RESEARCH

A fast parallel algorithm to reduce protein folding trajectories

Luis Garreta^{1†}, Mauricio Martinez² and Pedro A Moreno^{1*}

Abstract

Background: The simulations are one of the most important tools for studying and understanding the underlying mechanisms of the protein folding process. Protein folding simulations have experienced substantial progress in the last years, they are performed using diverse technologies and they are reaching the microseconds and greater timescales, which generates very long trajectories. As a result, the analysis of these trajectories entails to complications and is necessary to create tools to simplify them, so that both the main events and the temporal order in which they occur are preserved.

Results: We present an algorithm to reduce long protein folding trajectories in a fast and parallel way. The algorithm divides a trajectory into segments to be processed in parallel, and from each segment selects the most representative conformations using a rapid clustering strategy, which takes advantage of the temporal order of the conformations to compare them locally, avoiding an all-versus-all comparison. The algorithm reduces a trajectory in a high percentage, preserving both the patterns and the structure obtained by other more complex reduction techniques. In addition, its performance is close to that shown by other efficient reduction techniques, and this performance is improved when executed in parallel using more than one core.

Conclusions: The developed algorithm quickly reduces a protein folding trajectory by selecting its most representative conformations and thus preserving both its structure and its temporal order. The reduced trajectories can be used as input for more complex analysis techniques and even for other reduction techniques that become impractical when faced with long folding trajectories. The algorithm is fast and is designed to run in parallel on conventional PCs with multi-core technology, which are present in most typical research laboratories.

Keywords: Protein folding simulations; Protein structure comparison; Protein structure clustering

Background

We present a parallel algorithm to reduce protein folding trajectories which quickly obtains representative conformations, conserving both their three-dimensional structure (3D) and their temporal order. Proteins play a fundamental role in all living beings, but to be functional, they must fold from their linear amino acid (AA) sequence to a unique 3D or native state, which is known as the protein folding process. Understanding the mechanisms and rules of this process has been one of the most pursued objectives of computational biology, and an important theoretical tool to study it has been the simulations of protein

*Correspondence: pedro.moreno@correounivalle.edu.co

 $\mbox{{\bf Full}}$ list of author information is available at the end of the article

folding. These simulations generate folding trajectories (Figure 1), which describe the sequence of states that proteins follow as a function of time during their folding process.

Folding simulations mainly use the molecular dynamics (DM) method, which due to its computational cost is limited to small proteins (<100 AA) and very short times (picoseconds or microseconds). However, technological innovations have allowed significant advances in these simulations, both on time scales and technology to execute them. In 2011, using the Anton supercomputer, specially designed for protein folding [1], full simulations of 12 proteins were published, several on the order of milliseconds [2]. And more recently, in 2016, the Anton 2 supercomputer became operational [3], being up to ten times faster than its predecessor Anton. As an economic alternative, in 2014 graphic processing units (GPU) were used to

¹ Escuela de Ingeniería de Sistemas y Computación, Universidad del Valle, Santiago de Cali, Colombia

[†]Equal contributor

Garreta et al. Page 2 of 8

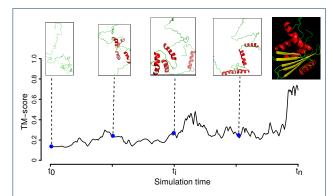


Figure 1 Protein folding trajectory. The evolution of the folding is measured at each step by comparing the structure at time t_i to the native structure (black image background) using a structure comparison metric, in this work we use the TM-Score (see Methods). X-axis: Simulation time from t_0 to t_n . Y-axis: TM-score value, from 0 (different) to 1 (similar).

simulate, on the order of microseconds, the folding of 17 proteins [4]. And years earlier, in 2007, the "folding@home" distributed computing platform utilized as many as 250,000 PCs, voluntarily available around the world, to simulate on the order of microseconds the folding of the villin-headpiece protein [5].

