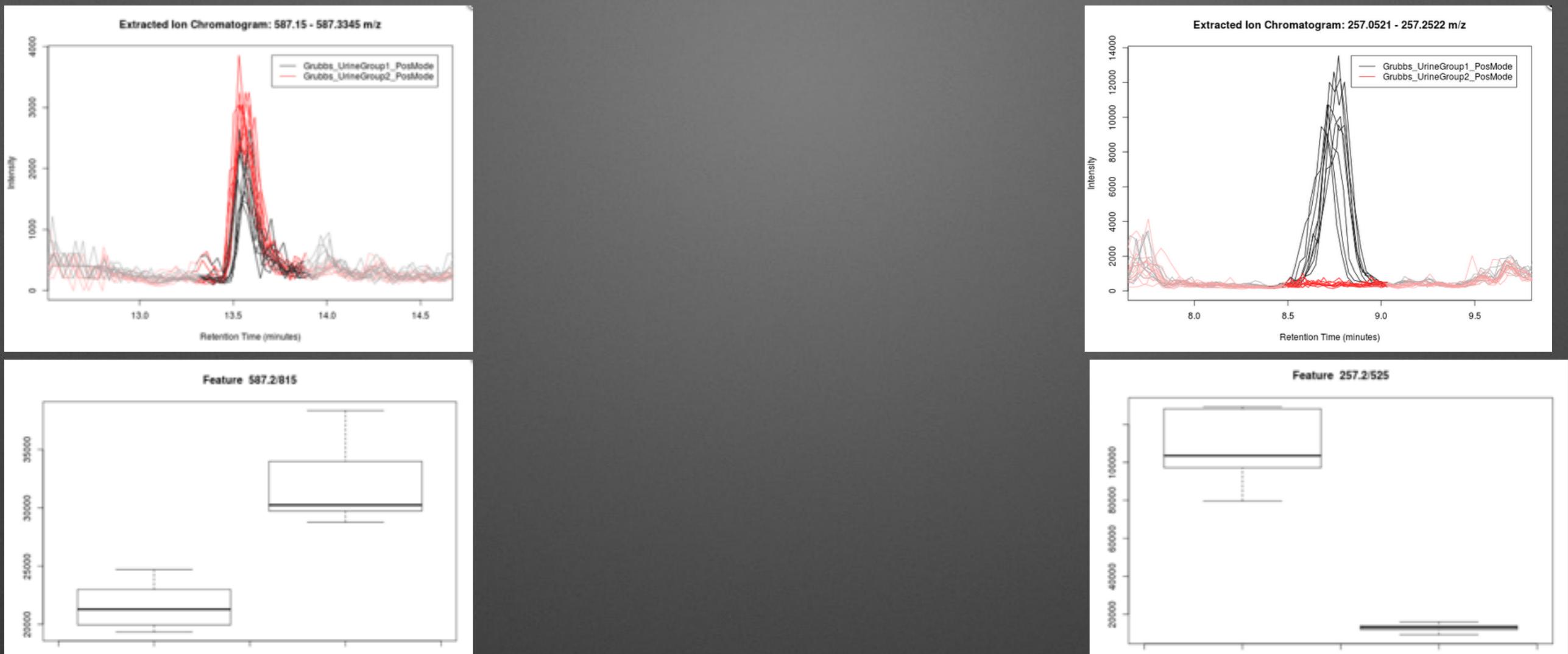
- Going back to each file to find any intensity that wasn't peak picked



Finally !!

