

Advanced 3D Monte Carlo Algorithms for Biophotonic and Medical Applications

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This thesis is submitted in partial fulfilment for the degree of
PhD
at the
University of St Andrews

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Declaration

I, Lewis McMillan, hereby certify that this thesis, which is approximately ***** words in length, has been written by me, that it is the record of work carried out by me, or principally by myself in collaboration with others as acknowledged, and that it has not been submitted in any previous application for a higher degree.

I was admitted as a research student in September 2015 and as a candidate for the degree of PhD in September 2015; the higher study for which this is a record was carried out in the University of St Andrews between 2015 and 2019.

Date Signature of candidate

I hereby certify that the candidate has fulfilled the conditions of the Resolution and Regulations appropriate for the degree of PhD in the University of St Andrews and that the candidate is qualified to submit this thesis in application for that degree.

Date Signature of supervisor

Date Signature of supervisor

Abstract

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Acknowledgements

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Abbreviations

CVD cardiovascular disease.

CVDs cardiovascular diseases.

EEM excitation emission matrix.

MCRT Monte Carlo radiation transfer.

NM Nelder-Mead.

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Chapter 1

Modelling Autofluorescence in Skin for Novel Biomarkers of Cardiovascular Diseases

1.1 Introduction

Cardiovascular diseases (CVDs) are the leading cause of death in the world [1]. It is estimated that around 18 million people died in 2016 from CVDs, accounting for 31% of global deaths [1]. Despite decreasing burden of CVDs in the UK, it was still the number two cause of death in the UK in 2014 [2].

Currently risk factors are used to try to determine if a patient has cardiovascular disease (CVD). However, these risk factors are only a “causal pathway leading to the disease” [3].

The risk factors such as high blood pressure, smoking, diabetes, physical inactivity, and dyslipidemia do not fully explain incidence of disease [4, 5]. Therefore research has moved to examining more novel biomarkers for detecting the disease. Among these novel biomarkers, the autofluorescence response of tissue is of much interest.

Autofluorescence is the natural fluorescence released by biological structures upon excitation by light. Autofluorescence is particularly attractive as a biomarker as it requires no exogenous dyes, which can be toxic, non-specific, or interfere with biological function [6]. In tissue there are several fluorophores responsible for this autofluorescent response, including: NADH (nicotinamide adenine dinucleotide), structural proteins like collagen and elastin, aromatic amino acids (tyrosine and tryptophan), porphyrins, and FAD (flavin adenine nucleotide) [7]. Changes in the autofluorescent response of tissue has been linked to cancer, Alzheimers, diabetes and CVDs [8–12]. Autofluorescence can be linked to these diseases as the fluorophores responsible for the autofluorescence either originate in the mitochondria, or are involved in important biochemical pathways that regulate apoptosis, free-radical generation, oxidative stress and bimolecular sensing of glucose, oxygen and nitric oxide.

NADH has recently been the subject of interest in particular as a biomarker for CVDs [13–15]. NADH is a intracellular co-enzyme that is a biomarker for metabolic activity and mitochondrial anomalies. NADH is involved in mitochondrial function, energy metabolism, calcium homoeostasis, gene expression, oxidative stress, ageing, apoptosis and glycolysis (see Fig. 1.1). Therefore if there is dysfunction, then the autofluorescent signal from NADH will be affected, thus it can be used as biomarker for disease.

Despite their appeal using these autofluorescence to diagnose and asses disease risk more research needs to be carried out on various unknowns: information on the location of fluorescence, which fluorophores contribute to the signal and how much, how much the optics of tissue affect the signal, and the variability of the signal over different location on the body. This chapter aims to determine how much the tissue optics affects the autofluorescent signal, which fluorophores contribute to the signal, and from which layer of the skin do they contribute from. Finally we introduce our ameboMCRT algorithm, created to determine the relative concentrations of the intrinsic fluorophores in the skin, to assess clinical outcomes.

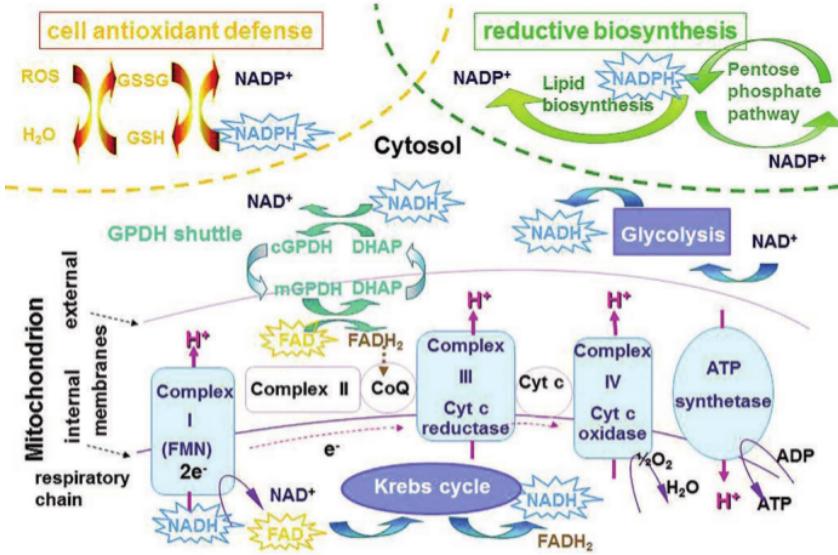


Figure 1.1: Simplified schematic showing the roles of NADH and FAD in various different metabolic pathways. The star boxes indicate fluorescing forms of NADH and FAD. Taken from Croce et al. [16].

1.2 Skin Model

So far in this thesis all tissue models have been simplified, by assuming that tissue is a homogeneous structure with uniform optical properties. However, this is not the case in reality. Tissue is un-homogeneous, with nonuniform optical properties. However, to create a one to one model of tissue in a simulation is impractical due to the resolution required to resolve all the constituent parts of the tissue down to the cell level. Therefore we need to make a compromise between reality and what is possible to model efficiently. This section presents a five layer model of human skin.

Dermatologists split the skin into several layers based upon the morphology, function and contents of each layer [17, 18]. The layers named from outer layer to inner most layer: Stratum corneum, Stratum lucidum, Stratum granulosum, Stratum spinosum, Stratum basale, Papillary dermis, Reticular dermis, and Hypodermis. As not all these layers are optically distinct, are too small to model or not present in a given location on the body. Therefore, we simplify the layers into just 5 layers: Stratum Corneum, Epidermis, Papillary Dermis, Reticular Dermis, and Hypodermis, with the epidermis comprising of Stratum lucidum, Stratum granulosum,

Stratum spinosum, Stratum basale. Figure 1.2 shows the geometry of this model.

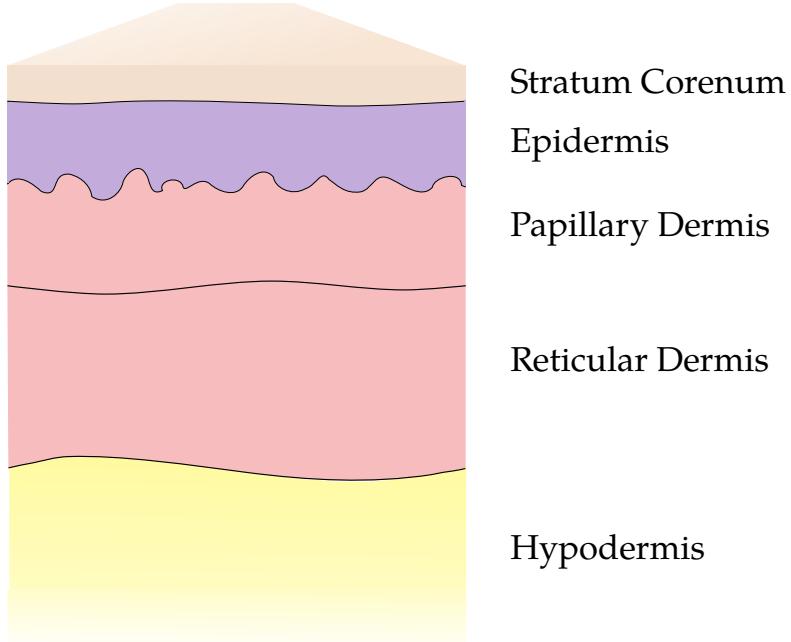


Figure 1.2: Illustration of our five layer skin model.

Each of these layers have various amounts of chromophores and scatterers. To accurately model these various chromophores and scatterers, and therefore the skin, we must discuss the biological make up and spatial structure of the skin.

Stratum Corenum

The outer most layer of the skin is the Stratum corenum. This layer mostly consists of dead skin cells (keratinocytes). Keratinocytes, which make up approximately 80 percent of the cells in the epidermis, are born in the stratum basale, and live for approximately 14 days. During this period they move upwards to the surface of the skin undergoing a series of complex morphological and metabolic events, which ends with the cells undergoing apoptosis by the time they reach the stratum corenum. The keratinocytes are flat in shape, cornified and stacked on top each other in this layer. The function of this layer is to be a protection barrier to prevent damage, infection and diffusion of unwanted chemicals further into the skin. The stratum corenum also prevents water loss and provides some UV protection [17,18].