These innovations show significant progress in protein folding simulations, both on time scales and technology to execute them, and as a result the generation of trajectories with millions of conformations. But due to their large number of conformations, their processing and analysis in conventional PCs is computationally expensive, and new algorithms are needed to efficiently simplify them, seeking to preserve as much information as possible.

Two approaches used to reduce these simulations have been the dimensionality reduction [6] and clustering [7]. In the dimensionality reduction approach, conformations are transformed into reduced sets of variables that represent them as well as possible. Here, both linear and non-linear techniques have been used (e.g. principal component analysis (PCA) and multidimensional scaling [8], Isomap [9], diffusion maps [10]). However, many of these techniques, instead of reducing a trajectory, analyze it, losing the structural information of the conformations (Figure 2, top) and making the results explainable only when observed together. In addition, many of these techniques require pairwise comparisons, which are computationally expensive when trajectories are very large.

In the clustering approach, the conformations are assigned to groups that share similar characteristics (e.g., similarity with the native structure), and from each group an average representative or its general characteristics can be taken. Here, hierarchical and

partitioned groupings have been used (e.g., k-means [11], link [12]). However, the groups lose their temporal order since they can include conformations that occur in very distant times (Figure 2, bottom). And also they require pairwise comparisons, which are computationally expensive when trajectories are very large.

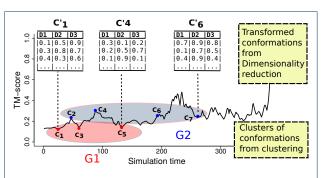


Figure 2 Loss of information in dimensionality reduction and clustering. The dimensionality reduction (above) transforms the conformations (c1, c2 and c3) into a new set of values (c'1, c'2 and, c'3), but their structural information is lost. While the clustering (bottom) forms two groups (G1 and G2), but their temporal order is lost since they contain conformations that overlap in time (c2, c3, c4, c5).

To reduce a folding trajectory, the proposed algorithm divides the path into segments that are processed in parallel. For each segment, characteristic events are quickly extracted using the rapid clustering strategy of Hobohm and Sander (1992) adapted for protein folding trajectories; and from these results, the most representative events are selected by a strategy of k-medoids [13]. The results of each segment are joined to form the reduced trajectory with the most representative conformations of the original trajectory, while retaining both its 3D representation as their temporal order.

The algorithm is implemented in the R language, except the function for pairwise structure comparison, the TM-score [14], which is the function executed more times and that is implemented in the Fortran language.

Methods

Data sets of protein folding trajectories

In this work, we used the trajectories from three protein folding simulations projects: Anton supercomputer [2] and folding@home [15] using Molecular Dynamics [15], and Parasol folding server using the Probabilistic Roadmap Method [16]. One trajectory was taken from each: the trajectory of the trp-cage

Garreta et al. Page 3 of 8

protein from the Anton supercomputer project, with a simulation time of 208 μs , a 200 ps time step, and 1044001 conformations. The trajectory of the villin-headpiece protein from the folding@home projec, simulated, with a simulation time of 8 μs , a 50 ps time step, and 15201 conformations. And the trajectory of the ribonuclease H protein from the Parasol folding server project, with 429 folding steps each corresponding to 429 conformations.

A time step in MD trajectories is the time length at which conformations are sampled or evaluated during the simulation. While a folding step, in the PRM and in our reduced trajectories, represents the most likely conformation occurring during a time interval or from a set of likely candidate conformations.

Pairwise comparison of protein structures using the TM-score

In this work, we used the TM-score metric for pairwise comparison of protein structures [14]. This metric is used in both the proposed algorithm and in the thecniques for reduction of protein folding used to compare its results. The TM-score is more sensitive to the global topology than local variations, and so it estimates the pairwise similarity of protein structures much more accurately than the Root Mean Square-Deviation (RMSD), a common metric used for the same purpose. The TM-score ranges from 0 to 1, where 1 is a perfect match. Based on statistics [17], a Tm-score lower than 0.17 indicates two random structures with no relation of similarity, and a Tm-score higher than 0.5 indicates that the structures have a degree of similarity that is not given by chance.