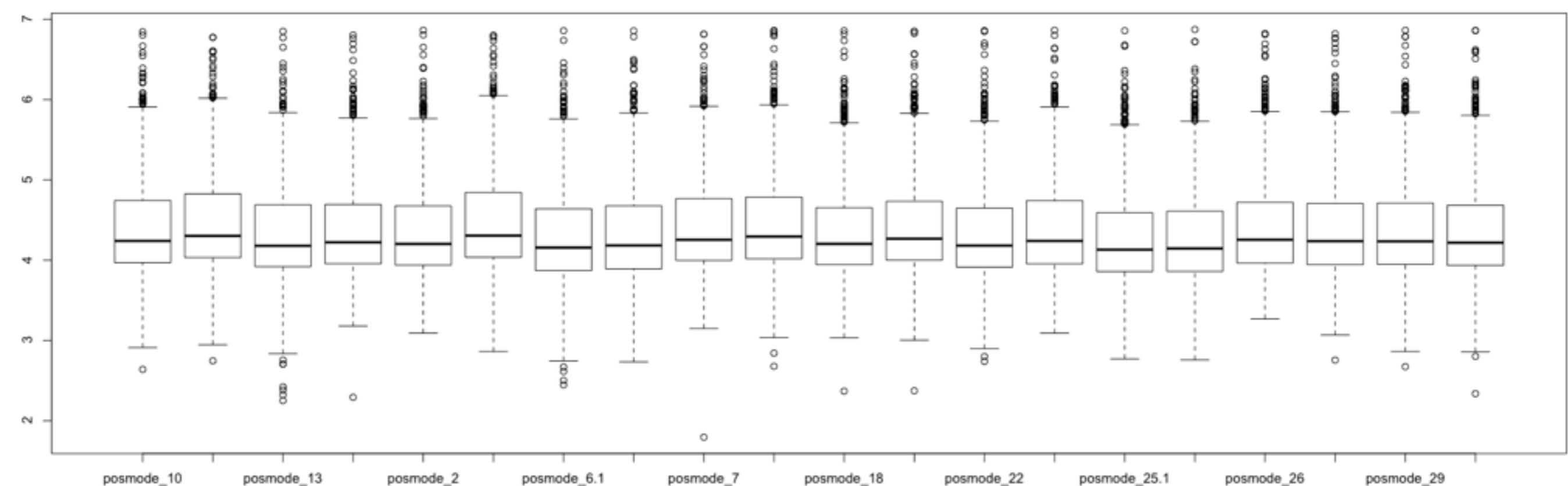
- We have all of our data corrected in a form we can use.