Epidermis

Below the Stratum corenum is the epidermis. The epidermis consist of several layers that are optically similar so we restrict our model to modelling this as one whole layer. The layers that make up the epidermis are the stratum basale, stratum spinosum, stratum granulosum, and the stratum lucidum*. Each of these layers are distinct from one another, in the fact that

*The stratum lucidum only appears in areas of the body where skin is thick, e.g the palm or the sole of the foot.

the keratinocytes in each layer are different from one another. In the stratum lucidum the keratinocytes are dead, and fairly flat. The stratum granulosum the keratinocytes are grainy and becoming fairly flat in comparison to the below layers. Stratum spinosum, the keratinocytes appear spiny (hence the name of the layer), and are polyhedral in shape. It is in the stratum spinosum that the cells begin to become keratinized and start the process of dying as they move upwards. Finally the last layer of the epidermis, the stratum basale, is the layer where the keratinocytes are born. Here the keratinocytes are columnar or cuboid in shape.

The purpose of the epidermis is as before to provide a protective barrier to the underlying layers. In the stratum basale there is also melanocytes which produce the pigment melanin which is responsible for the color of the skin, and providing some protection from harmful UV light. Other types of cells found in the epidermis are Langerhans cells and Merkel cells which are part of the immune system and nervous system respectively. Overall the epidermis provides protection from mechanical stress, flexibility to the skin, UV protection, retains water, and stops foreign bodies or chemicals from entering the skin [17, 18].

Dermis

The dermis is the layer of skin below the epidermis, and makes up the majority of the skin by size. The cellular makeup of the dermis is different from that of the epidermis, as the dermis is mostly made up of filamentous or fibrous proteins such as elastin or collagen. The dermis also contains various nerves and blood vessels. The dermis is split into three layers, the papillary dermis, the reticular dermis, and the hypodermis. The boundary between the papillary and reticular layers is demarcated by the subpapillary plexus, which is a horizontal plane of blood vessels. The boundary between the reticular and the hypodermis is marked by a abrupt change from fibrous to adipose tissue. The function of the dermis is to protect from mechanical injury, thermal regulation, contains receptors of sensory stimuli, gives the skin pliability, elasticity and tensile strength [17, 18].

Optical Properties

With a discussion of what makes up the skin, and what molecules contribute to the skins optical properties, this section gives an account of how our skin model models the optical properties of skin. First we will discuss the absorber that are found in the skin, and what their absorption properties are as a function of wavelength. Scattering properties of the several layers of tissue will then be discussed followed by a quick discussion on refractive indices and anisotropy values. Finally a discussion on the fluorophore found in the skin will be presented.

The first chromophore we will examine is blood. To model blood, we first split blood into its deoxygenated and oxygenated components. This is done as the absorption coefficient differs between the two types of blood. We mix these two groups using the tissue oxygenation coefficient S . Blood absorption spectra are taken from S. Prahl [19].

$$\mu_{a,Oxy/DeOxy} = 150 \ln 10 \frac{\epsilon}{64458} \quad (1.1)$$

$$\mu_{a,blood}(\lambda) = S\mu_{a,Oxy} + (1 - S)\mu_{a,DeOxy} \quad (1.2)$$

Where:

ϵ is the excitation coefficient of hemoglobin [cm^{-1}];

64458 is the molecular weight of hemoglobin [$g mol^{-1}$];

150 is the normal concentration of hemoglobin in blood [$g L^{-1}$];

and $\mu_{a,Oxy}$, $\mu_{a,DeOxy}$, and $\mu_{a,blood}$ are the absorption coefficients for oxygenated, deoxygenated and blood respectively [cm^{-1}].

We also include water in our skin model. Water's absorption spectrum is taken from the work of Wieliczka *et al* and Segelstein [20, 21].

The next chromophores are bilirubin and β -carotene. These chromophores are yellow/orange pigments. Bilirubin is usually responsible for the yellow skin colour seen in people with jaundice [22]. The spectra are taken from S. Prahl's compilation of PhotochemCAD data [23, 24].

$$\mu_{a,Bilirubin}(\lambda) = \frac{\epsilon_{bilirubin}}{585} \ln 10 C_{bilirubin} \quad (1.3)$$

$$\mu_{a,\beta-Caro}(\lambda) = \frac{\epsilon_{\beta-Caro}}{537} \ln 10 C_{\beta-Caro} \quad (1.4)$$

Where:

$\epsilon_{bilirubin}$ and $\epsilon_{\beta-Caro}$ are the excitation coefficients for bilirubin and β -carotene respectively [cm^{-1}];

585, and 537 are the molecular weights of bilirubin and β -carotene [g];

finally, $C_{bilirubin}$, and $C_{\beta-Caro}$ are the contaminations of bilirubin and β -carotene in the skin [$g L^{-1}$].

Melanin is the next chromophore we model. To model melanin's absorption coefficient we use Eq. (1.6), taken from [25].

$$\mu_{a,eumel}(\lambda) = 6.66 \times 10^{11} \times \lambda^{-3.33} \quad (1.5)$$

$$\mu_{a,phomel}(\lambda) = 2.9 \times 10^{15} \times \lambda^{-4.75} \quad (1.6)$$

Finally we use a base absorption coefficient to model the absorption due to the other parts of the skin that contribute to its optical properties, but individually do not have a large effect. The equation for modelling this was taken from I. Sahdi [26].

$$\mu_{a,base} = 7.84 \times 10^8 \times \lambda^{-3.255} \quad (1.7)$$

Figure 1.3 shows the absorption spectra for the above chromophores as a function of wavelength.

To create the five layer skin model we mix different amounts of the chromophores found in each layer, as described above. Equations (1.8) to (1.11) show the respective equations for the total absorption coefficient for each layer.

$$\mu_{a,strat} = ((0.1 - 0.3 \times 10^{-4} \cdot \lambda) + (0.125(\lambda/10.)) \times \mu_{a,b}(\lambda)) \times (1. - W) + W \cdot \mu_{H_2O}(\lambda) \quad (1.8)$$

$$\begin{aligned} \mu_{a,epi} = & (\nu_m \cdot (\mu_{phomel}(\lambda) + \mu_{eumel}(\lambda)) + (\mu_{a,b}(\lambda) + \ln 10 \cdot \mu_{a,\beta-caro}(\lambda) \cdot C_{caro}) \\ & \times (1. - \nu_m)) \times (1. - W) + W \cdot \mu_{H_2O}(\lambda) \end{aligned} \quad (1.9)$$

$$\begin{aligned} \mu_{a,pap/ret} = & ((S \cdot \mu_{a,oxy}(\lambda) + (1. - S) \cdot \mu_{a,deoxy}(\lambda) + \ln 10 \cdot \mu_{a,\beta-caro}(\lambda) \cdot C_{caro} + \\ & \ln 10 \cdot \mu_{a,bili}(\lambda) \cdot C_{bili}) \cdot B + (1. - B) \cdot \mu_{a,b}(\lambda)) \times (1. - W) + W \cdot \mu_{H_2O}(\lambda) \end{aligned} \quad (1.10)$$

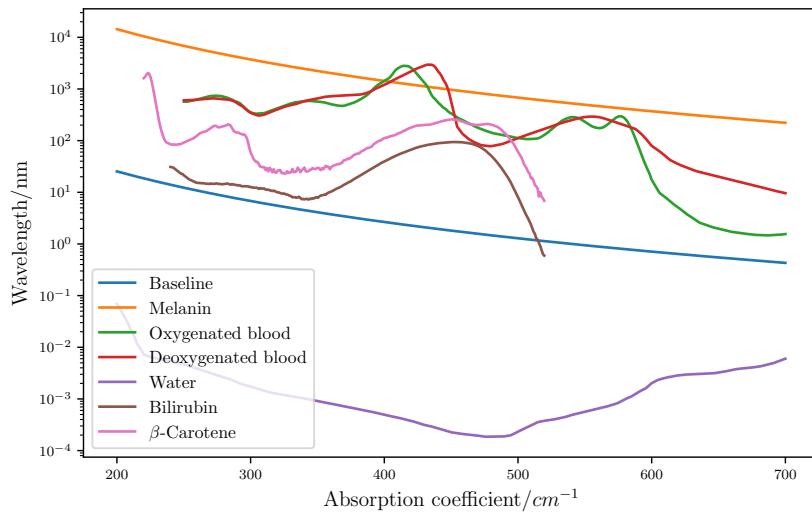


Figure 1.3: Absorption coefficients for the various chromophores found in skin.

$$\mu_{a,hypo} = ((S \cdot \mu_{a,oxy}(\lambda) + (1 - S) \cdot \mu_{a,deoxy}(\lambda)) \times B + \mu_{a,b}(\lambda) \cdot (1 - B)) \times (1 - W) + W \cdot \mu_{a,H_2O}(\lambda) \quad (1.11)$$

Where:

W is the volume fraction of water in a layer [-];

ν_m is the volume fraction of melanin in a layer [-];

B is the volume fraction of blood in each layer [-];

and all the other symbols have the same meanings as described above.

Table 1.1 shows the amount of each chromophore included in each layer, and the size adopted for each layer as well as the refractive index for each individual layer. Figure 1.4 shows the total absorption coefficient as a function of wavelength for the five layers.