Techniques for reduction of protein folding trajectories

To compare the results of the proposed algorithm we used three common techniques for protein folding reduction [11]: a non-metric multidimensional scaling method (nMDS), principal component analysis (PCA), and a complete-linkage clustering.

The nMDS reduction was carried out using the R-function monoMDS [18], taking as input the dissimilarity matrix obtained from the pairwise comparison of all the protein conformations of the folding trajectory. The PCA reduction was carried out using the R-function pca.xyz [19], taking as input a matrix with the 3D coordinates of the $C\alpha$ atoms of all the protein conformations of the folding trajectory. And the complete-linkage clustering reduction was carried out using the R-function hclust [20], taking as input a matrix with the first two principal components from the previous PCA reduction.

Implementation

The proposed algorithm reduces a trajectory of protein folding in three steps: partitioning, extraction, and selection. The first step runs only once, while the other two runs several times independently, allowing them to run in parallel. Each step involves a strategy to improve the efficiency of the algorithm when working with large protein folding trajectories. Figure 3 shows the overview of the algorithm and the steps are given below.

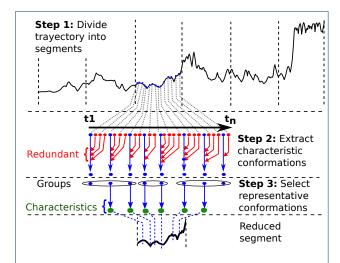


Figure 3 Overview of the proposed algorithm. Step 1: Divide the trajectory into segments (vertical dashed lines) and reduce each individually. Each segment has n conformations from time t_1 to t_n . Step 2: For each segment, extract characteristic conformations (blue dots) and remove redundant ones (red dots). Stage 3: Select the most representatives from the previous conformations.

Step 1: Partitioning

Divide the trajectory into segments to reduce them locally and in parallel (dotted vertical lines, Figure 3). This is carried out by dividing the trajectory from the start to the end in segments with N conformations each, where N is an input parameter. Local reductions allow to focus on the particular characteristics of each segment that will determine the global characteristics of the trajectory. And parallel reductions allow to improve the algorithm efficiency when it runs on machines with more than one processor (e.g. multi-core computers) (Figura 4).

Stage 2: Extraction

Quickly extract the characteristic conformations of each segment and eliminate the redundant ones. This

Garreta et al. Page 4 of 8

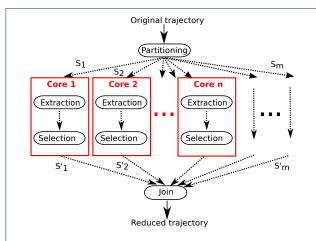


Figure 4 Parallel processing of trajectory segments. The trajectory is divided into m segments (S1, ..., Sm), and each segment is processed separately and in parallel on a machine with n processing cores. The results of the extraction and selection steps (S1', ..., Sm') are independent of each other and join to form the reduced trajectory.

is carried out efficiently by means of a rapid clustering approach that performs relatively few pairwise comparisons and, instead of grouping similar conformations of a segment, extracts the most dissimilar ones.

Here, we improved the fast clustering algorithm of Hobohm and Sander (1992) to work with a trajectory segment and exploit the implicit order of its conformations given by its simulation time (black horizontal line, Figure 3). The algorithm selects the initial conformation at time t_1 as the first characteristic. Then, the algorithm compares the previous characteristic with the following conformation. If dissimilar, then the conformation becomes a new characteristic, otherwise, the conformation is redundant and is removed (red dots, Figure 3). The process continues with the rest of conformations until finishing in the final one at time t_n , thus producing the set of representative characteristics of the segment (green dots, Figure 3)

Step 3: Selection

Take the conformations of previously extracted characteristics and cluster them to select the most representative characteristics. To find these representatives, the algorithm uses a k-medoids strategy (PAM algorithm [13]) that calculates the k conformations (medoids) whose average difference between all the other members of the group is minimal.