- Lets look at some data processing:
 - heatmaps
 - PCA
 - Some Stats

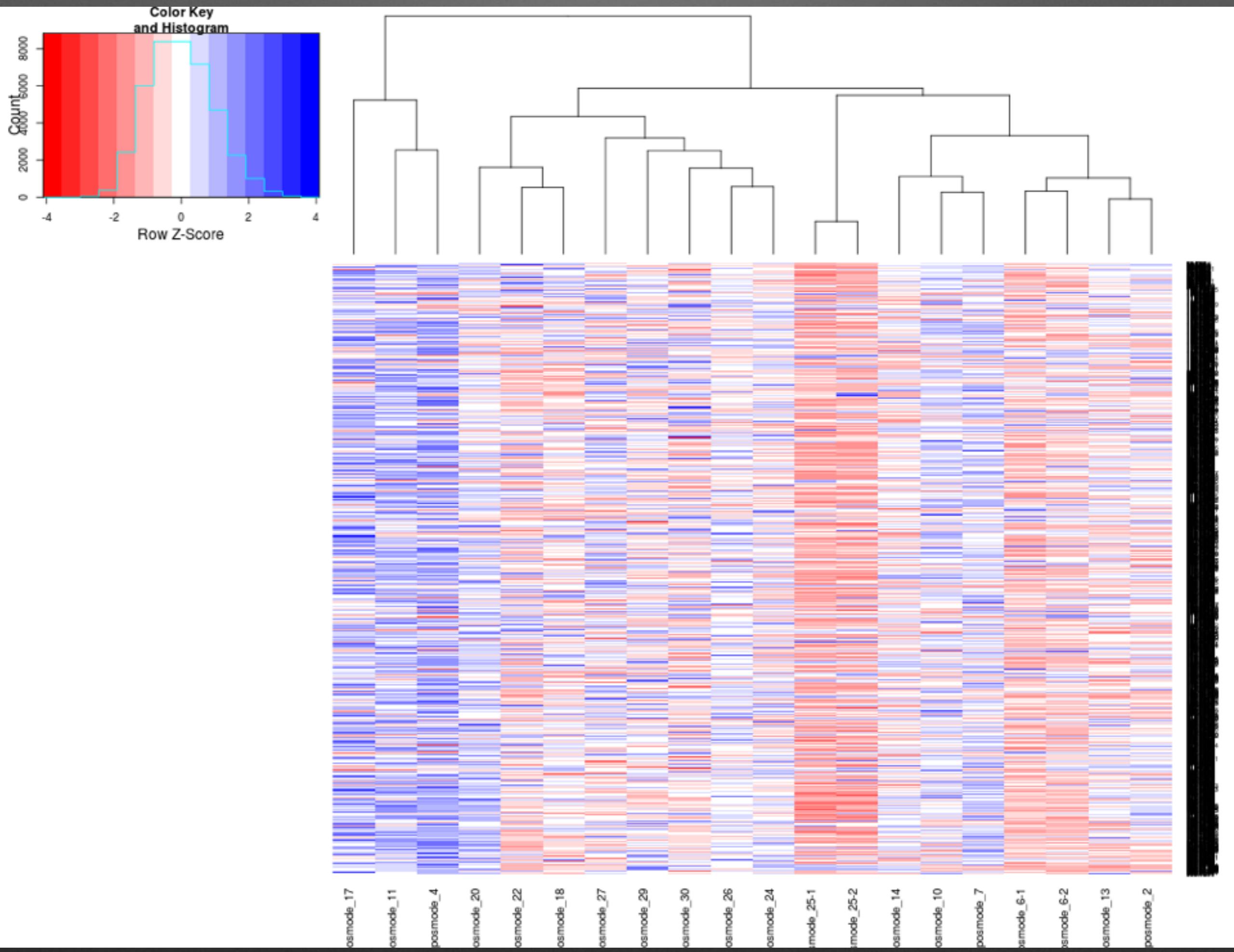
WAIT !!!!