Layer	Thickness/cm	Refractive index	Blood volume/%	Melanin volume/%	Bilirubin/gL ⁻¹	β -Carotene/gL ⁻¹	Water volume/%
Stratum Corneum	0.02	1.50	0.0	0.0	0.0	0.0	0.05
Epidermis	0.08	1.34	0.0	1.0	0.0	2.1e-4	20.0
Papillary Dermis	0.18	1.40	6.0	0.0	0.05	7e-5	50.0
Reticular Dermis	1.82	1.395	4.5	0.0	0.05	7e-5	70.0
Hypodermis	5.90	1.41	5.0	0.0	0.0	0.0	70.0

Table 1.1: Table of values used for the various concentrations and volumes fraction of the chromophores in the five layer skin model. Values taken from [25, 27–29].

With the absorption properties of the various chromophores in the skin defined we can now discuss the scattering properties of the skin. As the scattering properties do not vary from layer to layer by too much we use the same equation to describe the scattering coefficient [25, 30, 31].

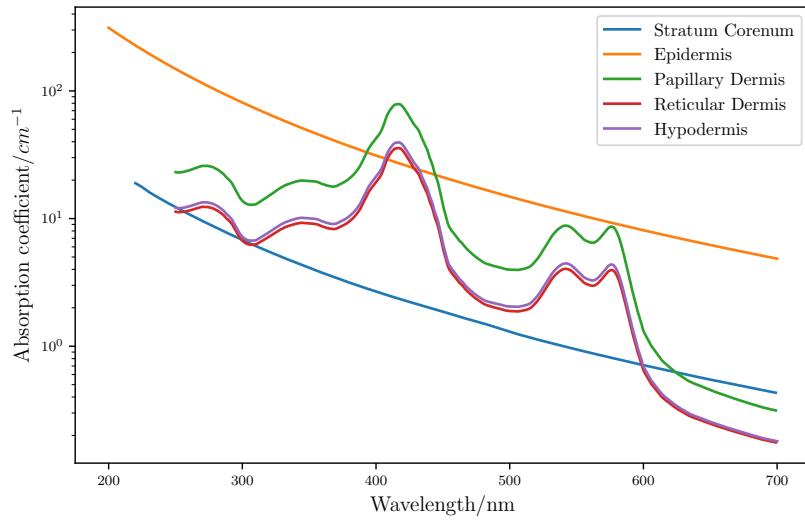


Figure 1.4: Absorption coefficients for the different layers in the skin.

$$\mu'_s(\lambda) = a' \left(f_{ray} \left(\frac{\lambda}{500(nm)} \right)^4 + (1 - f_{ray}) \left(\frac{\lambda}{500(nm)} \right)^{-b_{mie}} \right) \quad (1.12)$$

$$\mu'_s(\lambda) = 1050.60 \times \lambda^{-0.68} \quad (1.13)$$

Where:

μ'_s is the reduced scattering coefficient [cm^{-1}];

a' is a scaling factor [cm^{-1}];

f_{ray} is the fraction of Rayleigh scattering [-];

λ is the wavelength of light [m];

and b_{mie} is the “scattering power” [-].

This equation mixes both Mie and Rayleigh scattering into one equation. The first term represents the Rayleigh scattering terms, whilst the second represents the Mie scattering term. Figure 1.5 shows the reduced scattering coefficients for the different layers of the skin.

Layer	a'/cm^{-1}	f_{ray}	b_{mie}
Epidermis	66.7	0.29	0.69
Dermis	43.6	0.41	0.35

Table 1.2: Values of the constants for the scattering equations for the different layers of our skin model. Here epidermis represents the stratum corneum and the epidermis, and dermis represents the papillary, reticular and hypodermis in our model.

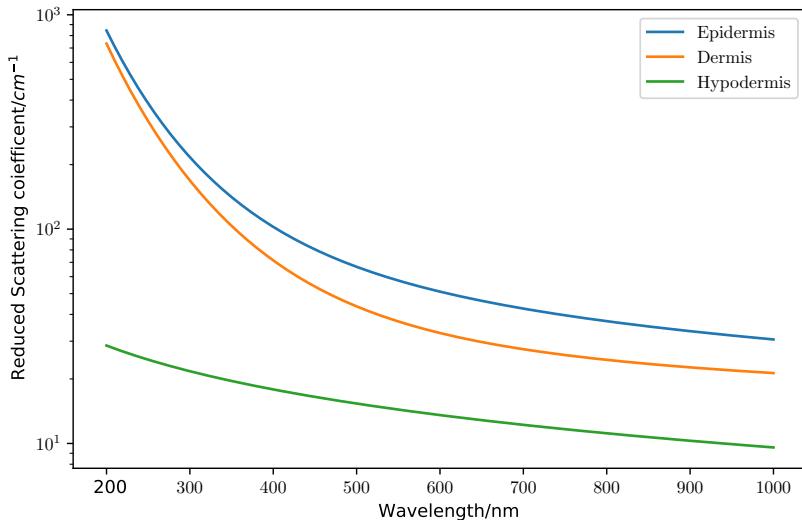


Figure 1.5: Figure shows the reduced scattering coefficient for the different layers of our skin model.

Finally we discuss the anisotropy values and refractive indices of our model.

First, the anisotropy g value is dependent on wavelength [31, 32].

$$g(\lambda) = 0.62 + 0.29\lambda \times 10^{-3} \quad (1.14)$$

Where:

g is the anisotropy value [—];

λ is the wavelength [nm].

The refractive indices of each layer is broadly the same and does not vary form layer to layer by a large amount. Table 1.3 shows the refractive indices adopted for our skin model.

Layer	Refractive index
Stratum corneum	1.5
Epidermis	1.34
Papillary dermis	1.40
Reticular dermis	1.395
Hypodermis	1.41

Table 1.3: Refractive indices used for the five layer skin model. Values are taken from [28].

As information on wavelength dependant refractive indices is not readily available, we assume that the refractive indices are the same for all wavelength used in this chapter.

Fluorophores in the Skin

The final optical property that makes up the five layer model is fluorescence. As mentioned in the introduction there are various molecules and proteins responsible for the autofluorescent repose of the skin. These include NADH (nicotinamide adenine dinucleotide), structural proteins like collagen and elastin, aromatic amino acids (tyrosine and tryptophan), porphyrins, and FAD

(flavin adenine nucleotide). Figure 1.6 shows the emission and absorption spectrum for these fluorophores. As NADH is found in cells, NADH is typically found in all layers of the skin. Elastin and collagen are only found in the dermis [33]. Tyrosine and tryptophan are found in all layers of the skin [33].

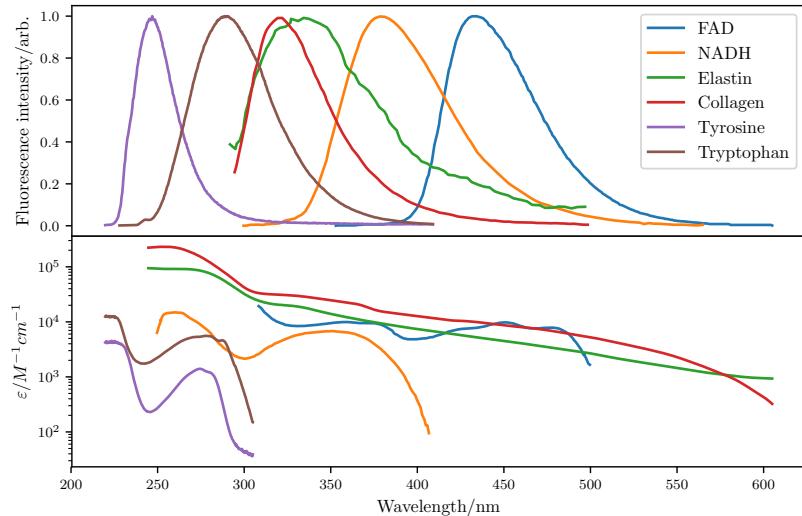


Figure 1.6: Top) Shows the fluorescent emission for the various different fluorophores. Bottom) Extinction coefficients for a selection fluorophores found in the skin [34–40].

1.3 Modelling Fluorescence

Fluorescence is the process in which light of a certain wavelength is incident on a molecule, the molecule absorbs the light and re-emits the light at a new longer wavelength. This process can be illustrated by a Jablonski diagram. A Jablonski diagram shows the electronic states of a molecule and the possible transitions between them. This allows the illustration of various radiative and nonradiative transitions that are possible for a give molecule, including fluorescence and phosphorescence. Figure 1.7 shows an example of Jablonski diagram for a molecule that absorbs light and re-emits it as fluorescence.