However, the PAM algorithm needs as input the dissimilarity matrix with the pairwise comparison of allversus-all conformations of the trajectory segment, which is an intensive computational task when the number of conformations is very large. But, this task

is feasible to perform since the algorithm is working here with a reduced set of characteristic conformations (previous step) of a trajectory segment and not of the complete trajectory.

Results and Discussion

To demonstrate the ability and performance of our algorithm in reducing protein folding trayectories, we carried out three tests: (1) Reduction of protein folding trajectories, (2) Comparison with other reduction techniques, and (3) Performance comparisons.

Reduction of three protein folding trajectories

We tested our algorithm on three folding trajectories from different simulation projects (see Methods). The figure 5 shows the results of these reductions, the details of the proteins are in the box on the left and both the original and reduced trajectories are in the box on the right.

Como se puede observar de la figura 5, los resultados de las reducciones son conformaciones de la misma trayectoria, las cuales siguen conservando tanto su estructura como su orden temporal. Este resultado es importante ya que estas reducciones, al ser un resumen de la trayectoria original, se pueden usar enteramente como entrada para análisis más complejos que pueden volverse imprácticos cuando tratan con trayectorias muy grandes. Otros técnicas de reducción usadas en análisis de trayectorias o bien transforman las conformaciones en estructuras de menos dimensiones, solo interpretables cuando se observan en conjunto, como el caso de MDS, Isomap, y diffusion maps [21, 10]; o crean grupos de ellas que resaltan alguna similaridad ya sea estructural o energética, sin importar su orden temporal, como en el caso de los agrupamientos [7]. Además, debido a que varias de estas técnicas se basan en el cálculo de las distancias entre pares de conformaciones, el alto costo computacional de realizar esos cálculos para millones o incluso miles de conformaciones, las puede volver imprácticas sino se utilizan trayectorias reducidas como las que produce nuestro algoritmo.

Sin embargo, aunque las conformaciones de las trayectorias reducidas conservan el orden temporal que tienen en la trayectoria original, el tiempo de simulación en que suceden no se conserva explícitamente. Es decir, las reducciones no describen pasos de tiempo sino pasos de plegamiento, que se refieren a la secuencia de eventos destacados que resumen el plegamiento

