

Level 4 Project

Week 3/16

Schedule for weeks 2-3 (15-16)

- Complete autoencoder tuning and image pre-processing.
 - Deliverables:
 - Code for an autoencoder model
 - Code for pre-processing
 - Explanation for the choices made
- Collate results obtained by research for evaluation
 - Deliverables:
 - Detailed evaluation plan
 - Report on current research methods for evaluating interactions
 - Excel tables of research metrics that can be methodically parsed
- Finalise image segmentation techniques
 - Deliverables:
 - Code for the image segmentation techniques, along with an explanation of why it was chosen

Autoencoder model

```
c = 3

input_img = Input(shape=(imw, imh, c))

x = Conv2D(64, (3, 3), padding='same')(input_img)
x = PReLU()(x)
x = MaxPooling2D((2, 2), padding='same')(x)
x = Conv2D(32, (3, 3), padding='same')(x)
x = PReLU()(x)
x = MaxPooling2D((2, 2), padding='same')(x)
x = Conv2D(32, (3, 3), padding='same')(x)
x = PReLU()(x)
x = MaxPooling2D((2, 2), padding='same')(x)
x = Conv2D(16, (3, 3), padding='same', strides=2)(x)
x = PReLU()(x)

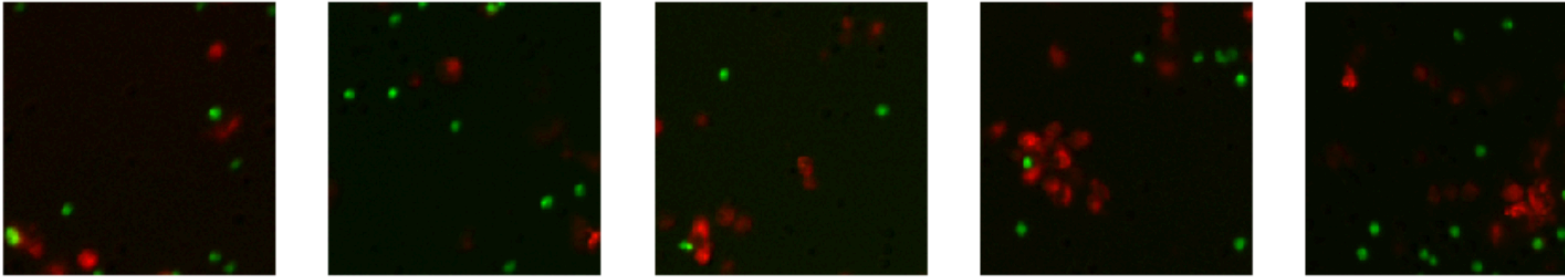
encoded = Flatten()(x)

x = UpSampling2D((2, 2))(x)
x = Conv2D(32, (3, 3), padding='same')(x)
x = PReLU()(x)
x = UpSampling2D((2, 2))(x)
x = Conv2D(32, (3, 3), padding='same')(x)
x = PReLU()(x)
x = UpSampling2D((2, 2))(x)
x = Conv2D(64, (3, 3), padding="same")(x)
x = PReLU()(x)
x = UpSampling2D((2, 2))(x)
decoded = Conv2D(c, (3, 3), activation='sigmoid', padding='same')(x)

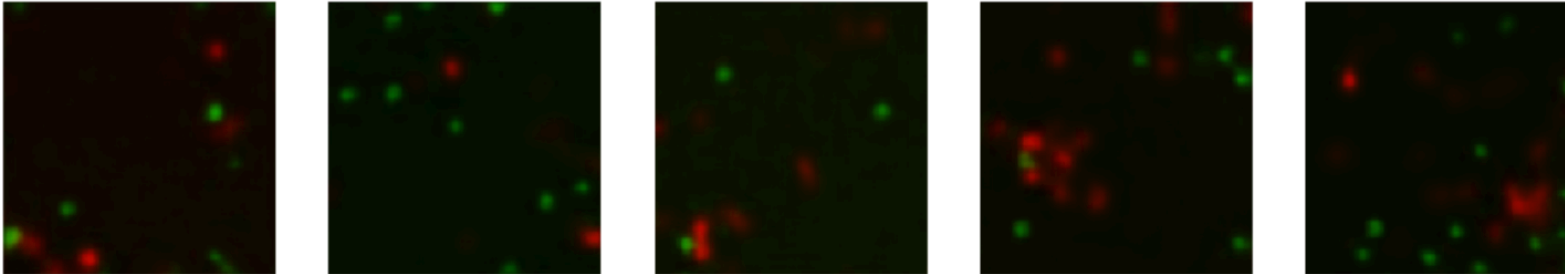
decoder = Model(input_img, decoded)
decoder.compile(optimizer='adam', loss='binary_crossentropy')
```