To model fluorescence from multiple fluorophores requires a change of the Monte Carlo radiation transfer (MCRT) code presented thus far. This change is to the interaction portion of the algorithm, so that it will now include the option for a packet to undergo fluorescence. To calculate whether a packet absorbs, scatters or fluoresces, first the probability of each of these events must be calculated. Discussion of scattering and absorption (by the bulk medium) was described in ???. To calculate the probability of fluorescence, we first assume that the quantum yield of the molecule is unity. This is physically unrealistic, however it does not affect the simulations accuracy, as modelling a realistic quantum yield would mean that more packets would be discarded, and thus the signal to noise ratio would be worse than if we assume a quantum yield of unity. To calculate the probability of fluorescence, the absorption coefficient of the fluorescent molecule must be calculated. This is shown in Eq. (1.15):

$$\mu_f = \ln(10) \epsilon C \quad (1.15)$$

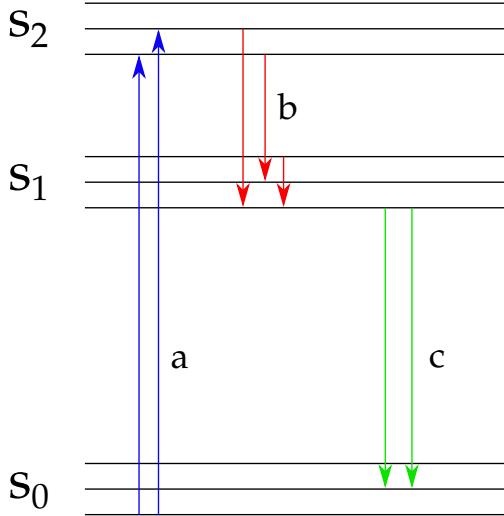


Figure 1.7: Jablonski diagram for PPIX. a) excitation of the ground state via absorption of a photon, b) non-radiative transition, and c) fluorescence.

Where C is the concentration of the fluorophore, ε is the extinction coefficient of the fluorophore, and $\ln(10)$ is the natural logarithm of 10^{\dagger} .

The next step is to calculate the total attenuation coefficient for a given species as in Eq. (1.16)

$$\mu_{t,i} = \mu_{s,i} + \mu_{a,i} + \mu_{f,i} \quad (1.16)$$

Where as usual μ_a and μ_s are the absorption and scattering coefficients, and μ_f is the fluorescence coefficient as defined in Eq. (1.15). As the absorption coefficient of fluorophores are small in comparison to the medium, and that the absorption coefficient of fluorescent molecules are generally much larger than that of their scattering coefficient, we assume that the scattering coefficient is negligible. Finally we calculate the probability of interacting with a given species using Eq. (1.17)

$$P_i = \frac{\mu_{t,i}}{\sum_{i=1}^N \mu_{t,i}} \quad (1.17)$$

Where P_i is the probability of interacting with the i^{th} species, the numerator is the attenuation coefficient for i^{th} species, and the denominator is the total attenuation coefficient for all the species.

Algorithm 1 shows the process used to determine which species to interact with.

[†]This factor appears as historically ε was measured in base 10 [30].

```

set  $\mu_{t_i}$ ;
set all  $P_i$ 's;
set  $\xi_1$ ;
if  $\xi_1 \leq P_1$  then
    set  $\xi_2$ ;
    if  $\xi_2 \leq a_m$  then
        | Scatter in medium;
    else
        | Absorb in medium;
    end
else if  $\xi_1 \leq P_1 + P_2$  then
    | Species 1 fluoresces;
else if  $\xi_1 \leq P_1 + P_2 + P_3$  then
    | Species 2 fluoresces;
else if  $\xi_1 \leq P_1 + P_2 + P_3 + \dots + P_n$  then
    | Species n fluoresces;
else
    | Error, no interaction;
end

```

Algorithm 1: An algorithm to determine which species to interact with. P_1 is the probability of interacting with the bulk medium, P_2 to P_n is the probability of interacting with a fluorescent species, a_m is the albedo of the bulk medium, ξ_i is a random number, and μ_{tot} is the total attenuation coefficient of all the species summed.

This method allows an arbitrary number of fluorophores to be modelled. To ensure that this method works as intended, the method is compared to experimental data taken by Campbell *et al.* [31].

The cuvette was filled with Intralipid 20% diluted with water, and a fluorescent agent Coproporphyrin III. The total volume of this mixture was 6 ml, consisting of 1 ml of Coproporphyrin III, 4.99 ml of water, and 0.01 ml of Intralipid 20%, of which 2 ml was pipetted into the cuvette. 405 nm light was shone on a cuvette of area 10 mm × 10 mm., and collected via the same fiber, of diameter 600 μm . The fiber has a NA of 0.22. The signal is recorded by a blah blah blah..

The simulation is setup to mimic the experimental setup. A medium of 10 mm^3 is used with 1 voxel to increase the speed of computation. As before Intralipid is assumed to be wholly scattering with no absorption, so an albedo of 1 is used. Conversely the Coproporphyrin III is wholly absorbing with no scattering. Coproporphyrin III absorption spectrum is as shown along side its emission spectrum in Fig. 1.8. If a photon packet leaves the top face of the simulated medium, within the radius of the fiber at an angle the fiber could accept, then the packet is recorded. The simulation is run with 10^7 photons which yielded Fig. 1.9.

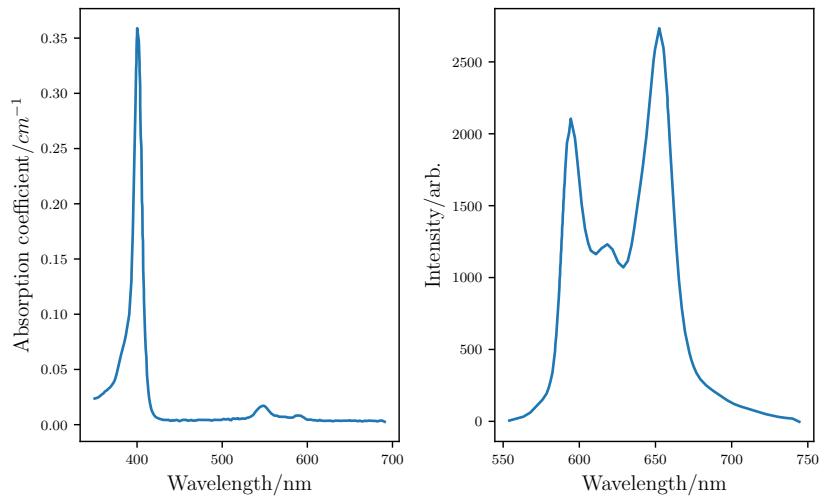


Figure 1.8: Optical properties of Coproporphyrin III. The figure on the left shows the absorption coefficient as a function of wavelength. The figure on the right shows the emission spectrum as a function of wavelength.

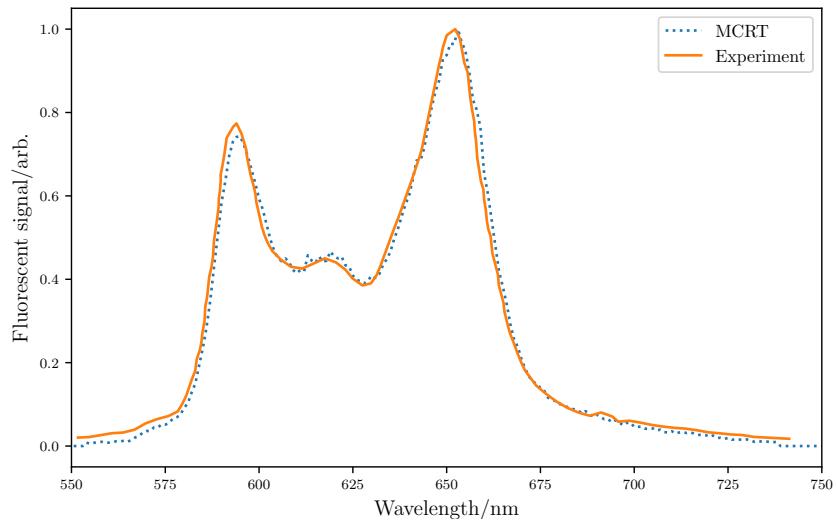


Figure 1.9: Validation of florescence modelling technique as described above. Figure shows that the MCRT method matches closely to the experimental results.

1.4 Nelder-Mead Method

The NM method is an algorithm for unconstrained optimisation. The algorithm is based upon iteratively updating a simplex. A simplex is a structure in *ndimensional* space, consisting of $n + 1$ points that are not in the same plane. Therefore in 1D, the simplex is a line, in 2D a triangle, in 3D a tetrahedron, etc. The Nelder-Mead method is a gradient free method, meaning that it does not require derivatives to be calculated and that the search space does not need to be smooth. This makes it ideal for problems where derivatives are not able to be computed easily, or the search space is not smooth. However, the NM method can also get stuck at local minima so care must be taken to avoid this. Genetic algorithms, which don't easily get stuck in local minima, were trialled but the computational cost of using them made it infeasible.

The NM algorithm works by removing the worst vertex of the simplex and replacing it with a “better” vertex calculated via a number of different operations. These operations can be seen in Fig. 1.10.

The first step of the NM method is to sort the initial vertices according to their fitness. For $n = 2$, we define x_w as the “worst” point, x_l and the “lousy” point, and x_b the “best” point, such that $f(x_b) \leq f(x_l) \leq f(x_w)$, where $f(x)$ is evaluating the ‘fitness’ of a point x . The fitness function varies from problem to problem, and usually takes the form of the function that is being optimised.