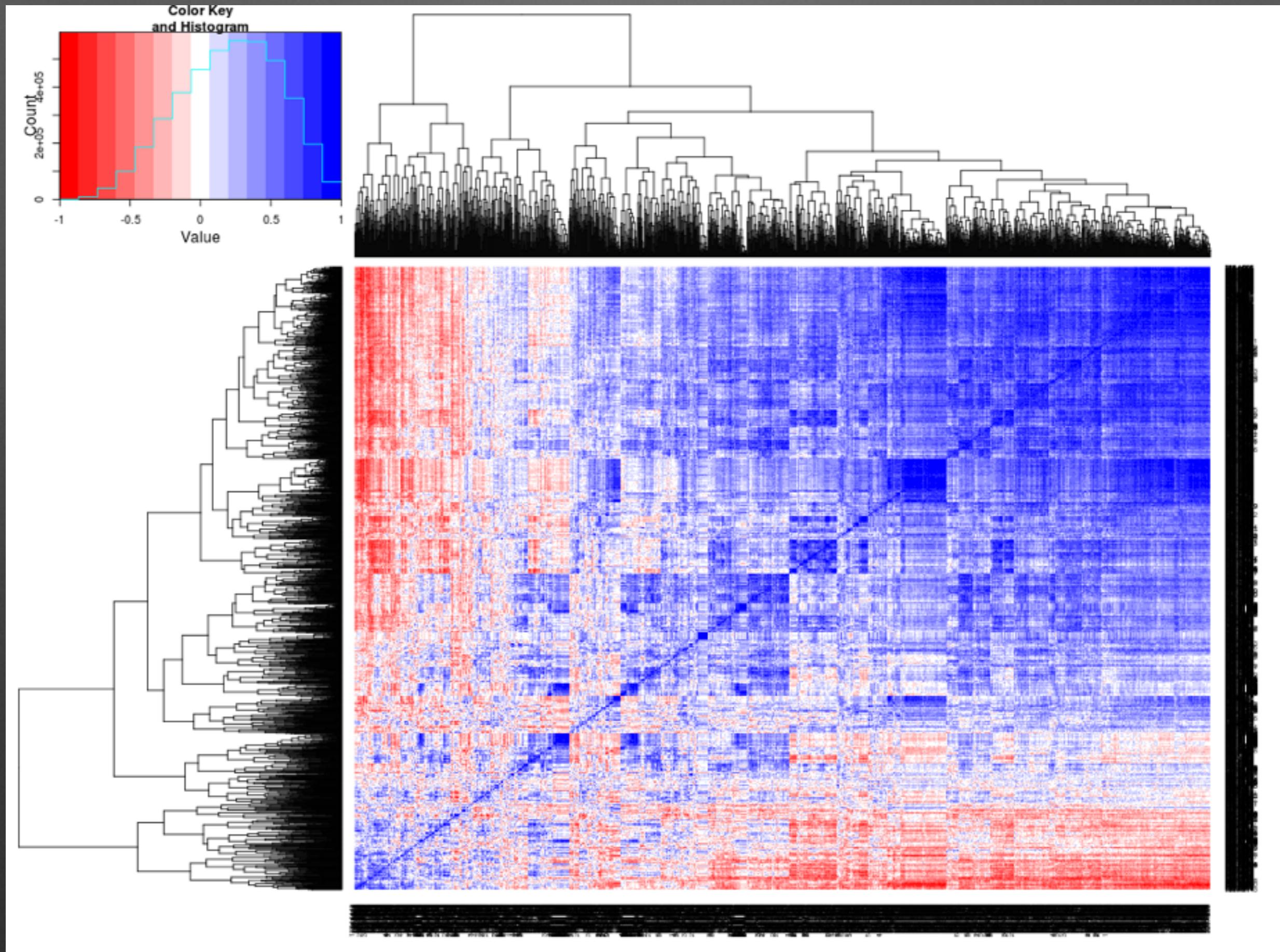


Job#1051415 : Grubbs_urine_pos_mmchg														View 1 - 100 of 1,801	
Feature	fold ch	p-value	q-value	m/z	retention time	MaxInt	Ctrl(sd)	Ctrl(\bar{x})	Exp(sd)	Exp(\bar{x})	isotopes	adducts	feature g		
1	3.0	5.33057e-8	0.00005	204.1446	11.55	8,367	8,385.9	61,606	3,635.3	20,590			35		
2	1.5	6.94626e-8	0.00005	587.2266	13.59	3,858	1,914.6	21,603	3,067.4	31,738	[M+Na] ⁺	54			
3	8.5	2.04304e-7	0.00008	257.1582	8.75	13,544	18,092.6	108,889	2,010.2	12,876	[M+H-H2O] ⁺	53			
4	3.8	2.44253e-7	0.00008	234.1863	11.74	8,264	14,440.5	71,894	10,301.1	19,157	[M+H-CH3] ⁺	57			
5	2.2	6.72076e-7	0.00018	345.1104	10.74	5,082	6,835.8	44,308	3,762.4	20,112			33		
6	1.8	1.03879e-6	0.00023	377.1435	11.91	160,143	116,893	501,472	135,537.2	909,410	[69][M] ⁺		18		
7	1.3	2.79905e-6	0.00054	181.0589	11.24	148,137	70,401.6	925,013	48,407.5	715,125			14		
8	1.5	4.03600e-6	0.00068	193.4786	13.59	4,468	2,356.9	20,927	4,613.7	32,018			4		
9	2.5	4.56496e-6	0.00069	249.1814	19.42	3,787	13,536.2	68,040	6,310.9	26,914			136		
10	1.8	5.57117e-6	0.00071	390.1744	12.87	3,106	7,077.7	43,763	4,776.8	23,898	[M+K+NH3] ⁺	48			
11	1.6	6.04996e-6	0.00071	425.1022	13.61	2,843	1,969.2	16,840	4,621.9	27,629	[2M+K] ⁺	114			
12	2.3	6.40652e-6	0.00071	549.3642	16.16	2,285	2,938.8	14,390	2,200.5	6,283	[134][M] ⁺	[3M+2Na] ⁺	24		

Normalisation needed?