Striding comparison

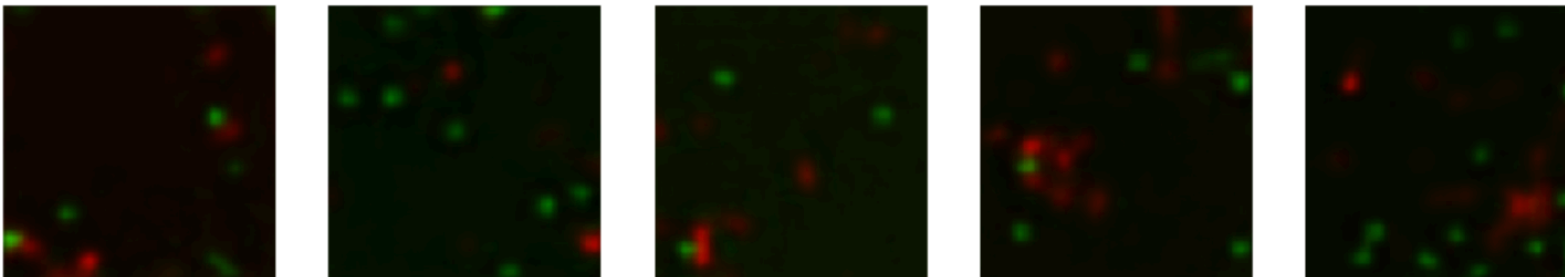
```
[46] plot_range(x_combined, rn=520)
```



```
[45] # structure with strides  
plot_range(decoded_imgs, rn=520)
```



```
# structure without strides  
plot_range(decoded_imgs, rn=520)
```

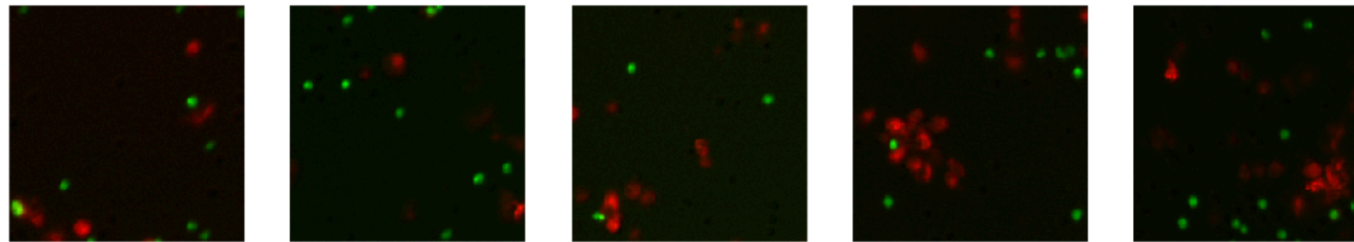


- Difference is minimal but seems slightly brighter
- Captures some overlap better

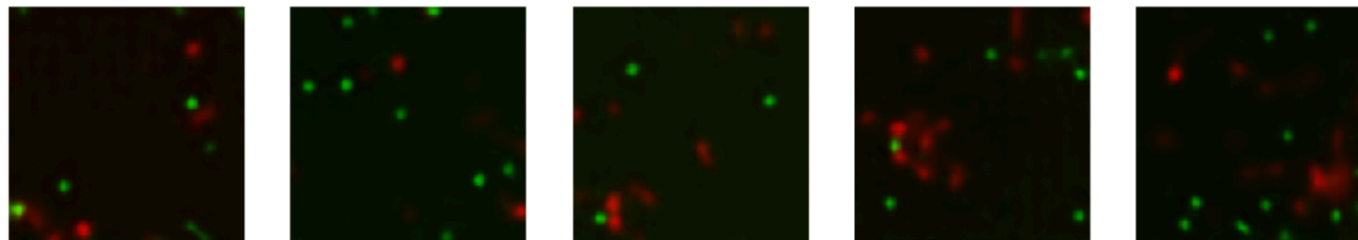
How big should the convolutional layers be?