With the vertices sorted, the centroid of the simplex is calculated as in Eq. (1.18). The centroid is the mean of all the vertices bar the “worst” point.

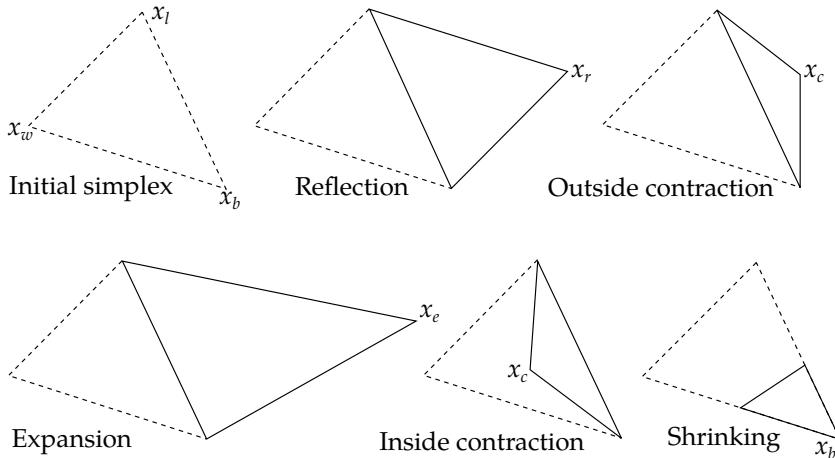


Figure 1.10: Operations that can be preformed on a simplex for $n = 2$.

The next step is to move the simplex via a reflection. To calculate the new vertex via reflection Eq. (1.19) is used, where α is the reflection factor. If this new point, x_r , is better[‡] than the current “best” point then we calculate a new point in the same direction but further using the expansion operation Eq. (1.20), where γ is the expansion factor. If this new point, x_e , is better than the “best” point then we replace x_w with x_e and start the process again. However, if x_e is not better than the “best” point, then we discard it and replace the worst point with x_r the reflected point.

[‡]Here better means the point has a lower fitness score

If when calculating x_r , we find that is worse than the “best” point, we then check if x_r is better than the ‘lousy’ point. If x_r is better than x_l then we replace the “worst” point and start the process again. However, if the x_r is worse than x_l , we then compare it to the “worst” point. If x_r is better than the “worst” point then we preform and inside contraction Eq. (1.22), where β is the contraction factor. If this new point, x_{ic} , is better than the “worst” point then we keep it, otherwise we preform the shrink operation, shrinking the whole simplex around the “best” point.

If x_r is not worse than the “worst” point then we preform an outside contraction Eq. (1.21). This computes a new point x_{oc} . If x_{oc} is better than x_w , then we keep it, otherwise again we shrink around the “best” point. The process described above is summarised in Fig. 1.11.

Standard values for the factors are: $\alpha = 1$, $\beta = \frac{1}{2}$, and $\gamma = 2$. Though in practice these values are adjusted for the problem at hand. For higher dimensions, i.e. where $n > 2$ F. Gao *et al.* suggest that the parameters should be changed based upon how many dimensions are used for the simplex [41]. The values F. Gao *et al* suggest are: $\alpha = 1$, $\beta = 1 + \frac{2}{n}$, $\gamma = 0.75 - \frac{1}{2n}$, and $\delta = 1 - \frac{1}{n}$. Where n is the order of dimensions.

$$c = \frac{1}{n} \sum_{i=1, i \neq w}^{n+1} x_i \quad (1.18)$$

$$x_r = c + \alpha(c - x_w) \quad (1.19)$$

$$x_e = c + \gamma(x_r - c) \quad (1.20)$$

$$x_{oc} = c + \beta(x_r - c) \quad (1.21)$$

$$x_{ic} = c + \beta(x_w - c) \quad (1.22)$$

As the Nelder-Mead method has no inbuilt convergence criteria, this must be added. We use two different criteria based upon simplex size, and vertex fitness. The criteria for the simplex size is as; The size of the simplex is calculated using Eq. (1.23):

$$\text{size} = \sum_{i=1}^{n+1} |p_i - p_{i+1}| \quad (1.23)$$

Where p_i and p_{i+1} are vertices in the simplex that are connected by an edge. If the size of the simplex falls below a pre-set value, then we preform a factorial test to see if the simplex should be restarted or if the algorithm should terminate. The factorial test checks the space around the current simplex to ensure that we have converged to a global minima. If the check fails then the algorithm is restarted with the current best point kept, and new vertices generated.

The other convergence criteria is a check to see if the best point is “good enough”. The current best point is compared to a pre-set fitness value. If the best point is better than the pre-set value then the algorithm terminates.

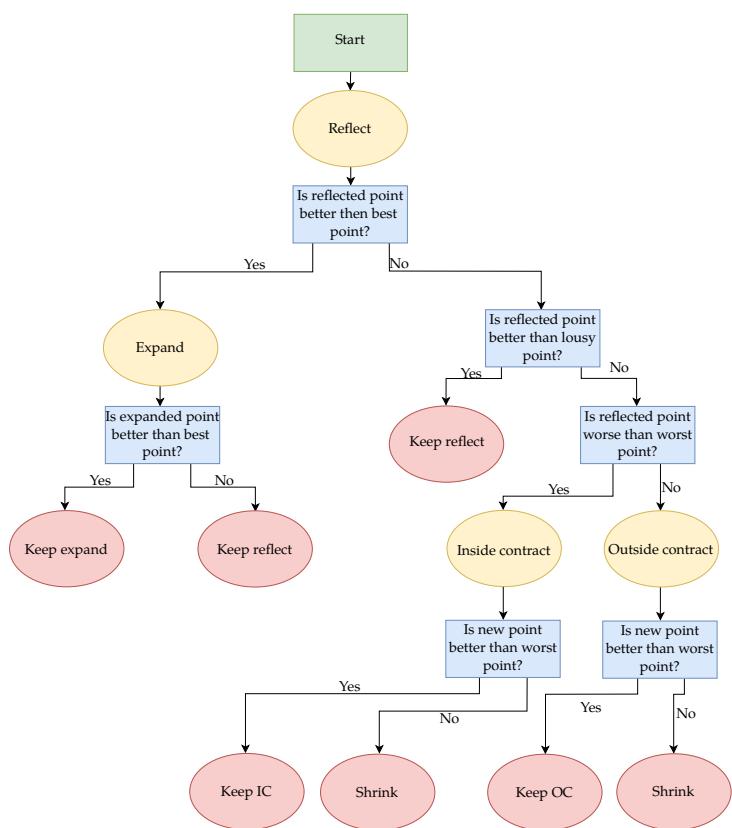


Figure 1.11: Nelder-Mead decision tree.

1.5 Validation

The NM method was coded in modern Fortran, so that it could be easily interfaced with the MCRT code developed as part of this thesis. To test that the NM method works as intended a number of trial optimisation functions were tested, see Table 1.4. This was achieved by selecting an initial simplex, and the method allowed to iterate until it converged. The results of this are shown in Fig. 1.12.

Name	Formula	Global Minimum
Sphere	$x^2 + y^2$	$f(0, 0) = 0.$
Rosenbrock	$(a - x)^2 + b(y - x^2)^2$	$f(1, 1) = 0.$
Ackely	$-20 \exp \left[-0.2\sqrt{0.5(x^2 + y^2)} \right] - \exp [0.5(\cos 2\pi x + \cos 2\pi y)] + e + 20$	$f(0, 0) = 0.$
Himmelblau's	$(x^2 + y - 11)^2 + (x + y^2 - 7)^2$	$f(3, 2) = 0.,$ $f(-2.805118, 3.131312) = 0.,$ $f(-3.779310, -3.283186) = 0.,$ $f(3.584428, -1.848126) = 0.$

Table 1.4: Table of standard test functions for numerical optimisation.

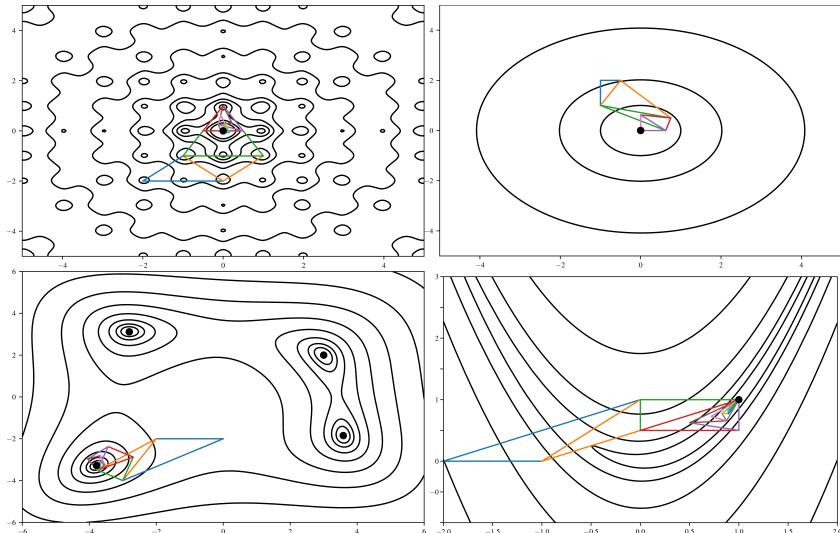


Figure 1.12: Contour plots of test functions with Nelder-Mead simplexes over plotted. Top left is the Ackely function, top right is the sphere function, bottom left is the Himmelblau's function, and the bottom right is the Rosenbrock function. Blue simplex is the initial simplex, and the large black dots represent the Global minima.