Garreta et al. Page 5 of 8

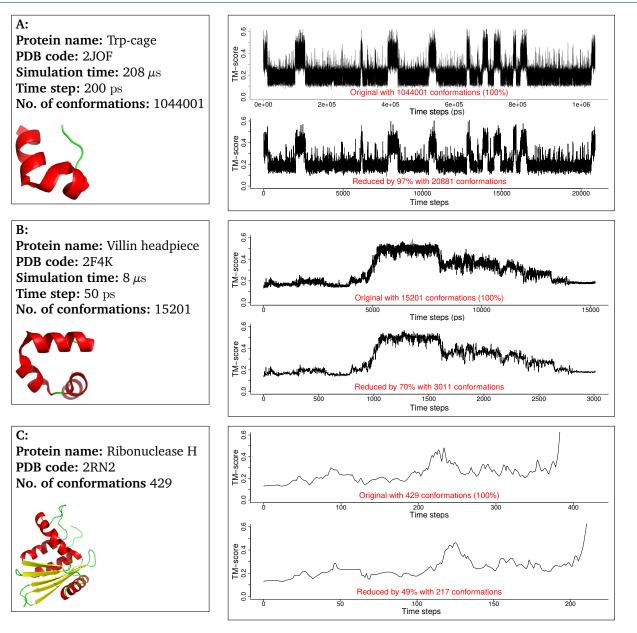


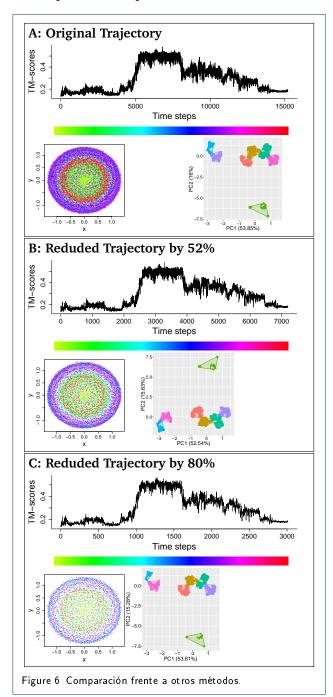
Figure 5 Plot images of the TM-score values for the original and reduced trajectories. Protein and simulation details on the left box. Trajectories on the right box: original (upper part), and reduced (lower part). (A) Trajectory of trp-cage protein, reduced by 98%, from 1044001 to 20881 conformations. (B) Trajectory of the villin-headpiece protein, reduced by 80%, from 15201 to 3011 conformations. (C) Trajectory of ribonuclease H protein, reduced by 49%, from 429 to 217 conformations. Folding steps refers to temporal ordering instead of simulation times.

de la proteína y no al tiempo exacto en que estos ocurren. No obstante, para obtener estos tiempos, se puede tomar el nombre o identificador de la conformación de interés en la trayectoria reducida y localizar su tiempo en la trayectoria original.

Comparación frente a otros métodos de reducción

Para las comparaciones utilizamos los datos de la simulación de plegamiento de la proteína villin-headpiece del proyecto folding@home [5]. Tomamos la trayectoria original y calculamos su reducción por los métodos de nMDS y PCA. Luego, calculamos dos reducciones con nuestro algoritmo sobre esta trayectoria y a los Garreta et al. Page 6 of 8

datos resultantes le calculamos nuevamente las reducciones por nMDS y PCA. Los resultados se muestran en la figura 6, donde cada fila contiene tres despliegues en 2D: de la trayectoria, del patrón resultante de la reducción por nMDS, y del agrupamiento al proyectar los dos primeros componentes del PCA.



Observamos que las reducciones de la trayectoria original producen un despliegue en 2D característico en ambos métodos de reducción: un patrón de círculos de puntos, para el nMDS; y una estructura de 7

grupos, para el agrupamiento por PCA (fila superior, figura 6). Así mismo, este mismo despliegue se repite en gran medida en las dos reducciones calculadas por nuestro algoritmo, la de compresión media del 52% y la de compresión alta del 80% (filas central e inferior de la figura 6, respectivamente).

Lo anterior nos indica que nuestras reducciones preservan en gran medida los eventos principales de la trayectoria al observar que tanto las reducciones con nMDS y PCA siguen conservando el mismo patrón y la misma estructura de grupos. Además, nuestro algoritmo presenta ventajas adicionales sobre los otros métodos de reducción. Primero, el cálculo de las reducciones es más eficiente que el de nMDS ya que no necesita la matriz de disimilaridades, que es sumamente costosa de calcular cuando el número de conformaciones es grande. Segundo, la interpretación de los resultados es directa ya que los resultados son conformaciones de la proteína y no transformaciones de los datos, como en el caso del nMDS y PCA, o grupos de conformaciones, como en el caso de los agrupamientos. Y tercero, el orden temporal se conserva ya que el resultado es una nueva trayectoria, a diferencia del agrupamiento en donde los grupos resultantes pueden contener conformaciones que ocurren en tiempos muy distintos.

Desempeño del algoritmo

El desempeño de nuestro algoritmo lo evaluamos en dos situaciones: comparándolo frente a otros métodos de reducción (Figura 7) y ejecutándolo en paralelo usando múltiples núcleos de procesamiento (Figura 8). Para esto utilizamos las 100K primeras conformaciones de la trayectoria de la proteína Trp-cage (ver Métodos). Para la primera evaluación ejecutamos los métodos con diferentes tamaños de subtrayectorias, desde 100 hasta 100K conformaciones, y en la segunda evaluación ejecutamos nuestro algoritmo con diferente número de núcleos de procesamiento.