- Tried both decreasing feature size and increasing feature size
 - Very similar performance but decreasing feature size has smaller dimensions once encoded, hence my choice
- Slightly bigger features has better performance
- Final dimensions are 2,302

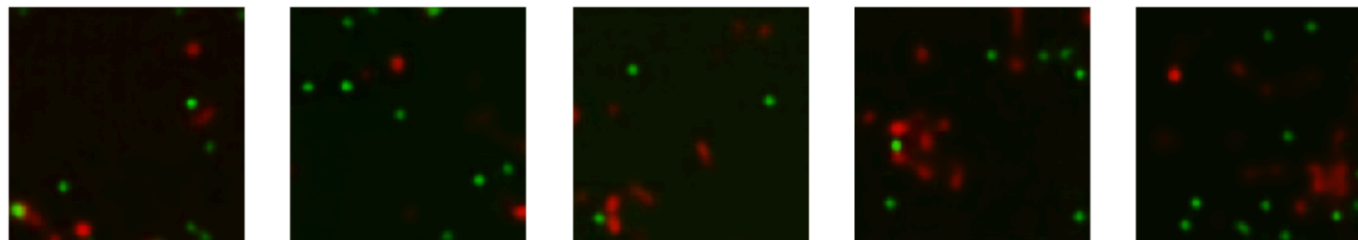
- original



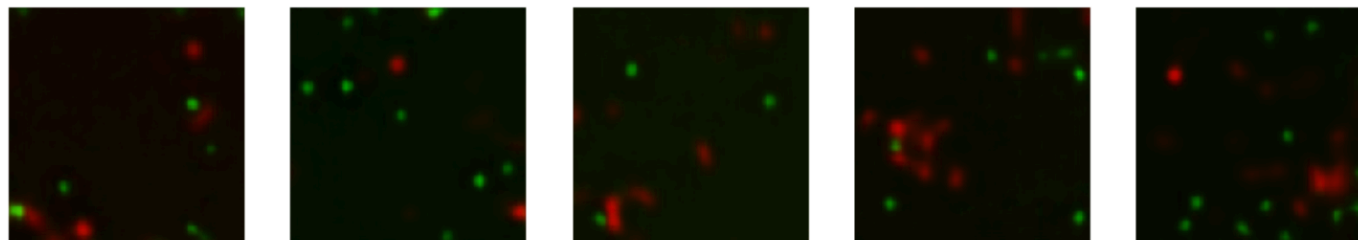
```
[66] # structure with strides, bigger  
plot_range(decoded_imgs, rn=520)
```



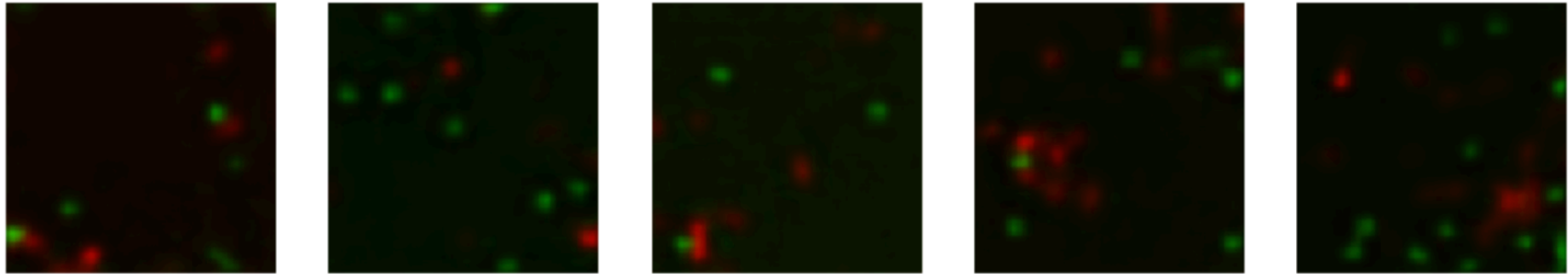
```
[59] # structure with strides, inverted features, bigger  
plot_range(decoded_imgs, rn=520)
```



```
[55] # structure with strides, inverted features  
plot_range(decoded_imgs, rn=520)
```