Some of these functions (Sphere, and Rosenbrock's) can also be extended to arbitrary dimensions. These functions were used to check that the NM method works as intended in these higher dimensions where the NM method will primarily be used in this thesis.

To ensure that the NM method can be used to find the unknown concentrations of the autofluorophores, we test the method with a two different toy models. The first model is a 2D model, with two different fluorophores evenly distributed over all 5 layers of our skin model. This

two different fluorophores are NADH, and a fictitious fluorophore that has similar properties to FAD and tryptophan, such that the excitation spectrum is that of NADH and the emission spectrum is that of tryptophan. The concentration in these layers is such that the bulk optical properties are not affected: NADH has a concentration of $1.0 \mu M$, and the fictitious fluorophore has a concentration of $2.5 \mu M$. To generate a spectrum to which the NM method can compare to, the MCRT code is run with the above configuration of fluorophores. This generated Fig. 1.13.

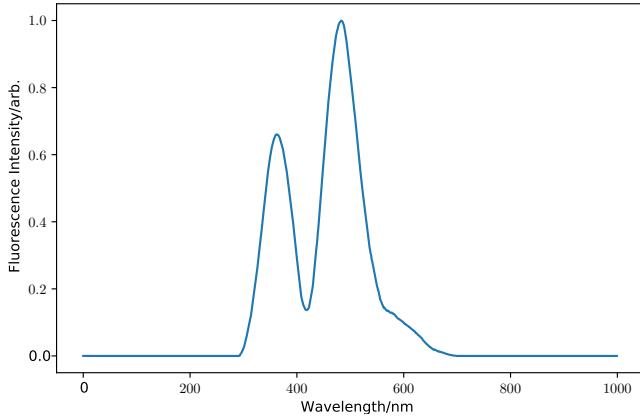


Figure 1.13: Example of toy model spectrum for testing NM method. The two peaks correspond to the fictitious fluorophore, and NADH.

The fitness function chosen to check whether the NM method is converging to the generated spectrum is as:

$$\text{fitness} = \sum_{i=1}^n (x_i - m_i)^2 \quad (1.24)$$

Where x_i is a data point at a wavelength λ_i produced by the MCRT, and m_i is a data point in the model at a wavelength λ_i .

As the NM can get stuck in local minima, we run the method for several different initial simplices to ensure that the method does not get stuck in a local minima. As many calls to the MCRT code need to be done, and the fluorophore concentration is low this means that many packets need to be run to achieve a good single to noise ratio. These two conditions result in a computational load that is infeasible to run. Therefore the MCRT algorithm has to be computationally efficient in order to arrive at an answer within a reasonable time. To this end the 3D skin model is shrunk to a 1D model in the z direction so that the optical integration routine can efficiently move the packet through the simulated medium. The optical properties of the incident wavelength are also stored so that when a new packet is started the optical properties can easily be adjusted without need for any calculation. Finally a filter is employed on the output fluorescence spectrum to smooth the noise out. The filter used is a Savitzky-Golay filter. This filter fits multiple low-degree polynomials to subsets of the data, thus smoothing the data [42]. Figure 1.14 shows the use of the Savitzky-Golay filter on sample output data from the MCRT simulation.

Using the above set-up with the Savitzky-Golay filter on the output spectrum, allowed the NM method to efficiently run many model of various different concentrations and “find” a set of concentration that resulted in a close match with the target spectrum. However, as the detected fluorescence spectra are normalised to their peak values we cannot use this method to determine

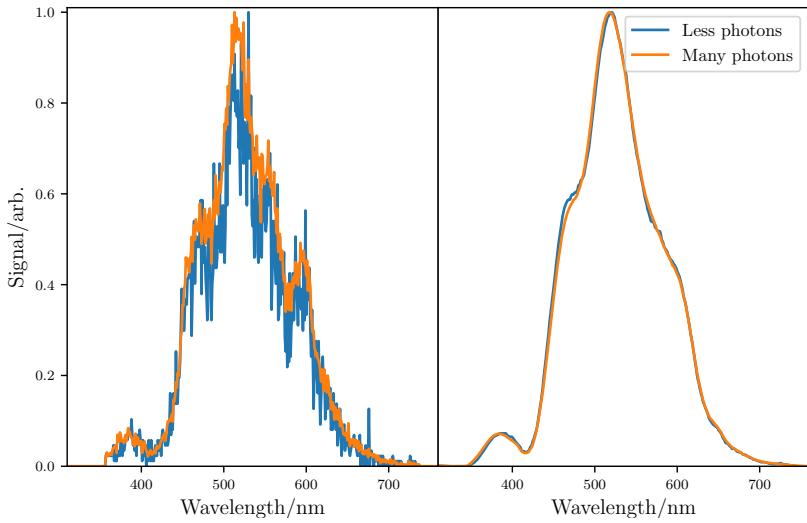


Figure 1.14: Illustration of how the Savitzky-Golay filter works on noisy data and recovers the roughly the same signal on the same dataset with less noise. Left image shows the raw signal for a simulation run with less photons and more photons. Right image shows the dataset after the Savitzky-Golay filter is applied.

the original concentration, but rather the ratio between the two concentration. Figure 1.15 shows the search space and the spread of the concentration values calculated by the NM method compared to the original target concentration. The spread of the concentrations calculated by the NM method follow a linear relationship as would be expected. Therefore a line of best fit was fitted to the concentrations. This yielded a line ($y = m x$) with $m = 2.49 \pm 0.05$. The expected relation between the concentration is where $m = 2.5$ therefore, the NM method can be used to determine the relative difference in concentrations within one standard deviation.

The NM method was also tested on a toy model for $n = 3$. Here the fluorophores used were: NADH, FAD, and a fictitious fluorophore with the absorption properties of NADH and the emission properties of Tyrosine. The fluorophores had concentrations of $1.05\mu M$, $525\mu M$, and $125\mu M$ respectively. The set-up is the same for the above $n = 2$ case, with the same filter and computational speed ups used. Figure 1.16 shows various concentrations as calculated by the NM method compared to the original concentration. Again a line of best fit was fitted to the calculated points [§]. As there is no closed form solution for the equation of a line in $n > 2$ dimensions, the equation of a line in 3D can be represented by Eq. (1.25):

$$\vec{r} = \vec{r}_0 + t \vec{v} \quad (1.25)$$

Where \vec{r} and \vec{r}_0 are position vectors for P and P_0 respectively, \vec{v} is a vector parallel to the line we are examining, and t is some real number. This equation also holds for an arbitrary number of dimensions. The line of best fit yielded $\vec{v} = [0.0013, 0.9858, 0.1682]$. Where as the “real” \vec{v} is equal to $[0.0019, 0.9728, 0.2316]$. This again shows that the NM method can be used to find the relative concentrations of the fluorophores in the skin, even in dimensions higher than 2.

[§]The line of best fit was calculated using the singular value decomposition (SVD) method of solving the least squares.

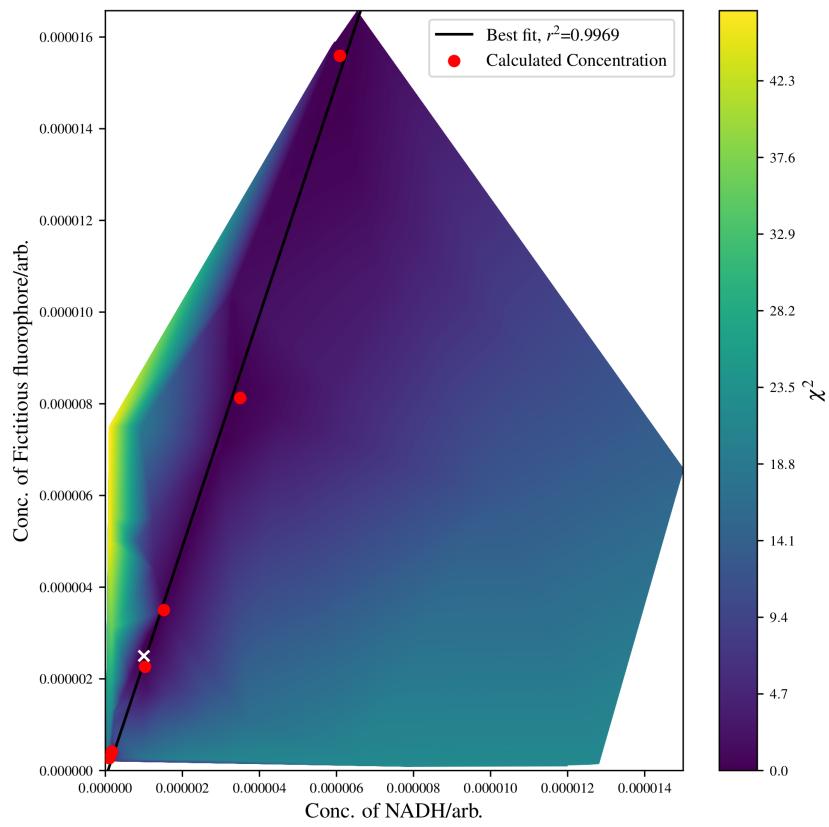


Figure 1.15: Figure shows the search space for the 2D toy problem outlined above. A line of best fit is fitted to the concentrations found by the NM method. Note also the valley of good fit where the line of best fit lies. The search space is also fairly smooth.