En la comparación con otros métodos de reducción, la figura 7 muestra que PCA es el más eficiente seguido de nuestro algoritmo FastReduction cuando se ejecuta con un solo núcleo de procesamiento. Sin embargo, si lo ejecutamos en paralelo con 2 núcleos, este se vuelve más eficiente que PCA. Por el contrario, nMDS y clustering se vuelven imprácticos con subtrayectorias medianamente largas. Ahora, si ejecutamos nuestro algoritmo en paralelo con 2 cores (FR2, línea azul), este se vuelve más eficiente que PCA.

Este comportamiento lo podemos ver más claramente en la figura 8, donde se muestran los tiempos y la aceleración que alcanza el algoritmo a medida que se ejecuta con más núcleos. Cada que duplicamos el

Garreta et al. Page 7 of 8

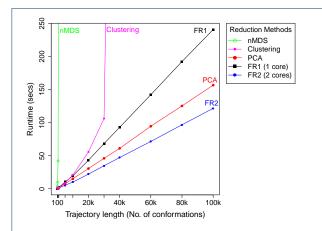


Figure 7 Desempeño del algoritmo frente a otros métodos. Comparación del nuestro algoritmo FR1 con nMDS, PCA, y agrupamiento. PCA y FR1 son los más eficientes, pero si nuestro algoritmo utiliza dos núcleos (FR2), el tiempo se disminuye a la mitad y se vuelve más eficiente que PCA. Por el contrario, nMDS y clustering toman demasiado tiempo, aún con trayectorias pequeñas.

número de núcleos, el tiempo de ejecución se disminuye casi a la mitad, hasta los 8 núcleos esta relación se conserva y luego la disminución es menor hasta volverse mínima pasados los 30 núcleos.

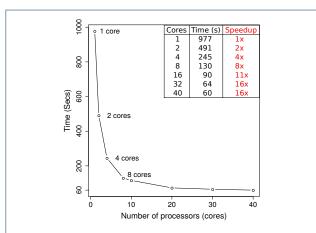


Figure 8 Desempeño del algoritmo cuando se ejecuta en paralelo con múltiples núcleos. Al duplicar el número de núcleos de procesamiento el tiempo se disminuye casi a la mitad y por lo tanto la aceleración crece casi de forma lineal, por lo menos hasta los 8 núcleos (8x). Después de esto, la aceleración sigue siendo apreciable hasta casi después de los 30 núcleos.

Todo lo anterior nos muestra que el algoritmo presenta un buen desempeño comparado con los otros métodos, y que este mejora más cuando aprovecha su paralelismo y se ejecuta con más de un núcleo. Como consecuencia, la aceleración de nuestro algoritmo escala de forma lineal con el número de núcleos que utiliza, por lo menos hasta 8x, es decir, la velocidad de ejecución cuando utiliza 8 núcleos es 8 veces más que cuando utiliza solo uno. Además, con 32 núcleos todavía se logra una aceleración de 16x, después de lo cual esta se mantiene sin mayor aumento (ver recuadro figura ??.B). Ahora, considerando que la tecnología multi-core ya está presente en muchas de los computadores de hoy día, el algoritmo tiene la capacidad de aprovechar esta tecnología para reducir trayectorias largas en tiempos cortos, cercanos e incluso mejores que los que toman algunos de los métodos cómunes usados en reducción de trayectorias de plegamiento.

Conclusiones

Las simulaciones de plegamiento de proteínas están avanzando significativamente y cada vez se realizan más para nuevas proteínas, con tiempos de duración más largos, y llevadas a cabo sobre diversas tecnologías. Como consecuencia, las trayectorias generadas por estas simulaciones cada vez son más extensas, del orden de millones de conformaciones, lo cual hace difícil su procesamiento y análisis. Para simplificarlas se han planteado diferentes técnicas que más bien son técnicas de análisis que transforman las conformaciones o crean grupos de ellas y sus resultados tienen sentido solo cuando se observan en conjunto.