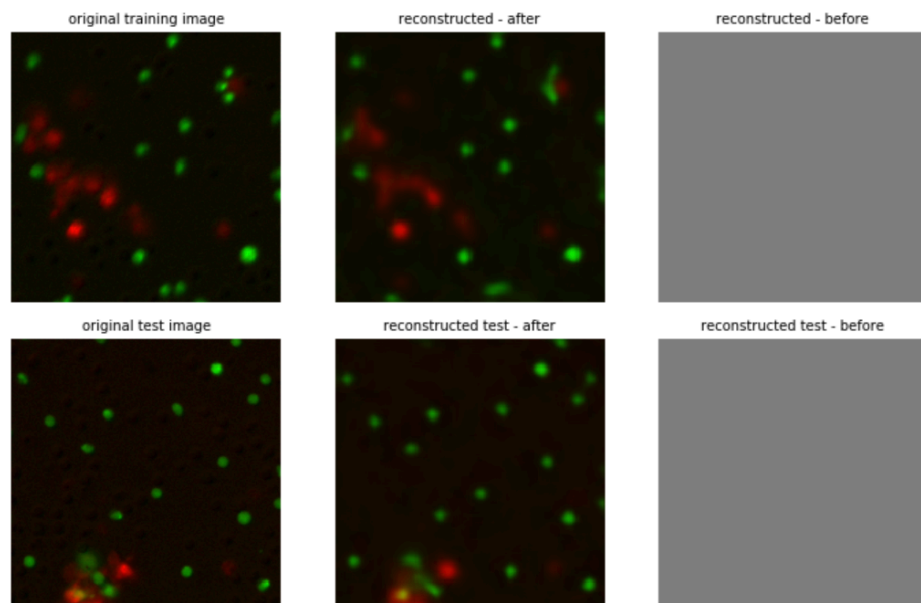
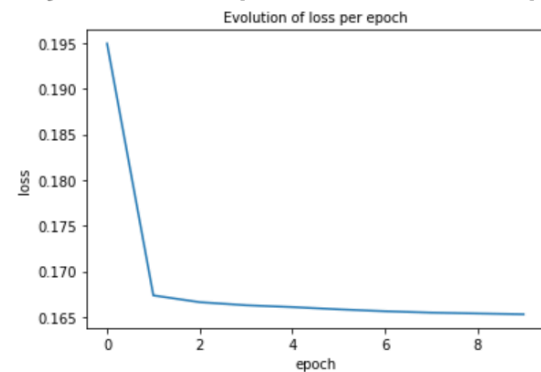
- No strides, smaller size



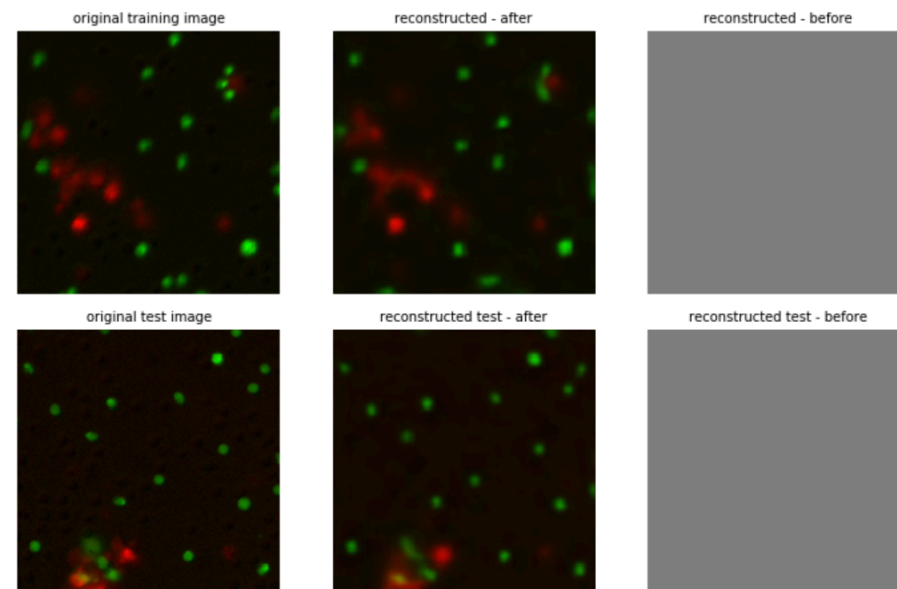
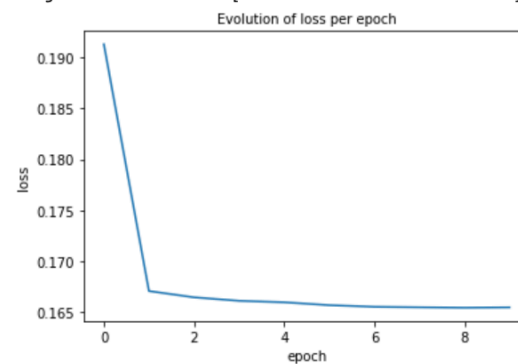
- Highlights improvements achieved