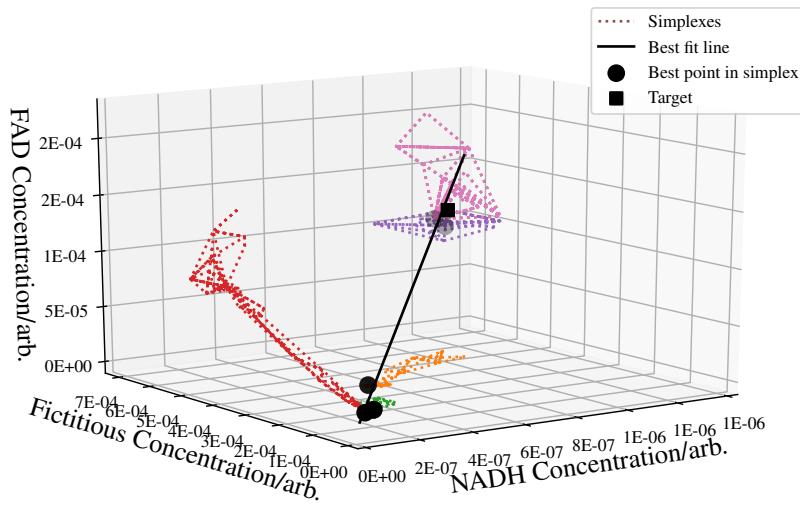


Figure 1.16: Figure shows the line of best fit for the case where $n = 3$. Figure also shows the simplices path over their whole lifetime, from initial guess to final simplex.

1.6 Results

1.6.1 Experimental Work

Experimental work discussed in this section was carried out by my collaborators S. Smirni *et al.* at the University of Dundee and Ninewells Hospital.

Autofluorescence spectra were taken from the volar forearms of volunteers using the lasima laser thing majig.

Spectra were taken before inflating a cuff, during inflation and after deflation. This is known as the PORH test

optical fiber delivers light and detects light blah blah pic of geometry?

1.6.2 Effect of Tissue Optics on Fluorescent Signal

As mentioned in the introduction to this chapter information about how tissue optics affects the fluorescent signal, which fluorophores contribute to the signal, the variability of the signal in different test sites, and location of the fluorescent signal are not well elucidated. Therefore, before running the NM method on the experimental data, the fluence of the input fluorescence light is analysed alongside the location as a function of depth of the fluorescent light. We also present results on how the excitation wavelength affects the signal.

Figure 1.17 shows the fluence as a function of depth for the incident UV light. The figure shows that most of the incident light is contained within the top three layers, with little getting to the Reticular dermis, with none reaching the Hypodermis. get measuerments of 50per and 10per as function of depth.

Figure 1.18 shows the fluence of detected fluorescent light (see ?? for discussion of how this is tracked.). The figure shows the fluence of light for each fluorophore,***add in elastin*** that is detected. The figure shows that the fluence is highest in the Papillary dermis and a number of peaks at the boundaries of the layers. ***show zoom in of top 4 layers*** First the refractive indices of the layers are different, this can lead to light getting “trapped” in a layer as it maybe

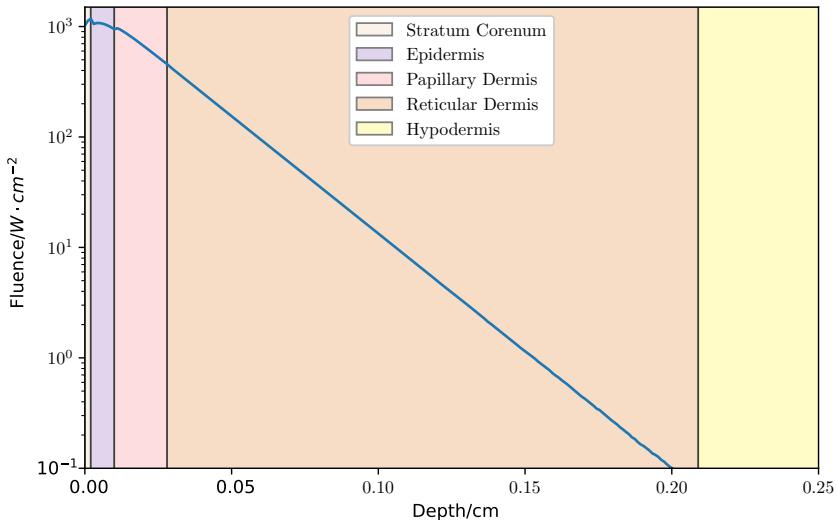


Figure 1.17: Penetration of UV radiation as a function of depth.

reflected off the layer boundary, thus leading to increased fluence. Second, fluorescent light is emitted isotropically which means that fluorescent light emitted in the upper layers of the skin, may be emitted in the direction of the Papillary dermis, that and light emitted from below the papillary dermis has to travel though the papillary dermis in order to be detected. Finally the optical properties also have an effect on the detected light fluence. The dermal layers have a minimum in their absorption spectra at the range of wavelengths that NADH and FAD emit at, thus light is more likely to “survive” though the dermal layers than the other layers such as the epidermis. Finally the geometry of the layers most likely has an effect as well. The stratum corneum’s and the epidermis’s thickness is small in comparison to that of the dermal layers, thus there will be less fluorescent light emitted from these layers when the concentration are the same.

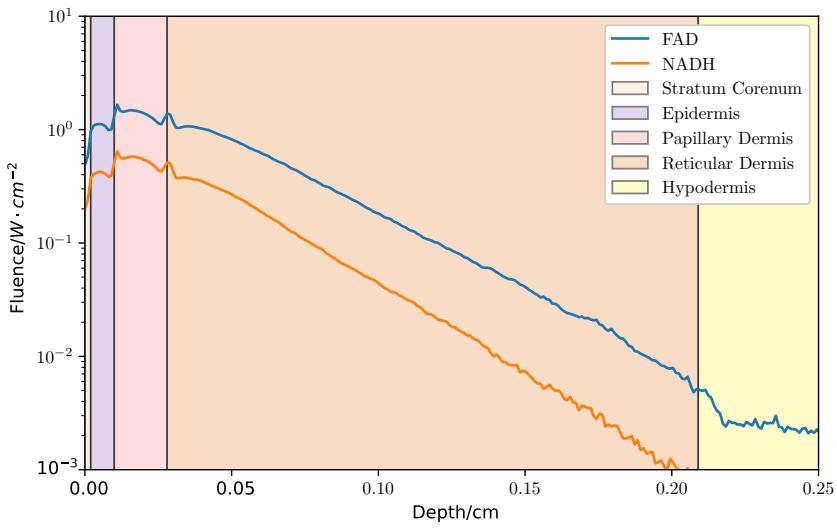


Figure 1.18: Detected fluence for FAD and NADH fluorescence.

Figure 1.19 shows the location of where the fluorescent light is emitted from. For both NADH, and FAD there peaks depth is just inside the epidermis. However, the vast majority of emitted Fluorescent light originates in the papillary dermis. This occurs as enough light gets to this layer, which allows more light to undergo fluorescence.

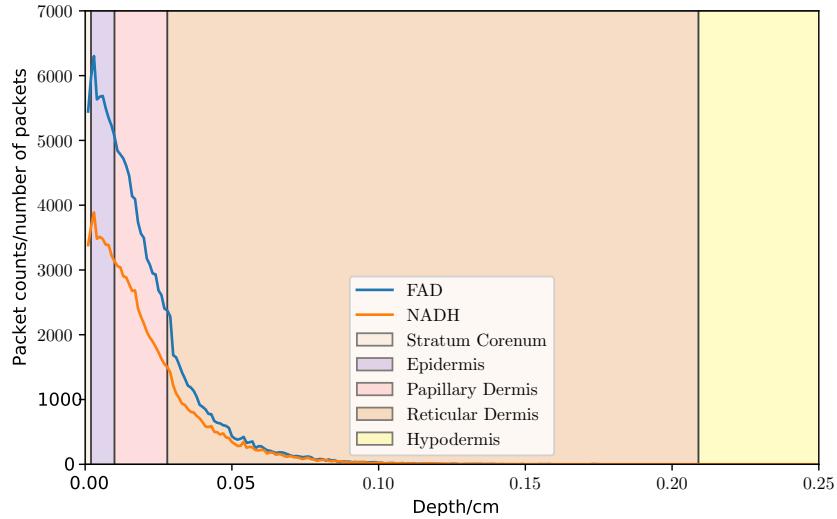


Figure 1.19: Amount of packets escaping as a function of depth for FAD and NADH fluorescence.