Aquí, nosotros hemos planteado un algoritmo para simplificar trayectorias de plegamiento que divide la trayectoria en segmentos y extrae de ellos sus eventos principales o conformaciones destacadas en dos fases: primero extrae rápidamente las conformaciones disimilares y luego una selecciona de estas a las más representativas. El algoritmo se caracteriza por ser rápido y fácilmente paralelizable, y por lo tanto ejecutable en máquinas ordinarias con múltiples cores, disponibles ya en la mayoría de laboratorios de investigación.

De acuerdo a los resultados, el algoritmo produce simplificaciones de las trayectorias originales con una compresión alta y con los eventos principales visualmente conservados. Así mismo, estos resultados conservan en gran medida los patrones y la estructura que producen las reducciones hechas por otras técnicas de reducción y análisis de trayectorias. En cuanto al desempeño del algoritmo, este se aproxima al mostrado por algunas de las técnicas más eficientes y mejora mucho cuando se ejecuta en paralelo.

Sin embargo, las simplificaciones producidas por el algoritmo están limitadas a crear resúmenes de las trayectorias sin realizarles ningún tipo de análisis, como lo hacen otras técnicas. Por esta misma razón, estas trayectorias resumidas pueden servir de entrada

Garreta et al. Page 8 of 8

tanto a técnicas de análisis complejas como a otras técnicas de reducción que empiezan a tener problemas a medida que las trayectorias se vuelven más grandes.

Author details

¹Escuela de Ingeniería de Sistemas y Computación, Universidad del Valle, Santiago de Cali, Colombia. ²The European Bioinformatics Institute (EMBL-EBI), Hinxton, Cambridgeshire, UK.

References

- Shaw, D.E., Chao, J.C., Eastwood, M.P., Gagliardo, J., Grossman, J.P., Ho, C.R., Lerardi, D.J., Kolossváry, I., Klepeis, J.L., Layman, T., McLeavey, C., Deneroff, M.M., Moraes, M.A., Mueller, R., Priest, E.C., Shan, Y., Spengler, J., Theobald, M., Towles, B., Wang, S.C., Dror, R.O., Kuskin, J.S., Larson, R.H., Salmon, J.K., Young, C., Batson, B., Bowers, K.J.: Anton, a special-purpose machine for molecular dynamics simulation. Communications of the ACM 51(7), 91 (2008). doi:10.1145/1364782.1364802
- Lindorff-Larsen, K., Piana, S., Dror, R.O., Shaw, D.E.: How fast-folding proteins fold. Science 334(6055), 517–520 (2011). doi:10.1126/science.1208351. arXiv:1011.1669v3
- Shaw, D.E., Grossman, J.P., Bank, J.A., Batson, B., Butts, J.A., Chao, J.C., Deneroff, M.M., Dror, R.O., Even, A., Fenton, C.H., Forte, A., Gagliardo, J., Gill, G., Greskamp, B., Ho, C.R., lerardi, D.J., Iserovich, L., Kuskin, J.S., Larson, R.H., Layman, T., Lee, L.-S., Lerer, A.K., Li, C., Killebrew, D., Mackenzie, K.M., Mok, S.Y.-H., Moraes, M.A., Mueller, R., Nociolo, L.J., Peticolas, J.L., Quan, T., Ramot, D., Salmon, J.K., Scarpazza, D.P., Schafer, U.B., Siddique, N., Snyder, C.W., Spengler, J., Tang, P.T.P., Theobald, M., Toma, H., Towles, B., Vitale, B., Wang, S.C., Young, C.: Anton 2: Raising the Bar for Performance and Programmability in a Special-Purpose Molecular Dynamics Supercomputer. In: Shaw2014 (ed.) SC14: International Conference for High Performance Computing, Networking, Storage and Analysis, pp. 41–53. IEEE, Los Alamitos, CA, USA (2014). doi:10.1109/SC.2014.9. http://ieeexplore.ieee.org/document/7012191/
- Nguyen, H., Maier, J., Huang, H., Perrone, V., Simmerling, C.: Folding simulations for proteins with diverse topologies are accessible in days with a physics-based force field and implicit solvent. Journal of the American Chemical Society 136 (40), 13959–13962 (2014). doi:10.1021/ja5032776
- Larson, S.M., Snow, C.D., Shirts, M., Pande, V.S.: Folding@Home and Genome@Home: Using distributed computing to tackle previously intractable problems in computational biology (2009). 0901.0866
- Duan, M., Fan, J., Li, M., Han, L., Huo, S.: Evaluation of Dimensionality-reduction Methods from Peptide Folding-unfolding Simulations. Journal of chemical theory and computation 9(5), 2490–2497 (2013). doi:10.1021/ct400052y
- Peng, J.-h., Wang, W., Yu, Y.-q., Gu, H.-l., Huang, X.: Clustering algorithms to analyze molecular dynamics simulation trajectories for complex chemical and biological systems. Chinese Journal of Chemical Physics 31(4), 404–420 (2018). doi:10.1063/1674-0068/31/cjcp1806147
- Rajan, A., Freddolino, P.L., Schulten, K.: Going beyond clustering in MD trajectory analysis: An application to villin headpiece folding. PLoS ONE 5(4), 9890 (2010). doi:10.1371/journal.pone.0009890
- Das, P., Moll, M., Stamati, H.: Low-dimensional, free-energy landscapes of protein-folding reactions by nonlinear dimensionality reduction. Proceedings of the ... 103(26) (2006)
- Kim, S.B., Dsilva, C.J., Kevrekidis, I.G., Debenedetti, P.G.: Systematic characterization of protein folding pathways using diffusion maps: Application to Trp-cage miniprotein. The Journal of Chemical Physics 142(8), 85101 (2015). doi:10.1063/1.4913322
- Doerr, S., Ariz-Extreme, I., Harvey, M.J., De Fabritiis, G.: Dimensionality reduction methods for molecular simulations (2017). 1710.10629
- 12. Shao, J., Tanner, S.W., Thompson, N., Cheatham, T.E.: Clustering Molecular Dynamics Trajectories: 1. Characterizing the Performance