Correlation heat map of the Bonferroni corrected ANOVA p-values

Summary

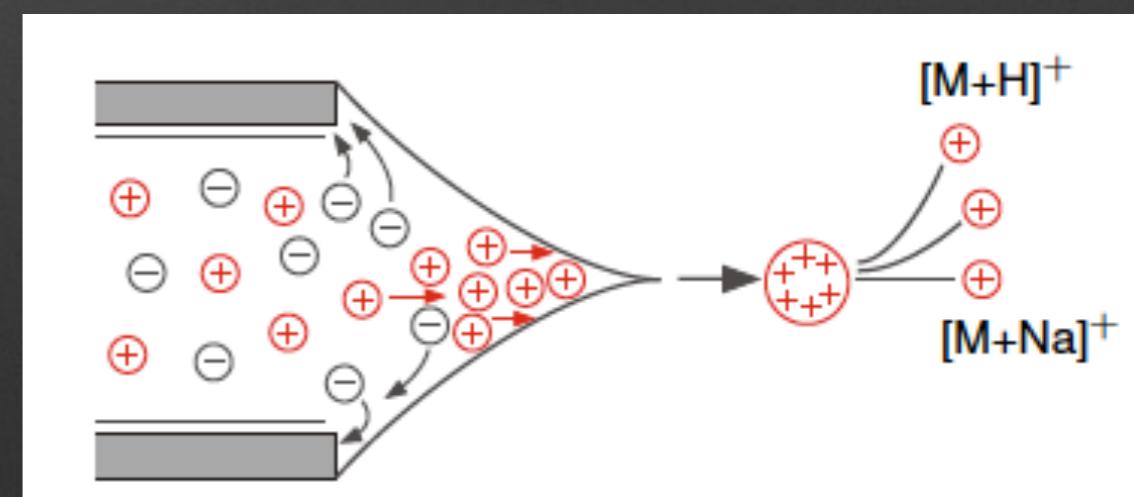
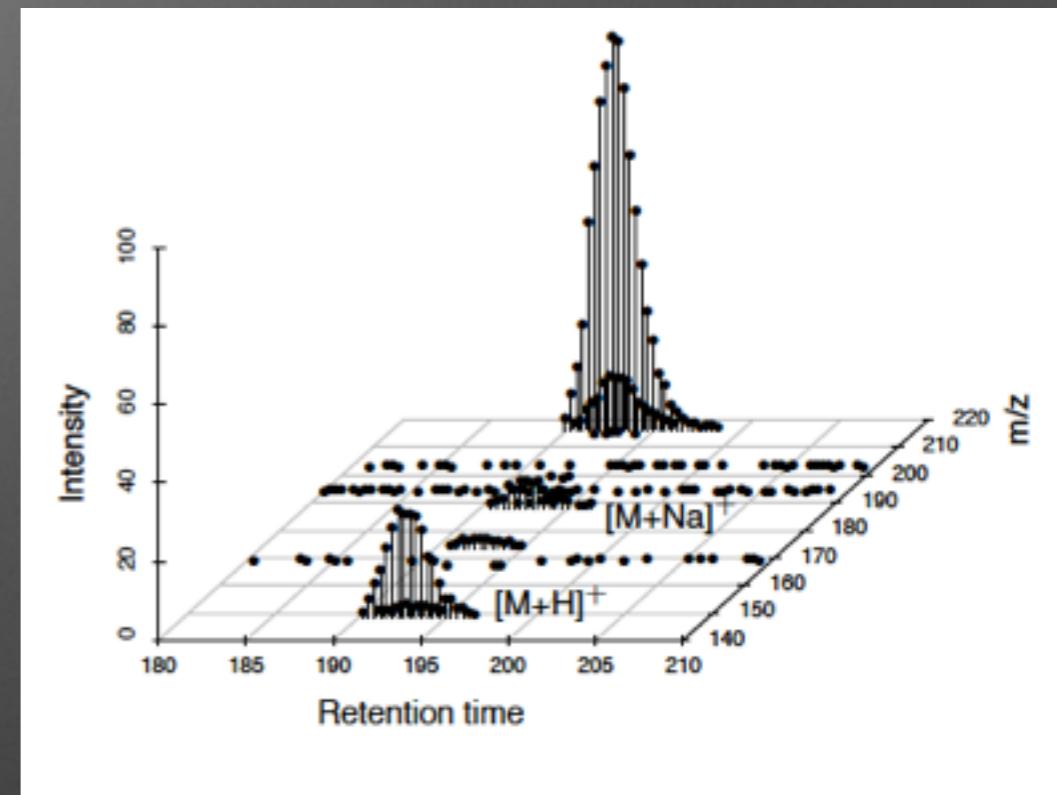
- XCMS processes LC-MS data and is complex
- XCMS processes LC-MS data and uses some simple algorithms. There are multiple algorithm for different jobs/data types.