Finally, we analyse the effect of excitation wavelength has on the detected signal of NADH and FAD. To achieve this we run several models in order to create an excitation emission matrix (EEM). A EEM is a way of investigating and displaying the effect of excitation wavelength has on the fluorescent output of a sample. Using the same setup as above, we vary the fluorophore concentration in each layer, such that only one layer at a time has any fluorophores in it. We then excite the model over a range of wavelengths. For NADH we use wavelength in the range of 250 to 400 nm, and for FAD we use 250 to 500 nm. Figures 1.21 and 1.22 show the EEM for NADH and FAD. The EEM for NADH shows that the strongest signal comes from the papillary dermis when compared to other layers. The EEM also shows a maximum of emission for excitation wavelengths corresponding to around 260 nm and 320-380 nm. For FAD the layer that yields the most fluorescence is the reticular dermis. The EEM shows that for excitation wavelengths of around the range 450-500 nm gives the most fluorescence.

For the case of NADH the optimal excitation wavelengths coincide with maximums in the absorption spectrum of NADH (see Fig. 1.20). However, for FAD the maximum in fluorescence does not coincide with the largest maxima in its absorption spectrum (see Fig. 1.20), but rather it falls within a smaller maxima. This is due to the fact that light at the maximal absorption peak (~ 300 nm) is highly absorbed by tissue, especially in layers such as the epidermis. Therefore, light at a longer wavelength is more likely to escape the tissue due to the tissue optical properties.

do literature search to see where people measure these fluorophores

1.6.3 Using the NM method

As before the simulation was setup to match the above experimental procedure.

Figure 1.23 show the output of the NM method during a run.

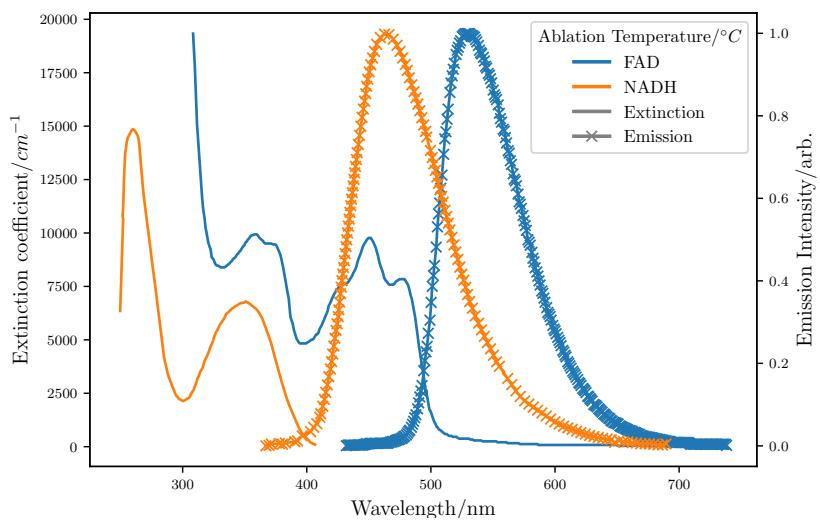


Figure 1.20: NADH and FAD absorption and emission spectra.

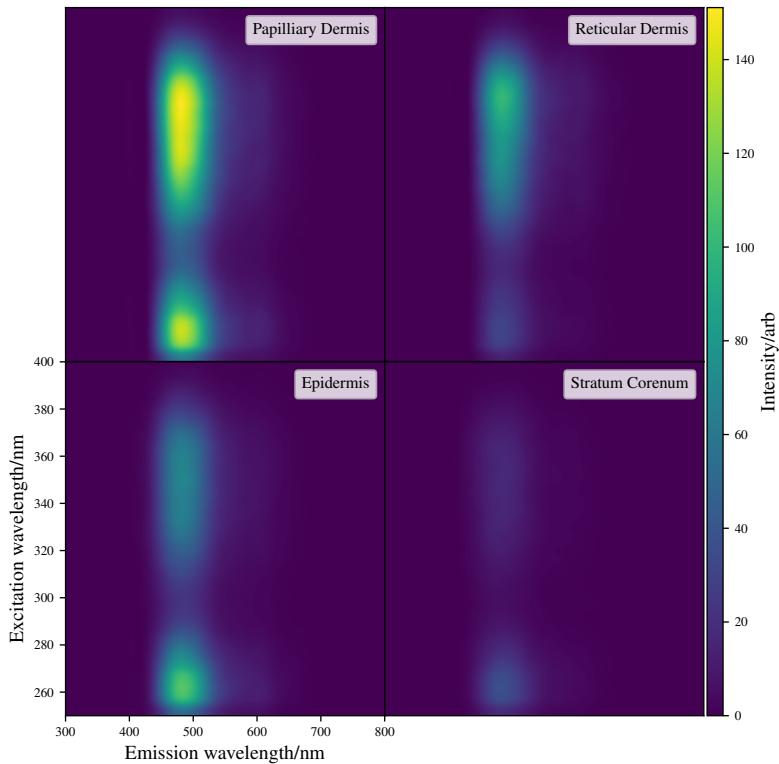


Figure 1.21: Excitation-emission matrix for NADH. Figure shows that the fluorescent signal from NADH is strongest in the papillary dermis.

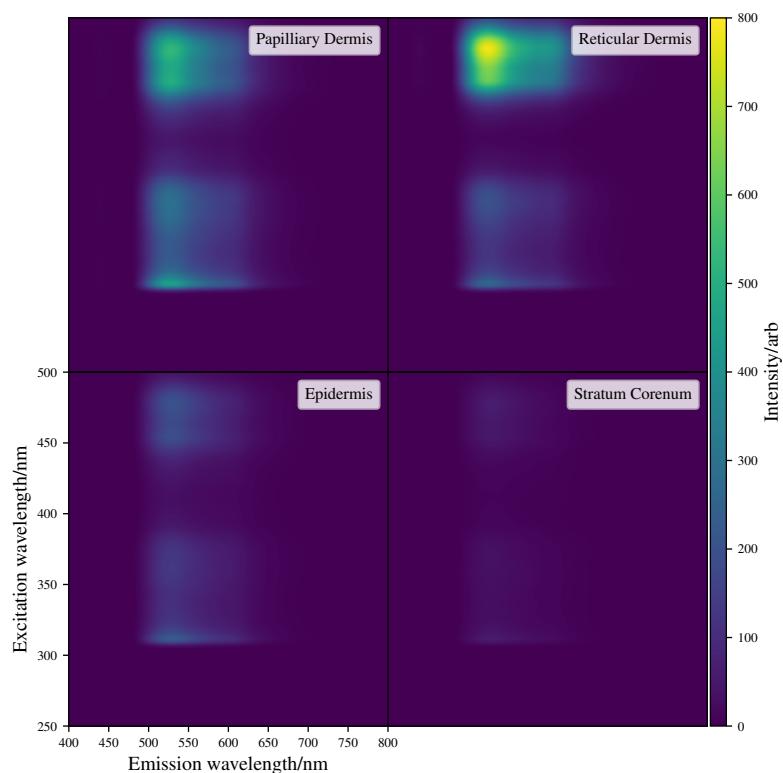


Figure 1.22: Excitation-emission matrix for FAD. Figure shows that the fluorescent signal from FAD is strongest in the reticular dermis.

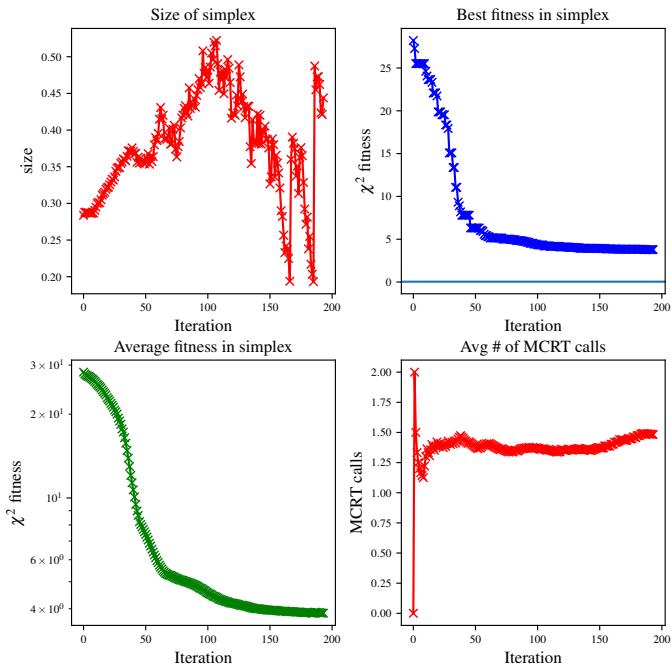


Figure 1.23: Figure shows the diagnostic information output by the NM method. Top left) shows the size of the simplex as it evolves. Top right) shows the best point in the simplex as it evolves. Bottom left) shows the average fitness of the simplex. Bottom right) shows the number of calls to the MCRT code per iteration of the simplex. The more call to the MCRT the simplex requires the higher the run time.

1.7 Discussion

1.8 Conclusion

We have presented our code, AmoebaMCRT, which combines the Nelder-Mead method and MCRT in order to determine the concentrations of naturally occurring fluorophores in human skin.

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