- of Different Clustering Algorithms. Journal of chemical theory and computation 3(6), 2312-34 (2007). doi:10.1021/ct700119m
- 13. Kaufman, L., Rousseeuw, P.: Finding Groups in Data. Wiley-Interscience; New York, ??? (1990)
- 14. Zhang, Y., Skolnick, J.: Scoring function for automated assessment of protein structure template quality. Proteins: Structure, Function, and Bioinformatics 68(4), 1020 (2007). doi:10.1002/prot.21643
- Ensign, D.L., Kasson, P.M., Pande, V.S.: Heterogeneity even at the speed limit of folding: Large-scale molecular dynamics study of a fast-folding variant of the villin headpiece. Journal of molecular biology 374(3), 806–816 (2007)
- Amato, N.M., Tapia, L., Thomas, S.: A Motion Planning Approach to Studying Molecular Motions. Communications in Information and Systems 10(1), 53-68 (2010). doi:10.4310/CIS.2010.v10.n1.a4
- 17. Xu, J., Zhang, Y.: How significant is a protein structure similarity with TM-score = 0.5? Bioinformatics 26(7), 889-895 (2010). doi:10.1093/bioinformatics/btq066
- Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R., O'Hara, R.B., Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., Wagner, H.: vegan: Community Ecology Package. (2019). https://cran.r-project.org/package=vegan
- Grant, B.J., Rodrigues, A.P.C., ElSawy, K.M., McCammon, J.A., Caves, L.S.D.: Bio3D: An R package for the comparative analysis of protein structures. Bioinformatics 22(21), 2695–2696 (2006). doi:10.1093/bioinformatics/btl461
- R Core Team: R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria (2018). R Foundation for Statistical Computing. https://www.r-project.org/
- Duan, M., Han, L., Rudolph, L., Huo, S., Carlson, G.H.: Geometric Issues in Dimensionality Reduction and Protein Conformation Space. (2014)