Boxes and Foxes

- XCMS is all about boxes
 - Boxes are sly and slippery and are the main problem in data analysis
 - If you're having issues try changing alignment methods and thinking about how much deviation in m/z or RT the data has before and post alignment

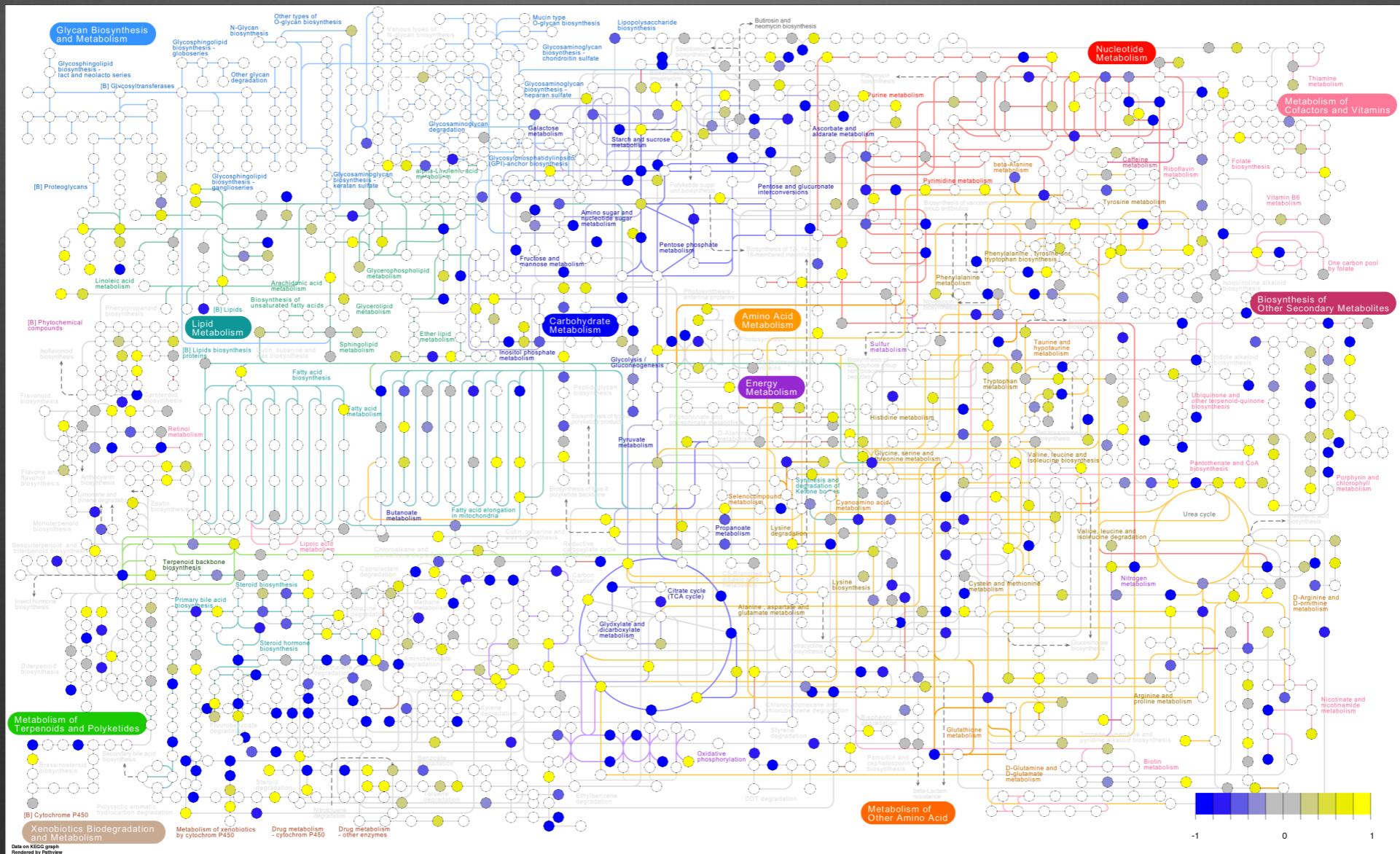
CAMERA

- Same compound should be at the same retention time
- Same compound should have a linear relationship
- Using linear correlation and RT windows adducts/isotopes are labeled



On-wards to biology

- Network maps from related metabolites



Thank You!

- Questions?
- Many more updates coming soon including speed and more stats



Prof. Gary
Siuzdak

The whole
xcms team



Dr. Colin Smith