PReLU vs. ReLU

9600/9600 [=====] - 37s 4ms/step - loss: 0.1653
Weight difference: [0.25939307 -0.04599345]



Epoch 10/10
9600/9600 [=====] - 28s 3ms/step - loss: 0.1655
Weight difference: [0.19250129 -0.05974737]



PReLU vs. ReLU

- Very similar
- PReLU seems slightly better
- PReLU has been shown to outperform ReLU

“Delving Deep into Rectifiers: Surpassing Human-Level Performance on ImageNet Classification,” He et al.

“Empirical Evaluation of Rectified Activations in Convolutional Network,” Xu et al.)

Why this autoencoder?

- Not many details in the images to capture means we can reduce the size of feature maps
 - Similar research used similarly structured CNNs
 - “Robust and accurate quantification of biomarkers of immune cells in lung cancer micro-environment using deep convolutional neural networks”
 - “Deep learning as a tool for increased accuracy and efficiency of histopathological diagnosis”
 - Simpler has been shown to work well
 - “Deep learning nuclei detection: A simple approach can deliver state-of-the-art results”
- Did testing to see what worked well for *this* dataset

Processing code

```
def preprocess(data, labels, mask=False):
    # avoid changing the dataset directly
    data = np.copy(data)

    # initialise arrays for filling in
    x_data = np.ndarray(shape=(len(data)//2, 192, 192, 3))
    y_data = np.ndarray(shape=(len(data)//2))

    # initialise index values
    idx = 0
    i = 0

    # loop through images and process
    while idx < (len(data)):
        # ignore 100, 300, etc. values as they will already have been processed
        if (idx % 100 == 0) and (idx % 200 != 0):
            idx += 100
        else:
            # if the image is "faulty" we cannot low_clip and apply min
            max -> NaN
            if is_faulty(data[idx]) or is_faulty(data[idx+100]):
                tcell = minmax(data[idx])
                dcell = minmax(data[idx+100])
                y_data[i] = 3
            else:
                tcell = minmax(low_clip(data[idx]))
                dcell = minmax(low_clip(data[idx+100]))
                y_data[i] = labels[idx]

            # mask out the background
            if mask:
                x_data[i, ..., 0] = dcell*get_mask(dcell) # red-coloured
                x_data[i, ..., 1] = tcell*get_mask(tcell) # green-coloured
            else:
                x_data[i, :, :, 0] = dcell
                x_data[i, :, :, 1] = tcell

            idx+=1
            i+=1

        # try and save memory
        tcell = None
        dcell = None

    print('Images preprocessed. Size of dataset: {}'.format(len(x_data)))
    return x_data, y_data
```

Collating results for evaluation

- DMSO_metrics
 - DMSO from CK19, CK21, CK22
- CK19_metrics
 - 6 sets of each unstimulated, OVA, ConA
- CK22_metrics
 - Mix of unstimulated and OVA
 - Can the algorithm differentiate between two sets, if not three?
- Question
 - Difference between compound concentration and drug?
 - E.g. Compound Conc. μM at 10 vs. ConA (5 $\mu\text{g/ml}$)

Evaluation plan

- https://github.com/leonore/l4-project/blob/master/data/evaluation_plan.md
- Artifacts to evaluate:
 - Autoencoder
 - Clustering
 - Image segmentation

Next steps for this week

- Work on getting all datasets finalised
 - Try reducing memory footprint by changing the datatype
 - Might have to reduce dataset by half (much bigger than DMSO)
- Immunology side:
 - Organise session to sit at GE software and take notes
 - Read provided articles
- Look at practical uses of UNet

Next steps for the coming week

- Ahead of schedule
- Jupyter notebooks → Python files
- Live visualisation for outlier images on a graph
- Start evaluating, possibly